SUPPLEMENTAL MATERIAL

TABLE S1. Primers used in this study

Description	Primer name	Sequence (5' to 3')[a]	Position	Reference
1st LA PCR for XbaI-digested fragments of SeRI*fusB*-3692 and SeRI*fusB*-85	fusB 437–465F[b]	GAGAAATTTCTAATCAGGTTGTAAAGGGG	*fusB* 437–465	1
	fusB 282–253R[b]	AAGTTTTTGCGGACTAGGTAGTTCAAAAGG	*fusB* 282–253	
2nd LA PCR for XbaI-digested fragments of SeRI*fusB*-3692 and cSePI*fusB*-857 and inverse PCR for AclI-digested	fusB 553–580F[c]	GGTGACTATATATGTCGAGATAGCATTC	*fusB* 553–580F	
	fusB221–192 R[c]	GCTTCAATTTCTTTGTTTGATAATCTGATG	*fusB* 221–192	

fragments of cSePI*fusB*-857				
Inverse PCR for PacI-digested fragments of SeRI*fusB*-3692	3692-EcoRI-LAF	GGAACGGATGATAGTTACACC	*aj2* downstream 102–122	This study
	3692-PacI-IR	CGATTGAATAACTTTGACGG	*fusB* 114–95	
Inverse PCR for BsmI-digested fragments of SeRI*fusB*-3692	fusB 389–361R[d]	TTCCGATTTGATGCAAGTTCATTCCATCC	*fusB* 389–361	
Detection of SeRI*fusB*-3692 upstream	S.epi ssrA-binding protein(F)	AAGATACAATTGAAGCGGG	*smpB* 74–92	This study
	8-7 BsmI-UR	TTCGTGCCTTACCTTCTG	SeRI*fusB*-3692 ORF13 -82_-99	
Inverse PCR	S. epi inverse	AGGTGCGAAGATTG	SePI*fusB*-857 △aj3	This study

for Spe- and	1990–2007F	CAGG	189–206
XbaI-digested	S. epi inverse	GTTTCACTCATCGC	SePI$_{fusB\text{-}857}$ LP
fragments of	21–1R	AACACAG	-113_-133
SePI$_{fusB\text{-}857}$			
Inverse PCR	S. epi	GGTATCTATTCAAG	SePI$_{fusB\text{-}857}$ *tnp*
for	BclI–1F	AGGTATGG	-18_4
BclI-digested	S. epi	CATTTTTTAGTCACT	SePI$_{fusB\text{-}857}$ ORF8
fragments of	BclI–1R	TCACGG	494–474
SePI$_{fusB\text{-}857}$			
Inverse PCR	S. epi	TCGTTAACTTTATC	SePI$_{fusB\text{-}857}$
for	hindIII–3F[e]	CACCCG	ORF17 -246_-227
PacI-digested			
fragments of			
SePI$_{fusB\text{-}857}$			
Detection of	S.epi	ATACGTTGTGTGAA	SePI$_{fusB\text{-}857}$ ORF7
SeRI$_{fusB\text{-}857}$	BclI–2R[f]	ATACCC	357–338
Upstream and			
downstream			

	S.epi PacI–3F	CATTTATATGGGCTATGCTGG	SePI*fusB*-857 ORF20 -7_14	This study
	S.epi sodium transporter 1146–1168F	TCTCACTATGGATTTAACTTCCG	SePI*fusB*-857 ORF24 1146–1168	
Detection of *vapE*	VapE 93F	WTCWGCYACWACACAAGC	*vapE* 93–110	This study
	VapE 1246R	GCCATACATAAGCACCTGG	*vapE* 1246–1228	

[a] K = G or T, and Y = C or T.

[b]Each of the two primers were paired with C1 provided by LA-PCR in vitro cloning kit (Takara Shuzo Co. Ltd., Japan) for the first LA PCR.

[b]Each of the the two primers were paired with C2 provided by LA-PCR in vitro [c]loning kit (Takara Shuzo Co. Ltd., Japan) for the nested LA PCR.

[d]The primer was paired with fusB 437–465F.

[e]The primer was paired with fusB 282–253R.

[f]The primer was paired with S.epi ssrA-binding protein(F) for upstream detection.

REFERENCES

1. **Chen HJ, Tsai JC, Hung WC, Tseng SP, Hsueh PR, Teng LJ.** 2011. Identification of *fusB*-mediated fusidic acid resistance islands in *Staphylococcus epidermidis* isolates. Antimicrob. Agents Chemother. **55:**5842-5849.