

## Supplementary Tables

Supplementary Table 1. Strains and plasmids used in this study

	Description	Source
Strains		
R6	Wild type	ATCC BAA 255
R6M2B	R6 clone selected from CIP resistance	This work
T5-R6M2B	R6 clone transform with gDNA from R6M2B selected for CIP resistance	This work
T1-60827	R6 clone transformed with gDNA from 60827 selected for CIP resistance	This work
14599	clinical isolate	Dr. GG Zhanel
14635	clinical isolate	Dr. GG Zhanel
14636	clinical isolate	Dr. GG Zhanel
64933	clinical isolate	Dr. GG Zhanel
45089	clinical isolate	Dr. GG Zhanel
51597	clinical isolate	Dr. GG Zhanel
59774	clinical isolate	Dr. GG Zhanel
60827	clinical isolate	Dr. GG Zhanel
60911	clinical isolate	Dr. GG Zhanel
45693	clinical isolate	Dr. GG Zhanel
60827	clinical isolate	Dr. GG Zhanel
R6 pFF60043	R6 transformed with plasmid pFF60043, KAN <sup>r</sup>	This work
R6M2B pFF6patA	R6 transformed with plasmid pFF6patA, KAN <sup>r</sup>	This work
R6M2B pFF6patB	R6 transformed with plasmid pFF6patB, KAN <sup>r</sup>	This work
R6M2B pFF60043	R6M2B transformed with plasmid pFF60043, KAN <sup>r</sup>	This work
R6M2B pFF6patA	R6M2B transformed with plasmid pFF6patA, KAN <sup>r</sup>	This work
R6M2B pFF6patB	R6M2B transformed with plasmid pFF6patB, KAN <sup>r</sup>	This work
T5-R6M2B pFF60043	T5-R6M2B transformed with plasmid pFF60043, KAN <sup>r</sup>	This work
T5-R6M2B pFF6patA	T5-R6M2B transformed with plasmid pFF6patA, KAN <sup>r</sup>	This work
T5-R6M2B pFF6patB	T5-R6M2B transformed with plasmid pFF6patB, KAN <sup>r</sup>	This work
T1-60827 pFF30043	T1-60827 transformed with plasmid pFF30043, CM <sup>r</sup>	This work
T1-60827 pFF3patA	T1-60827 transformed with plasmid pFF3patA, CM <sup>r</sup>	This work
T1-60827 pFF3patB	T1-60827 transformed with plasmid pFF3patA, CM <sup>r</sup>	This work

60827 pFF30043	60827 transformed with plasmid pFF30043, CM <sup>r</sup>	This work
60827 pFF3patA	60827 transformed with plasmid pFF3patA, CM <sup>r</sup>	This work
60827 pFF3patB	60827 transformed with plasmid pFF3patB, CM <sup>r</sup>	This work
R6 <sup>parC-R6M2B</sup>	R6 transformed with <i>parC</i> PCR fragment from R6M2B	This work
R6 <sup>spr1902-T5R6M2B,smR</sup>	R6 transformed with <i>spr1902</i> PCR fragment from T5-R6M2B and <i>rpsL</i> <sup>+</sup> streptomycin resistance marker, SM <sup>r</sup>	This work
R6 <sup>parC, gyrA-R6M2B</sup>	R6 <sup>parC</sup> transformed with <i>gyrA</i> PCR fragments from R6M2B	This work
R6 <sup>parC, gyrA, spr1902-T5R6M2B,smR</sup>	R6 <sup>parC,gyrA-R6M2B</sup> co-transformed with <i>spr1902</i> PCR fragment from T5-R6M2B and <i>rpsL</i> <sup>+</sup> , SM <sup>r</sup>	This work
R6 <sup>parC, gyrA, spr1544-T5R6M2B,smR</sup>	R6 <sup>parC,gyrA-R6M2B</sup> co-transformed with <i>spr1544</i> PCR fragment from T5-R6M2B and <i>rpsL</i> <sup>+</sup> , SM <sup>r</sup>	This work
R6 <sup>patB,T1-60827,smR</sup>	R6 co-transformed with <i>patB</i> fragment from T1-60827 and <i>rpsL</i> <sup>+</sup> ( contain mutations <b>T142G</b> ;T357C; T618C;C1260T), SM <sup>r</sup>	This work
R6 <sup>patA,T1-60827,smR</sup>	R6 co-transformed with <i>patB</i> fragment from T1-60827 and <i>rpsL</i> <sup>+</sup> ( contain mutations <b>G335T</b> ; T537C; A540C; C660T) SM <sup>r</sup>	This work
T5-R6M2B <sup>spr0335R6,smR</sup>	T5-R6M2B co-transformed with <i>spr0335</i> PCR fragment from R6 and <i>rpsL</i> <sup>+</sup> , SM <sup>r</sup>	This work
R6M2B <sup>spr0129R6,smR</sup>	R6M2B co-transformed with <i>spr0129</i> PCR fragment from R6 and <i>rpsL</i> <sup>+</sup> , SM <sup>r</sup>	This work
Plasmids		
pFF3	<i>S. pneumoniae</i> non-replicative vector that contains a CM <sup>r</sup> resistance marker	Unpublished data
pFF6	pFF3 in which the CM <sup>r</sup> resistance marker was replaced with a KM <sup>r</sup> resistance marker.	Unpublished data
pFF60043	plasmid pFF6(KAN <sup>r</sup> ) containing a fragment of <i>spr0043</i> amplified from R6	This work
pFF6patA	pFF6(KAN <sup>r</sup> ) containing a fragment of <i>patA</i> ( <i>spr1887</i> ) amplified from R6	This work
pFF6patB	pFF6(KAN <sup>r</sup> ) containing a fragment of <i>patB</i> ( <i>spr1885</i> ) amplified from R6	This work
pFF30043	pFF3(CM <sup>r</sup> ) containing a fragment of <i>patA</i> ( <i>spr1887</i> ) amplified from 60827	This work
pFF3patA	plasmid pFF3(CM <sup>r</sup> ) containing a fragment of <i>patA</i> ( <i>spr1887</i> ) amplified from 60827	This work
pFF3patB	Plasmid pFF3(CM <sup>r</sup> ) containing a fragment of <i>patB</i> ( <i>spr1885</i> ) amplified from 60827	This work

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Abbreviations: CIP: Ciprofloxacin; KAN: Kanamycin; CM: Chloramphenicol; SM: streptomycin; CRI; Centre de Recherche en Infectiologie

Supplementary Table 2. Primers used in this study

<b>Primers</b>	<b>Primer sequences</b>
<b>Genes inactivations</b>	
<i>patBF</i> -KO	CCAACCTCCAGCAGAAAGAG
<i>patBR</i> -KO	CCTGACTAGCATCTGGCACA
<i>patAF</i> -KO	GGGGTACCGTTGGTTCATCGCTTCTTT
<i>patAR</i> -KO	CGGGATCCATCCTTTGTTTTGTCCACC
spr0043F-KO	GGGAGATATGACCTTCAAGC
spr0043R-KO	AAACTGCTAGTCGCCTCATC
<b>Mutants reconstruction</b>	
parC-F	TGGGTTGAAGCCGGTTCA
parC-R	CAAGACCGTTGGTTCTTTC
gyrA-F	TTCTCTACGGAATGAATG
gyrA-R	GATATCACGAAGCATTTCAG
spr0129(L)-F	TGTTGCTTGCGTTTATGGAG
spr0129(L)-R	TGATAGAGAAATTTTTATGA
spr1902-F	ATGGAAAACAAACCGTCGCCGT
spr1902-R	TTAAGACCACTCATTTTCTGC
spr1902(L)-F	CCATATTGAAGGCCAAGTCCT
spr1902(L)-R	GTTGCAGCTGCCTTTGATATGG
spr0335(L)-F	TGACCGTTTTGGCGCTTGTATC
spr0335(L)-R	ATTTTGTAGCTGACCATTGA
spr1544(L)-F	TTCAGTTCTGTTTTTCAATATCG
spr1544(L)-R	TGGTACGACCTGTAAATTGGT
<i>uppatA</i> -F	AAACCAAGACTCACTAGTTA
<i>patB</i> -R	TTATTTCGAAAACAAATTGATTGTG
<b>qRT-PCR</b>	
RT16sRNA-F	CCTATTGTTAGTTGCCATCATTTCAG
RT16srRNA-R	GACTCGTTGTACCAGCCATTGT
<i>RTpatA</i> -R	GGTGGCAAGGATATTCGAGA
<i>RTpatA</i> -R	AGAATGGCACGTTGGAGAAC
<i>RTpatB</i> -F	GCCTTTACTGGAGCTGAACG
<i>RTpatB</i> -R	AGGTTGGAGCCTTTTCAGGT
RTspr0081-F	CCTACTACGAAGCAGCGACAG
RTspr0081-R	TACAAGCGGAGTCAACTGAGG
RTspr1884-F	CGGTGTCCAACCTTGTC AATG
RTspr1884-R	GAGTGAGATGTCCCGAAAGG

Abbreviation: F: Forward; R: Reverse;(L): Long fragment; up: upstream; qRT-PCR: quantitative real-time PCR.

Supplementary Table 3. Reversion of the efflux phenotype by reserpine.

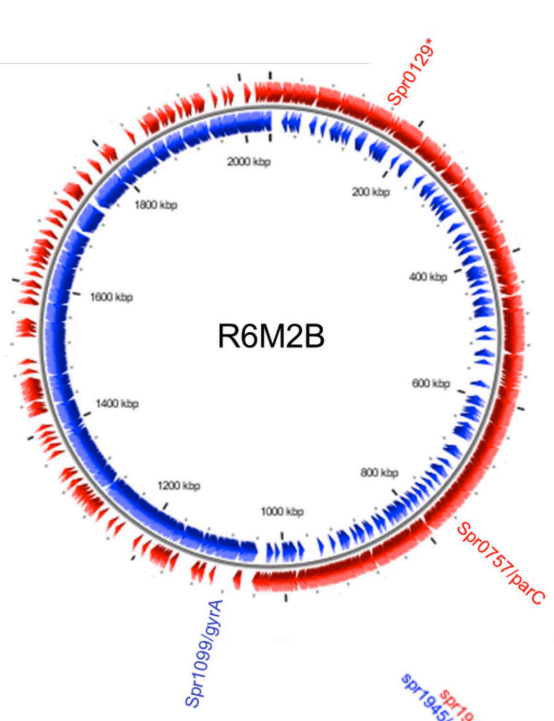
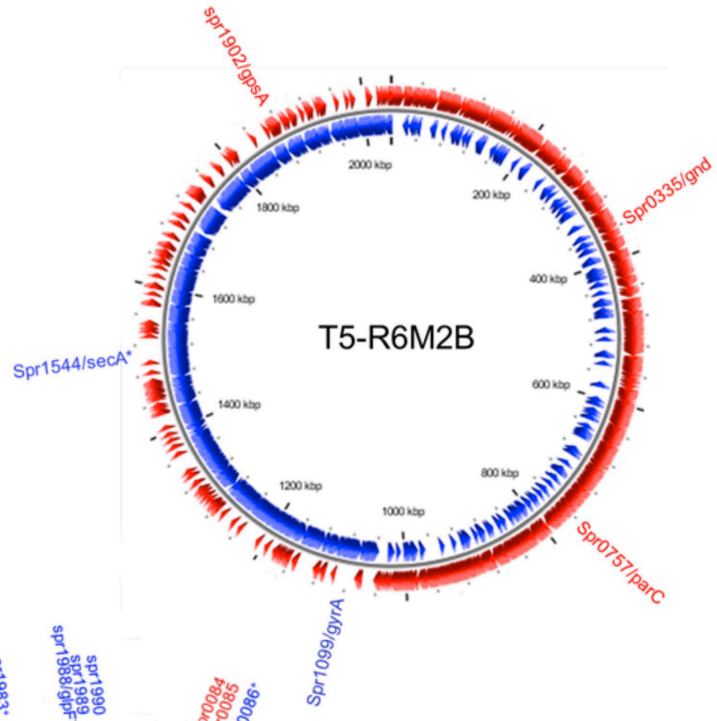
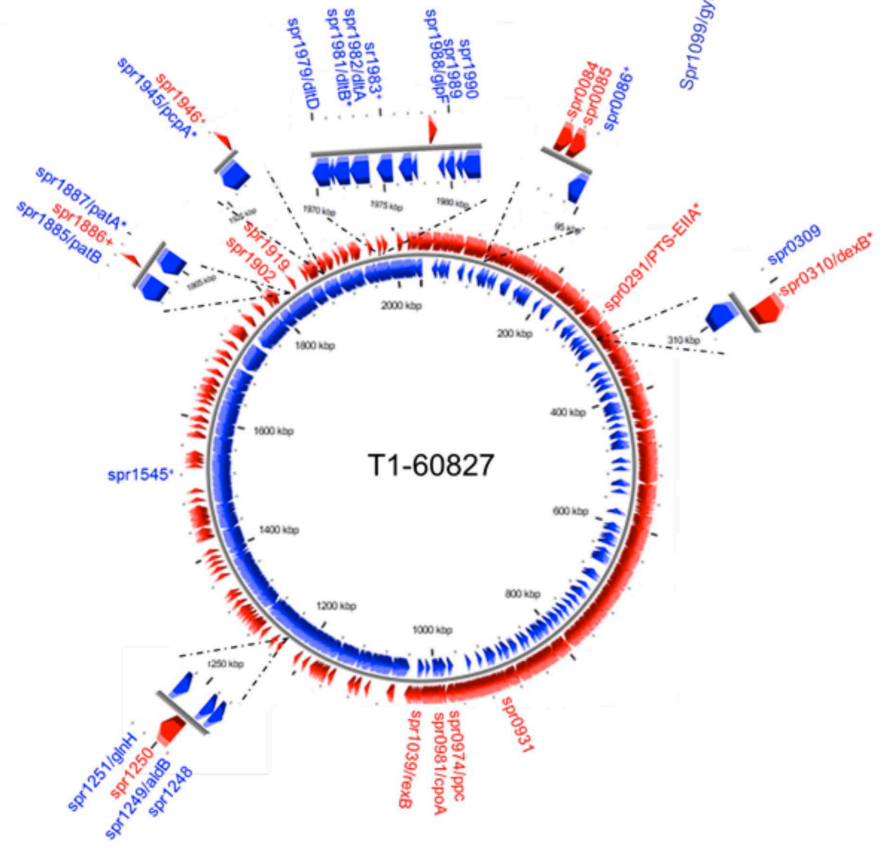
<b>Strains</b>	<b>CIP MIC <math>\mu\text{g/mL}</math></b>				
	<b>0</b>	<b>2.5</b>	<b>5</b>	<b>10</b>	<b>20</b>
<b><math>\mu\text{g/mL}</math> reserpine</b>					
R6	0.5	0.5	0.5	0.5	0.5
T1-60827	2	0.5	0.5	0.5	0.5
60827	2	1	1	0.5	0.5
R6M2B	128	64	64	32	32
T5-R6M2B	128	64	64	32	32

Supplementary Table 4: QRDRs mutations, *patA* expression and CIP MIC in *S. pneumoniae* clinical isolates.

Strains	QRDRs mutations		<i>patA</i> expression <sup>a</sup>	MIC(ug/mL) <sup>b</sup>	
	<i>gyrA</i>	<i>parC</i>		CIP	CIP+R
14599			1.1±0.25	0.5	0.5
14635			1.4±0.3	0.5	0.5
14636			0.734±0.169	0.5	0.5
64933			1.13±0.04	1	1
45089		<i>K137N</i>	1.14±0.234	1	1
51597		<i>S79F</i>	8.87±1.356	8	2
59774			11.13±2.347	4	0.5
60911			11.09±2.67	4	0.5
45693	<i>S81F</i>	<i>S79F</i>	6.98±0.03	128	32

<sup>a</sup>As measured by qRT-PCR

<sup>b</sup>CIP(ciprofloxacin); R(reserpine 20ug/mL). MICs have been measured at least in triplicates.

**A****B****C**

**Figure S1. Schematic representation (in CG view) of the genomic loci mutated in *S. pneumoniae* R6M2B (A), T5-R6M2B (B) and T1-60827 (C).** Genes indicated in red are encoded on the forward DNA strand and genes in blue are encoded on the reverse strand. Genes with an asterisk (\*) have mutation(s) in the region upstream of the ATG and genes with a plus symbol (†) have mutation(s) downstream of their coding region. Abbreviations: *parC*: DNA topoisomerase IV subunit A, *gyrA*: DNA gyrase subunit A, *gnd*: 6-phosphogluconate dehydrogenase, *secA*: preprotein translocase subunit SecA, *gpsA*: NAD(P)H-dependent glycerol-3-phosphate dehydrogenase, *PTS-EIIA*: PTS system IIA component, *dexB*: glucan 1,6- $\alpha$ -glucosidase, *ppc*: phosphoenolpyruvate carboxylase, *cpoA*: glycosyl transferase CpoA, *rexB*: second subunit of major exonuclease, *aldB*: alpha-acetolactate decarboxylase, *glnH*: amino acid ABC transporter amino acid-binding protein, *patB*: ABC transporter ATP-binding protein/permease, *patA*: ABC transporter ATP-binding protein/permease, *pcpA*: choline binding protein PcpA, *dltD*: D-alanine transfer from undecaprenol-phosphate to the poly(glycerophosphate) chain, *dltC*: D-alanine--poly(phosphoribitol) ligase subunit 2, *dltB*: D-alanine transfer from Dcp to undecaprenol-phosphate, *dltA*: D-alanine--poly(phosphoribitol) ligase subunit 1, *glpF*: glycerol uptake facilitator protein.