

1 **Isolation of butanol- and isobutanol-tolerant bacteria and physiological characterization**  
2 **of their butanol tolerance**

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5 **SUPPLEMENTAL MATERIAL**  
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7 **Supplemental TABLE S1.** Primers used for the amplification, direct sequencing, and cloning  
8 of the *cfa* gene of strain CM4A.  
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Primer name	Use	Primer sequence (5' to 3')
<i>cfa</i> -F <sup>a</sup>	Amplification	GAGGGAATGCAATGTTAG
<i>cfa</i> -R <sup>a</sup>	Amplification	TCTATTAACCAATCCGG
<i>cfa</i> 362R <sup>a</sup>	Sequencing	CCTAGATCGTAATGGCTGTG
<i>cfa</i> 871F <sup>a</sup>	Sequencing	GGTGGCTATATTCCTGGTG
<i>cfa</i> -5EF <sup>b</sup>	Cloning	<u>CCGAATTC</u> GAAATGCAATGTTAG
<i>cfa</i> -5XR <sup>b</sup>	Cloning	<u>CCCTCGAGATTAACCAATCCG</u>

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11 <sup>a</sup>The primers were designed from the *cfa* gene sequence in the *Enterococcus faecalis* V583  
12 genome (NC004668). <sup>b</sup>Primers containing *Eco*RI and *Xho*I sites, as underlined, were used to  
13 amplify the *cfa* gene for cloning into the pET-28b expression vector (Novagen) to produce an  
14 N-terminal His 6-tagged fusion protein.  
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**Supplemental TABLE S2.** Cell surface hydrophobicity of strain CM4A grown with or without 2.0% butanol.

CM4A cells	BATH (%) <sup>a</sup>				
	Butanol	<i>n</i> -Hexane	<i>n</i> -Tetradecane	Toluene	Xylene
without butanol	11.6 ± 2.94	22.3 ± 0.26	15.9 ± 0.45	30.5 ± 0.45	21.9 ± 1.58
with 2.0% butanol	0.8 ± 4.11	4.3 ± 1.62	6.5 ± 1.39	5.3 ± 1.39	2.6 ± 1.87

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<sup>a</sup>The value represents the percentage of cells adhering to a given solvent in three independent measurements.  $[1 - (\text{OD}_{600} \text{ of aqueous phase after mixing}) / (\text{OD}_{600} \text{ of initial suspension})] \times 100$ .

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**Supplemental TABLE S3.** Changes in the membrane fatty acid compositions of the strain *E. coli*/pCFA and the control strain *E. coli*/pET28 in response to the presence of 0.1 mM IPTG<sup>a</sup>.

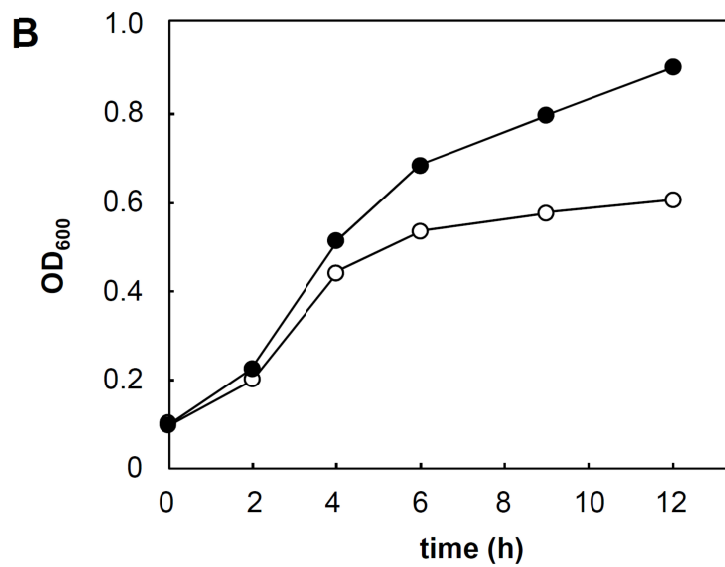
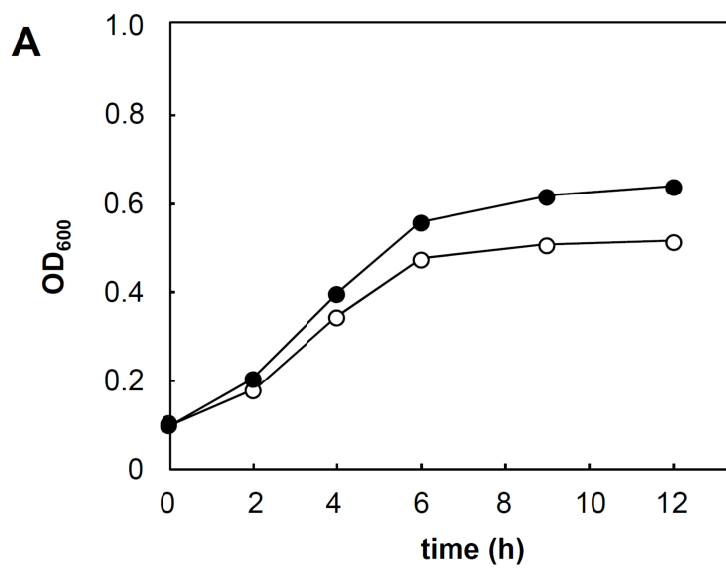
	<i>E. coli</i> /pET28	<i>E. coli</i> /pCFA
C12:0	4.1 ± 0.1	4.3 ± 0.3
C14:0	8.8 ± 0.5	11.1 ± 0.1
C14:0-3OH	3.4 ± 0.3	3.4 ± 1.4
C14:1 $\omega$ 7c	ND	0.1 ± 0.1
C15:0	0.2 ± 0.0	0.1 ± 0.1
C16:0	43.2 ± 0.2	45.2 ± 0.6
C16:1 $\omega$ 7c	16.3 ± 0.1	17.6 ± 0.6
cyclo-C17:0	3.9 ± 0.1	5.6 ± 0.7
C18:0	0.5 ± 0.1	0.2 ± 0.2
C18:1 $\omega$ 7c	19.4 ± 0.7	6.1 ± 0.6
cyclo-C19:0	0.3 ± 0.0	6.5 ± 1.3

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<sup>a</sup>Each fatty acid composition is described as a percentage of the total fatty acids. The values are the means ± standard deviations of three independent measurements. Abbreviations: X:Y $\omega$ Zc, fatty acid containing X carbon atoms with Y double bonds at position Z, counted from the methyl terminus in the *cis* configuration; C14:0-3OH, 3-hydroxy tetradecanoic acid; cyclo-C17:0, *cis*-9,10-methylene hexadecanoic acid; cyclo-C19:0, *cis*-11,12-methylene octadecanoic acid; ND, not detected.

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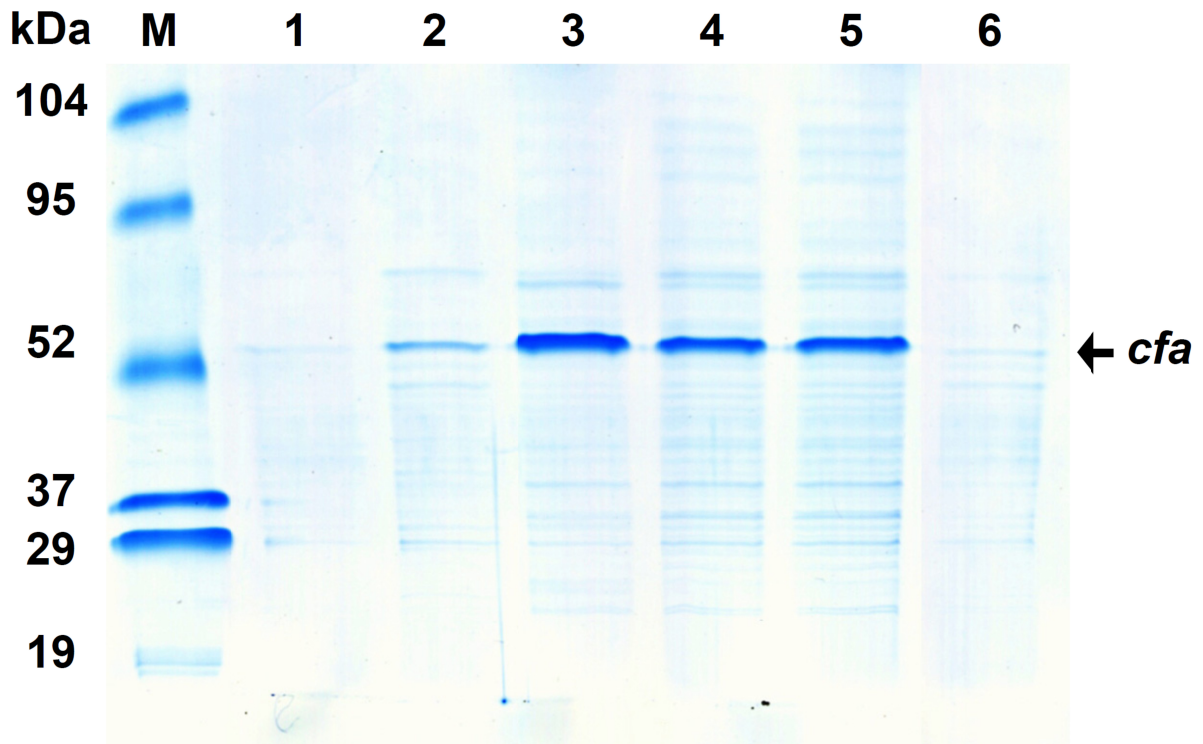
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46 **Supplemental FIG. S1.** Growth of *E. coli/pCFA* (●) and the control strain *E. coli/pET28* (○)  
47 in the presence of 0.8% butanol (A) or 0.8% isobutanol (B). The values represent the mean of  
48 triplicate experiments.

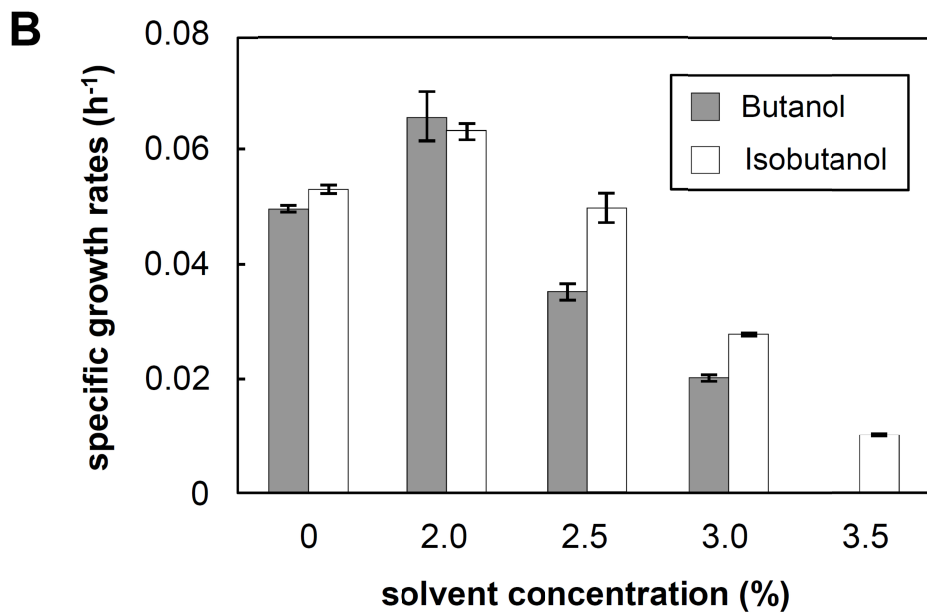
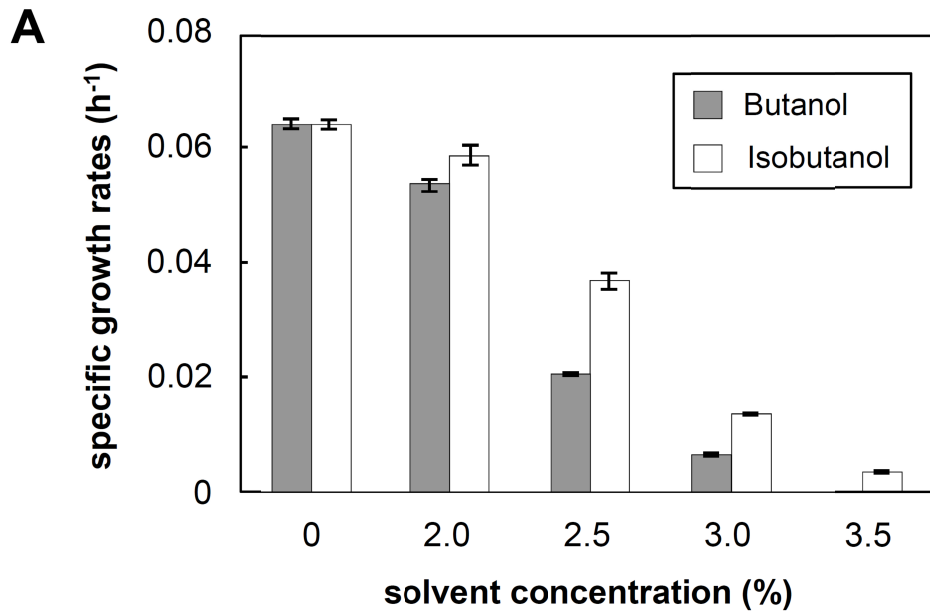
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**Supplemental FIG. S2.** SDS-PAGE analysis of *cfa* gene expression in *E. coli*/pCFA (lane 1-5) and *E. coli*/pET28 (lane 6) at different concentration of IPTG. In total, 60  $\mu$ g of protein in the sonicated supernatant was purified by His-selective nickel affinity gel chromatography and analyzed by SDS-PAGE. Lane M, molecular weight marker; lane 1, 10 mM glucose without IPTG (negative control); lane 2, no IPTG; lane 3, 0.01 mM IPTG; lane 4, 0.1 mM IPTG; lane 5, 1.0 mM IPTG; lane 6, *E. coli*/pET28 grown without IPTG (negative control).



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66 **Supplemental FIG. S3.** Butanol and isobutanol tolerance of strain GK12. The specific  
 67 growth rates of non-adapted (A) and butanol-adapted (B) cells following different butanol or  
 68 isobutanol challenges. Cell adaptation was previously achieved by 15 consecutive passages  
 69 with 2.0% butanol. The values and error bars represent the mean and SD of triplicate  
 70 experiments.

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