# Divergent evolution of protein conformational dynamics in dihydrofolate reductase

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## **Supplementary Figures**



Supplementary Figure 1. X-ray structures and omit maps for ligands built into hDHFR structures. (a) Overlay of ecDHFR (PDB code  $1RX2^1$ ) and hDHFR bound to NADP<sup>+</sup> and folate, showing that hDHFR and ecDHFR are highly conserved at the structural level. ecDHFR is shown in purple with green ligands, and hDHFR is shown in red with yellow ligands. (b-f)  $2F_0$ - $F_c$  omit maps are shown for each ligand, contoured to  $\sigma = 1.0$ . (b) Folate from the hE–NADP<sup>+</sup>–FOL structure. (c) NADP<sup>+</sup> from the hE–NADP<sup>+</sup>–FOL structure. (d) NADPH from the hE–NADP<sup>+</sup>–ddTHF structure. (e) ddTHF from the hE–NADP<sup>+</sup>–ddTHF structure. (f) NADP<sup>+</sup> from the hE–NADP<sup>+</sup>–ddTHF structure. (g-h) As discussed in the text, the density for this NADP<sup>+</sup> ligand was discontinuous, indicating disorder and/or low occupancy. (g) Ligand occupancy was refined using PHENIX, and the ligand is

modeled in at an occupancy of ~0.8.  $2F_O$ - $F_C$  map ( $\sigma = 1$ ; blue) and  $F_O$ - $F_C$  map ( $\sigma = 3$ ; red, negative density; green positive density) after refinement with NADP<sup>+</sup> modeled into the hE–NADP<sup>+</sup>-ddTHF structure. (**h**)  $2F_O$ - $F_C$  map ( $\sigma = 1$ ; blue) and  $F_O$ - $F_C$  map ( $\sigma = 3$ ; red, negative density; green positive density) after refinement with 2 phosphates, 2 glycerols and waters modeled into the NADP-binding site in the hE–NADP<sup>+</sup>-ddTHF structure. Remaining positive density is observed in the adenine region and the nicotinamide region, showing that the density is not satisfied upon modeling in buffer molecules. (**i**)  $2F_O$ - $F_C$  map ( $\sigma = 1$ ; grey) and  $F_O$ - $F_C$  map ( $\sigma = 3$ ; red, negative density; green positive density) after refinement with 6S-ddTHF (blue sticks). (**j**)  $2F_O$ - $F_C$  map ( $\sigma = 1$ ; grey) and  $F_O$ - $F_C$  map ( $\sigma = 3$ ; red, negative density) after refinement with 6S-ddTHF (blue sticks).



Supplementary Figure 2. Hydrogen bonding networks that support the hinge-open conformation. (a-c) Overlay of hE–NADP<sup>+</sup>–FOL (cyan) and hE–NADPH (light pink), aligned on the "Met20" loop (a), residues 71-114 (b) and helix  $\alpha F$  (c). NADP<sup>+</sup> corresponding to hE– NADP<sup>+</sup>–FOL is in blue, and NADPH corresponding to the hE–NADPH structure is in pink. Ligands and side chains are colored by element: nitrogen blue, oxygen red, phosphorus orange, carbon as described for each ligand or structure. Hydrogen bonds are shown as blue dashed lines for E–NADP<sup>+</sup>–FOL and pink dashed lines for E–NADPH. Hydrogen bonds stabilizing the nicotinamide moiety (a) and the adenosine moiety (b) are present in both conformations. (c) Hydrogen bonds from the diphosphate groups of NADPH to Ser119 (pink dashed lines) stabilize the "hinge-open" conformation. These hydrogen bonds are broken in the "hinge-closed" conformation, as other hydrogen bonds are formed to folate (Supplementary Table 2). (d-e) Hydrogen bonds for Hinge 1 in hDHFR (d) and ecDHFR (e). Many hydrogen bonds stabilize Hinge 1 in human DHFR. Residues in the loop subdomain are colored purple, and residues in the adenosine-binding subdomain are coloured green. (f). The hinge-open conformation is stabilized in C. albicans DHFR. An overlay of caDHFR bound to NADPH (purple, PDB code: 1AI9<sup>2</sup>) with hE–NADP<sup>+</sup>–FOL (green, hinge-closed) and hE–NADPH (cyan, hinge-open) show that the αF helix in cDHFR is aligned with that in hE–NADPH. The structures are aligned on the loop subdomain. Ser119, which stabilizes the hinge-open conformation in hDHFR, has been substituted with Glu116 in caDHFR, which can also stabilize the hinge-open conformation via a water-mediated hydrogen bond to NADPH.



**Supplementary Figure 3. Supporting NMR data for hDHFR. (a-c)** Numerous residues in hE–NADP<sup>+</sup>–FOL show <sup>15</sup>N R<sub>1p</sub> dispersion at 280 K, but <sup>15</sup>N R<sub>2</sub> dispersion is not observed in hDHFR. (a) Selected residues showing <sup>15</sup>N R<sub>1p</sub> dispersion in regions A, B and C as well as a

region that likely "gates" NADP flux are shown in red on the hE–NADP<sup>+</sup>–FOL structure. Ligands are shown as sticks. (**b**) Examples of  $R_{1\rho}$  dispersion curves. Residue numbers are indicated in the top right for each curve. (**c**) Representative data for hE–NADP<sup>+</sup>–FOL <sup>15</sup>N R<sub>2</sub> dispersion experiments at 500 MHz (black) and 800 MHz (red). These data were collected at 303 K. No dispersion was observed for data collected at 280 K, 292K, 298 K, or 309 K. (**d**-e) <sup>15</sup>N <sup>1</sup>H chemical shift differences in hDHFR binary and ternary complexes. (**d**) Overlay of hE–NADP<sup>+</sup>– FOL (black) and hE–NADPH (magenta) <sup>15</sup>N HSQC spectra at pH 8.0 and a temperature of 300 K. (**e**) Weighted average <sup>1</sup>H, <sup>15</sup>N chemical shift differences between hE–NADP<sup>+</sup>–FOL and hE–NADPH (shown in A), mapped onto the hDHFR structure. Chemical shift differences are colored onto the structure using a gradient of thickness and red to white, with red, thick regions representing the largest chemical shift differences between the two complexes; ligands are shown as dark gray sticks. Large changes are observed in helix  $\alpha$ F and hinge 2. Chemical shift differences were calculated as follows:

$$\sqrt{\left(\Delta^{1} \mathrm{H}\right)^{2} + \left(\frac{\Delta^{15} \mathrm{N}}{5}\right)^{2}}$$



**Supplementary Figure 4. Supporting NMR data for** <sup>21</sup>**PWNAL**<sup>24</sup> **ecDHFR mutant. (a)** The <sup>21</sup>**PWNAL**<sup>24</sup> *E. coli* DHFR mutant is structurally similar to N23PP (or <sup>21</sup>PWPPL<sup>24</sup>) ecDHFR. Overlay of <sup>21</sup>**PWNAL**<sup>24</sup> ecDHFR <sup>15</sup>N HSQC spectrum (red) with N23PP ecDHFR <sup>15</sup>N HSQC spectrum (black), with both enzymes bound to NADP<sup>+</sup> and FOL, showing that resonances do not shift significantly. (**b-c**) <sup>15</sup>N R<sub>2</sub> dispersion is present, but considerably dampened, for a few active site residues in <sup>21</sup>**PWNAL**<sup>24</sup> ecDHFR. (**b**) Residues showing <sup>15</sup>N R2 dispersion in relaxation compensated constant time CPMG experiments are plotted as spheres on the wild type ecDHFR–NADP<sup>+</sup>–FOL structure (3QL0<sup>3</sup>). Dispersion profiles for C-terminal associated residues (blue) are unaffected by the mutation. All other residues exhibiting dispersion are located in the active site and are shown in red. (**c**). <sup>15</sup>N R<sub>2</sub> dispersion curves for active site residues in the mutant, and corresponding curves for the wild type enzyme. The few residues that do show dispersion in the mutant have significantly reduced R<sub>ex</sub> values.

а	Region A	Conformational flexibility	v across h-transfer step	<sub>b</sub> , d
E. coli (WT)	PWN-LPAD	Yes Reduced	1	
E. COII ("PWPPL")	PWPPLPAD	Reduced		
E. COII ("PWNAL")	PWNALPAD	Yes		
B. anthracis	PWR-LPSE	Yes		
6 proumoniae	PHR-LF-HD	Yes		
V cholorao	PHE-DPAE	Yes		
C elegans	PWR-LPAD	Yes		
D. rerio	DWNDIDI SNP	No		Region C
R. norvegicus	PWPLIP-NP	No		
S. scrofa	PWPPI.RNE	No		Begin
B. taurus	DWDDI.D NP	No		I A A A A A A A A A A A A A A A A A A A
H. sapiens	PWPPLRNE	No		
C	Region B	с	Region C	
E. coli (WT)	FKRNTLNKPVIM	E. coli (WT)	YEQFLPKAQKLYLT	
E. coli (21PWPPL24)	FKRNTLNKPVIM	E. coli (21PWPPL24)	YEOFLPKAOKLYLT	
E. coli (21 PWNAL24)	FKRNTLNKPVIM	E. coli ( <sup>21</sup> PWNAL <sup>24</sup> )	YEQFLPKAQKLYLT	
B. anthracis	VKKTTMGHPLIM	B. anthracis	YDLFLPYVDKLYIT	
S. aureus	VKKLSTGHTLVM	S. aureus	FEEMIDKVDDMYIT	
S. pneumoniae	FRETTLNHAILM	S. pneumoniae	FQAFEPYLDEVIVT	
V. cholerae	FKRCTLGKPVVM	V. cholerae	YAACLPMAHKLYIT	Region
C. elegans	FASVTKNVSDQSKRNAVLM	C. elegans	YDLALRENLVDEIHLT	
D. rerio	FQKMIMIPSDEGKKNVVIM	D. rerio	YKEVMERSGHRRLFVT	
R. norvegicus	FURMITISSVEGRÜNLVIM	R. norvegicus	YQEAMNQPGHLRLFVT	3
S. scrofa	FORMITISSVEGRONLVIM	S. scrofa	YKEAMNKPGHIRLFVT	
B. taurus	FORMITVSSVEGRONLVIM	B. taurus	YKEAMNKPGHVRLFVT	
H. sapiens	FORMTTISSVEGRONLVIM	H. sapiens	YKEAMNHPGHLKLFVT	



**Supplementary Figure 5. Sequence analysis of DHFRs from different species. (a-d)** Sequence alignments of DHFR enzymes for which NMR data were obtained. (e) Tree of eukaryotic DHFR sequences showing lengths of Regions A, B and C. *E. coli* DHFR is also shown for comparison.



Supplementary Figure 6. <sup>15</sup>N HSQC spectra for bacterial DHFRs (a-d) and eukaryotic DHFRs (e-i). Spectra show DHFRs in the E-NADP<sup>+</sup> – FOL (black) or E-NADP<sup>+</sup> –THF (red) complexes. Spectra are shown for *S. aureus* (a), *S. pneumoniae* (b), *B. anthracis* (c), *V. cholerae* (d), *S. scrofa* (e), *B. taurus* (f), *R. norvegicus* (g), *C. elegans* (h), and *D. rerio* (i).

# **Supplementary Tables**

	hE–NADP <sup>+</sup> –FOL		hE–NADPH				
Donor	Acceptor	Distance	Donor	Acceptor	Distance		
A9-N	NAP-NO7	2.9	A9-N	NDP-NO7	3.0		
K54-N	NAP-AO4*	3.0	K54-N	NDP-AO4*	3.0		
K54-NZ	NAP-AOP2	2.8	K54-NZ	NDP-AOP2	2.5		
K55-N	NAP-AO5*	3.1	K55-N	NDP-AO5*	3.2		
T56-N	NAP-AO2	2.9	T56-N	NDP-AO2	3.0		
T56-OG1	NAP-AO2	2.7	T56-OG1	NDP-AO2	2.6		
S76-OG	NAP-AOP2	2.6	S76-OG	NDP-AOP2	2.5		
R77-N	NAP-AOP1	2.7	R77-N	NDP-AOP1	2.8		
G117-N	NAP-AO1	3.2	G117-N	NDP-AO1	3.1		
			G117-N	NDP-AO2	3.1		
S118-N	NAP-NO2	3.0	S118-N	NDP-NO5*	2.9		
S118-OG	NAP-NO2	3.3					
			S119-OG	NDP-AO1	2.8		
			S119-N	NDP-NO2	2.8		
			S119-OG	NDP-NO2	3.1		
			NDP-NN7	A9-0	2.8		
NAP-AN1	R91-O	3.4					
NAP-NN7	А9-О	2.8					
NAP-NN7	I16-O	3.0	NDP-NN7	I16-O	3.2		
NAP-AN7	S119-OG	3.2					
N64-ND2	FOL-O	2.9					
R70-NH1	FOL-O1	2.9					
R70-NH2	FOL-O2	2.8					
FOL-N3	E30-OE1	2.8					
FOL-NA2	E30-OE2	2.8					

# Supplementary Table 1. Ligand-protein hydrogen bonds in hDHFR

**Supplementary Table 2**. **Table of hydrogen bonds in both hinge regions for human and** *E. coli* **DHFRs.** The extensive network of hydrogen bonds in the long hinge 1 of hDHFR supports the rigid body hinge-twisting motion that opens the active site.

HINGE 1										
Human I	OHFR, E–NADP-	<i>E. coli</i> DHFR, E–NADP+–FOL								
Donor	Acceptor	Distance	Donor	Acceptor	Distance					
T39-N	Q35-O	2.9	L36-N	K32-O	2.9					
T39-OG	Q35-O	2.7	D37-N	G56-O	2.8					
T40-N	R36-O	3.1	K38-N	Т35-О	3.2					
T40-OG1	R36-O	2.7	K38-NZ	E90-OE1	2.8					
S41-OG	V43-O	2.8	K38-NZ	N34-O	2.9					
S41-OG	K46-O	3.2								
S41-OG	D110-OD2	3.5								
S42-N	D110-OD2	2.8								
S42-OG	D110-OD2	2.7								
V43-N	S41-OG	3.0								
K46-N	V43-O	3.2								
Q47-N	K108-O	3.0								
N48-ND2	Т38-О	2.9								
N48-ND2	Т40-О	3.0								
L49-N	M111-O	2.9								
R70-NH1	T38-OG1	2.9								
I71-N	N48-O	2.9								
D110-N	Q47-O	2.8								
M111-N	N48-OD1	2.9								
W113-N	L49-O	2.8								
		HING	E 2							
Human I	OHFR, E–NADP-	E. coli D	HFR, E–NADI	P+-FOL						
H127-N	A124-O	3.0	K106-N	F103-O	3.0					
H127-ND1	P128-O	2.8								
G129-N	D186-O*	2.8	A107-N	L104-O	3.1					
L131-N	K184-O	3.1	R158-NE	A107-O	2.8					
K184-NZ	H127-O	2.7								
K184-NZ	G129-O	2.7	R158-NE	A107-O	3.2					
K184-N	L131-O	3.0								

\*C-terminal residue

# **Supplementary Table 3. Complete list of fully sequenced genomes analyzed.** An excel spreadsheet is also provided.

Metazoa Acystosiphon pisum	pea aphid	Ecdysczca/Arthropoda/Insects	Baylor	07.05.2010	Acyt_10()	NCBI Build 1	protein fa gz	fip.ifip.ncbi.nim.nih.govigenomes/Acyrthosiphon_pisumiprotein/
Aedez aegypti Amphimedon cueenzlandica	Yellowfever mosquito Soonge	Ecdysczca/Arthropoda/Insecta Potfera	Ensembl JGI	12.07.2007	mil 45 v1	vl	Andex_segyptiAaegL145.pep.al.ts Acuerniandica_v1.pep.fata	http://www.ensembil.org/infoldataldowniced.html ftp://tip.io-out.org/au/JCI_data/Amphimedon_pueenalandica/annotation/
Anoliz carolinensis	Green anole	Vertebrata/Amniota	Broad Institute of Harvard and MIT	07.05.2010	nel 57		Anola, carolinensia AnoCar1.0.57 pep all fa gr	fp://tp.ensembl.org/publicument_fastulanola_carolinemia.jpp/
Apiz melifera	Honeybee	Ecdysczoa/Arthropoda/Insecta	HGSC at Baylor College of Medicine	16.11.2007	Release1 of the OGS		Ame_release1_OGS_pep.ts	htp://www.man.edu/download/ASTA.html
Bonbyx mori Boz teuruz	Sik moth Cow	Ecdysczoa/Arthropoda/Insecta Vertebrata/Mammalia	Beijing Genomics Institute Ensembl	16.11.2007 21.05.2010	nel 57		SW_ge2k_BGF.pep Bos_teurus.Btau_4.0.57.pep.al.fs.gz	htp://sikkwom.genomics.org.or/jepidowniosd.jep ftp://tip.ensembi.org/publicurent_fastarbos_taurus/pepi
Branchiostome floridae Brugia malayi	Florida lancelet, Amphioxus Filarial nematode worm	Cephalochordata Ecdysczoa/Nematoda	JGI Wornbase	12.07.2007	1	v 1.0 (March 2006) 9/29/2007	Brafi fasts bra 1.pep.fa.gz	htip://genome.jpi.paf.org/Braff1/Braff1.home.html fip://genome.jpi.paf.org/Braff1/Braff1.home.html
Caenorhabdiliz briggzae Caenorhabdiliz eleganz	Roundworm	Ecdysczca/Nematoda Ecdysczca/Nematoda	Wornbase Ensembl	12.11.2009	506 mi 45	8/29/2009	brigpep.WS206.fa.gz Caenorhabdita: elegana.WB170.45.pep.all.fa	ftp.//Tp.sanger.ac.ukipub/wormbasei.tbrjggasei.tb25.agp8 http://www.ensembi.org/infoidataidowriced.html
Caniz familiariz	Dog	Vertebrata/Mammalia	Ensembl	07.05.2010	nel 57	- 1.0 (Deleter 2005)	Canis, familiaris BROADD2.57 pep all fa gz	fp://tp:ensembl.org/publicurrent_fastalcanis_familiaria/pep/
Cavia porceituz	Guinea pig	VertebrataMammalia	Ensembl	07.05.2010	rel 57	e 1.9 (ocable 2000)	Cavia_porcellus cavPor3.57 pep al fa gz	fp:/fp.ensembl.org/publicament_fastalcavia_porcelus/pep/
Ciona intestinalis Ciona zavignyi	Transparent sea squirt Pacific transparent sea squirt	Urochordata/Aacidiacea Urochordata/Aacidiacea	Ensembl	07.05.2010 07.05.2010	nel 57 nel 57		Cona_intestinals.JGI2.57 pep.all.fs.gz Cona_savignyi.CSAV2.0.57 pep.all.fs.gz	ftp://tp.ensembi.org/publcurset_fts/sita/cons_intestivalisipep/ ftp://tp.ensembi.org/publcurset_fts/sita/cons_sav/gty/ipep/
Culex pipiens Danio rerio	House mosquito Zebrafish	Ecdysczca/Arthropoda/Insecta Vertebrata/Euteleostel	Broad Institute of Harvard and MIT Ensembl	18.01.2008 07.05.2010	rel 57	39434	culex_pipiene_3_proteins.fasta gz Danio_rerio Zvili 57.pep.ali.fa.gz	http://www.broad.mit.edu.brnotationigenomeiculex_pipiens/Dowrloads.html ftp://tp.ensembl.org/pub/current_fastaldanio_reno/pep/
Daphnia pulex Desemblia melanoramier	Water fea	Ecdysczca/Arthropoda/Crustacea	JGI Enverthi	12.07.2007	1 ml 57	v.1.0 (September 2005)	DAPPU_ProzenGeneCatalog_2007_07_03.as fasta Drawohila melanorasiar BDCPS 13 57 ose al fa or	htp://genome.jgi-paf.org/Dappu1/Dappu1.htms.html ftp://genome.jgi-paf.org/Dappu1/barse.html
Felz cetuz	Cat	Vertebrata/Mammalia	Ensembl	14.05.2010	rel 57		Fels_catus CAT 57 pep all fs.gz	fip iftp ensembl org/publicument_fasta/fela_catus/pep/
Gasterosteux aculeatux	Three-spined stickleback	Vertebrata/Euteleostel	Ensembl	07.05.2010	nel 57		Gasterosteus, acuteatus BROADS1 57 pep all tagz	fp:/fp.enembl.org/pubcurrent_testa/gasterosteux_aculeatus/pep/
Heichdelle robuste Homo sepiens	Californian leech Human	Lophotrochozoa/Annelida Vertebrata/Mammalia	JGI Ensembl	22.09.07 07.05.2010	mil 57	v 1.0 (July 2007)	proteins.Helto1_FilteredModels3.fasta Homo_sapiens.GRCh37.57.pep.all.fs.gz	http://genome.jgi-paf.org/Helro1/Helro1 / Journiaed Rp.html ftp://tp.ensembi.org/publcurrent_fasta/homo_xapiens/pep/
Hydra magnipapillata Ixodez scapulariz	Freshwater polyp Black-legged tick	Chidata Ecdyspopa/Arthropoda/Chelicerata	JCVI VectorBase	07.05.2010	1 (1/25/2009)	NCBI Build 1	protein fa gz iscasularis PEPTIDES-iscaWI.1 fa oz	fip.//Tp.ncbi.nin.nih.govjenomes/Hydra_magnipapilataiprotein/
Lottia gigardea	Owl Impet	Lophotrochozoa/Mollusca	JGI	29.07.2007	1	v1.0 (08/24/2006)	Logit_Genellodeix_Fileredliodeix1_ax fasta	http://genome.goi.pdf.cog/Lfl.cog/Lfl.com/codd ftp://www.initiation
Muz muzculus	Mouse	Vertebrata/Mammalia	Ensembl	07.05.2010	rel 57		Nus_musculus.NCBIN37.57.pep all fa gr	ttp:/Ttp.ensembil.org/publicsment_fastalmus_musculus/pep/
Nazonia vitripenniz Nematoziella vectenziz	Parasitic wasp Starlet sea anemone	Ecdysczca/Arthropoda/Insects Cnidatia	HGSC at Baylor College of Medicine JGI	29.01.2008 12.07.2007	1	12 September 2007	protein ta gz Nenwe 1FilteredModele 1 faata	ftp://tp.ncbi.nim.nih.gov/genomes/Nasonia_vibtpennis/protein http://genome.jgi-paf.org/Nerrive f.Nerrive f.homa.html
Okopleure disice Omthorhynchus anatinus	Tunicate Duckbill platypus	Urochordata/Appendicularia Vertebrata/Mammalia	Genoscope/Sats centre/MPING Ensembl	14.11.2008 07.05.2010	rel 57	25-Mar-2008	Okopieura_dioica SNAP.Od.faata Omithorhynchua_anatinus OANAS 57 pep all fa.gz	htp://www.genoscops.cns.trixdemeDownloadProjetsProjet_/Gidatatemotation/ fp://tp.ensembi.org/publicsment_tastaiomithorhynchus_anatinus/pep/
Cryziez latipez	Japanese ricefish Chimnannes	Vertebrata/Euteleostei Vertebrata/Marroralia	Ensembl	07.05.2010	rel 57 rel 57		Oryzias, latipes.MEDAKA1.57.pep.all.fa.gz Den. trockristes CHMP2.1.57.rep.all.fa.gz	ftp://tp.ensembl.org/pub/current_fastaloyzias_latipes/pep/ ftp://tp.ensembl.org/pub/current_rest_free/deal/statistics/
Prizionchuz pecificuz	Roundworm	Ecdysczca/Nematoda	Worrbase	12.11.2009	205	7/30/2009	ppapep.WS205 fa gz	fp://tp.wombase.org/pub/wombase/genomes/p_pacificua/sequences/protein/
Saccogitazuz kowalevzki	Acorn worm	Deuterostomia/Hemichordata	Baylor	29.10.2010	NCBI Build No 1 Ver 1	27 July 2010	protein fa gz (annotated proteins)	fp://p.ncb.nim.nih.gov/penomes/Saccoglossus_kovaisvaki/protein/
Schatozoma manzoni Strongylocentroluz purpuratuz	Blood fluke Purple sea urchin	Lophotrochozoa/Platyhelminthes Echinodermats	TIGR HGSC at Baylor College of Medicine	07.11.2007 05.09.2007		5/14/2007 03/09/2007	Smanaoni_preliminary_annotation.pep.gz STRPU_GLEAN3.fa	ftp://tp.tgr.org/pub/data/Eukaryotic_Projecta/a_mansonipreliminary_annotation ftp://tp.tgsc.bcm.tmc.edu/pub/data/Spurpuratus/fasta/Annotation
Teeniopygia guttata Tekifugu rubripez	Zebra finch Japanese cufferfish	Vertebrata/Amniota Vertebrata/Euteleoatei	WashUlEnsembl Ensembl	07.05.2010	rel 57 rel 57		Taeniopygia_guttatataeGut3.2.4.57.pep.all.fa.gz Takfuou_rubrices.FUGU4.57.pep.all.fa.gz	fp://tp.ensembi.org/publicsment_fasta/aeriopygia_guitata/pep/ fip://tp.ensembi.org/publicsment_fasta/fakitus_statorea/pep/
Tetraodon nigrovindia Tetralum canteneum	Green puffer Red four heads	Vertebrata/Euteleostei	Ensembl	07.05.2010	rel 57	210/2009	Tetracidor, nigrovinida. TETRACIONE.57 pep all fa gz TBY: A ruentine fa	ftp://tp.ensembl.org/publicarient_fasta/letracidon_nigrovintial.pep/ ftp://bioinformatics.iou.edu/ar/bi0asta/fasta/fasta/fasta/fasta/fasta/fasta/fasta/fasta/fasta/fasta/fasta/fasta/
Trichoplax adheerens		Placezoa	101	01.10.2007	-	v1.0 (30 August 2006)	Triad 1_best_proteins fasts	http://genome.jgi-paf.org/Triad1/Triad1.download.fp.html
Chosnofiagellates	www.bim caweo frog	verse/808/NTC/IDI8	uran Di	35.11.2009			Annapan_mapa288.3644.1.57.080.88.02	op. no. en
Monoarga brevicolla Salpingoeca rosetta/Protercapongia ap. ATCC 5081	uncanoflagellate 8	uncanofagelida Chcanofagelida	Jul BROAD	12.07.2007 02.12.2010	1	v 1.0 (July 2006)	voror _oext_proteins.faata salpingoeca_roseita_1_proteins.faata.gz	mp : genome, ge-per, org whork't Monte't home, html http://www.broadmathute.org/annotation/genome/multice/lu/antly_project/Mult/Downloads.html
Pilastorea Capsaspora cieczarzaki		ichthyaspores	BROAD	02.12.2010			capaaspors_owczarzaki_stoc_30804_1_proteins.fasta.gz	http://www.broadinatiluie.org/annotation/genome/multice/lulatity_project/lulatiCownloads.html
Fungi Acremonium alcaloohlum		Ascomucota/Sordariom-relea	.g	05.04.2011		1	Acrall cla Generatalog proteina 20100903.av fuela re-	hts:/benome.ici.cet.org/paper/dvamic/transm/Dowrload.it/Torganiam=Actail =1=
Agericus bisporus ver. burnetti JB137-58		Basidonycota?ionobasidonycetes	IGI IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	19.05.2010	2.0		Abisporus_varburnetii.v2.Fiteredilodeis1 proteins.fasta.gz	http://genome.goi.pef.org/Agabi_verbur_11Agabi_verbur_1 download fip.html
Ashbya gozzypil		Ascomycola/Saccharomycolina	University of Basel	02.12.2010 07.11.2007	AGD version 3		anoryone_readogram_ator_ator_aproxens.fasta.gz + alonyone_macrogram_ator_38327_m 982 A_gonsypii.fasta.gz	n ny - www.aukanniana.org.annotason.genomentustos.usany_project suss.com/oads.html http://www.ebi.ac.uk/rtegr/IPpGearch.do?ogProtecmeld=982
Aspergilluz flavuz Aspergilluz niger	Black mold	Ascomycota/Eurotiales Ascomycota/Eurotiales	BROAD BROAD	02.01.2008	2			http://www.broad.mit.edu.lennotationigenomelaspengikar_group/MultDownloads.html http://www.broad.mit.edu.lennotationigenomelaspengikar_group/MultDownloads.html
Candida albicanz WO-1 Candida tropicaliz		Ascomycola/Saccharomycolina Ascomycola/Saccharomycolina	BROAD	02.11.2007	1 (05/07/2007) 3 (05/23/2014)		candida_abicana_wo1_1_proteins.fasta candida trocicale 3 enterna.fasta.cz	http://www.broad.mit.edu/annotation/genome/candida_abicana/Downloads.html http://www.broad.mit.edu/annotation/senome/candida_broadca/atml
Ceriporiopaiz aubvermiapora B		BasidionycotaYlomobasidionycetes	101	02.12.2010	40514	v 1.0 (July 15, 2010)	Ceriporiopsis_subvernispora proteina fasta gr	htp://www.doc.in.in.incomments.com/optimized/comments.incom
Chaetomium globosum Cocoldioides immitis RS	Soil fungus Valley fever fungus	Ascomycota/Sordariomycetes Ascomycota/Onygenales	BROAD BROAD	07.11.2007 02.01.2008	1 (04/25/2005) 2		chaetonium_globosum_1_proteins fasts	http://www.broad.mit.edu/annotation/genome/chaetonium_globoeum/Downloads.html http://www.broad.mit.edu/annotation/genome/coccidioides_groupMultDownloads.html
Cochloboluz helerostrophus C5 Conlophora putean		Ascomycota/Dothideomycetes Basidiomycota/Homobasidiomycetes	101 101	01.07.2008	1	v1.0 (December 2007 1	CocheC5_1_GeneNiddels_FilteredModels1_sea fasta.gz Conpu1_GeneCatalog_proteins_20101116.aa fasta.gz	http://genome.jpi.pef.org/CocheC5_11CocheC5_1.download.tlp.html http://genome.jpi.pef.org/pages/dynamicOrganiamOownload.j#?organiam=Conput
Coprinuz cinereuz Cnotococcus reoformany A	Inky cap fungus	Basidomycota/tomobasidomycetes Basidomycota/TremeTomoretes	Broad Institute of Harvard and MIT Broad Institute of Harvard and MIT	01.11.2007	1 (05/04/2005) 1 (04/25/2014)		coprinus_cinereus_1_proteins.fasta cryptococcus neoformans 1 proteins.fasta	htip //www.broad mit.edu/annotation/genome/coprinus_chereus/Downloads.html http://www.broad.mit.edu/annotation/senome/cov/couccus_needommans/Downloads.html
Dictomitur aqualenz		Basidonycota/Homobasidomycetes	101	08.04.2011		1	Dicard _GeneCatalog_proteins_20101110 as fasts gz	htp://penome.jp/peli-op/pagesidynamicOrganismDownload.jp/?organism=Dicaq1
Dothistroma septosporum NZE10 v1.0Mycosphaen Emericella nidulanz	alla pini	Ascomycota/Dothideomycetes Ascomycota/Eurotiales	JGI BRDAD	02.12.2010 14.07.2007	1.0 V.1 05/19/2005	v 1.0 (September 30, 2010)	Dotse1_GeneCatalog_proteins_20100018.as/fasts.gz aspergilus_nidularus_1_proteins.fasts	htp://genome.jgi-pal.org/Dotes1.Dotes1.download.ttp.html http://www.broad.mt.edu/annotation/genome/aspergilua_prosp/MultiForms.html
Encephalitozoon cuniculi Fomitocala pinicole 551 y1.0	Microsporidian parasite	Mcrosporidia Basidomycota?fomobasidomycetes	Genoecope	29.01.2008	(7/24/2007) 1 (12/15/2010)	v 1.0	11 files for 11 chromosomes Fomoi1 GeneCatalog proteins 2010/1006.as.fasta.oz	fip.i/tp.ncbi.nim.nih.govigenomes/Fungi/Encephalitazoon_cunicul/ http://penome.ioi.cef.ors/Fornoi/i/Fornoi/i.downiced.ftp.html
Gibberella zeae Ghanshullum trabaum	Wheat head blight fungus	Asconycota/Sordariomycetes	BRDAD	07.11.2007	3	v 1.0 (November 1. 2010)	Girtri 1 GeneCatalon notating 20100021 as fasts or	http://www.broad.mit.edulannotation/genome/tuanum_group/AutiDownloads.html
Neterobasidion annosum	Root rot fungus	BasidionycotaYlomobasidionycetes	101	27.11.2009	1	v 1.0 (May 2009)	Hannosum Fillered Models 1. proteins. fasta	htp:/genome.jgi-pal.org/Netan1Netan1.download.fp.html
Ruyversmyces lectis Ruyversmyces webi		Ascomycola/Saccharomycolina Ascomycola/Saccharomycolina	Génolevures BROAD,SGD	01.11.2007 19.10.1910	GL2: 2 (2006/08/17) (Wed Sep 21 2005)		Kla-GL2/2 aa Klayveronyces_walti.SNAP.as.gz	http://cbi.labri.u-bordeaux.th/Genolevures/download/KLLA_annot.php http://tungal.genome.duke.edu/annotations/kwallaa/
Laccarla bicolor Mechacorthe orizes	Bicoloured deceiver Rice blast fungus	Basidionycota/Homobasidionycetes Ascomycota/Sordarionycetes	JGI BROAD	12.07.2007	1 5 (05/07/2007)	v 1.0 (March 2005)	LACEL_BestModelsv1 as tests magnaporthe prices 5 proteins tests	http://genome.jpi-pal.org/Lacbif/Lacbif.home.html http://genub.totad.mit.edu/antotation/senome/magnapothe_criaes/Downloads.html
Melampoora laricit-populina		Basidionycota/Puccinionycotina	101	17.11.2008	v1.0	v.1.0 (November 2007)	Maricia, populina Filiered Modelal as fasta gr	http://genome.jpi.pef.org/Meltp1/Meltp1.download.ftp.html
Mycellophthora thermophila/+Sporotrichum thermop	thie)	Ascomycola/Sordariomyceles	351	08.04.2011	20	2	Spoth2_GeneCatalog_proteine_20101221 as fasts gz	mp i genome, gi par orgenizozzenizozz downowa tip zmil http:/genome.jgi-paf.org/pagesidynamicOrganiamDownicad.je?organiam=Spoth2
Mycozphaerella fijenziz Mycozphaerella graminicola		Ascomycota/Dothideomycetes Ascomycota/Dothideomycetes	101 101	01.11.2007 01.11.2007	1	v.1.0 (November 2005) v.1.0 (Nov 2005)	FilteredModelsv1.0.xx fasts proteins.FilteredModels1.fasts	http://genome.jpi-pef.org/Mycd11.Mycd11.download.ftp.html http://genome.jpi-pef.org/Mycg11.Mycg11.download.ftp.html
Neczartorya fizcheri Neumanora crassa		Ascomycota/Eurotiales	BROAD	02.01.2008	1		neurosnova crassa Z novisios fasta	http://www.broad.mit.edu/annotation/genome/aspergiliua_group/MultDownloads.html
Phaecaphaeria nodorum	Glume blotch fungus	Ascomycota/Dothideomycetes	Broad Institute of Harvard and MIT	01.11.2007	1 (3/14/2005)		stagonospora_nodorum_1_proteina.faeta	http://www.broad.mit.edu/amnotation/genome/stagonospora_nodorum/Downloads.html
Phycomyces blakesleeanus Pichia angusta/Hansenula polymorpha NCYC 435 k	na1.1/	Mucoromycolina (Zygomycola) Ascomycola/Saccharomycolina	101 101	28.08.2007 02.12.2010	2	v.1.0 (September 2005) v.2.0 (November 17, 2010)	Phybl1_best_proteins.fasta Hanpo2_GeneCatalog_proteins_20100927.ax.fasta.gz	http://genome.jgi-pat.org/PhybitSPhybitIdownload.ftp.html http://genome.jgi-pat.org/Hanpo2/download.ftp.html
Pleurotus catveetus	Oyster mushroom	Basidionycota/Homobasidiomycetes	JGI U Granizati	07.05.2010	2.0		PiecePC9_1_GeneModels_Filteredmodels2_as/tasts.gz	http://genome.jpi-paf.org/Piece/PC9_1/Piece/PC9_1 download ftp.html
Posta placenta		BasidiomycotaNomobasidiomycetes	1GI	01.11.2007	1	v 1.0 (September 2005)	Poopi1_FilteredModels2_proteins.fasts	htp://genome.jgi.pat.org/Pospi1/Pospi1.download.ttp.html
Pyconia gramina Pyrenophora tritici-repentia	stack elem rust fungus	Ascomycota/Dothideomycetes	Broad Institute of Harvard and MIT	02.01.2008	2 (06/06/2007)		puccha_gramma_r_ap_moc_x_proteina.tasta.gz pyrenophora_tritici-repentis_1_proteins.tasta.zip	http://www.broad.mit.edu/annotation/genome/procina_graminia.uowinoada.ntmi http://www.broad.mit.edu/annotation/genome/pyrenophora_tritid_repentix.3MultiDownloads.html
Rhizopuz oryzae Rhodotorule graminiz strain WP1		Mucoromycotina (Zygomycota) Basidomycota/Pucciniomycotina	Broad Institute of Harvard and MIT JGI	02.11.2007	3 (04/21/2005)	v 1.1 (November 17, 2010)	rhizopus_oryzae_3_proteins.testa Rhoba1 1 GeneCataloo proteins 20100913.as.testa.oz	http://www.broad.mit.edu/annotation/genome/htizopus_oryzae/Downloads.html http://www.broad.en/Rhoba1_I/Rhoba1_I.download.ito.html
Saccharomycez caztelli	Yeast	Ascomycola/Saccharomycolina	Washington University	08.01.2008	(104/2003)		of_transfasts.gz	ftp://genome-ftp.stanford.edu/pub/yeast/sequence/fungal_genomes/5_castelli/WashUlorf_protein/
Schizosaccharomycez japonicuz	Fission yeast	Ascomycola/Saconalomycolina Ascomycola/Taphrinomycolina	Broad Institute of Harvard and MIT	14.07.2007	V.1 2/20/2007		yeast_ntpep tasta schizosaccharomyces_japonicus_1_profeins.fasta	np : genome-np. stantoro, edu puo yeaatisequence: versaank http://www.broad.mit.edu/annotation/genome/schizosaccharomyces_japonicus/Downloads.html
Schizosaccharomyces pombe Septoria musiva SO2202Mycosphaerella populorur	Fission yeast	Ascomycota/Taphrinomycotina Ascomycota/Dothideomycetes	Sanger JGI	16.07.2007 02.12.2010	v 19 07/09/2007 1.0	v 1.0 (November 16, 2010)	pompep.tasta Sepmu1_GeneCatalog_proteins_20100915.as.fasta.gz	ftp://tp.sanger.ac.uk/pub/yeast/pombe/Protein_data/pompep http://genome.jgi-paf.org/Sepmu1.Sepmu1.download.ftp.html
Serpula lacrymana Spizellomycez ourchetus	Dry rot fungus	Basidionycota/Homobasidionycetes Chytridiomycota	JGI BRDAD	14.05.2010	2.0		Serpula_lacrymans_57_3_v2.proteins.fasta.gz spizeliomyces punctatus daom bri17_1 proteins.fasta.gz + spizeliomyces punctatus daom bri17_1	htip iligenome (gi-pat.org/SerlaS7_3_2/SerlaS7_3_2/download.ftp.html http://www.broadinatitute.org/annotation/commerimutical/ularity_project/MultiCownloads.html
Sporobolomycez rozeuz Thielevia terrentrin		Basidiomycota/Pucciniomycotina	101	21.08.2007	1	v.1.0 (Apr 2006) v. 1.0 (Santamber 2009)	Stoseus Filered/Iodels1 as fasts Third Grand/Iodels Ellerer/Models2 as fasts	http://genome.jgi.paf.org/Sporo1/Sporo1/home.html
Trametez versicolor		Basidiomycota/Homobasidiomycetes	101	08.04.2011		1	Trave1_GeneCatalog_proteins_20101111 aa.faata.gz	http://genome.jpi-paf.org/pagesidynamicOrganismDownload.jaf?organism+Trave1
Tremela mezenterica Trichoderma vinenz	Jelly fungus	Basidiomycota/Ternellomycetes Ascomycota/Sordariomycetes	101 101	27.11.2009 01.01.2008	1	v1.0 (June 2009) v1.0	Treme 1_beat_proteins.faata proteins.FMD_Trive1.faata.gz	http://genome.jpj-paf.org/Treme1/freme1/download.ftp.html http://genome.jpj-paf.org/Trive1/Trive1/download.ftp.html
Uncinocarpuz reezil Uztilezo mevdiz	Smut funcua	Ascomycota/Onygenales Basidomycota/Ustiacinomycotina	BROAD Broad institute of Harvard and MT	02.01.2008	2 (05/12/2005) 1 (04/25/2005)		uncinocaspus_reesil_2_proteins.fasta.gz ustilaco mardia 1 proteins.fasta	http://www.broad.mit.edu/annotation/genome/uncinocerpus_reesil/Downloads.html http://www.broad.mit.edu/annotation/genome/ustilaco_mavda/Downloads.html
Walersia aeb/ Wolfponia cocca MD-104 5510		BasidomycotaWalemiomycetes BasidomycotaNomobasidomycetes	IDL IDL	02.12.2010	1.0 40514	v 1.0 (October 11, 2010) v 1.0 (November 24, 2010)	Walke 1_GeneCatalog_proteins_20100910 as fasts gr Wolco1_GeneCatalog_proteins_20100915 as fasts or	http://genome.jg/.pdf.org/Wake1/Wake1.download.ftp.html http://genome.ig/.pdf.org/Wokc01/Wokc01.download.ftp.html
Yarowia lipolytica		Ascomycola/Saccharomycolina	Génolevures	01.11.2007	GL2: 2 (2006/05/22)		Ya6-GL22.as	http://tbi.labri.u-bondeaux.ht/Genolevurealdownload/YALI_annot.php
Apusozoa Thecamonaz trahenz		Apuezzoa Apuezmonadidae	BROAD	02.12.2010			thecamonas_trahens_ator_50062_1_proteins.fasta.gz	http://www.broadinatitute.org/annotation/genome/multice/lularity_project/Mult/Downloads.html
Dictyozlellum dizcoideum	Sime mold	Nycetozoa	dictyEase/HGSC at Eavier College of Med	ci 29.07.2007	39278.0875		dicly_primary_protein	http://dictybase.org/db/gj-biv/dictyBase/download/blast_databases.pl
Dictyostelium purpureum QSDP1 Entemoeba histolytica	Sime mold	Mycetozoa Entempebidae	JGI TIGR	19:09:2008	1.0 (12(22)2004)	v1.0 (August 18 2008)	Dicpu1_beat_proteins.fasta.gz ENA1.pep	htp://genome.joj-pal.org/Dicpu1/Dicpu1.download flp.html flp://tp.fgr.org/publidata/Dukaryolic_Projectale_hatolytica/whole_genome_wequencing/DHA1.pep
Archaeplastida Apulegia coerulea	Rocky mountain columbine	VirdiplantaelEmbryophyta	19	100500	16.0	199919	Accerules 151 pecide/a cz	fig. (to joi on project) (a) data john score (v6.0)
Arabidopaix Iyrata Arabidopaix Ihaliana	Lyre-leaved rock-cress Mouse-ear cress	Viridplantae/Embryophyta Viridplantae/Embryophyta	JGI TAIR	03.10.2008	v1.0 TAIR7	v1.0 (April 7, 2008)	Araly1_GeneModels_FilenedModelsE_ax.texts TAR7_pep_20070425	http://genome.jg/.pdf.org/Analy1/Analy1.download.ftp.html ftp://tb.anabidoosis.org/homes/tais/Genes/TAR7_biasteet=
Brachypodium diatachyon	Purple false brome	Viridiplantae/Embryophyta	101	04.12.2011	v6.0	8/30/10	Ediatechyon_114_peptide.fa.gz	ftp://tp.jp.pd.org/pub/JGL_data/phylozome/sG_0
Chiamydomonas reinhardhi		Chlorophyta/Chlorophyceae		12.07.2007	3		China3_1.GeneCatalogProteina.6.ULD6.fasta	http://genome.jgi-pail.org/Chire&I.html
Chionelle ap. NC64A Citrus clementine		Chlorophyta/Trebouxiophyceae ViridiplantaelEmbryophyta	101	15.05.2008	1 v6.0	1.0 (June 13, 2008) 1/15/11	Chlorella_NC64A.best_proteins.fasta gz Colementina_165_peptide.fa.gz	http://genome.igi-pat.org/ChINC64A_11ChINC64A_1.download.ttp.html ftp://ttp.jgi-pat.org/publ/3G1_data/phytozome/x6.0/
Citrus ainenais Coccomyxe zp. C-169/Chlorelle vulgeris C-140	Sweet orange	Viridiplantae/Embryophyta Chlorophyta/Trebouxiophyceae	101	04.12.2011 02.12.2010	v6.0 2	1/15/11 2.0 (December 18, 2006)	Cainensis_154_peptide.ts.gz Coccernyos_C102_v2_titered_proteins.texts.gz	ftp://tp.jpi.psf.org/pub/3Gl_data/phytozome/x6.0/ http:/jpimsome.jpi.psf.org/Coc, C160_15/Coc, C160_16.download.ftp.html
Cucumis sativis	Garden cucumber	Viridplantae/Embryophyta	JGI National Institute of Country, Inc.	07.05.2010	1.0 (17.5ep.2005)		Cucas_v1 pep fa.gz	http://genome.jg/pail.org/cucumber/cucumber/coverload.ftp.html
Eucelyptus grandis		VirdiplantaeEmbryophyta	JGI	04.12.2011	v6.0	915/11	Egrandis_162_peptide.ts.gz	fip :rtp ;g-pat org/pub/JGI ; data/phy/coores/vG (D)
seycine max Manihot exculente	Soytean Tapicca	vinopiantae/Embryophyta Viridiplantae/Embryophyta	3GI	09.11.2009 07.05.2010	1.0 1.1	(12/17/2008)	commit pepiningz commin_v1.1 pep filter fa	mp:rmp.gp-pat.org/pubuuudata/Glycine_max/Glyma1tannotation/ http://genome.jgi-pat.org/cassava/cassava.download ftp.html
Medicago truncatula Micromonaz pusilla COMP1545	Barrel medic	ViridiplantaelEmbryophyta Chlorophyta/Prasinophyceae	101 101	04.12.2011 07.03.2008	v6.0 2	8/25/11 v2.0(Feb, 2008)	Mhuncatula_135_peptide.fa.gz MicromonasCCMP1545_ProzenGeneCatalog_20082205 as fasta.gz	fip:/fip.gi-paf.org/pub/JGL_data/phytozome//6.0/ http://penome.ioi-paf.org/Mcou/22/Mcou/22.downioad.fip.html
Meromonas strain RCC299 Member cambur	Valing monkey for	Chlorophyta/Prasinophyceae	101	07.03.2008	2	v2.0 (Feb 2008)	MicromonaeRCC299. ProzenGeneCatalog_20080205 as fasta gz Microlea cultura v1.0 curcilia fa cr	http://genome.jpi-pal.org/McpuN2/McpuN2.download /tp.html
Ciyza zativa	Rice	Viridiplantae/Embryophyta	TIGR	14.07.2007	Release 5		ous_chr1_12_igrants	http://www.ligr.org/idbi/2.k1/coas1/data_download.ahtml
Listreococcus lucimaritus Cistreococcus sp. RCC809		CricrophytaPtasinophyceae ChicrophytaPtasinophyceae	3GI	12.07.2007 14.08.2008	2 1	v 1.0 (August 2008)	O scorennus / M as fasta OuRCC000 FM1 as fasta gr	mspungenome.gp-per.org/3x89601_3/Cel9901_3.home.html http://genome.jpi-pel.org/Cel9CC609_11/Cel9CC609_11.ntb.html
Catreococcus teuri Physicomitrelle patens sap patens	Moss	Chlorophyta®tasinophyceae ViridiplantaelEmbryophyta	101 JGI	22.09.2007 22.09.2007	2 1.1	v1.1 (March 2007)	O.tauri FM.aa.faata proteins.Phype1_1.FilteredModels3.tasta	http://genome.jp/pet.org/Detta4/Detta4.download.ftp.html http://genome.jp/pet.org/Ptypet_1_1Ptypet_1.download.ftp.html
Populus hichocarpa Phunus persica	Western balsam poplar Peach	Viridplantae/Embryophyta Viridplantae/Embryophyta	101 IQI	10.02.2007	1.1 v6.0	v1.0 (June 2004) 8/25/10	proteins.Poptr1_1.JamboreeModels.fasta Poemica 139 peotide.fs.o	http://genome.jg/pef.org/Poptr1_1/Poptr1_1.download.ftp.html ftp://tb.loi.ord.corpub/261.data/shrkazone/45.01
Ricinus communis	Castor bean	VirdiplantaelEmbryophyta	121	04.12.2011	v6.0	8/30/10	Roommunik_110_peptide.fx.g	fip if p jg-psf or physiolicities (data physicareme Vid D)
Selaginella moellendorffi Setaria italica	Spikemosa Foxtail millet	ViridplantaelEmbryophyta ViridplantaelEmbryophyta	101 JGI	21.12.2007 04.12.2011	1 v6.0	v1.0 (March 2007) 8/30/10	Selmo 1_GeneModels_FilemedModels3_aa Stalica_155_peptide.fs.gz	http://genome.jgi-pat.org/Selmo 1./Selmo 1./Se
Sorghum bicolor Vitiz vinifera	Sorghum Grape	Viridiplantae/Embryophyta Viridiplantae/Embryophyta	JGI Genoecope	21.04.2008 23.11.2007	1.0 v1	v1.0 (March 2007) (05-Sep-2007)	Sorbi1_GeneModels_FilteredModels6_ass.fasta.gz Vita_vinflera_peptide_v1.fa	http://genome.jgi-pat.org/Sobil/Sobil/Sobil/Sobil/Sobilisten/ http://genoscope.cns.thindeme/English/Projets/Projet_ML/data/annotation/
Volvax carteri f. nagariensis		Chlorophyta/Chlorophyceae	10	12.07.2007	1	v 1.0 (March 2007)	VOLCA proteins Filtered Nodels2 fasta.gz	http://genome.jcji.pdf.org/Volca1/Volca1/Norme.html
Chromalveolates				25.14.2011				
Aureococcus anophagefferens Cryptosporidium hominis TU502		stramenopiles/Pelagophyceae Alveolata/Apicomplexa/Coccidia	NHNADApD8	10.01.2007 07.11.2007	1 rel 3.0 (11-May-2007)	v.1.0 (June 2007)	proteina Auran'i "FilteredNodela's fasta ChominiaAnnotatedProtein fas	mp urgenome, ga-per, organiuran 1 Auran 1. download, fip html http://apidb.org/common/downloads/indexeads/.0.ChyptoPlasmoToso_downloads/ChominisProteins/
Cryptosporidium parvum IOWA Emilania husleyi CCMP1516		Alveolata/Apicomplexa/Coccidia Haptophyta	NHNADApDB JGI	07.11.2007 29.04.2008	rel 3.0 (11-May-2007) V 1.0	v 1.0 (3 November 200%)	CpanvumAnnotatedProtein.fsa Emihu1_best_proteins.fsata.gz	http:/lipidb.org/common/downloads/inlease3.0 CryptoPlasmoTooc_downloads/CparvumProteina/ http:/igenome.igi-pat.org/Emihu1/Emihu1 download /tp.html
Pregilericpaiz cylindruz		stramenopiles/Bacillariophyta (datoms)	JGI	27.11.2009	1	(September 30 2009)	Fracy1_GeneNodels_FilteredNodels1_ass Itatis	http://genome.jgi-pel.org/Tracy1.Fracy1.download.ftp.html
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**Supplementary Table 4**. **Sequence conservation of Hinge 1.** The first two residues shown are the bending residues in this hinge.

hDHFR	Т	Т	S	S	V	Ε	G	К	Q	N	L
% identity	31.4	38.8	18.2	43.0	21.5	24.8	45.5	85.1	32.2	100.0	14.9
% conservation	57.0	42.1	69.4	51.2	22.3	28.1	45.5	85.1	32.2	100.0	31.4

#### Supplementary note

#### **Expanded discussions**

#### The human DHFR active site

For *E. coli* DHFR, a large chemical shift change in the <sup>15</sup>N dimension is observed for Ala6 between THF complexes and FOL complexes. This difference is the result of a hydrogen bond to Ile5, which is formed when THF is bound, but not when FOL is bound<sup>1</sup>. Based on the crystal structures, a similar hydrogen bond would be expected for hDHFR involving Ile7. However, there is no evidence for its formation from our NMR data. From the near identity of the E-NADP<sup>+</sup>-FOL and E-NADP<sup>+</sup>-THF spectra, we conclude that hDHFR does not discriminate as effectively as ecDHFR with respect to FOL and THF. Consistent with this notion, the dissociation constants for FOL, DHF and THF are very similar for hDHFR, while ecDHFR binds two orders of magnitude more tightly to THF than it does to FOL<sup>4-7</sup>. The <sup>15</sup>N HSCO spectrum of hE–NADPH–THF shows significant broadening and some shifting of resonances compared with that of the  $hE-NADP^+-THF$  complex. However, we were unable to prepare a stable sample for this complex, and the <sup>15</sup>N HSQC spectrum began changing within a few hours. Therefore, reliable assignments could not be made, and we could not study this complex in detail. A chemical shift analysis of hE–THF and hE–NADP<sup>+</sup>–THF shows that, in the product binary complex, the differences in chemical shifts can be attributed to the presence or absence of NADP, without additional changes in backbone conformation. Thus, X-ray structures and <sup>15</sup>N HSQCs show that the backbone conformation of hE–NADP<sup>+</sup>–FOL, hE–NADP<sup>+</sup>–THF and hE– THF are all very similar and in the "closed" conformation. The "closed" conformation in ecDHFR is stabilized by hydrogen bonds between the backbone amide and sidechain carboxylate of Asp122 and the backbone carbonyl and amide of Gly15 and Glu17, respectively. In the

"occluded" conformation, these hydrogen bonds are broken, and new ones are formed between backbone carbonyl and amide of Asn23 and the backbone amide and side-chain hydroxyl of Ser148, respectively<sup>1</sup>. In hDHFR, this hydrogen-bonding pattern is conserved for the observed "closed" conformation, but the hydrogen bonds stabilizing the "occluded" conformation cannot be formed as Asn23 of ecDHFR is replaced by two prolines in hDHFR. As mentioned in the main text, the active site of hDHFR appears to be better packed than that of ecDHFR. The key side chains that contribute to the efficient active site packing in the human enzyme are Leu22, Pro26, Phe31, Ile60 and Pro61. In ecDHFR, Leu22 is replaced by the more flexible Met20, Phe31 is substituted by a smaller Leu28, and Pro61 by the highly flexible Gly51, resulting in a more loosely packed active site.

### Chemical shift analysis of hE–NADPH and hE–NADP<sup>+</sup>–FOL

To assess the presence of the hinge motion in solution, we turned to NMR experiments. A comparison of the <sup>15</sup>N HSQC spectra of hE–NADPH and hE–NADP<sup>+</sup>–FOL shows chemical shift differences that are consistent with the hinge movement observed in the crystal structures. As hE–NADPH is not stable at pH 6.5, the pH at which the chemical shift analysis was carried out for other complexes, spectra of both hE–NADPH and hE–NADP<sup>+</sup>–FOL were collected at pH 8.0 to facilitate an analysis that is not influenced by pH-dependent chemical shift changes in either complex (Supplementary Fig. 3d,e). Several chemical shift changes are observed, of which only a subset reflect ligand (FOL) binding, as expected. Gly129, one of the hinge-bending residues shows a very large chemical shift difference in both <sup>1</sup>H and <sup>15</sup>N positions. Other residues in the region, Leu131 and Lys132 also show significant <sup>15</sup>N and <sup>1</sup>H chemical shift differences. These data are consistent with a conformational change in the region, and we conclude that the

difference in chemical shifts most likely reflects the hinge movement, with the hinge being closed in the hE–NADP<sup>+</sup>–FOL complex, and open or averaging between open and closed in the hE–NADPH complex. In the region of hinge 1, the backbone amide cross peak of Gly45 is broadened beyond detection, and resonances of residues 42 and 44 are weak, indicative of flexibility in that region. We also observe a chemical shift difference for Asn107. Asn107 N forms a hydrogen bond with Pro103 CO, stabilizing a short 1.5 turn helix. In the ternary hE–NADP<sup>+</sup>–FOL complex, Pro103 is slightly shifted out of register, and the distance between Asn107 N and Pro103 CO increases from 3.1 Å in hE–NADPH to 4.5 Å. The hydrogen bond between Asn107 N and Pro103 CO is, therefore, broken, and the secondary structure in this region is loosened in the ternary (hinge-closed) complex.

#### Hinge movements in hDHFR

A comparison of the hydrogen bonding patterns in the hE–NADP<sup>+</sup>–FOL and hE–NADPH structures sheds considerable light on how these movements are driven by ligand binding and release in human DHFR. The NADP cofactor in both hE–NADPH and E–NADP<sup>+</sup>–FOL is firmly anchored to the loop subdomain in the region of the nicotinamide ring, and to the adenosine binding subdomain in the region of the adenosine moiety. Thus, the hydrogen-bonding network to the protein is maintained at both ends of NADP. However, in the pyrophosphate region of NADP, the interactions with the  $\alpha$ F helix differ between the binary hE–NADPH and ternary hE– NADP<sup>+</sup>–FOL structures. The phosphate groups form hydrogen bonds to the hydroxyl group and backbone amide of Ser119 in hE–NADPH, and an additional hydrogen bond to the backbone amide of Gly117 (Supplementary Fig. 2a-c). Upon folate binding, several new hydrogen bonds are formed from FOL to both subdomains of the protein (Supplementary Table 1), pulling both subdomains towards each other. In the hinge-closed state, the hydrogen bonds between NADPH and Ser119 are broken, allowing helix  $\alpha$ F to slide 2.5 Å away from the active site as the rest of the adenosine binding subdomain closes in towards the loop subdomain. The sliding of helix  $\alpha$ F introduces a twisting motion into the hinge movement, and allows sufficient space for the nicotinamide ring of NADP<sup>+</sup> to be rearranged slightly to avoid steric clash with the pterin ring of folate. In the absence of substrate, the NADP to Ser119 hydrogen bonds stabilize the hinge-open conformation, explaining why this conformation is not observed when a ligand is bound in the substrate-binding site. The hinge motions are, therefore, driven by ligand binding, and the two conformations are stabilized by hydrogen bonding interactions and hydrophobic packing interactions between the ligands and the enzyme.

Thr40, a classical helix C-cap and one of the bending residues in hinge 1, forms a hydrogen bond to the backbone CO of Arg36 in the substrate-binding helix, and also to the Asn48 side chain at the end of the hinge. Additionally, Asn48 forms a hydrogen bond to the backbone amide of Met111 in the adenosine-binding subdomain, and to Thr38, the last residue in the substrate binding helix preceding hinge 1. The bending of Thr39 and Thr40 results in a rigid body movement of a subset of residues in hinge 1 and the adenosine binding subdomain (Thr41-I114) moves as a rigid body; the length of this hinge provides the framework for the rigid body movement. The hinge 1 region in ecDHFR is 7 residues shorter than that of hDHFR, and forms a network of hydrogen bonds to both subdomains, restricting the extent of possible movements in this region; accordingly, hinge-bending is of much smaller magnitude in ecDHFR<sup>1</sup> than in the human enzyme (Supplementary Fig. 2a-c).

A comparison of the hE–NADPH complex with mouse DHFR (mDHFR, mE) in complex with NADPH shows that the hinge-open conformation is only observed in the hE–NADPH

structure, not in the mE–NADPH structure, although the sequences are highly conserved. The two structures are in different space groups, and both have lattice contacts in areas that could influence the stabilization of the hinge-open versus hinge-closed conformation in the crystal structures. Most importantly, the mE–NADPH structure (PDB code:  $3D84^8$ ) has a glycerol molecule bound in the substrate-binding site, forming key hydrogen bonds to the substrate binding helix, adenosine binding domain and core  $\beta$ -sheet, which would be expected to stabilize the hinge-closed conformation in a similar manner as FOL.

#### Further discussion of PWNAL ecDHFR mutant

The <sup>15</sup>N HSQC spectrum of the <sup>21</sup>PWNAL<sup>24</sup> E–NADP<sup>+</sup>–FOL is very similar to that of the wild type enzyme, and even more so to the N23PP/S148A mutant, indicating that it is structurally similar (Supplementary Fig. 4a). Due to the very small amount of dispersion, which is detectable in <sup>15</sup>N R2 dispersion experiments only at the higher field, the data cannot be fit to obtain an accurate rate, but indicate the presence of limited flexibility in these residues on the  $\mu$ s-ms timescale, at a much faster rate (estimated to be ~3000 s<sup>-1</sup>) than for the wild type enzyme. Motions are altered in key loop residues, such as Gly121 and His149, which show large fluctuations in wild type E–NADP<sup>+</sup>–FOL, but no observable <sup>15</sup>N R<sub>2</sub> dispersion in the mutant (Supplementary Fig. 4b,c). C-terminal-associated residues maintain millisecond timescale dynamics similar to wild type and <sup>21</sup>PWPPL<sup>24</sup> ecDHFR, with a rate (k<sub>ex</sub>) of 600 (± 16) s<sup>-1</sup>, and an excited state population of 3.2%.

#### Relationship between Regions A, B & C in DHFR

The length of Region 1 relates to conformational flexibility of the active site "Met20" loop, and the length and composition of Regions 2 and 3 are likely determinants of whether hinge movements can occur in the enzyme. Based on our sequence analysis, we find that the combination of lengths in these three regions show certain preferences. The vast majority of bacterial sequences contain 7 residues in Region A, 12 residues in region B, and 14 residues in Region C, and thus have flexible Met20 loops and short hinges that appear unable to support the large scale opening of the active site observed in hDHFR. All Region B sequences containing 19 residues exactly can be aligned accurately to the hDHFR sequence and structure, on the assumption that the flanking secondary structures ( $\alpha B$  and  $\beta B$ ) are conserved in length. Longer hinges are present in most eukaryotes, but lower eukaryotes retain flexible Met20 loops (short Region A), while higher eukaryotes contain an insertion in Region A, limiting the conformational change after hydride transfer. It seems likely that, while almost all prokaryotic DHFRs cannot accommodate the hinge movements, most eukaryotic DHFRs have already developed the length of hinge that would be required for the hinge movements, but only a few eukaryotic DHFRs have eliminated conformational flexibility influenced by Region A. In a minor group of bacterial sequences, the length of Region C (and therefore likely hinge 2) is similar to hDHFR, but the length of Region B (and likely hinge 1) is identical to the majority of bacterial sequences.

A comprehensive analysis of all DHFR structures in the PDB was carried out, paying special attention to loop and hinge conformations in Regions A, B and C. Although several structures of DHFR are available, analysis of these is not as insightful as one would expect for two main reasons: A) Most structures are of either the *E. coli* or human enzyme bound to an inhibitor and NADP<sup>+</sup> in a ternary complex. In the ternary complex, the active site loops are

preferentially closed, and do not give much insight into the conformations relevant to catalysis. B) An analysis of all structures shows most to be exactly the same as the "closed" conformation of ecDHFR. Very few structures differ significantly from this conformation. For the structures that shed insights on understanding the dynamic mechanism, we have included our observations below or in the main text.

It is interesting to note that, in a small subset of bacterial sequences, the length of Region B is increased by one residue compared with the consensus length for bacterial DHFRs. This subset represents thermophilic bacteria including Thermatoga maritima DHFR (tmDHFR), as well as a few enterobacteria. Since structures are available for tmDHFR, we can assess the differences between tmDHFR and ecDHFR further. tmDHFR is a dimer, in which the active site loop ("Met20 loop" assessed in region 1) forms part of the dimer interface, and is stabilized in an "open" conformation, which is different from the "closed" and "occluded" conformations of ecDHFR. The "open" conformation has been observed in some crystal structures of ecDHFR, wherein it is stabilized by lattice contacts, but is not a stable ground state in any intermediate complex for ecDHFR. While this conformation is conducive to NADP binding, it is not optimal for catalysis, as the "open" loop does not shield the active site from bulk solvent<sup>1</sup>. The additional residue in Region B of tmDHFR forms an extra turn in the substrate binding helix, and does not add to the length of the connecting loop. However, we note that the dynamic mechanism in tmDHFR is likely to be quite different from that of ecDHFR due to the dimerization interface of the enzyme at the active site, and the fact that the enzyme functions at much higher temperatures.

*Gallus gallus* (chicken) DHFR (cDHFR) is the vertebrate DHFR that is most divergent from hDHFR and for which a structure is available (PDB codes: 1DR1, ternary complex with thioNADP<sup>+</sup> and biopterin and 8DFR, binary complex with thioNADP<sup>+</sup>)<sup>9,10</sup>. A comparison of both cDHFR structures with hE–NADP<sup>+</sup>–FOL shows that the  $\alpha$ F helix superimposes exactly with hDHFR in the hinge-closed conformation. When the structures are aligned on the loop subdomain, only Gly53 and K68 in the cDHFR adenosine binding domain are in a slightly more "open" conformation than hE–NADP<sup>+</sup>–FOL. In the presence of substrate or product, this region would be expected to form hydrogen bonds to the ligand, and close the active site. It is likely that helix  $\alpha$ F cannot stabilize the hinge-open conformation, as Ser119 of hDHFR is substituted with Ala119 in cDHFR, and the hydrogen bonds between this residue at the N-terminus of helix  $\alpha$ F forms stabilizing hydrogen bonds with NADPH in the binary hE–NADPH hinge-open conformation. There is no notable difference in the conformation of the binary cE–thioNADP<sup>+</sup> and ternary cE–thioNADP<sup>+</sup>:biopterin, showing that the hinge-open conformation is not present, and providing further evidence that the interaction of the Ser119 residue in  $\alpha$ F with NADP is important in maintaining the hinge-open conformation in hDHFR.

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