Supplemental Material



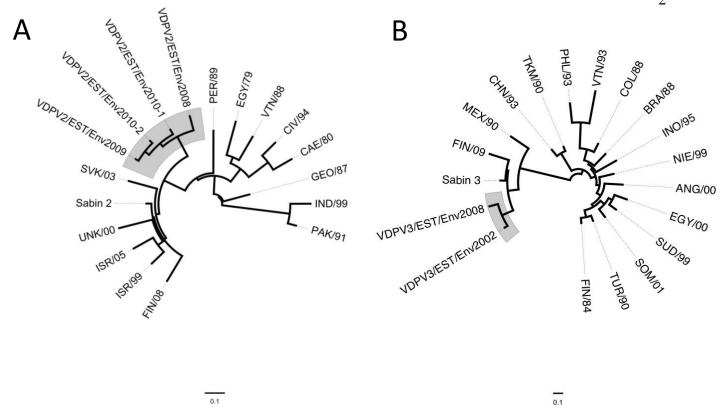


FIG S1 Maximum-likelihood trees depicting the sequence relationships in the VP1 region of the Estonian vaccine-derived poliovirus (VDPV) isolates of serotypes 2 (VDPV2) (A) and 3 (VDPV3) (B) to other environmental VDPV isolates and wild poliovirus (WPV) isolates representing different genotypes found in different parts of the world, 1979–2001. Trees with highest likelihood were inferred after a heuristic search under a general time-reversible substitution rate matrix, including the proportion of invariable sites and gamma rate heterogeneity, as determined by MODELTEST (1). Phylogenetic analyses were performed using the PAUP (2) plugin in the bioinformatics package Geneious v5.6 (http://www.geneious.com). Country abbreviations: ANG, Angola; BRA, Brazil; CAE, Cameroon; CHN, China; CIV, Côte d'Ivoire; COL, Colombia; EGY, Egypt; EST, Estonia; FIN, Finland; GEO, Georgia; IND, India; INO, Indonesia; ISR, Israel; MEX, Mexico; NIE, Nigeria; PAK, Pakistan; PER, Peru; PHL, Philippines; SOM, Somalia; SUD, Sudan; SVK, Slovakia; TKM, Turkmenistan; TUR, Turkey; UNK, United Kingdom; VTN, Vietnam. VP1 sequences of diverse VDPV2, VDPV3, WPV2, and WPV3 isolates were obtained from GenBank (PV2: CAE/80, AF551799; CIV/94, HQ286320; EGY/79, AF551798; FIN/08, KC784371; GEO/87, AF551802; IND/99, HQ286321; ISR/99, AM040036; ISR/05, AM158276; PAK/91, AF551805; PER/89, AF551803; SVK/03, JX913645; UNK/00, AJ544513; VTN/88, AF551804. PV3: ANG/00, AY221237; BRA/88, HQ286294; CHN/93, HQ286295; COL/88; HQ286296; EGY/00, GU562888; FIN/84, X04468; FIN/09, KC784373; INO/95, HQ286297; MEX/90, HQ286298; NIE/99, AY221233; PHL/93, HQ286302; SOM/01, AY221241; SUD/99, AY221243; TKM/90, KC769998; TUR/90, FJ839689; VTN/93, HQ286307).

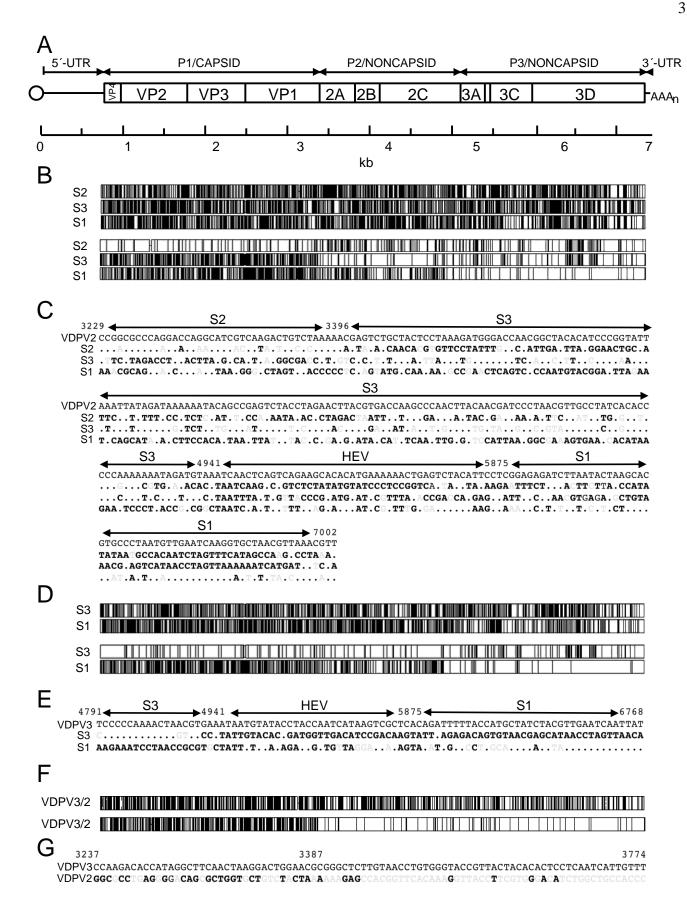


FIG S2 Approximate location of recombination sites in the Estonian VDPV2 and VDPV3 isolates. Panel A: In the schematic of the poliovirus genome, the single open reading frame is represented by an open rectangle flanked by the 5'- and 3'- untranslated regions (UTRs), represented by lines. Panel B: Alignment of the VDPV2/EST/Env2008 sequence (reference) with blocks marking sequence differences from Sabin 2, Sabin 3, and Sabin 1 (S1, S2, and S3) (upper trio: total nucleotide differences; lower trio: transversion differences). Panel C: Alignment of VDPV2/EST/Env2008 sequence (top, reference) showing nucleotide sequence differences from Sabin 2, Sabin 3, and Sabin 1 at sites of transversions (boldface type) relative to at least one of the Sabin strains. Transitions are shown in grey type. Panel D: Alignment of the VDPV3/EST/Env2002 sequence (top, reference) with blocks as described in panel B marking sequence differences from Sabin 3 and Sabin 1. Panel E: Alignment of the VDPV3/EST/Env2002 sequences (reference) showing nucleotide sequence differences from Sabin 3 and Sabin 1 at sites of transversions (boldface type). Symbols are as described in panel C. Panel F: Alignment of pairwise differences between VDPV3/EST/Env2002 (reference) and VDPV2/EST/Env2008 in total nucleotides (top) and transversions (bottom). Panel G: Pairwise transversion differences between VDPV3/EST/Env2002 (reference) and VDPV2/EST/Env2008 near the S2/S3 recombination site. Nucleotide positions are numbered according to Toyoda et al. (3) with the polymorphic sites distributed non-uniformly along the genome.

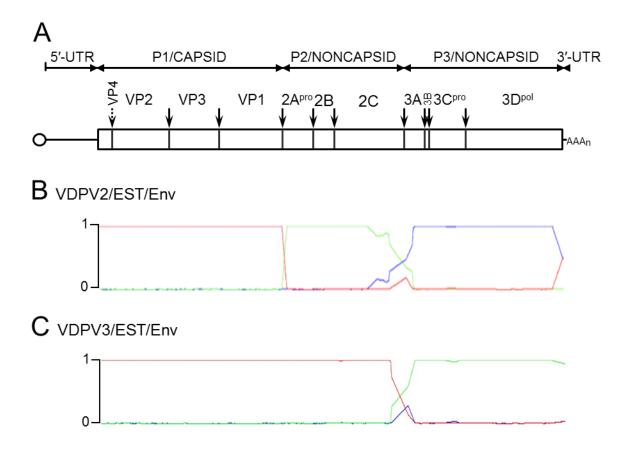


FIG S3 Schematic of the poliovirus genome (A) aligned with DualBrothers (4, 5) recombination detection profiles for VDPV2/EST/2008 (B) and VDPV3/EST/2002 (C). In each profile, as the scan moves from 5' to 3', color coding of the donor sequence in the recombinant blocks shifts from red to green to blue. Therefore, a particular donor sequence may be assigned a different color code in different plots. Profile B (5' to 3'): Sabin 2 (red)—Sabin 3 (green)—non-Sabin—Sabin 1 (blue). Profile C (5' to 3'): Sabin 3 (red)—non-Sabin—Sabin 1 (green). Plots are scaled to the posterior probability of the recombinant topology. The profiles scored only for transversional differences across donor sequences, which greatly reduced noise in the genetic signal.

		NAgS1										NAgS2											NAgS3						AgS4							
	VP1										VP2						VP1						VP3					VP2	VF	23						
	91	92	93	94	95	96	97	98	99	100	101	102	164	165	166	168	169	170	270	221	222	223	224	226	58	8 5	9 60	71	73	_	72	7				
Sabin 2	E	٧	D	N	D	Α	Р	Т	K	R	Α	s	т	N	Α	N	Р	Α	R	Α	s	т	Е	D	-	Γ 8	Q	Е	s	_	R		A .			
VDPV2/EST/Env2008	-	-	-	-	-	-	-	-	Q	-	-	-	N	-	-	-	-	т	-	s	Т	s	-	-			-	-	-		s		-			
VDPV2/EST/Env2009	-	-		-		-	-		Q	-	т	-	*	D	-	s		-	-	s	т	s		-			-	-	-		N					
VDPV2/EST/Env2010-1	-	-	-	-	-	-	-	-	Q	-	-	-	N	-	-	s	-	-	-	s	т	s	-	-			-	-	-		N					
VDPV2/EST/Env2010-2	-	-	-	-	-	-	-	-	Q	s	-	-	N	-	-	s	-	-	-	s	Т	s	-	-			-	-	-		N					
		NAgS1										NAgS2													NAgS3											
	VP1																NAg	S2									NAg	S3						NAgS	4	
														V	P2		NAg	S2	\	P1				v	P3		NAg	S3		VP1			VP2	NAgS	4 VP3	
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Sabin 3	89 E	90 V	91 D	92 N	93 E	VI	P1	96 T	97 T	98 R	99 A	100 Q	164 N			172 E					226 S	 	58 E		60 7	1 7	3				289 L	290 D			VP3	7
Sabin 3 VDPV3/EST/Env2002	E	90 V -	D	N		94 Q	P1 95	96 T	97 T A	R		Q	N	166	167 T			221	222 2 Q	23 224 I G				59 S	50 7 T		3	286 R		288 N	L		72	76	VP3	79 S

FIG S4 Substitutions in neutralizing antigenic (NAg) sites in selected Estonian VDPV strains as compared to the parental Sabin 2 and 3 strains. *, amino acid deletion.

References

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