

## Supplementary Material

### Raw datasets

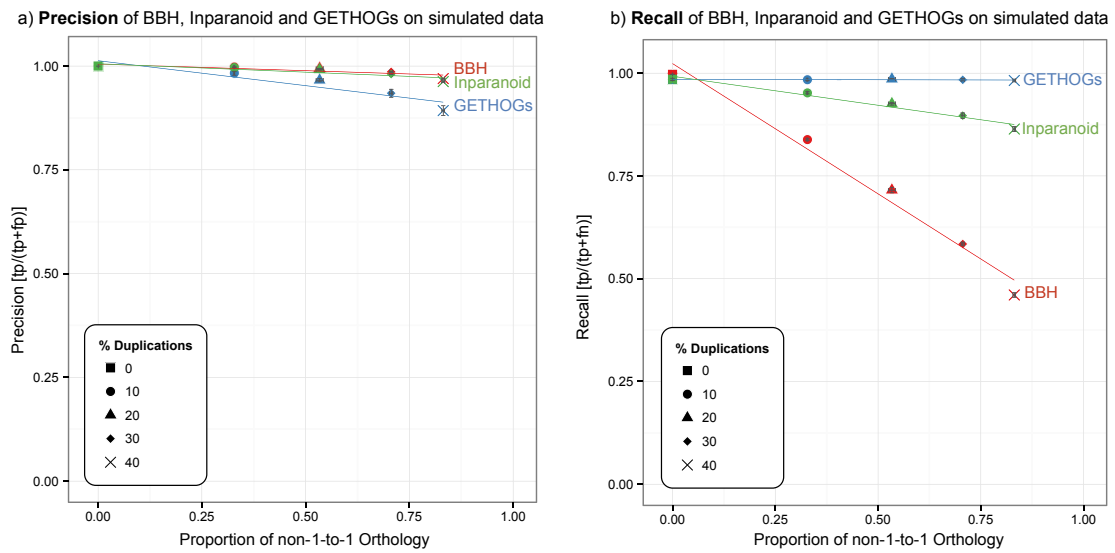
All raw datasets can be downloaded from the following URL:

- [http://lab.dessimoz.org/13\\_bbh](http://lab.dessimoz.org/13_bbh)

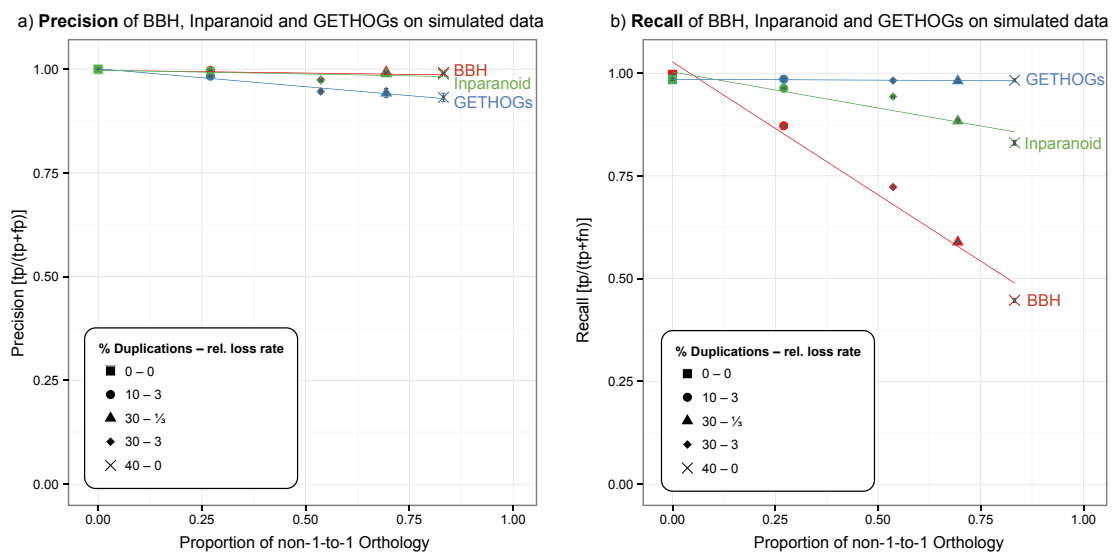
### Additional experiments on simulated data (bacteria tree)

	% Duplications						
	equal loss rate			varying relative loss rate			
	20	30	40	10	30	30	40
<hr/>							
<b>parameters values</b>							
# of sequences				1000			
distr. of seq. length				$\Gamma(k = 2.4, \theta = 133.8)$			
min. sequence length				50			
substitution model				WAG			
insertion and deletion rate				0.000125			
gene duplication rate	0.006	0.0105	0.017	0.0026	0.0087	0.009	0.0125
gene loss rate	0.006	0.0105	0.017	0.0078	0.0029	0.027	0
# of species				30			
<hr/>							
<b>key statistics</b>							
seq. length - mean	321.6	324.4	323.7	323.1	317.9	320.9	324.6
seq. length - stderr	205.5	210.9	207.1	208.9	201.2	205.8	207.2
avg. % gap chars in MSA	22.5	21.44	20.35	19.54	26.48	12.08	31.11
variance of % gap chars	68.4	79.4	90.7	65.2	74.6	56.8	74.9
total tree length				763.6			
minimum tree height				31.70			
maximum tree height				77.80			
average tree height				41.36			
average pairwise distance				72.60			
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**Table 3:** Key statistics for datasets of additional experiments (bacteria tree)

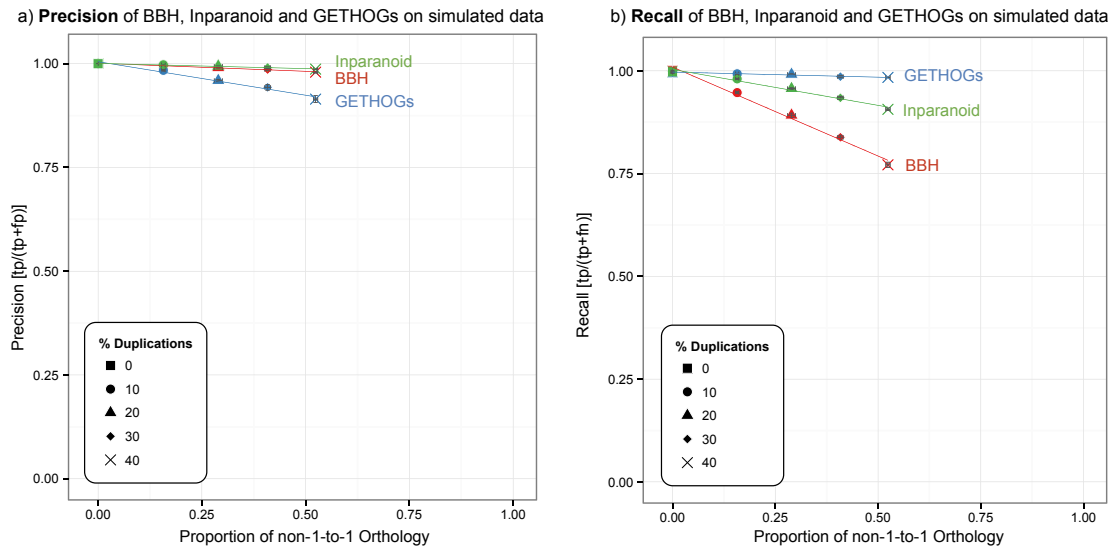


**Supplementary Figure 1: Equal duplication and loss rates.** Relationship between the proportion of non-1-to-1 orthology and precision/recall for BBH (in red) on simulated datasets with different proportions of genes with a history of duplications. Loss rates were equal to duplication rates. Results for Inparanoid (green) and OMA/GETHOGs (blue) are given for comparison. Each point corresponds to the mean value of five replicates. Error bars give the 95% confidence interval of the mean values in both dimensions.

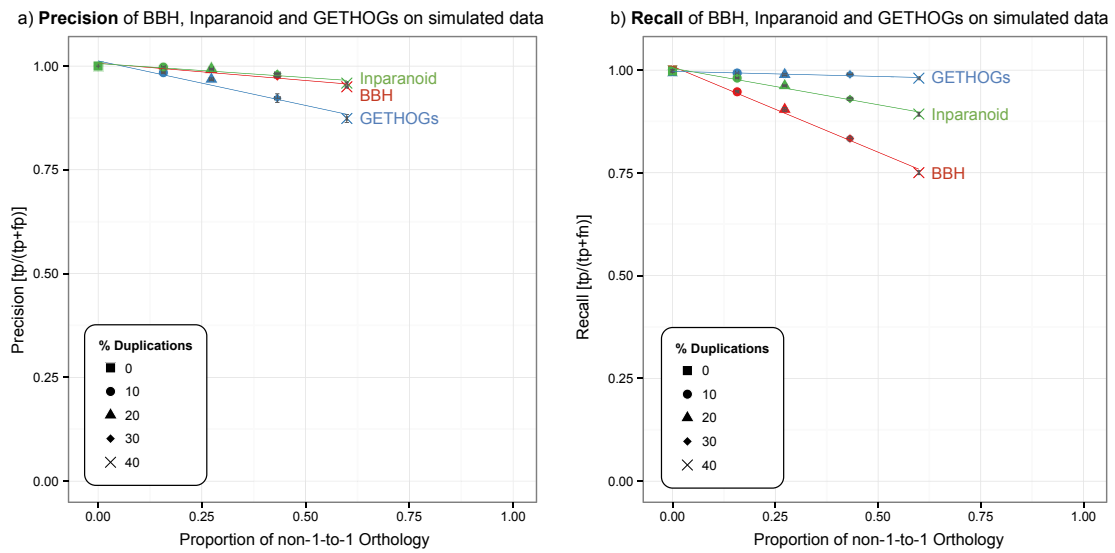


**Supplementary Figure 2: Varying relative loss rates.** Relationship between the proportion of non-1-to-1 orthology and precision/recall for BBH (in red) on simulated datasets with different proportions of genes with a history of duplications. Datasets with 10% duplications had a loss rate that was three times the duplication rate. For the datasets with 30% duplications, loss rate was either a third of or three times the duplication rate. For datasets with 40% duplications, loss rate was set to 0. Results for Inparanoid (green) and OMA/GETHOGs (blue) are given for comparison. Each point corresponds to the mean value of five replicates. Error bars give the 95% confidence interval of the mean values in both dimensions.

## Additional experiments on simulated data (mammalia-like tree)



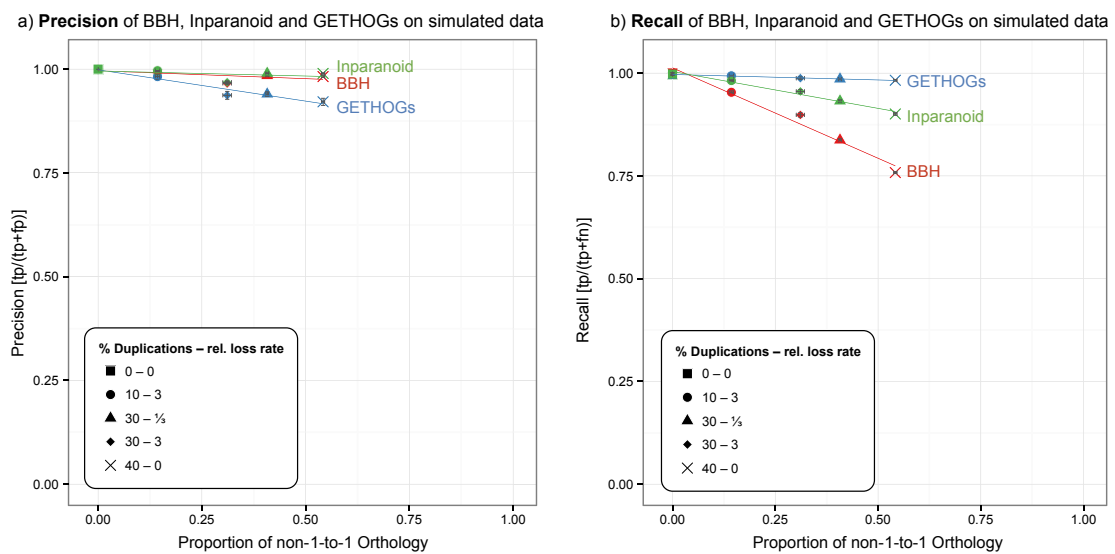
**Supplementary Figure 3: Constant loss rates.** Relationship between the proportion of non-1-to-1 orthology and precision/recall for BBH (in red) on simulated datasets with different proportions of genes with a history of duplications. Loss rate for all datasets was equal to the duplication rate of datasets with 10% duplication. Results for Inparanoid (green) and OMA/GETHOGs (blue) are given for comparison. Each point corresponds to the mean value of five replicates. Error bars give the 95% confidence interval of the mean values in both dimensions.



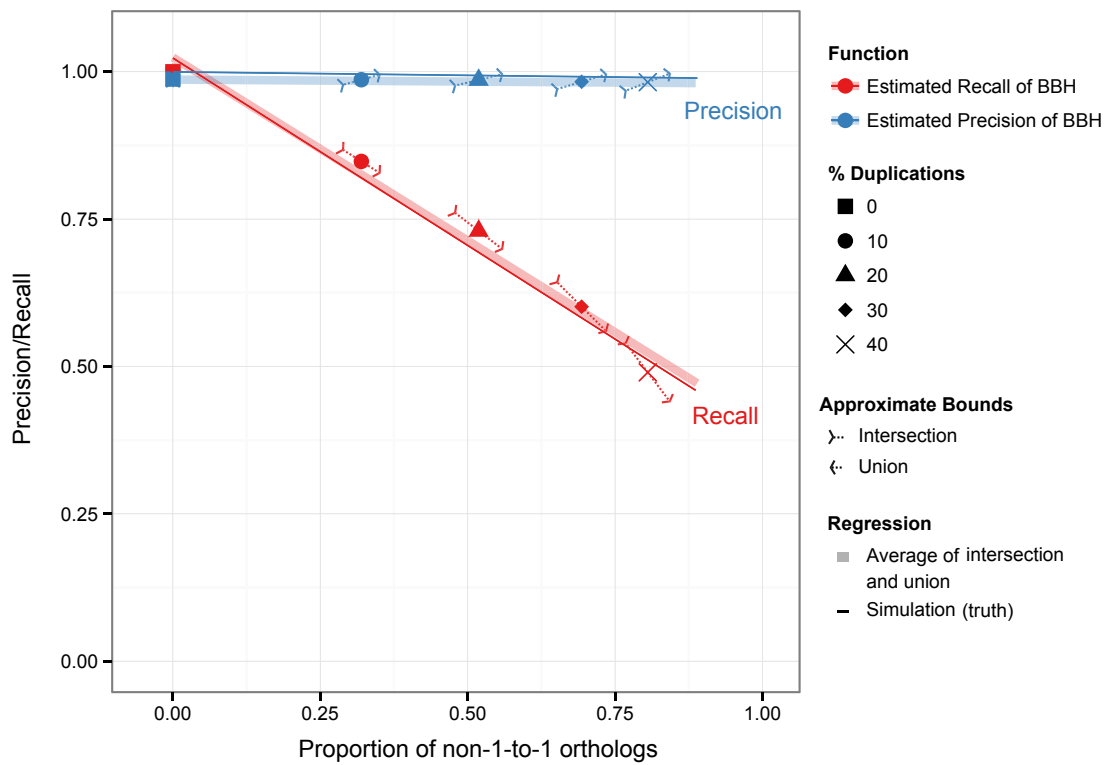
**Supplementary Figure 4: Equal duplication and loss rates.** Relationship between the proportion of non-1-to-1 orthology and precision/recall for BBH (in red) on simulated datasets with different proportions of genes with a history of duplications. Loss rates were equal to duplication rates. Results for Inparanoid (green) and OMA/GETHOGs (blue) are given for comparison. Each point corresponds to the mean value of five replicates. Error bars give the 95% confidence interval of the mean values in both dimensions.

	const. loss rate					% Duplications							
	0	10	20	30	40	equal loss rate	20	30	40	varying relative loss rate	10	30	40
<b>parameters values</b>													
# of sequences						1000							
distr. of seq. length						$\Gamma(k = 1.8, \theta = 274.1)$							
min. sequence length						50							
substitution model						WAG							
insertion and deletion rate						0.000125							
gene duplication rate	0	0.0065	0.013	0.0205	0.0295	0.013	0.025	0.0455	0.0065	0.0201	0.021	0.03	
gene loss rate	0		0.0065			0.013	0.025	0.045	0.0195	0.0067	0.063	0	
# of species						20							
<b>key statistics</b>													
seq. length - mean	487.6	485.8	483.0	487.6	482.8	488.6	488.1	495.5	483.9	483.9	484.2	481.1	
seq. length - stderr	363.2	363.0	363.6	373.1	350.9	367.9	362.5	369.5	358.5	364.4	365.5	356.8	
avg. % gap chars in MSA	4.0	3.87	4.27	4.74	5.29	3.8	3.86	3.75	3.26	4.72	2.35	5.89	
variance of % gap chars	12.6	13.4	15.2	18.6	21.9	15.1	16.4	18.6	12.0	18.7	9.7	23.2	
total tree length						101.2							
minimum tree height						14.70							
maximum tree height						19.18							
average tree height						17.48							
average pairwise distance						14.50							

**Table 4:** Key statistics for datasets of additional experiments (mammalia-like tree)

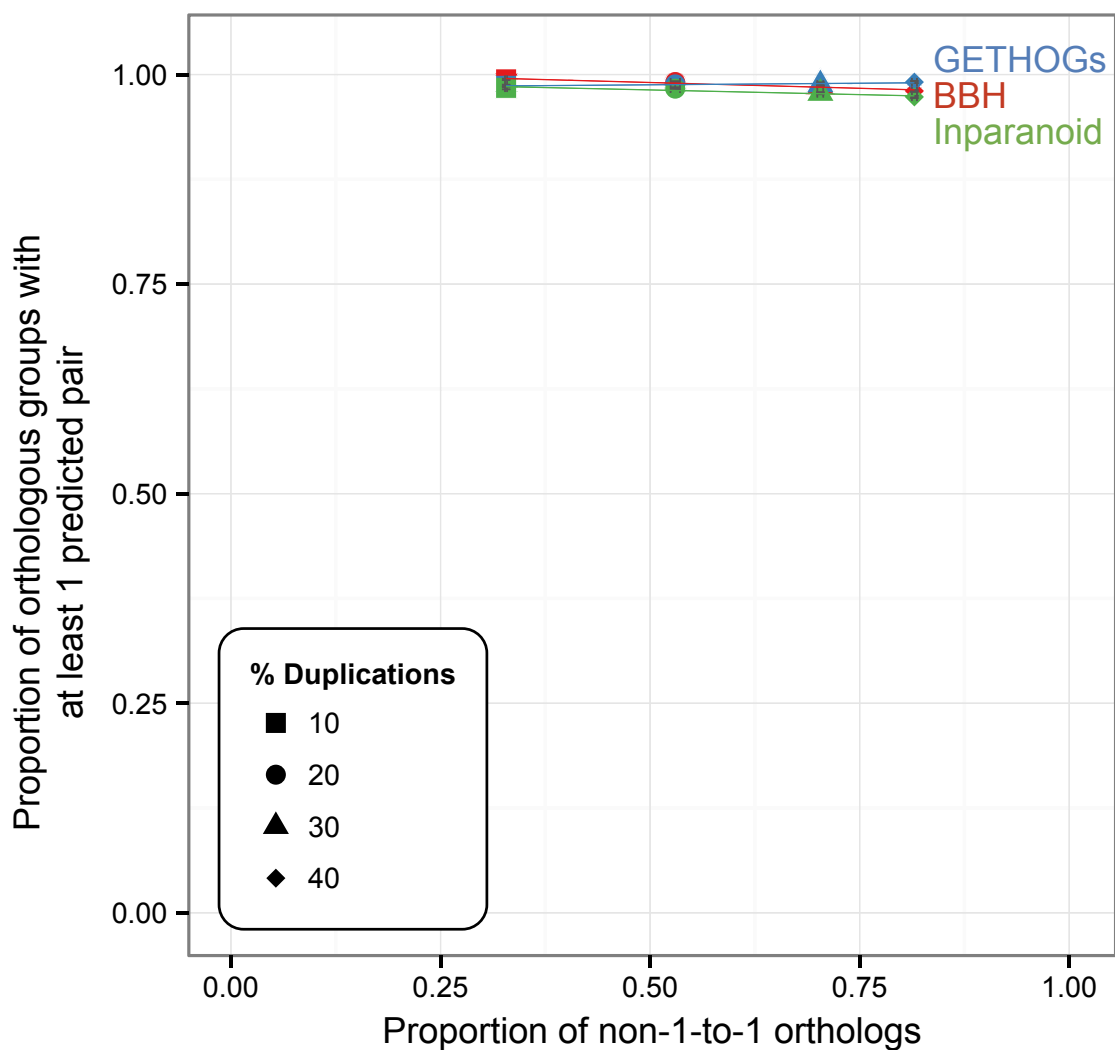


**Supplementary Figure 5: Varying relative loss rates.** Relationship between the proportion of non-1-to-1 orthology and precision/recall for BBH (in red) on simulated datasets with different proportions of genes with a history of duplications. Datasets with 10% duplications had a loss rate that was three times the duplication rate. For the datasets with 30% duplications, loss rate was either a third of or three times the duplication rate. For datasets with 40% duplications, loss rate was set to 0. Results for Inparanoid (green) and OMA/GETHOGs (blue) are given for comparison. Each point corresponds to the mean value of five replicates. Error bars give the 95% confidence interval of the mean values in both dimensions.

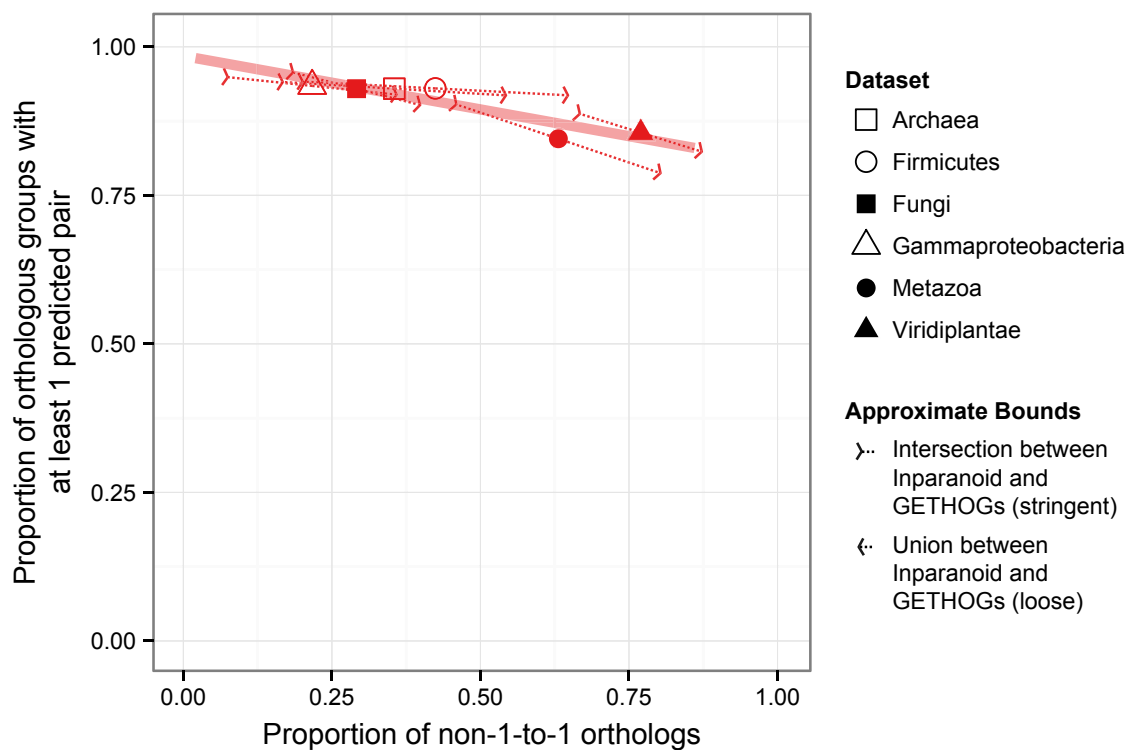


**Supplementary Figure 6: Testing the intersection/union-bound approach on simulated data** Precision and recall of BBH on simulated biological datasets, estimated from the intersection and union sets of orthologs inferred by Inparanoid and GETHOgs—the intersection yielding a lower bound and the union yielding an upper bound for precision and recall. Visually, the estimated trendlines (thick lines) coincide well with the true trendlines (slim solid lines).





**Supplementary Figure 7:** Suitability of BBH to identify ortholog seeds (simulation). Proportion of orthologous groups—defined here as sets of n:m orthologs—for which at least one pair is predicted by BBH (red), as a function of the proportion of non-1-to-1 orthologs. Analysis on simulated datasets, with different proportions of genes with a history of duplications. Loss rate for all datasets was equal to the duplication rate of datasets with 10% duplication. Results for Inparanoid (green) and OMA/GETHOGs (blue) are given for comparison.



**Supplementary Figure 8:** Suitability of BBH to identify ortholog seeds (real data). Proportion of orthologous groups—defined here as sets of n:m orthologs—for which at least one pair is predicted by BBH, as a function of the proportion of non-1-to-1 orthologs. Analysis on real biological datasets, estimated from the intersection and union sets of orthologs inferred by Inparanoid and GETHOGs.

## List of Species

<u>species</u>	<u># of sequences</u>
Aciduliprofundum boonei (strain DSM 19572)	1539
Aeropyrum pernix (strain ATCC 700893)	1699
Ferroglobus placidus (strain DSM 10642)	2463
Haloferax volcanii (strain ATCC 29605)	3986
Ignisphaera aggregans (strain DSM 17230)	1929
Ignicoccus hospitalis (strain KIN4/I)	1434
Korarchaeum cryptofilum (strain OPF8)	1602
Methanosarcina acetivorans (strain ATCC 35395)	4463
Methanopyrus kandleri (strain AV19)	1690
Methanococcus maripaludis	1849
Methanoplanus petrolearius (strain DSM 11571)	2779
Methanothermobacter marburgensis (strain DSM 2133)	1755
Nanoarchaeum equitans (strain Kin4-M)	536
Nitrosopumilus maritimus (strain SCM1)	1795
Picrophilus torridus (strain ATCC 700027)	1535
Pyrococcus furiosus (strain ATCC 43587)	2052
Staphylothermus marinus (strain ATCC 43588)	1570
Sulfolobus islandicus (strain M.16.4)	2729
Thermofilum pendens (strain Hrk 5)	1876
Vulcanisaeta distributa (strain DSM 14429)	2492

**Supplementary Table 1:** List of species in archaea dataset.

<b>species</b>	<b># of sequences</b>
Alicyclobacillus acidocaldarius subsp. acidocaldarius (strain ATCC 27009)	3059
Alkaliphilus metalliredigens (strain QYMF)	4463
Anaerococcus prevotii (strain ATCC 9321)	1795
Caldicellulosiruptor saccharolyticus (strain ATCC 43494)	2619
Clostridium beijerinckii (strain ATCC 51743)	5003
Clostridium cellulolyticum (strain ATCC 35319)	3286
Clostridium phytofermentans (strain ATCC 700394)	3889
Coprothermobacter proteolyticus (strain ATCC 35245)	1481
Desulfitobacterium hafniense (strain Y51)	5015
Desulfotomaculum reducens (strain MI-1)	3214
Finegoldia magna (strain ATCC 29328)	1813
Geobacillus sp. (strain Y412MC10)	6237
Halothermothrix orenii (strain H 168)	2324
Lactobacillus acidophilus (strain ATCC 700396)	1860
Leuconostoc mesenteroides subsp. mesenteroides (strain ATCC 8293)	2002
Moorella thermoacetica (strain ATCC 39073)	2450
Natranaerobius thermophilus (strain ATCC BAA-1301)	2836
Symbiobacterium thermophilum (strain T)	3312
Syntrophomonas wolfei subsp. wolfei (strain Goettingen)	2471
Thermoanaerobacter sp. (strain X514)	2322

**Supplementary Table 2:** List of species in firmicutes dataset.

<b>species</b>	<b># of sequences</b>
Agaricus bisporus	10366
Ashbya gossypii (strain ATCC 10895)	4707
Neosartorya fumigata (strain ATCC MYA-4609)	9832
Botryotinia fuckeliana (strain B05.10)	16299
Candida albicans (strain WO-1)	5695
Candida glabrata (strain ATCC 2001)	4752
Cryptococcus neoformans	6231
Debaryomyces hansenii (strain ATCC 36239)	6263
Emericella nidulans (strain FGSC A4)	10510
Encephalitozoon cuniculi (strain GB-M1)	1895
Kluyveromyces lactis (strain ATCC 8585)	5230
Laccaria bicolor	20052
Lodderomyces elongisporus (strain ATCC 11503)	5772
Magnaporthe grisea	12637
Mycosphaerella graminicola	10932
Neurospora crassa (strain ATCC 24698)	7569
Phaeosphaeria nodorum (strain SN15)	16489
Scheffersomyces stipitis (strain ATCC 58785)	5799
Puccinia graminis f. sp. tritici (strain CRL 75-36-700-3)	20364
Schizosaccharomyces pombe (strain 972)	4958
Ustilago maydis (strain 521)	6510
Yarrowia lipolytica (strain CLIB 122)	6603
Saccharomyces cerevisiae (strain ATCC 204508)	6328

**Supplementary Table 3:** List of species in fungi dataset.

<b>species</b>	<b># of sequences</b>
<i>Acinetobacter baumannii</i> (strain AYE)	3715
<i>Acidithiobacillus ferrooxidans</i> (strain ATCC 23270)	3119
<i>Alcanivorax borkumensis</i> (strain SK2)	2752
<i>Baumannia cicadellinicola</i> subsp. <i>Homalodisca coagulata</i>	595
<i>Buchnera aphidicola</i> subsp. <i>Acyrtosiphon pisum</i> (strain APS)	571
<i>Buchnera aphidicola</i> subsp. <i>Baizongia pistaciae</i> (strain Bp)	507
<i>Buchnera aphidicola</i> subsp. <i>Cinara cedri</i> (strain Cc)	365
<i>Carsonella ruddii</i> (strain PV)	182
<i>Coxiella burnetii</i> (strain Dugway 5J108-111)	2110
<i>Dichelobacter nodosus</i> (strain VCS1703A)	1273
<i>Francisella tularensis</i> subsp. <i>novicida</i> (strain U112)	1719
<i>Legionella pneumophila</i> (strain Paris)	3059
<i>Marinomonas</i> sp. (strain MWYL1)	4415
<i>Pseudomonas aeruginosa</i> (strain UCBPP-PA14)	5886
<i>Ruthia magnifica</i> subsp. <i>Calyptogena magnifica</i>	976
<i>Shewanella baltica</i> (strain OS195)	4616
<i>Thiomicrospira crunogena</i> (strain XCL-2)	2183
<i>Thioalkalivibrio</i> sp. (strain HL-EbGR7)	3272
<i>Wigglesworthia glossinidia brevipalpis</i>	612
<i>Xanthomonas campestris</i> pv. <i>campestris</i> (strain ATCC 33913)	4123

**Supplementary Table 4:** List of species in Gammaproteobacteria dataset.

<b>species</b>	<b># of sequences</b>
<i>Acyrtosiphon pisum</i>	33992
<i>Aedes aegypti</i>	15129
<i>Apis mellifera</i>	10378
<i>Bombyx mori</i>	14577
<i>Branchiostoma floridae</i>	28464
<i>Caenorhabditis remanei</i>	31096
<i>Capitella</i> sp. 1	31562
<i>Ciona intestinalis</i>	15464
<i>Daphnia pulex</i>	30088
<i>Drosophila virilis</i>	14351
<i>Gasterosteus aculeatus</i>	21832
<i>Helobdella robusta</i>	23263
<i>Ixodes scapularis</i>	20454
<i>Lottia gigantea</i>	23514
<i>Nematostella vectensis</i>	26036
<i>Pediculus humanus</i> subsp. <i>corporis</i>	10733
<i>Pristionchus pacificus</i>	29363
<i>Schistosoma mansoni</i>	11404
<i>Strongylocentrotus purpuratus</i>	26882
<i>Trichoplax adhaerens</i>	11466

**Supplementary Table 5:** List of species in Metazoa dataset.

<b>species</b>	<b># of sequences</b>
<i>Arabidopsis lyrata</i>	32085
<i>Arabidopsis thaliana</i>	27489
<i>Chlamydomonas reinhardtii</i>	16900
<i>Hordeum vulgare</i> var. <i>distichum</i>	29214
<i>Zea mays</i>	107258
<i>Manihot esculenta</i>	32737
<i>Oryza sativa</i> subsp. <i>japonica</i>	58246
<i>Ostreococcus lucimarinus</i> (strain CCE9901)	7410
<i>Ostreococcus tauri</i>	7860
<i>Populus trichocarpa</i>	41297
<i>Sorghum bicolor</i>	35447
<i>Vitis vinifera</i>	26207

**Supplementary Table 6:** List of species in Viridiplantae dataset.