**Supplementary Information** 





**Fig. S1:**  $IC_{50}$  chart of the *P. falciparum* growth curve at 48h in presence of either fosmidomycin, Salt **2** or conjugate **3**. Each data point represents the mean and standard deviation of three replicates. Graph was generated by non-linear one site  $IogIC_{50}$  fit using the software Graphpad Prism.



Figure S2

Fig. S2: Diethyl ester of fosmidomycin does not inhibit growth of *Mycobacterium bovis* BCG.

(A) *M. bovis* BCG was grown in presence of isoniazid, fosmidomycin, conjugate 3, or the diethyl ester derivative of fosmidomycin (compound 9 in the synthesis scheme). Similar to fosmidomycin compound 9 did not show any significant inhibitory effect on mycobacterial growth. Each data point represent the mean (± S.D.) of three biological replicates.

## **General methods**

All reactions were monitored by TLC using silica gel 60  $F_{254}$  plates. Visualization of the reaction components was achieved using UV fluorescence (254 nm) and Hanessian's stain. Silica gel chromatography was carried out over Kieselgel 60. The yields reported are after purification. <sup>1</sup>H, <sup>13</sup>C NMR spectra were recorded in deuterated solvents and chemical shifts ( $\delta$ ) are quoted in parts per million (ppm) calibrated to solvent residual peak (<sup>1</sup>H and <sup>13</sup>C). Coupling constants (*J*) are measured in Hertz (Hz). The following abbreviations are used to describe multiplicities: *s* = singlet, *d* = doublet, *t* = triplet, *q* = quartet, *b* = broad, *m* = multiplet. Purification of final compounds was done by RP-HPLC (*Supelco Discovery BIO Wide Pore C18 column* (25 cm x 21.2 mm, 10 µm *Sigma-Aldrich*) flow rate: 10 ml/min, detection wavelength: 220 nm).

## Chemistry:

Synthesis of the targeted derivative of the fosmidomycin compound **9** (Scheme 1) was achieved starting from the commercially available 3-Bromopropylphosphonic acid diethylester **5**. Compound **5** was first transformed to the corresponding *O*-benzylhydroxylamino compound **6** by the nucleophilic substitution ( $S_N^2$ ) reaction of the bromide group by *O*-benzylhydroxylamine at refluxing acetonitrile. *N*-acetylated compound **7** was achieved by the treatment of acetyl chloride (AcCl) in the presence of triethylamine (Et<sub>3</sub>N). Subsequent hydrogenolysis with H<sub>2</sub> gas in

presence of catalytic Pd-C (10%), removed the benzyl protection to give the free N-hydroxylamine compound **8**.





Treatment of compound **4** with glutaric acid anhydride in presence of  $Et_3N$  and 4-*N*,*N*-dimethylaminopyridine (DMAP) yielded the targeted derivative of fosmidomycin.





Coupling of the fosmidomycin derivative **9** with the resin-bound carrier (octaarginine) was performed on a PAL-PEG solid support using *O*-(7-Aza-1H-

benzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate (HATU) as the coupling agent and diisopropyl ethyl amine (DIPEA) as the base in anhydrous DMF. Subsequent Pbf deprotection and cleavage from the resin with a cocktail mixture of TFA:H<sub>2</sub>O:TIPS provided the fosmidomycin-Arg<sub>8</sub>-conjugate **3** (Scheme 2).

$$EtO \xrightarrow{P} N_{O}^{H}$$

(3-Benzyloxyamino-propyl)-phosphonic acid diethylester (6): To a mixture of Obenzyl-hydroxylamine hydrochloride (3.07 g, 19.25 mmol), K<sub>2</sub>CO<sub>3</sub> (19.25 g, 19.25 mmol) and KI (0.63 g, 3.85 mmol) in CH<sub>3</sub>CN (50 mL) was added the 3-Bromopropylphosphonic acid diethylester 5 (1 g, 3.85 mmol) at room temperature and the reaction mixture was allowed to reflux for overnight. The reaction was quenched by the addition of H<sub>2</sub>O and the aqueous phase was extracted with CHCl<sub>3</sub> (3 x 30 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated under reduced pressure to give the crude compound. The residue was purified by column chromatography on silica gel using pure EtOAc as an eluent to give (3-benzyloxyamino-propyl)-phosphonic acid diethyl ester 6 as colourless oil (790 mg, 68%).<sup>1</sup>**H-NMR** (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.30 (*t*, *J* = 7.2, 6H), 1.74-1.85 (*m*, 4H), 2.96 (*t*, *J* = 6.4, 2H), 4.03-4.12 (*m*, 4H), 4.68 (*s*, 2H), 5.58 (*bs*, 1H), 7.28-7.34 (*m*, 5H). <sup>13</sup>**C-NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  16.4 (*d*, <sup>3</sup>*J*<sub>*C-P*</sub> = 5.9), 20.6 (*d*, <sup>2</sup>*J*<sub>*C-P*</sub> = 4.9), 23.2  $(d, {}^{1}J_{C-P} = 141.2), 52.1 (d, {}^{3}J_{C-P} = 16.6), 61.4 (d, {}^{2}J_{C-P} = 6.5), 76.4, 127.8,$ 128.4, 137.8. **HR-MS-ESI:** *m/z* Calcd. for C<sub>14</sub>H<sub>24</sub>NO<sub>4</sub>P: 302.1521 [M+H]<sup>+</sup>: *m/z* Found: 302.1518 [M+H]<sup>+</sup>.



[3-(acetylbenzyloxy-amino)-propyl]-phosphonic acid diethyl (7): ester Acetylchloride (0.12 mL, 1.55 mmol) was added dropwise to a solution of 6 (390 mg, 1.29 mmol) and triethylamine (0.22 mL, 1.55 mmol) in dichloromethane. The reaction mixture was stirred at room temperature for 4 h. The reaction mixture was quenched with saturated NaHCO<sub>3</sub> and then washed with H<sub>2</sub>O. The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated under reduced pressure to give the crude compound. The residue was purified by column chromatography on silica gel using CH<sub>2</sub>Cl<sub>2</sub>-MeOH (90:10) as an eluent to give 7 as a yellow oil (390 mg, 88%).<sup>1</sup>**H-NMR** (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.31 (*t*, *J* = 7.2, 6H), 1.69-1.78 (*m*, 2H), 1.90-2.01 (*m*, 2H), 2.10 (*s*, 3H), 3.72 (*t*, *J* = 6.8, 2H), 4.05-4.11 (*m*, 4H), 4.83 (s, 2H), 7.37-7.40 (m, 5H). <sup>13</sup>**C-NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  16.4 (d, <sup>3</sup>J<sub>C-P</sub> = 5.9), 20.2  $(d, {}^{2}J_{C-P} = 4.7)$ , 20.5, 22.3, 23.7, 61.6  $(d, {}^{2}J_{C-P} = 6.5)$ , 76.4, 128.7, 129.0, 129.2, 134.3. HR-MS-ESI: m/z Calcd. for C<sub>16</sub>H<sub>27</sub>NO<sub>5</sub>P: 344.1627 [M+H]<sup>+</sup>: m/z Found: 344.1621 [M+H]<sup>+</sup>.



[3-(acetyl-hydroxyamino)-propyl]-phosphonic acid diethyl ester (8): To a solution of the benzyl protected compound 7 (3.1 g, 9.02 mmol) in MeOH (100 mL) was added 10% Pd/C (184 mg) and hydrogenated at atmospheric pressure for 3h. The mixture was filtered over celite, and the solvent was removed by rotary evaporation to yield compound 8 as colorless oil (1.9 g, 84%).<sup>1</sup>H-NMR (400 MHz,

CDCl<sub>3</sub>):  $\delta$  1.29 (*t*, *J* = 6.8, 6H), 1.75-1.83 (*m*, 2H), 1.87-1.95 (*m*, 2H), 2.12 (*s*, 3H), 3.69 (*t*, *J* = 6.0, 2H), 4.01-4.06 (*m*, 4H), 9.69 (*bs*, 1H). <sup>13</sup>**C-NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  16.3 (*d*, <sup>3</sup>*J*<sub>*C-P*</sub> = 6.0), 19.1 (*d*, <sup>2</sup>*J*<sub>*C-P*</sub> = 5.2), 20.5, 22.2 (*d*, <sup>1</sup>*J*<sub>*C-P*</sub> = 139.9), 47.4 (*d*, <sup>3</sup>*J*<sub>*C-P*</sub> = 9.2), 62.1 (*d*, <sup>2</sup>*J*<sub>*C-P*</sub> = 6.7), 172.6. **HR-MS-ESI:** *m*/*z* Calcd. for C<sub>9</sub>H<sub>21</sub>NO<sub>4</sub>P: 254.1157 [M+H]<sup>+</sup>: *m*/*z* Found: 254.1154 [M+H]<sup>+</sup>.

5-((N-(3-(diethoxyphosphoryl)propyl)acetamido)oxy)-5-oxopentanoic acid (9): A mixture of compound **8** (253 mg, 1.0 mmol), Glutaric acid anhydride (111 mg, 0.98 mmol), DMAP (122 mg, 1.0 mmol) and Et<sub>3</sub>N (0.08 mL, 1.2 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) were stirred at room temperature for overnight. The reaction was quenched by the addition of saturated NaHCO<sub>3</sub> (~ pH-9). The organic layer was separated and the aqueous layer was washed with CH<sub>2</sub>Cl<sub>2</sub>. The aqueous layer was acidified by the addition of 6N HCl (~ pH-2) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 x 30 mL). The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated under reduced pressure to give the crude compound as faint pink liquid (140 mg, 38%) which was pure enough for the next step. <sup>1</sup>**H-NMR** (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.28 (*t*, *J* = 6.8, 6H), 1.74-2.02 (*m*, 9H), 2.40 (*t*, *J* = 7.2, 2H), 2.53 (*t*, *J* = 7.2, 2H), 3.72 (*t*, *J* = 6.4, 2H), 4.03-4.09 (*m*, 4H), 9.69 (*bs*, 1H). <sup>13</sup>**C-NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  16.3 (*d*, <sup>3</sup>*J*<sub>C</sub>-*p* = 6.0), 19.1, 19.6, 20.3, 20.1 (*d*, <sup>2</sup>*J*<sub>C-P</sub> = 17.1), 22.4 (*d*, <sup>1</sup>*J*<sub>C-P</sub> = 141.8), 32.8, 30.7,

62.0 (*d*, <sup>2</sup>*J*<sub>*C-P*</sub> = 6.6), 170.9, 172.7, 174.8. **HR-MS-ESI:** *m/z* Calcd. for C<sub>14</sub>H<sub>27</sub>NO<sub>8</sub>P: 368.1474 [M+H]<sup>+</sup>: *m/z* Found: 368.1468 [M+H]<sup>+</sup>.

 $H_2N-(Arg(Pbf))_8$ -PAL-PEG-resin: Fmoc-(Arg(Pbf)\_8-PAL-PEG-resin was prepared on an AB synthesiser using *FastMoc®* chemistry and Fmoc-Arg(Pbf)-OH (4.0 eq.), HBTU (3.9 eq.) on a 0.25 mmol scale of PAL-PEG-resin. Fmoc deprotection was performed by shaking the resin (220 mg, 0.21 mmol/g loading, 0.05 mmol) with three 10 mL-portions of 20% piperidine in DMF (10 min per treatment, draining between treatments), washed with DMF (3 x 10 mL), CH<sub>2</sub>Cl<sub>2</sub> (3 x 10 mL).



6-Fam-octaarginine amide (1): A small amount of the above Fmoc-deprotected resin was coupled to the isomerically pure 6-carboxyfluorescein (6-FAM) on solid support. The peptide was cleaved from the resin by treatment with TFA:H<sub>2</sub>O:TIPS (95:2.5:2.5) and the crude product was purified by HPLC on *Supelco Discovery BIO Wide Pore C18 column* (25 cm x 21.2 mm, 10  $\mu$ m *Sigma-Aldrich*) with MeCN (0.1 % TFA) : H<sub>2</sub>O (0.1 % TFA) ratio of 20 : 80 for 10 min and subsequently with a gradient from 20 : 80 to 25 : 75 in 5 min: t<sub>R</sub> 12 min; **HR-MS-ESI:** *m/z* Calcd. for C<sub>69</sub>H<sub>119</sub>N<sub>33</sub>O<sub>14</sub>: 1624.8905 [M+H]<sup>+</sup>: *m/z* Found: [M+H]<sup>+</sup>.



6-Fam-octaarginine-Fosmidomycin-Salt (2): A solution of 6-Fam-Arg<sub>8</sub>-NH<sub>2</sub> (1) (1.25  $\mu$ mol) in H<sub>2</sub>O (5 mL) was treated with a solution of fosmidomycin (5.00  $\mu$ mol) in H<sub>2</sub>O (5 mL) and the mixture was lyophilized to yield the 6-Fam-octaarginine-fosmidomycin salt (2).



Fosmidomycin-Arg<sub>8</sub>-conjugate (**3**): The above H<sub>2</sub>N-(Arg(Pbf))<sub>8</sub>-PAL-PEG-resin was shaken with a mixture of **9** (55 mg, 0.15 mmol), HATU (57 mg, 0.15 mmol) and DIPEA (0.05 mL, 0.3 mmol) in DMF (5 mL) for 5 h. The solution was then drained, washed with DMF (3 x 10 mL), MeOH (3 x 10 mL) and CH<sub>2</sub>Cl<sub>2</sub> (3 x 10 mL). The Pbf deprotection and cleavage from the resin was achieved by treatment with a cocktail mixture of TFA:H<sub>2</sub>O:TIPS for 5 h. The crude product was purified by RP-HPLC on *Supelco Discovery BIO Wide Pore C18 column* (25 cm x 21.2 mm, 10  $\mu$ m *Sigma-Aldrich*, eluent system MeCN (0.1 % TFA): H<sub>2</sub>O (0.1 % TFA)) by applying a linear gradient of 0%-18% (MeCN) in 15 min followed by an isocratic gradient of 18 : 80 up to 30 min: t<sub>R</sub> 25.3 min. **HR-MS-ESI:** *m/z* Calcd. for C<sub>62</sub>H<sub>123</sub>N<sub>34</sub>O<sub>15</sub>P: 1615.9723 [M+H]<sup>+</sup>: *m/z* Found: 1615.9696 [M+H]<sup>+</sup>.