

# Kinetic Mechanism of Human Histidine Triad Nucleotide Binding Protein 1 (Hint1)

Xin Zhou<sup>1</sup>, Tsui-Fen Chou<sup>1</sup>, Brandon E Aubol<sup>3</sup>, Chin Ju Park<sup>2</sup>, Richard Wolfenden<sup>4</sup>,  
Joseph Adams<sup>3</sup> and Carston R. Wagner<sup>1\*</sup>

<sup>1</sup>Department of Medicinal Chemistry and <sup>2</sup>Minnesota NMR Facility,  
University of Minnesota, Minneapolis MN

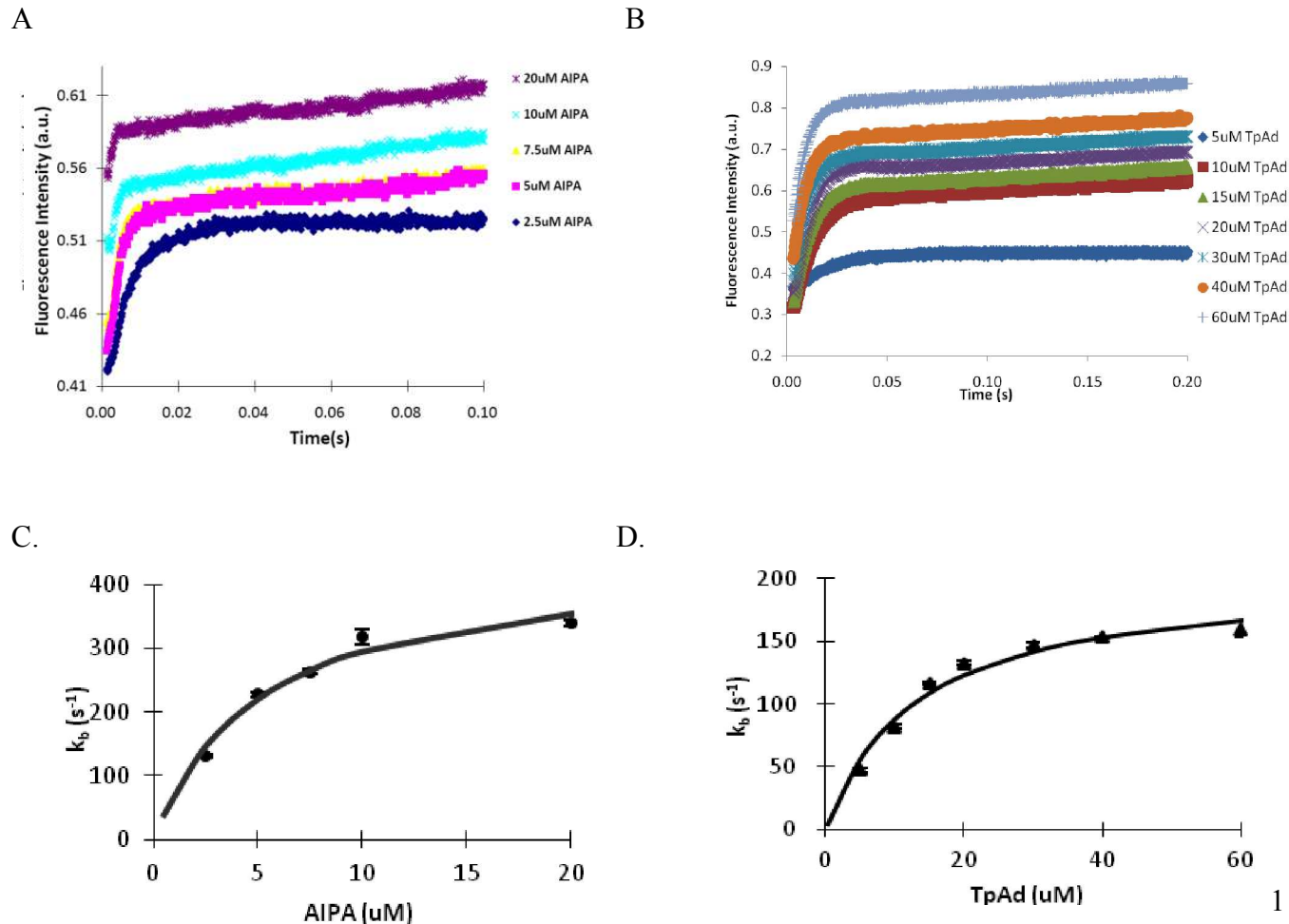
<sup>3</sup>Department of Pharmacology, University of California-San Diego, San Diego, CA

<sup>4</sup>Department of Biochemistry, University of North Carolina, Chapel Hill NC.

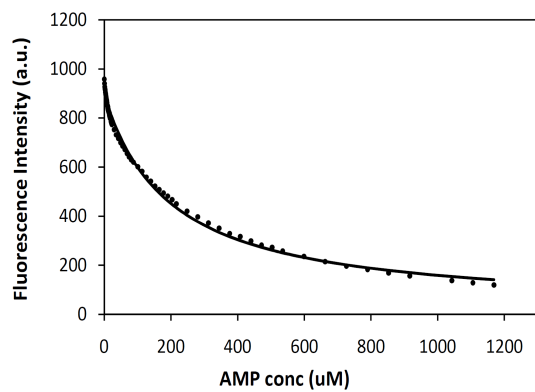
## Supporting information

### Supplemental Figure S1-S6

**Figure S1** Pre-steady state kinetics of formation of the adenylated enzyme intermediate. (A) Stopped flow traces using varied concentration of AIPA exhibited a biphasic manner, a burst phase followed by a linear phase. (B) Stopped flow traces using varied concentration of TpAd. (C) Plot of the determined burst rate constants as a function of the AIPA concentration according to Eq 2. (D) Plot of the determined burst rate constants as a function of the TpAd concentration according to Eq 2.



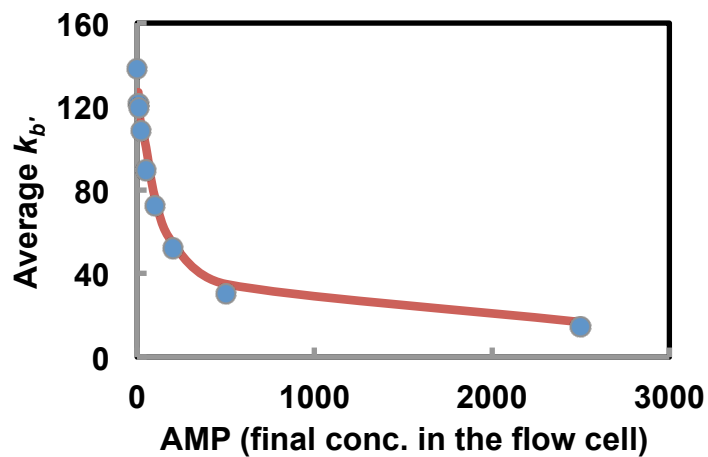
**Figure S2** Dissociation constant ( $K_d$ ) of AMP determined by fluorescence titration of hH1. The fluorescence of WT human Hint1 (6 $\mu$ M) was titrated by addition of AMP. Data were fitted using eq 15 and  $K_d = 194 \pm 9$   $\mu$ M.



**Figure S3** Determination of  $K_i$  for AMP in catalytic trapping experiments.  $k_b'$  and  $k_b$  represent the burst rates in the presence and absence of AMP, respectively. A  $K_i$  of  $136\mu\text{M}$  was estimated by fitting the curve with eq. 18.

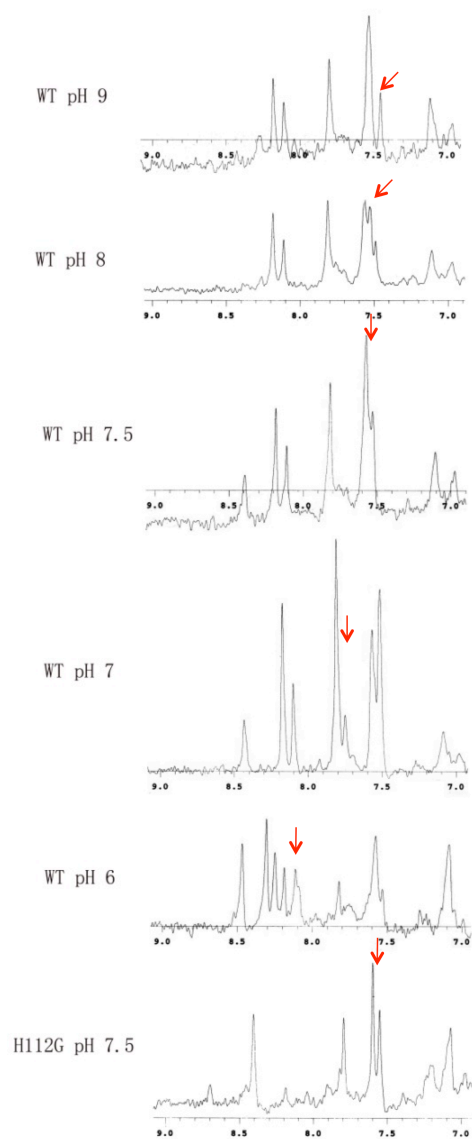
$$k_b' = k_b / (1 + [\text{AMP}] / K_i)$$

Eq. 18



**Figure S4** NMR analysis of histidine pKa in human Hint1. (A) Representative 1D proton TOCSY spectra for WT and H112G Hint1; (B) pKa titration curve for H112.

A.



B.

