Supporting Information

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Fig. S1. Broad-spectrum HIV-1-neutralizing activity of breast milk of uninfected women and purified milk proteins. Neutralization activity of breast milk of uninfected women and breast milk proteins TNC, lactoferrin, and mucin-1 in the TZM-bl reporter cell assay, reported as ID₅₀ or IC₅₀, respectively.



Fig. S2. Size-exclusion fractionation of breast milk reveals that HIV-1-neutralizing activity is solely contained in the high molecular mass fraction. Breast milk fractions (peaks 1–4) were tested for neutralization against the chronic clade C HIV-1 variant C.Du156 (tier 2 neutralization sensitivity) in the TZM-bl neutralization assay. Only the highest molecular mass fraction (peak 1, >500 kDa) had detectable neutralizing activity, with an inhibitory concentration 50% (IC₅₀) of 407 μ g/mL. The IC₅₀ of each size-fractionated protein peak is listed in the table.



Fig. S3. Distinct N-linked glycosylation pattern of Tenascin-C (TNC) produced in different cell lines. TNC purified from breast milk, purified from a glioma cell line (Millipore), and recombinantly produced by HEK293T cells have distinct banding patterns on an anti-TNC Western blot after deglycosylation with 100 U/μL PGNase (*Right*).



Fig. S4. Spliced short (TNC-S) and long (TNC-L) isoforms of TNC bind to HIV-1 Envelope (Env) gp120 and gp140 proteins. Recombinant TNC-L and TNC-S were covalently coupled to the surface plasmon resonance (SPR) chip, and HIV-1 Env gp120 and gp140 were flowed over the chip. TNC isoforms bound to both clade B and C and consensus Env gp120 and gp140 proteins, including the purified gp140 trimer of the transmitted/founder (T/F) virus CH0505. TNC-S and TNC-L bound with similar affinity to B.MN gp120 (54.8 and 58.2 nM, respectively).



Fig. S5. Binding of TNC to Env gp140 has a slower off-rate than the binding of TNC to Env gp120. Recombinant TNC-L and TNC-S were covalently coupled to the SPR chip, and HIV-1 Env gp120 and gp140 were flowed over the chip. TNC isoforms bound to both gp120 and gp140 proteins of the T/F HIV-1 variants B. CH0505 and C.1086, but the binding to gp140 proteins had a slower off-rate.



Fig. S6. CD4 binding to TNC does not account for the increased binding of soluble CD4-preincubated gp120 than gp120 alone to TNC. Recombinant TNC-S was covalently coupled to the SPR chip, and HIV-1 B.MN gp120 was flowed over the chip both before and after soluble CD4 preincubation. As a control, soluble CD4 alone was flowed over the TNC-S chip and no binding was detected.

DNAS



Fig. S7. Env CD4 triggering mildly increases TNC HIV-1 virion capture but not neutralization, and TNC does not antagonize the neutralizing activity of an antigp120 CD4-inducible mAb isolated from colostrum. (*A*) Preincubation of HIV-1 B.MN virions with soluble CD4 increases the efficiency of TNC virion capture by ~1.5-fold. (*B*) Preincubation of HIV-1 virions (B.Du156) with TZM-bl cells for 10 min on ice to allow virion–CD4 interaction before addition of TNC does not enhance the neutralizing potency of TNC. Graphs represent data from two assays performed in duplicate; lines indicate SD. (*C*) Preincubation of T/F HIV C.1086 virions with 100 µg/mL anti-C1 Env mAb A32, which is known to induce a conformational change of the HIV-1 Env similar to that of CD4 binding, for 1 h before the addition of TNC (200 µg/mL) and TZM-bl cells does not enhance TNC neutralization. (*D*) TNC does not antagonize the neutralizing potency of the CD4inducible, colostrum mAb CH08 against HIV-1 C.MW965 at a range of concentrations performed in duplicate.