Supporting Information

Simpson-Lavy and Johnston 10.1073/pnas.1304839110

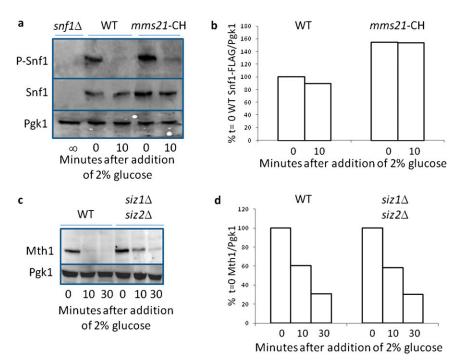


Fig. S1. Other small ubiquitin-like modifier protein (SUMO) ligases do not regulate Mth1 stability in glucose. (A) Cells were grown in 2% galactose at 30 °C overnight. The temperature was elevated to 34 °C for 1 h before addition of preheated glucose to 2%. Samples were processed for immunoblots at the indicated times. (B) Quantification of Snf1 levels in A. Amount of Snf1 is shown relative to wild-type in galactose-grown cells (t = 0). (C) Cells were grown in 2% galactose at 30 °C overnight. Glucose was added to 2% and samples were processed for immunoblots at the indicated times. (D) Quantification of C. Amount of Mth1 is shown relative to each strain in the absence of glucose (t = 0).

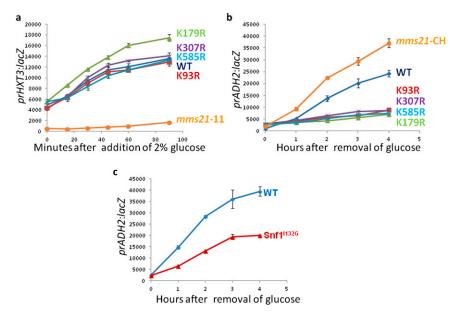


Fig. 52. Other SUMO motifs in Snf1 are nonfunctional. (*A*) $snf1\Delta$ cells with plasmids bearing wild-type SNF1 or SNF1 with the indicated mutations, or $snf1\Delta mms21-11$ cells with a plasmid bearing wild-type SNF1 were grown overnight at 30 °C in 2% galactose. Glucose was added to 2%, and samples were taken for β -galactosidase assays at the indicated times. n = 3. Plasmids are listed in Table S3. (*B*) $snf1\Delta$ cells with plasmids bearing wild-type SNF1 or SNF1 with the indicated mutations, or $snf1\Delta mms21-CH$ cells with a plasmid bearing wild-type SNF1 were grown overnight at 30 °C in 4% glucose, then for 1 h at 34 °C. Cells were washed three times with water at 34 °C, and resuspended in medium containing 3% glycerol at 34 °C. Samples were taken for β -galactosidase assays at the indicated times. n = 3. Plasmids are listed in Table S3. (*C*) $snf1\Delta$ cells with plasmids bearing wild-type SNF1 were grown overnight at 30 °C in 4% glucose, then for 1 h at 34 °C. Cells were washed three times with water at 34 °C, and resuspended in medium containing 3% glycerol at 34 °C. Samples were taken for β -galactosidase assays at the indicated times. n = 3. Plasmids are listed in Table S3. (*C*) $snf1\Delta$ cells with plasmids bearing wild-type SNF1 or $SNF1^{1132G}$ were grown overnight at 30 °C in 4% glucose. Cells were washed three times with water and resuspended in medium containing 3% glycerol at 30 °C. Samples were taken for β -galactosidase assays at the indicated times. n = 3.



Fig. S3. Functional domains and amino acid residues of Snf1. Schematic diagram of Snf1 illustrating locations of domains and residues discussed herein.

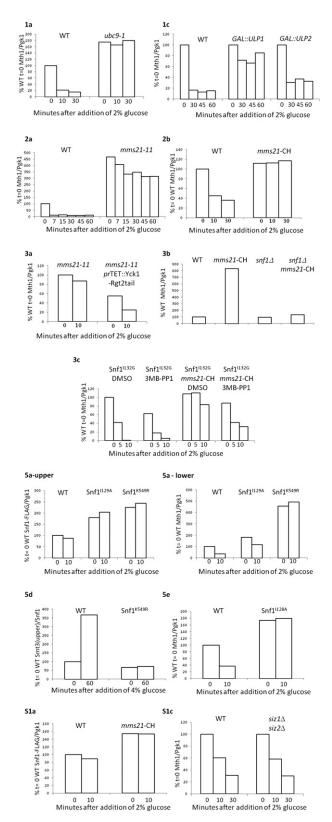


Fig. 54. Quantifications of blots. Quantifications of blots shown in this article were performed using ImageJ. The boldface numbers in the upper left corner of each panel signifies the figure and panel referenced in this article (i.e., "1a" represents Fig. 1A in this article).

DNAS

Table S1. Yeast strains

Name	Genotype	Source	
W303	leu2-3,112 trp1-1 can1-100 ura3-1 ade2-1 his3-11,15 [phi ⁺] rad5-535, bud4, ssd1-d2	Lab collection	
G16	W303 <i>MAT</i> a RAD5	(1)	
Т79	W303 MATa RAD5 mms21-11::LEU2	(1)	
T80-14	W303 MATa RAD5 mms21-11::URA3	(1)	
M21-CH	W303 MATa RAD5 mms21-CH::URA3		
M21-CHu3	W303 MATa RAD5 mms21-CH::URA3::ura3		
Snf1KAN (CKY18)	W303 $MAT\alpha$ snf1 Δ ::KAN	(2)	
Snf1U3	W303 MATa snf1∆::URA3	J. Rutter	
Snf1KANm21-CH	W303 MATα snf1Δ::KAN mms21-CH::URA3		
Snf1KANm21-CHura3	W303 MATα snf1Δ::KAN mms21-CH::URA3::ura3		
BY4741	MATa his3∆1 leu2∆0/LYS2 met15∆0 ura3∆0		
BY4742	MATα his3 Δ 1 leu2 Δ 0 ura3 Δ 0 lys2 Δ 0 MET15		
ubc9-1 (ts)	BY4741 ubc9-1::NAT	T. Ravid	
YAC391	MATα <i>siz1</i> ∆::HygB <i>siz2</i> ∆::KAN, <i>lyp1</i> ∆::STE3pr:LEU2, his3∆1 leu2∆0 ura3∆0	(3)	
EY1274	BY MATα ulp1-ts::MET15 LYS2 STE3pr::LEU2 his3Δ1 ura3Δ0	(4)	
Ubp8	BY4741 ubp8::KAN		
Ubp8M21CH	BY4741 ubp8::KAN mms21-CH::URA3::ura3		
Ubp8slx8	BY4741 ubp8::KAN s/x8::URA3		

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1. Zhao X, Blobel G (2005) A SUMO ligase is part of a nuclear multiprotein complex that affects DNA repair and chromosomal organization. Proc Natl Acad Sci USA 102(13):4777–4782.

2. Ratnakumar S, Kacherovsky N, Arms E, Young ET (2009) Snf1 controls the activity of adr1 through dephosphorylation of Ser230. Genetics 182(3):735–745. 3. Chavez A, George V, Agrawal V, Johnson FB (2010) Sumoylation and the structural maintenance of chromosomes (Smc) 5/6 complex slow senescence through recombination

intermediate resolution. J Biol Chem 285(16):11922–11930.

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4. Wykoff DD, O'Shea EK (2005) Identification of sumoylated proteins by systematic immunoprecipitation of the budding yeast proteome. Mol Cell Proteomics 4(1):73–83.

Table S2. Plas	mids			
Name	Genotype	Backbone*	Mutations	Source
pBM4560	Mth1-9myc	pRS 313		Lab collection
pBM5020	Mth1-Stag	pRS 313		Lab collection
pBGM18	prADH2::lacZ	pRS 313, 315, 316		(1)
pBM2819	pr <i>HXT3</i> :: <i>lacZ</i>	2 μ URA3		Lab collection
pBM3118	pr <i>HXT3</i> :: <i>lacZ</i>	2 μ LEU2		Lab collection
pBM4924	prTET ^{on} ::YCK1-Rgt2tail	2 μ TRP1		Lab collection
Mms21-47	mms21-CH	pUC19 URA3	C200A H202A	(2)
pBM3926	pr <i>GAL1</i> :His6-FLAG-Smt3(G98)	pRS 313, 315 , 316		(3)
S12F	Snf1-2FLAG	pRS 314, 315, 316	I129A, K549R	(4)
pYL199 (S18M)	Snf1-8myc	pRS 313 , 314, 316	I129A, K549R	(5)
Snf1-as1	Snf1 ^{1132G}	pRS 316		(6)
Snf1	Snf1	pRS 316 , 313		(6)
GALulp1	pr <i>GAL1</i> :GST-Ulp1	2 μ URA3		M. Hochstrasser
GALulp2	pr <i>GAL1</i> :Ulp2-FLAG	2 μ TRP1		M. Hochstrasser

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1. Dombek KM, Voronkova V, Raney A, Young ET (1999) Functional analysis of the yeast GIc7-binding protein Reg1 identifies a protein phosphatase type 1-binding motif as essential for repression of ADH2 expression. *Mol Cell Biol* 19(9):6029–6040.

2. Santa Maria SR, Gangavarapu V, Johnson RE, Prakash L, Prakash S (2007) Requirement of Nse1, a subunit of the Smc5-Smc6 complex, for Rad52-dependent postreplication repair of UV-damaged DNA in Saccharomyces cerevisiae. Mol Cell Biol 27(23):8409–8418.

3. Johnson ES, Blobel G (1999) Cell cycle-regulated attachment of the ubiquitin-related protein SUMO to the yeast septins. J Cell Biol 147(5):981–994.

4. Nayak V, et al. (2006) Structure and dimerization of the kinase domain from yeast Snf1, a member of the Snf1/AMPK protein family. Structure 14(3):477-485.

5. Raines RT, McCormick M, Van Oosbree TR, Mierendorf RC (2000) The S.Tag fusion system for protein purification. Methods Enzymol 326:362-376.

6. Shirra MK, et al. (2008) A chemical genomics study identifies Snf1 as a repressor of GCN4 translation. J Biol Chem 283(51):35889–35898.

Table S3. Supplemental plasmids								
Name	Genotype	Backbone	Mutations	Source				
\$12F	Snf1-2FLAG	pRS 316	K93R, K179R, K307R, K585R	(1)				

1. Nayak V, et al. (2006) Structure and dimerization of the kinase domain from yeast Snf1, a member of the Snf1/AMPK protein family. Structure 14(3):477-485.

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