

Supplementary data

The *Arabidopsis* *MHX* gene includes an intronic element that boosts translation when localized in a 5' UTR intron

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Table S1

Element	IMEter v1.0
E1	0.831
E2	1.665
E3	-0.793
E4	3.549
E5	6.101

Table S1. IMEter scores of individual LI elements.

The scores were calculated (without deleting any nucleotides from the 5' or 3' ends of each element) with IMEter v1.0 (<http://korflab.ucdavis.edu/cgi-bin/web-imeter.pl>) (only IMEter v1.0 was available when this study was initiated). We divided the internal LI sequence into regions having, as much as possible (considering also the length of each region), roughly comparable IMEter scores.

Table S2

Constructs of the UTR or CDS series	IMEter v2.0 scores of <i>CAT1</i> introns containing the modified LI elements	IMEter v2.0 scores of putative <i>CAT1</i> introns containing the original LI elements	% Difference in IMEter v2.0 scores of <i>CAT1</i> introns containing the modified sequence
U1 or C1	5.95	5.95	0
U0 or C0	6.56	6.56	0
U12 or C12	8.41	9.02	-6.8
U2 or C2	9.43	10.13	-6.98
U123 or C123	12.89	13.49	-4.48
U3 or C3	14.28	14.28	0
U23 or C23	14.63	15.33	-4.6
U1-5 or C1-5	15.16	16.09	-5.8

Table S2. IMEter scores of the modified LI elements inside the *CAT1* compared to putative similar constructs containing the comparable non-modified LI elements.

The constructs are presented in the order of their increasing IMEter scores. The modifications in the LI sequence resulted in only 4-7% reduction in the IMEter scores of most constructs compared to putative similar constructs containing the comparable non-modified LI elements. The scores of the introns of constructs U1-C1 (in which E1 was not modified) and U3-C3 were identical to that of a putative *CAT1* intron containing the original LI sequence. For constructs U0-C0, which did not include any LI elements, the value reflects the score of the *CAT1* intron alone. The scores were calculated with IMEter v2.0 (<http://korflab.ucdavis.edu/cgi-bin/web-imeter2.pl>) using the full sequence of each intron, including the *CAT1* sequence. The second version of the IMEter algorithm was found to be a better predictor than the first version of how well any intron will enhance mRNA accumulation (Parra et al., 2011).

Fig. S1

A

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Orig: 1 AACGCTTGACCGATTCCAATCAGCTCCTCTCGATTTCCGTTTGTTCGGAAAATCTCTCCCG 60
      |
Modif: 1 AACGCTTGACCGATTCCAATCAGCTCCTCTCGATTTCCGTTTGTTCGGAAAATCTCTCCCG 60

                                     Intron
                                     ↓
Orig: 61 TGATCGGCGTATTGTGAATGCCGCTCACCGAGATATTCTCCGATTCTTTTCCCAGTGAG 120
      |
Modif: 61 TGATCGGCGTATTGTGAAGGCCGCTCACCGAGATATTCTCCGGTTCTTTTCCCAGTGAG 120

Orig: 121 GACAAGTGTTTCAGTTGACTTATTAGGAGGTGGGGTTTGAATAAGTTACC 169
      |
Modif: 121 GACAAGTGTTTCAGTTGACTTATTAGGAGGAGGGGTTTGAATAAGTTACC 169
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B

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Orig: 1 gtaaatttctagtttttctccttcattttcttggttaggacccttttctctttttat 60
      |
Modif: 1 gtaaatttctagtttttctccttcattttcttggttaggacccttttctctttttat 60

      NheI                                     HpaI AflII
Orig: 61 tttgagctttgatcttttctttaaactgatctattttttaattgat-tggttatggtgtaa 119
      |
Modif: 61 tttgagctagcatcttttctttaaactgatctattttttaattg_taaccttaagctc 120
                                     ↑
                                     LI elements

Orig: 120 atattacatagctttaactgataatctgattactttatttcggtgtgtctatgatgatgat 179
      |
Modif: 121 atattacatagctttaactgataatctgattactttatttcggtgtgtctgattgattgga 180

Orig: 180 gatagttacag 190
      |
Modif: 181 aataattacag 191
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Fig. S1. The sequences of the 5' UTR of *AtMHX* and the *CAT1* intron.

(A) The modifications introduced into the 5' UTR of *AtMHX* in the UTR and CDS series compared to the DEL series. Origi: – original sequence, Modif: – modified sequence. The T to G mutation (bold, underlined letter) is for eliminating the upstream AUG codon of the 5' UTR. The two other modifications (highlighted in gray) are for elimination of potential cryptic splice sites. The position of the LI of *AtMHX* or the *CAT1* intron in the 5' UTR is indicated by the arrow.

(B) The modified *CAT1* intron used throughout this study compared to the original sequence of this intron in *Ricinus communis*. Nucleotides introduced to create restriction enzyme sites are underlined. The modifications indicated by bold letters are for elimination of ATG triplets (by keeping the same nucleotides in a different order). The modifications highlighted in gray are for elimination of potential cryptic splice sites. The position into which the LI elements were subsequently introduced (in the middle of the HpaI site) is indicated by the arrow. Prediction of potential cryptic splice sites was carried out using the NetPlantGene Server (<http://www.cbs.dtu.dk/services/NetPGene/>).

Fig. S2

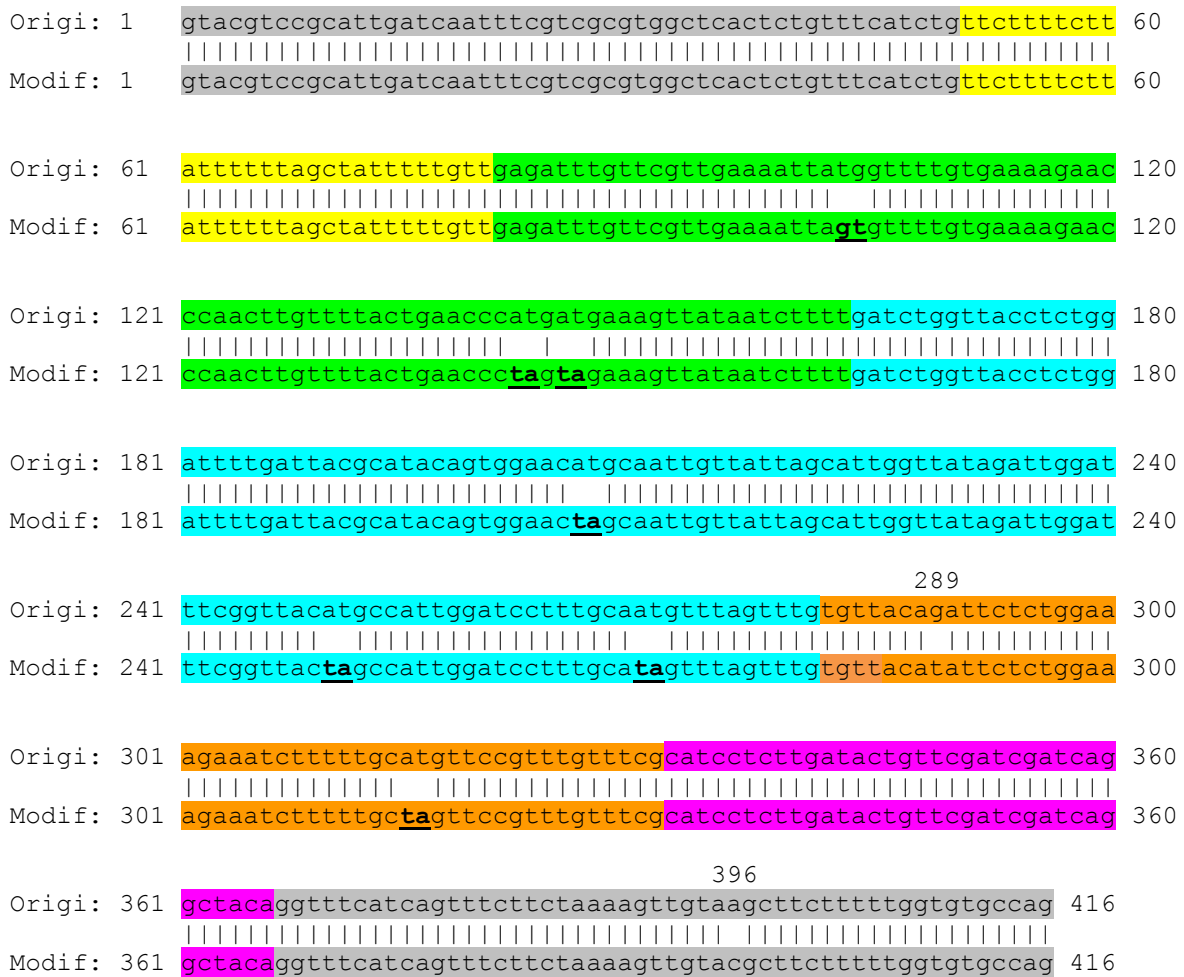


Fig. S2. The modifications introduced into the LI elements in the UTR and CDS series compared to the DEL series.

The different elements were color highlighted as follows: E1 – yellow, E2 – green, E3 – blue, E4 – orange, and E5 – pink. The two terminal 50 nt regions were highlighted in gray. Orig: – original sequence. Modif: – modified sequence. The modifications indicated by bold, underlined letters were for elimination of all ATG triplets (by keeping the same nucleotides in a different order). The G to T substitution at position 289 was for elimination of a potential cryptic splice site. The A to C substitution at position 396 was for elimination of a HindIII site that interfered with cloning into the binary vector. The modifications resulted in less than 7% reduction in the IMETER scores of the LI elements (Supplemental Table 2).

Fig. S3

Intron
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ATGGTAGACCTGAGGAACCGACGACTCGTCCGTCTGTAGAAACCCCAACCCGTGAAATCAAAAACTCGACGGCCT
GTGGGCATTCAGTCTGGATCGCGAAAACTGTGGAATTGATCAGCGTTGGTGGGAAAGCGCGTTACAAGAAAGCCGGG
CAATTGCTGTGCCAGGCAGTTTTAACGATCAGTTCGCCGATGCAGATATTCGTAATTATGCGGGCAACGTCTGGTAT
CAGCGCGAAGTCTTTATACCGAAAGGTTGGGCAGGCCAGCGTATCGTGCTGCGTTTTCGATGCGGTCACATCATTACGG
CAAAGTGTGGGTCAATAATCAGGAAGTGATGGAGCATCAGGGCGGCTATACGCCATTTGAAGCCGATGTCAGCCGT
ATGTTATTGCCGGGAAAAGTGACGTATCACCGTTTTGTGTGAACAACGAACGAACGAACTGGCAGACTATCCC GCCGGGA
ATGGTGATTACCGACGAAAAACGGCAAGAAAAAGCAGTCTTACTTCCATGATTTCTTTAACTATGCCGGAATCCATCG
CAGCGTAATGCTCTACACCACGCCGAACACCTGGGTGGACGATATCACCGTGGTGACGCATGTCGCGCAAGACTGTA
ACCACGCGTCTGTTGACTGGCAGGTGGTGGCCAATGGTGTATGTCAGCGTTGAACTGCGTGATGCGGATCAACAGGTG
GTTGCAACTGGACAAGGCACACTAGCGGGACTTTGCAAGTGGTGAATCCGCACCTCTGGCAACCGGGTGAAGGTTATCT
CTATGAACCTCGAAGTCACAGCCAAAAGCCAGACAGAGTCTGATATCTACCCGTTTCGCGTCGCGATCCGGTCAGTGG
CAGTGAAGGCCAACAGTTCCTGATTAACCACAAAACCGTTCTACTTTACTGGCTTTGGTTCGTCATGAAGATGCGGAC
TTACGTGGCAAAGGATTCGATAACGTGCTGATGGTGCACGACCACGCATTAATGGACTGGATTGGGGCCAACTCCTA
CCGTACCTCGCATTACCCTTACGCTGAAGAGATGCTCGACTGGGCAGATGAACATGGCATCGTGGTGATTGATGAAA
CTGCTGCTGTCGGCTTTCAGTGTCTTTAGGCATTGGTTTTCGAAGCGGGCAACAAGCCGAAAGAACTGTACAGCGAA
GAGGCAGTCAACGGGAAACTCAGCAAGCGCACTTACAGGCGATTAAGAGCTGATAGCGCGTGACAAAAACCACC
AAGCGTGGTGATGTGGAGTATTGCCAACGAACCGGATACCCGTCGCAAGGTGCACGGGAATATTTGCGGCCACTGG
CGGAAGCAACGCGTAAACTCGACCCGACGCGTCCGATCACCTGCGTCAATGTAATGTTCTGCGACGCTCACACCGAT
ACCATCAGCGATCTCTTTGATGTGCTGTGCCTGAACCGTTATTACGGATGGTATGTCCAAAGCGGGCATTGGAAC
GGCAGAGAAGGTACTGGAAAAAGAACTTCTGGCCTGGCAGGAGAACTGCATCAGCCGATTATCATCACCGAATACG
GCGTGGATACGTTAGCCGGGCTGCACTCAATGTACACCGACATGTGGAGTGAAGAGTATCAGTGTGCATGGCTGGAT
ATGTATCACCGCTCTTTGATCGCGTCAGCGCCGTCGTCGGTGAACAGGTATGGAATTTCCCGGATTTTGCACCTC
GCAAGGCATATTGCGCGTTGGCGGTAACAAGAAAGGGATCTTCACTCGCGACCGCAAACCGAAGTCGCGCGGCTTTTC
TGCTGCAAAAACGCTGGACTGGCATGAACTTCGGTGAAAAACCGCAGCAGGGAGGCAAACAATGA
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Fig. S3. The coding sequence of *GUS*.

The position of the introns introduced in the CDS series is indicated by the arrow. The *CATI* intron derivatives introduced here included elimination of a potential cryptic splice site (Fig. S1B). This elimination was shown to result in full and accurate splicing of this intron when introduced into this position of *GUS* (Ma *et al.*, 2011).

Fig. S4

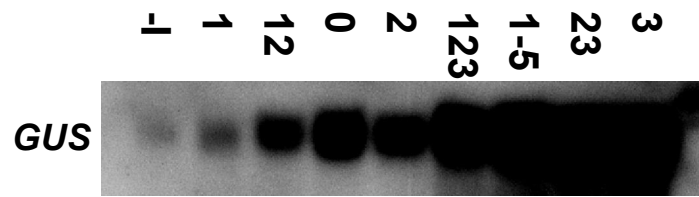


Fig. S4. The Northern blot shown in Fig. 3D of the manuscript exposed for a longer period. The levels of *GUS* mRNA in constructs -I and U1 relative to construct U12 were determined using the ImageJ program (NIH). The values obtained were used to relate *GUS* mRNA levels in the constructs that had low levels of *GUS* mRNA to the values of other constructs, which were quantified from gels exposed for shorter periods.