Gene	Forward 5'-3'	Reverse 5'-3'	Probe 5'-Fam Tamra-3'
IFNγ	AACAGTAAAGCAAAAAAGGATGCATT	TTCATTGACAGCTTTGTGCTGG	CGCCAAGTTCGAGGTGAACAACCC
IL-2R	CACATGCTGTGTACCAGGAGAACCT	CCACGAAGTGGTAGATTCTCTTGG	CAGGTCACTGCAGGGAGCCCC
IL-6	TCAACTCCATCTGCCCTTCAG	AAGGCAACTGGCTGGAAGTCT	AACAGCTATGAAGTTTCTCTCCGCA
β-Actin	GTACAACCTCCTTGCAGCTCCT	TTGTCGACGACGAGCGC	CGCCACCAGTTCGCCATGGAT
CCR2	CACTTAGACCAGGCCATGCA	ACTTCTCACCAACAAAGGCATAAAT	TGACAGAGACTTGGAATGACACACTGCTG
CXCL9	TTGCCCCAAGCCCTAACTG	ACCCTTGCTGAATCTGGGTCTAC	CATCGCTACACTGAAGAACGGAGATCA
CXCL10	GAAGACCCTCTGGATACAGCTGC	TGCTCCACTGCCTTGCTTTT	CGCTGTCATCGATTTCTCCCCTGTGA
TLR2	AGAACTAAGAGATACTAACTTG	ACAGCTTCAGGAGTTCATTAAA	TTTCTGAACTGTCTGTAGACGAAAT
TLR4	CCTGAAGATCTTAAGAAGCTAT	CCTTGTCTTCAATTGTCTCAAT	TTCACCAATTTCTCACAACTTCAGT
TLR7	TCAGCCACAACCAGCTGACAA	AATTGCAAAGCATCTTCTAGAAA	CCTGCGAGATTGGCCAACTGTT
TLR9	CTGGACCTGTCCTATAAGAA	ACAGATTGGCCAGAAAACTGA	ACCTGTACCATTCGAAATCGTTCA
PD-L1	TGGAGTATGGCAGCAATGTC	CCTCCACAAACTGAATAACT	ATGCAGATTCCCAGTAGAACAGA
IDO	CAGGTTACAGCGCCTGGCAC	TCGCAGTAGGGAACAGCAAT	ACATCACCATGGCGTATGTGTGGAA
eNOS	CTGTGCATGGATGAATACGAT	TGCTGCAAAGCTCTCTCCAT	ATCCCTAGAGCATGAGGAATTG
iNOS	TTCCCATCGCTCCGCTG	CCGGAGCTGTAGCACGCA	AACACAGTAATGGCCGACCTGATGTTGC
FoxP3	NP_001101720.1 (Life Technologies, Carlsbad, CA)		

Table S1. RT PCR primer design.

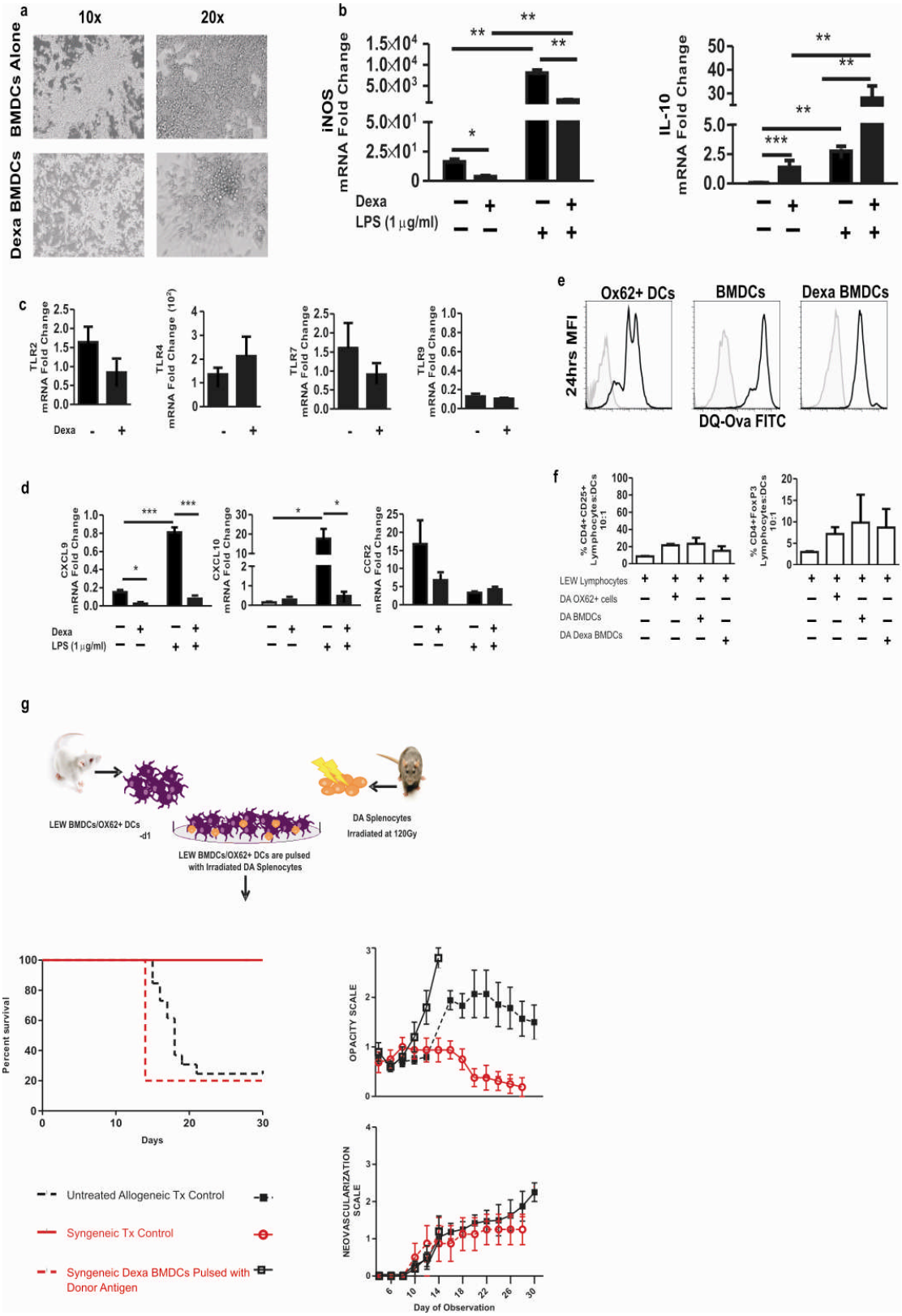
a
10x
20x
BMDCs Alone
Dexa BMDCs
b
iNOS mRNA Fold Change
IL-10 mRNA Fold Change
Dexa
LPS (1 μg/ml)
c
TLR2 mRNA Fold Change
TLR4 mRNA Fold Change
TLR7 mRNA Fold Change
TLR9 mRNA Fold Change
Dexa
d
CXCL9 mRNA Fold Change
CXCL10 mRNA Fold Change
CCR2 mRNA Fold Change
Dexa
LPS (1 μg/ml)
e
Ox62+ DCs
BMDCs
Dexa BMDCs
24hrs MFI
DQ-Ova FITC
f
% CD4+CD25+ Lymphocytes:DCs 10:1
% CD4+Fox P3 Lymphocytes:DCs 10:1
LEW Lymphocytes
DA OX62+ cells
DA BMDCs
DA Dexa BMDCs
g
LEW BMDCs/OX62+ DCs -d1
DA Splenocytes Irradiated at 120Gy
LEW BMDCs/OX62+ DCs are pulsed with Irradiated DA Splenocytes
Percent survival
Days
Untreated Allogeneic Tx Control
Syngeneic Tx Control
Syngeneic Dexa BMDCs Pulsed with Donor Antigen
OPACITY SCALE
NEOVASCULARIZATION SCALE
Day of Observation

Figure S1.

(a) Representative bright field microscopy images of BMDCs and Dexa BMDCs. (b) BMDCs and Dexa BMDC cultures mRNA expression (normalised to β-actin and fold change relative to Ox62+ DCs) of immunomodulatory molecules iNOS and IL-10 were analysed. (c) As were expression levels of TLR2, 4, 7 and 9 for BMDCs and Dexa BMDC on day 10 and (d) chemokines CXCL9, CXCL10 and the chemokine receptor CCR2 (mean ± SEM *p≤0.05, **p≤0.01 and ***p≤0.001 two-tailed Student's t test n=3). (e) Time point analysis (MFI 24hr time point illustrated) of BMDC and Dexa BMDC antigen uptake and processing was analysed using DQ OVA assay (50μg/ml, Ox62+ DCs were used as a control cell population). (f) LEW lymphocyte stimulation with DA Ox62+ DCs, BMDCs or Dexa BMDC percentage population of activated CD4+CD25+ cells and regulatory CD4+FoxP3+ve cells within these co-cultures (mean ± SEM, n=4 two-tailed Student's t test) combined analysis of two independent experiments. (g) Illustration of experimental design to pulse LEW BMDCs with DA donor derived alloantigen and graft survival curves of untreated allogeneic control (n=26), syngeneic control (n= 8) and syngeneic Dexa BMDCs pulsed with donor antigen (n=5, Kaplan-Meier survival analysis and Mantel-Cox log rank test used for comparisons between curves). Opacity scores and neovascularization day 4-day 30 for control groups and syngenic Dexa BMDCs pulsed with donor antigen.

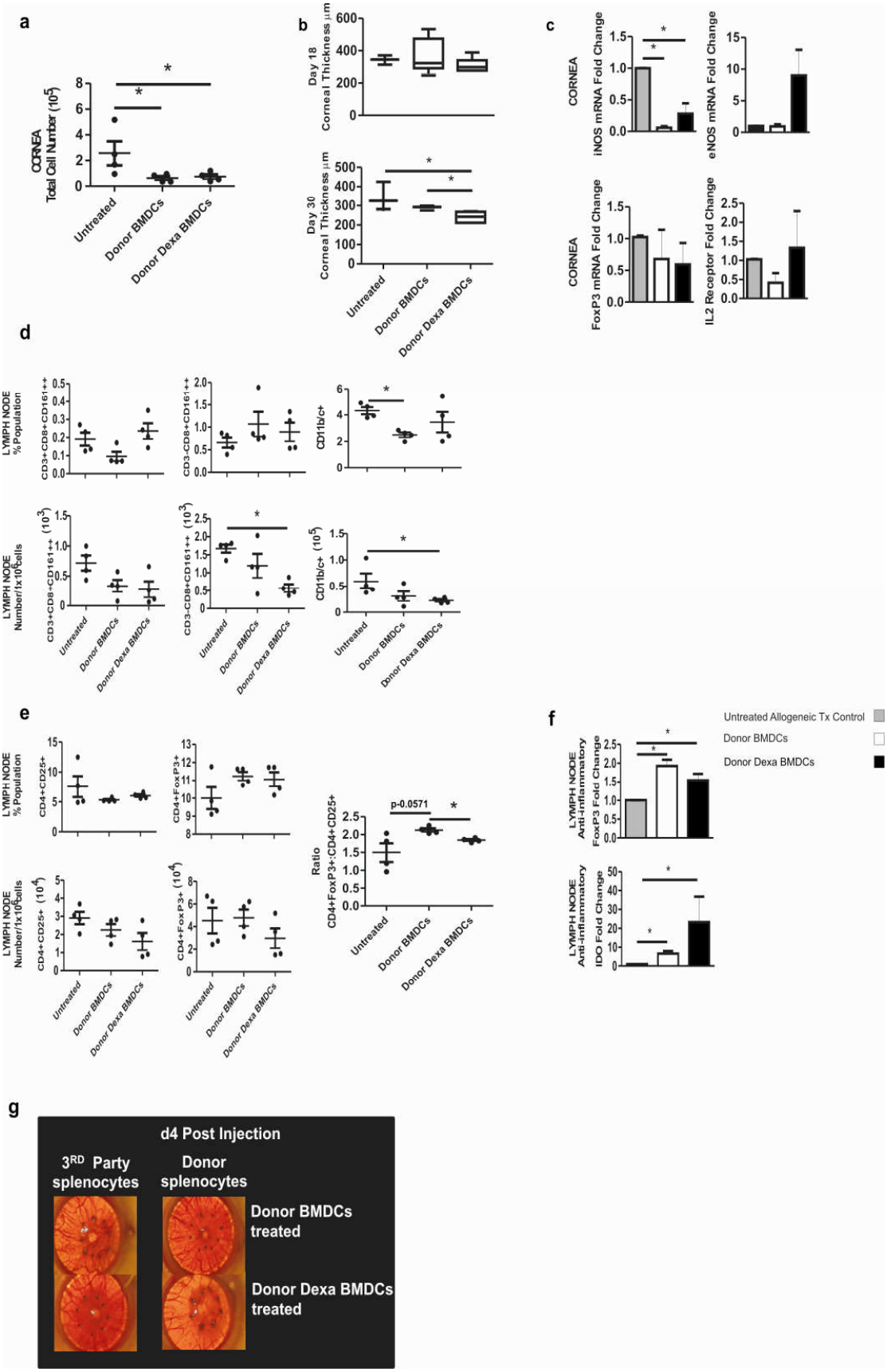
a
CORNEA
Total Cell Number (10^5)
Untreated
Donor BMDCs
Donor Dexa BMDCs
b
Day 18
Corneal Thickness µm
Day 30
Corneal Thickness µm
c
CORNEA
iNOS mRNA Fold Change
eNOS mRNA Fold Change
FoxP3 mRNA Fold Change
IL2 Receptor Fold Change
d
LYMPH NODE
% Population
CD3+CD8+CD161++
CD3-CD8+CD161++
CD11b/c+
Number/1x10^6cells
e
CD4+CD25+
CD4+FoxP3+
CD4+CD25+ (10^4)
CD4+FoxP3+ (10^4)
Ratio
CD4+FoxP3+:CD4+CD25+
p-0.0571
f
Untreated Allogeneic Tx Control
Donor BMDCs
Donor Dexa BMDCs
Anti-inflammatory
FoxP3 Fold Change
IDO Fold Change
g
d4 Post Injection
3RD Party splenocytes
Donor splenocytes
Donor BMDCs treated
Donor Dexa BMDCs treated

Figure S2.
(a) Total cell number in corneal allograft (mean ± SEM *p≤0.05 two-tailed Mann-Whitney test n=4 per group). (b) Corneal thickness was also evaluated for all groups at both time points (mean ± SEM *p≤0.05 two-tailed Mann-Whitney test n=2-5 per group). (c) mRNA analysis of intragraft cytokine expression (normalised to β-actin, fold change relative to untreated allogeneic Tx controls) of iNOS, eNOS, Foxp3 and IL2 receptor (CD25) (mean ± SEM *p≤0.05 two-tailed Mann-Whitney test n=4 per group). (d) Analysis of the same cell populations for draining LNs as described for corneal allograft, percentage cell population, cell numbers/$1x10^6$ cells and (e) ratio of lymph node regulatory CD4+FoxP3+ cells. (f) Analysis of mRNA expression (normalised to β-actin, fold change relative to untreated allogeneic Tx controls) of immunomodulatory molecules and anti-inflammatory molecules within the draining LNs (mean ± SEM *p≤0.05 two-tailed Mann-Whitney test n=4 per group). (g) Clinical evaluation of corneal allograft opacity in long-term allograft graft survivors day 4 post injection of donor derived splenocytes (n=3-4 per group).