Stem Cell Reports, Volume 1

Supplemental Information

High Yields of Oligodendrocyte Lineage Cells from

Human Embryonic Stem Cells at Physiological Oxygen

Tensions for Evaluation of Translational Biology

Sybil R.L. Stacpoole, Sonia Spitzer, Bilada Bilican, Alastair Compston, Ragnhildur Karadottir, Siddharthan Chandran, and Robin J.M. Franklin

Inventory of Supplementary Information

- 1. Supplementary Figure 1 oligodendrocyte lineage cells were derived from the HUES-9 cell line with similar efficiency; linked to figures 2 and 3
- 2. Supplementary Figure 2 hESC-NPC derived OPCs do not express neuronal or astrocytic markers; linked to figure 2
- 3. Supplementary Table 1 PCR Primers; linked to methods
- 4. Supplementary Table 2 Details of Primary Antibodies used for Immunolabelling; linked to methods

Supplementary Information:



Supplementary Figure One: Oligodendrocyte lineage cells were derived from the HUES-9 cell line with similar efficiency; related to Figures 2 and 3.

(a) OLIG2 induction at D28-35 in HUES-9 hESC-derived NPCs via the FGF-2/SHH ventral forebrain route was equivalent to that achieved with the H9 cell line ($53.1 \pm 4.2\%$). (b) After 5 weeks of terminal differentiation of D100 hESC-NPCs, large numbers of O4+ oligodendrocytes were generated from the HUES-9 line ($46 \pm 6\%$) (c) MBP expression was also observed in oligodendrocytes derived from HUES-9 hESCs. Scale bar = 50 µm.



Supplementary Figure Two: hESC-NPC derived OPCs do not express neuronal or astrocytic markers; related to Figure 2.

(a&b) There was no overlap in expression between β -III tubulin and O4, nor of NG2 and GFAP, indicating that cells labeled with NG2 or O4 were indeed oligodendroglial lineage cells rather than neurons or astrocytes. (b&c) NG2 expression did overlap with NESTIN, but only in the NESTIN+ population that did not express GFAP. b&c show the same field of view for clarity of comparison of marker co-expression. Scale bar = 50 µm.

β -ACTIN F:	5'-GTTACAGGAAGTCCCTTGCCATCC-3'
β -ACTIN R:	5'-CACCTCCCCTGTGTGGACTTGGG-3'
GSX2 F:	5'-AGCCTGAGCCGAGCGGTACTC-3'
GSX2 R:	5'-CTGGTCCTCACGTCCCCGC-3'
HOXB4 F:	5'-GTGAGCACGGTAAACCCCAAT-3'
HOXB4 R:	5'-CGAGCGGATCTTGGTGTTG-3'
<i>NKX2.1</i> F:	5'-AACCAAGCGCATCCAATCTCAAGG
<i>NKX2.1</i> R:	5'-TGTGCCCAGAGTGAAGTTTGGTCT -3'
<i>PAX6</i> F:	5'-ATGTGTGAGTAAAATTCTGGGCA-3'
<i>PAX6</i> R:	5'-GCTTACAACTTCTGGAGTCGCTA-3'

Supplementary Table 1: PCR Primers; related to methods and Figure 1.

Supplementary Table 2: Details of Primary Antibodies used for Immunolabelling; related to methods.

Primary Antibody	Isotype	Concentration	Manufacturer
β-III TUBULIN	Mouse	1 in 1000	Sigma
	IgG2b		T8660
GALC	Mouse	1 in 100	Chemicon
	IgG3		MAB342
GFAP	Rabbit	1 in 1000	Dako
	polyclonal		Z0334
MBP	Rat	1 in 100	Abcam
			AB7349
NESTIN	Mouse	1 in 500	Chemicon
(human specific)	IgG1		MAB5326
NG2	Mouse	1 in 250	BD Pharmingen
	IgG2a		554275
NG2	Rabbit	1 in 200	Chemicon
	polyclonal		AB5320
NKX2.2	Mouse	1 in 75	DSHB
	IgG2b		74.5A5
01	Mouse	1 in 1000	R&D systems
	IgM		MAB1327
O4	Mouse	1 in 1000	R&D systems
	IgM		MAB1326
OLIG2	Rabbit	1 in 500	Chemicon
	polyclonal		AB9610
PDGF-Ra	Rabbit	1 in 250	Santa Cruz
	polyclonal		SC-338
VIMENTIN	Chick	1 in 500	Chemicon
			AB5733