

SUPPORTING INFORMATION

Inhibition studies of cytochrome bo_3 (cbo_3) using a novel enzyme assay

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Simulations, methods

Cyclic voltammograms were modeled using a finite difference procedure programmed in C according to Hirst et al.[1] The model is based on a kinetic scheme as shown in Figure S11. Due to number of parameters in this already simplified model, we cannot obtain values for these parameters by fitting the cyclic voltammogram data nor can we use the experimental data to independently verify the whole of the model (but it is possible to verify parts of the model, see below). Instead, these models have been used to support the hypotheses made in the main article and parameters are fixed at physical relevant values.

Rates and Parameters

The interfacial electron transfer rate between the electrode and ubiquinol-10 (UQ-10) are modeled using the Butler-Volmer equations with the following parameters: electron transport rate at zero overpotential (k_0), reduction potential of UQ-10 (E_0^{UQ}) and n_{app} to represent cooperativity of the two electron transfer process ($n = 1$ for non-cooperative and $n=2$ for fully cooperative). The values used for these and other parameters are given in Table 1. Rate k_1 is calculated using Michaelis-Menten assuming the two consecutive reductions by ubiquinol – required for the reduction of single oxygen molecule – with equal rates. K_M^{UQ} was estimated from the data shown in Figure 1B of the article and k_1^{cat} , like k_2 was set to an appropriate value for cbo_3 and varied to simulate inhibition. As all inhibition experiments were performed at > 10 times the apparent K_M for oxygen, the oxygen reduction rate was assumed to be constant modeled with a single rate. The surface coverage of ubiquinol-10 (Γ_{UQ}) was varied as shown in the results below. The coverage of the enzyme (Γ_{cbo}) is fixed and given in Table 1. Note that for each reduction of ubiquinol-10 (UQ-10), 2 electrons are transferred and for each catalytic step (k_2) oxygen is thus reduced by 4 electrons to water.

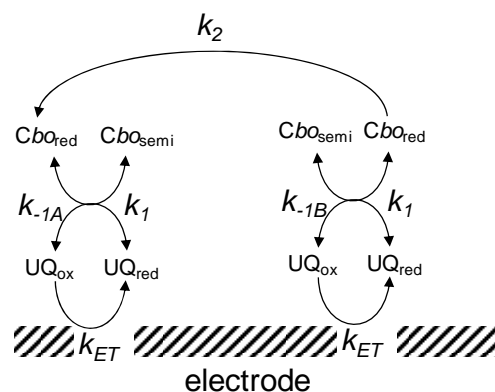


Figure S11: Schematic representation of the model used to simulate the cyclic voltammograms. See text for more explanation.

Parameter	Value
k_0	0.004 s^{-1}
k_1^{cat}	$< 500 \text{ s}^{-1}$
k_2	$< 250 \text{ s}^{-1}$
K_M^{UQ}	5 pmol/cm^2
E_0^{UQ}	0.05 V
n_{app}	1.33
Γ_{UQ}	Variable
Γ_{cbo}	50 fmol/cm^2

Table 1: Parameters used in the simulation

Results

Substrate inhibition

Figure SI2A shows the simulated cyclic voltammograms for the model outlined above. In this simulation, in which no substrate or product inhibition is modeled, three observations can be made. First, the onset of the wave shifts to more positive potentials upon addition of UQ- $\mu\text{mol}/\text{cm}^2$, similar to the experimental results. The major increase in enzyme activity is seen when increasing the UQ-10 coverage till $20 \mu\text{mol}/\text{cm}^2$, due to the chosen K_M^{UQ} of $5 \mu\text{mol}/\text{cm}^2$. Finally, at UQ-10 coverages above $20 \mu\text{mol}/\text{cm}^2$, a quinone reduction peak becomes visible that appears superimposed on the catalytic wave.

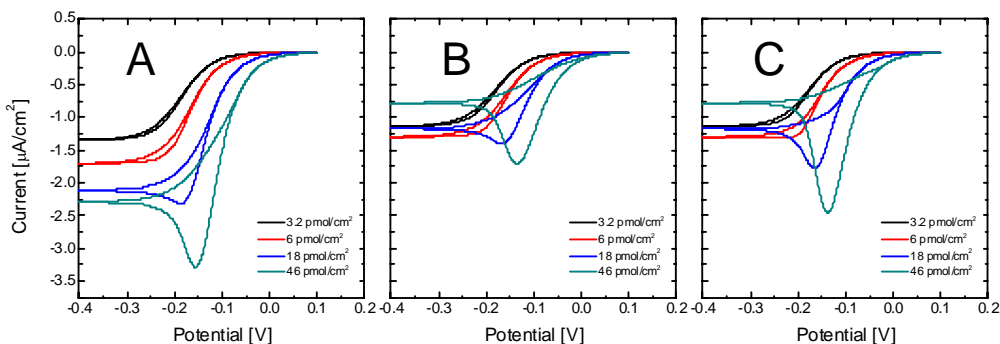


Figure SI2: Simulations of cyclic voltammograms using the models explained in detail in the text. (A) No inhibitions, (B) Substrate and product inhibition, (C) Substrate inhibition. Parameters used: $k_1^{\text{cat}} = 500 \text{ s}^{-1}$; $k_2 = 250 \text{ s}^{-1}$; Γ_{UQ} as indicated in the graphs.

In order to simulate the observed inhibition at high UQ-10 coverage (Figure 1 of the article), two models have been evaluated for their effectiveness to qualitatively describe the experimental data. In the first model, the UQ-10 oxidation activity (k_1) was inhibited as a function of the total UQ-10 concentration (oxidised or reduced) with a K_I of $12.5 \mu\text{mol}/\text{cm}^2$. Rate k_1 is thus multiplied by $K_I/(K_I + \Gamma_{\text{UQ}})$. This simulation is shown in Figure SI2B and the inhibition effect is clearly visible, as expected. Comparing Figure SI2A and SI2B shows that the ‘superimposed’ peaks at high UQ-10 coverage, due to the reduction of UQ-10 on the surface, are similar in magnitude. The (normalised) peak area is thus $\leq \Gamma_{\text{UQ}}$. In contrast, the experimental results indicate that the peak area exceeds Γ_{UQ} . To explain this, the simulation was adjusted so that catalytic rate (k_1) is only inhibited by reduced UQ-10 (substrate inhibition, Figure SI2C). In this case it is observed that the area underneath the peak at high Γ_{UQ} exceeds Γ_{UQ} supporting the hypothesis in the article that cbO_3 is inhibited by its ubiquinol (substrate), but not ubiquinone (product).

Other inhibitors

Figure SI3 shows simulations performed to model the inhibitory effects of NaCN, 2-n-Heptyl-4-hydroxyquinoline N-oxide (HQNO) and Zn(II) ions. In Figure SI3A, the NaCN behaviour is modelled by decreasing the rate k_2 as NaCN binds to the binuclear active site of cbO_3 and impairs oxygen reduction. In the bottom row of Figure SI3, the first derivatives of the simulations are shown which can be compared to the experimental data in the article (Figure 2). Similar to the experimental data, the midpoint potential (peak in the first derivative) of the catalytic wave shifts to higher potential upon lowering the oxygen reduction rate, k_2 .

In order to simulate the inhibitory effect of HQNO, the rate k_1 was systematically reduced (Figure SI3B). In contrast to the experimental data, this did not produce a set of cyclic

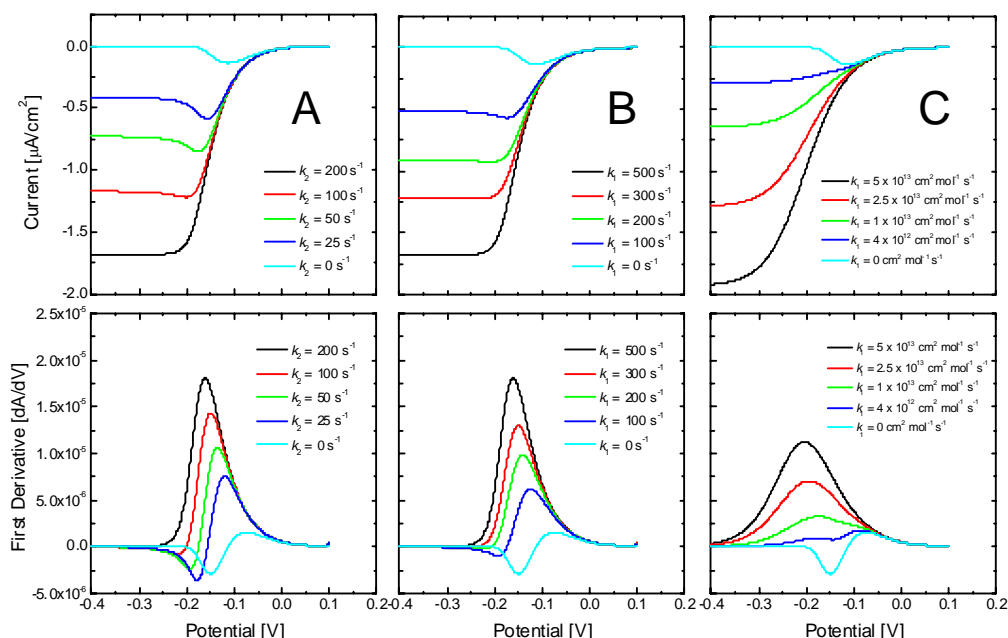


Figure SI3: Simulations of cyclic voltammograms (top) and the first derivatives (bottom) using the models explained in detail in the text. (A) Parameters as shown in Table 1, $k_1 = 500 \text{ s}^{-1}$, $\Gamma_{\text{UQ}} = 8 \text{ pmol}/\text{cm}^2$, k_2 as indicated. (B) Parameters as shown in Table 1, $k_2 = 200 \text{ s}^{-1}$, $\Gamma_{\text{UQ}} = 8 \text{ pmol}/\text{cm}^2$, k_1 as indicated. (C) Alternative model in which rate k_1 is modelled as a reversible second order rate with rate k_1 as given and rate k_{-1A} and k_{-1B} (Figure SI1) calculated using the equilibrium using the reduction potential $E_0^{\text{cho3A}} = 0.025 \text{ V}$ and $E_0^{\text{cho3B}} = 0.075 \text{ V}$, respectively.

voltammograms in which the midpoint of the catalytic wave remained largely unchanged. Detailed analysis of the modelling data indicated that this is due to the fact that the Michaelis-Menten kinetics does not take the reverse reaction into account (i.e., enzyme reduction by ubiquinone). We therefore repeated the calculations assuming the quinone/ubiquinol redox reactions are reversible assuming reduction potentials of the enzyme close to that ubiquinone/ubiquinol ($\Delta E = 0.025$ and 0.075 V for the first and second redox reaction respectively). This second model is shown in Figure SI3C, which now more closely follows the experimental observations in that the midpoint of the wave remains largely unaltered upon reducing the rate k_1 . Another feature that changes is that the catalytic wave is significantly broadened, more closely representing the experimental data. The exact wave shape of the latter model is dependent on many parameters used for the simulations, but is generally a consequence of the fact that the enzyme's oxidation state is in equilibrium with the redox state of the ubiquinol/one pool. Finally, we have repeated all the simulations shown in Figure SI2 and SI3 using the reversible quinol/quinone reaction and the conclusions made above are still valid.