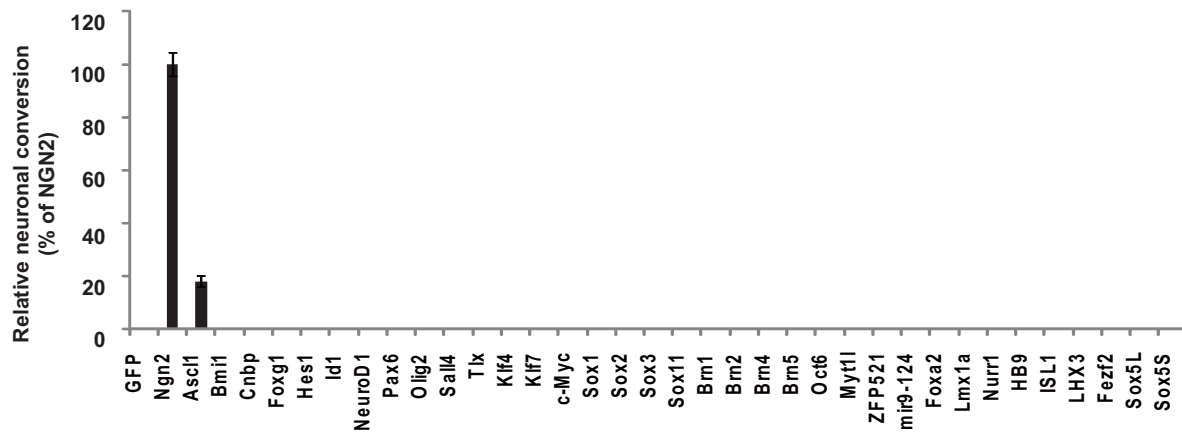
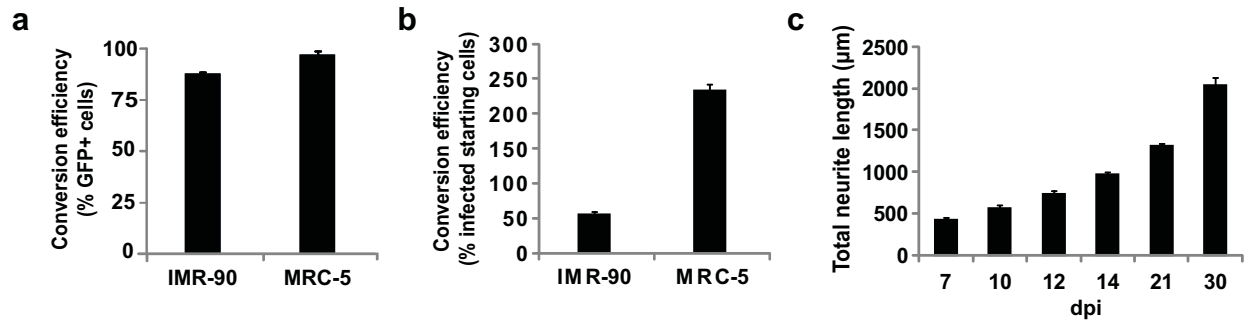


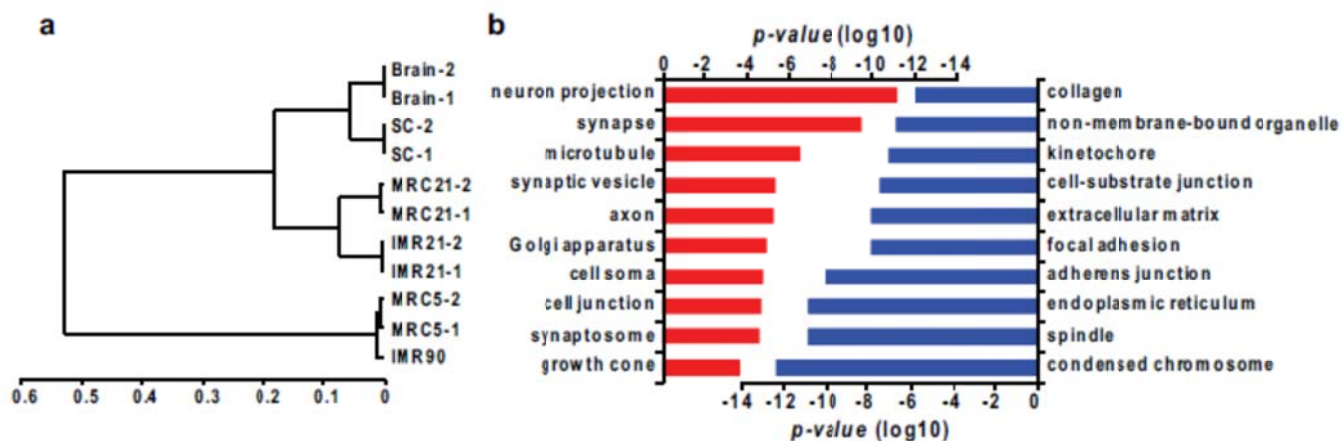
Supplementary Figure S1. Neuronal conversion in response to different dosage of forskolin (FSK). Tuj1⁺ cells were examined 5 days post treatment of IMR-90 fibroblasts (means ± s.e.m., n=20 randomly selected 20x fields from triplicate samples). Tuj1⁺ cells were undetectable in non-infected or control virus-infected fibroblasts under the same culture condition.



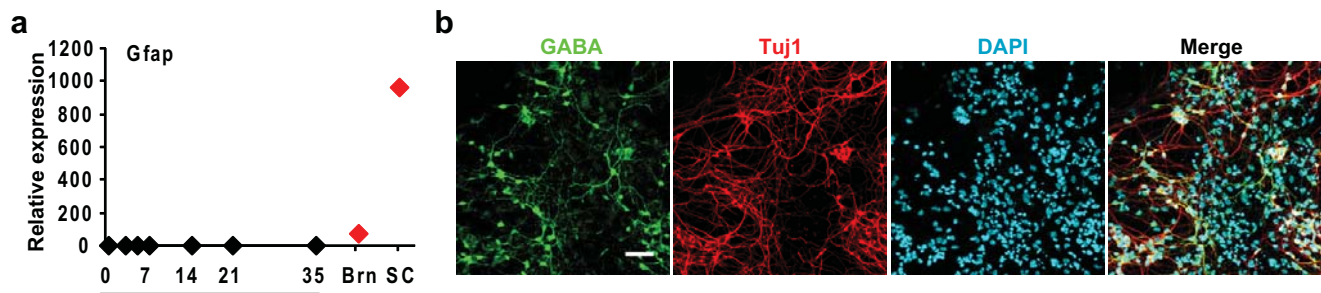
Supplementary Figure S2. FSK and DM uniquely synergize with NGN2 to reprogram human fetal fibroblasts to neurons. Tuj1⁺ cells from IMR-90 fibroblasts were quantified after 5 days of incubation with FSK and DM (means ± s.e.m., n=20 randomly selected 20x fields from triplicate sam) |[^]•[Ⓓ]



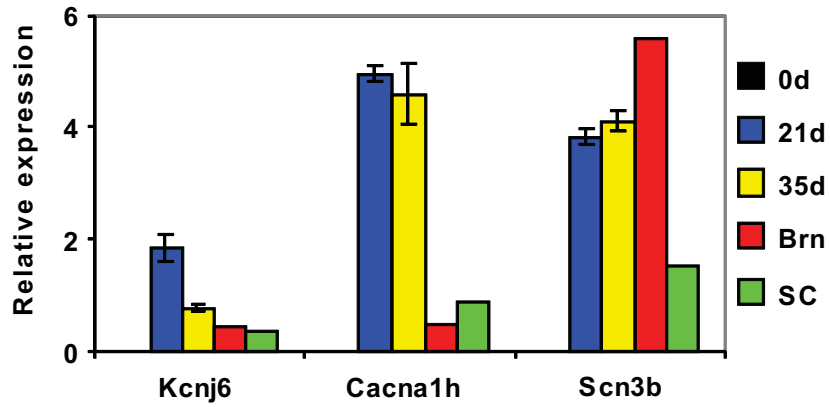
Supplementary Figure S3. Small molecule-mediated efficient conversion of human fetal fibroblasts by NGN2. **(a-b)** Conversion efficiency at 14 dpi. Total number of Tuj1⁺ cells was normalized to that of either NGN2-expressing (GFP⁺) or infected starting cells (means \pm s.e.m., n=3). **(c)** Neurite growth of converted neurons over time (means \pm s.e.m., n=10 cells).



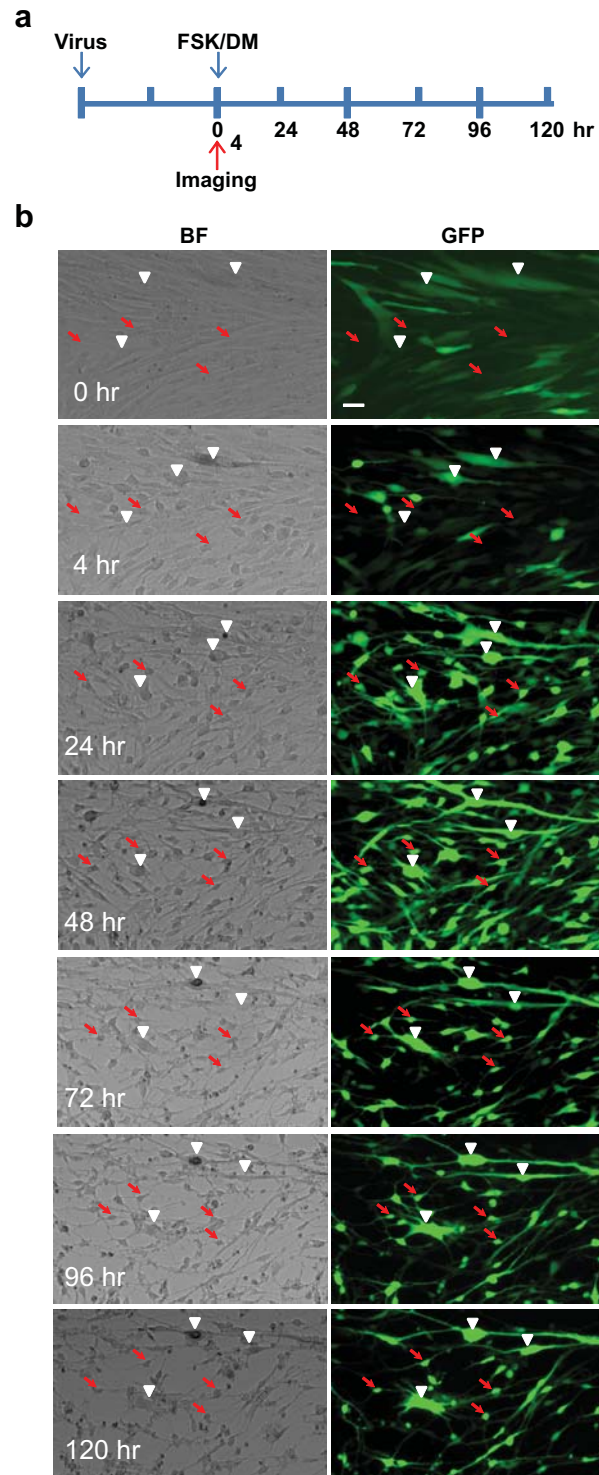
Supplementary Figure S4. Gene expression analysis of induced neurons by NGN2. **(a)** Hierarchical clustering of global gene expression. MRC21 and IMR21 represent reprogrammed cells from MRC-5 and IMR-90 fibroblasts in duplicate at 21 dpi, respectively. IMR-90 and MRC-5 are non-reprogrammed human fetal lung fibroblasts. Adult human brain and spinal cord (SC) were used as controls. **(b)** Top ten highly enriched gene sets by DAVID ontology analysis. Red and blue bars indicate significantly up-regulated and down-regulated genes, respectively.



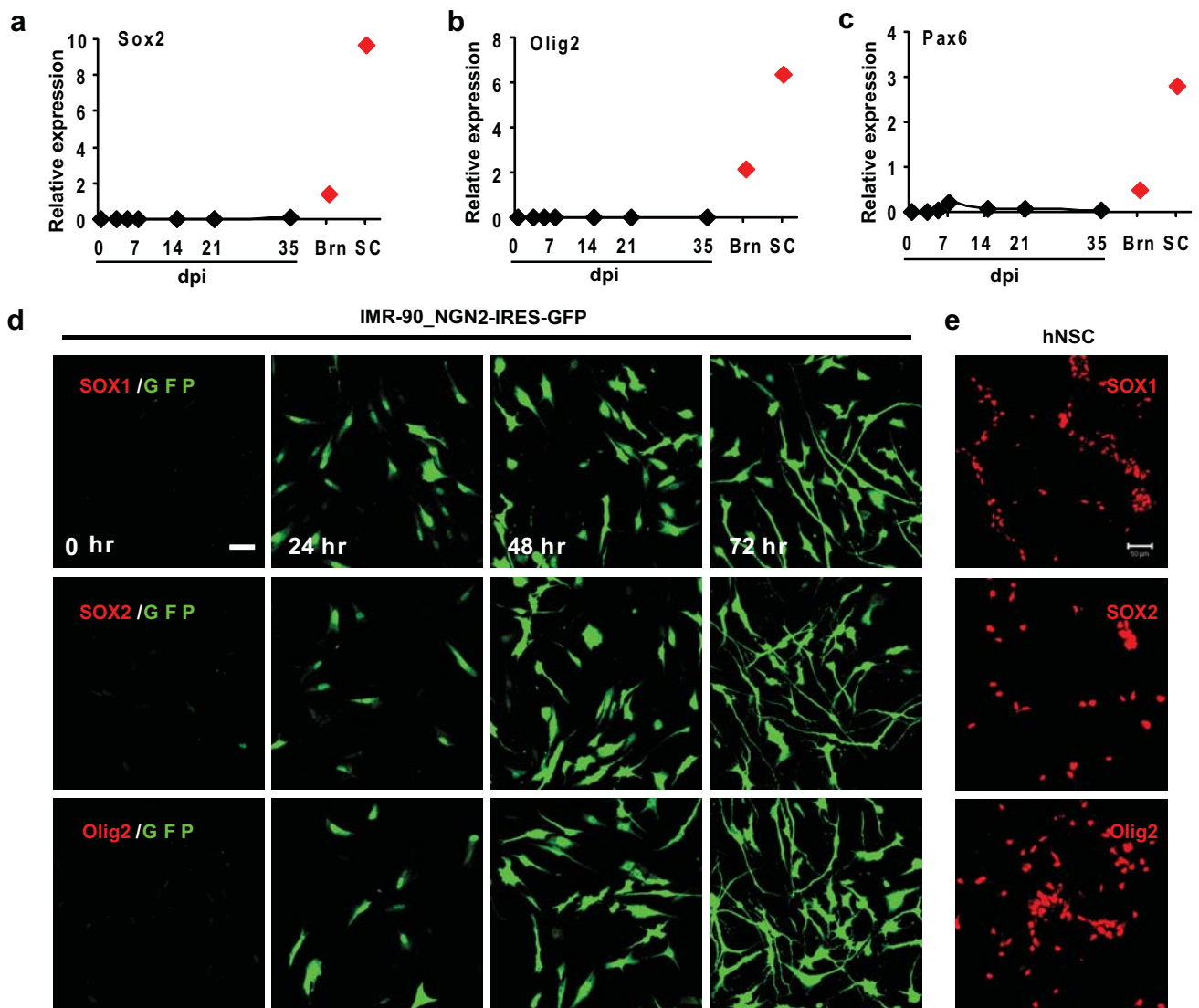
Supplementary Figure S5. Gene expression analysis of converted neurons from human fetal fibroblasts. **(a)** qRT-PCR analysis of a marker for glia during reprogramming (n=3 independent samples at the indicated time points). Samples from adult human brain (Brn) or spinal cord (SC) were used as controls (n=1). **(b)** GABA expression in neurons differentiated from human neural stem cells. This was a positive control for GABA antibody used in **Figure 1o**. Scale, 50 μ m.



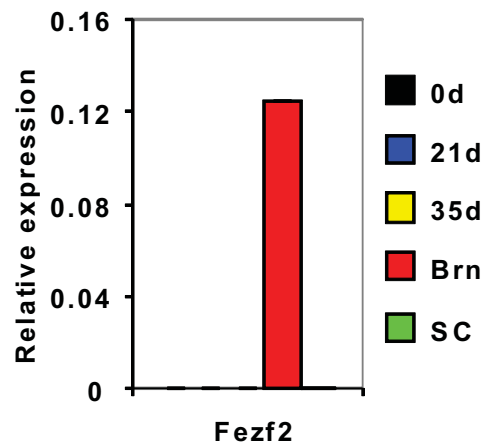
Supplementary Figure S6. Channels for potassium (Kcnj6), calcium (Cacna1h) and sodium (Scn3b) were induced in converted neurons (means \pm s.e.m., $n=3$). Samples from adult human brain (Brn) and spinal cord (SC) were used as controls ($n=1$).



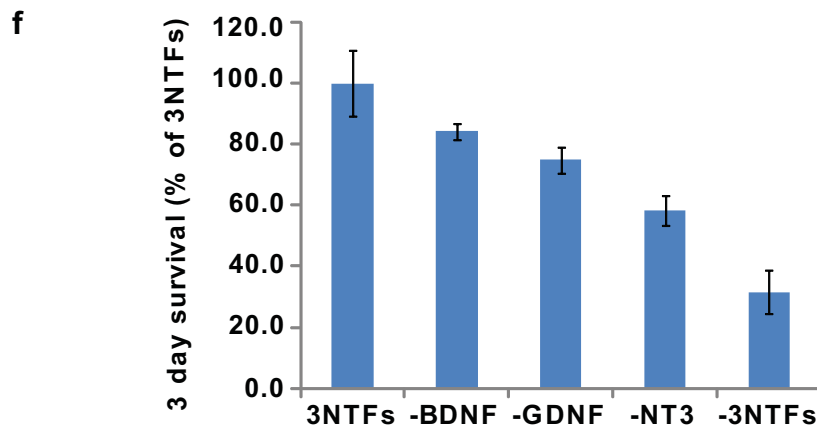
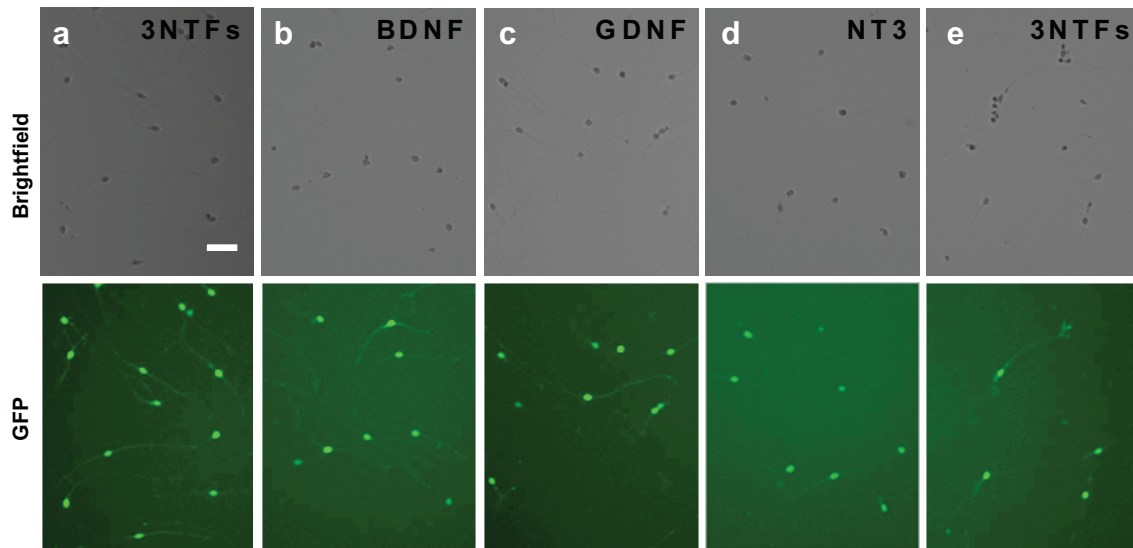
Supplementary Figure S7. Small molecules enable NGN2 to induce rapid morphological change of fibroblasts into neuron-like cells. (a) Experimental design. (b) Time-lapse live cell images taken at the indicated time points after treatment with FSK/DM. The same imaging fields are indicated by white arrowheads, whereas red arrows point to cells rapidly changing shape into neuron-like morphology without going through cell divisions. Scale, 50 μ m.



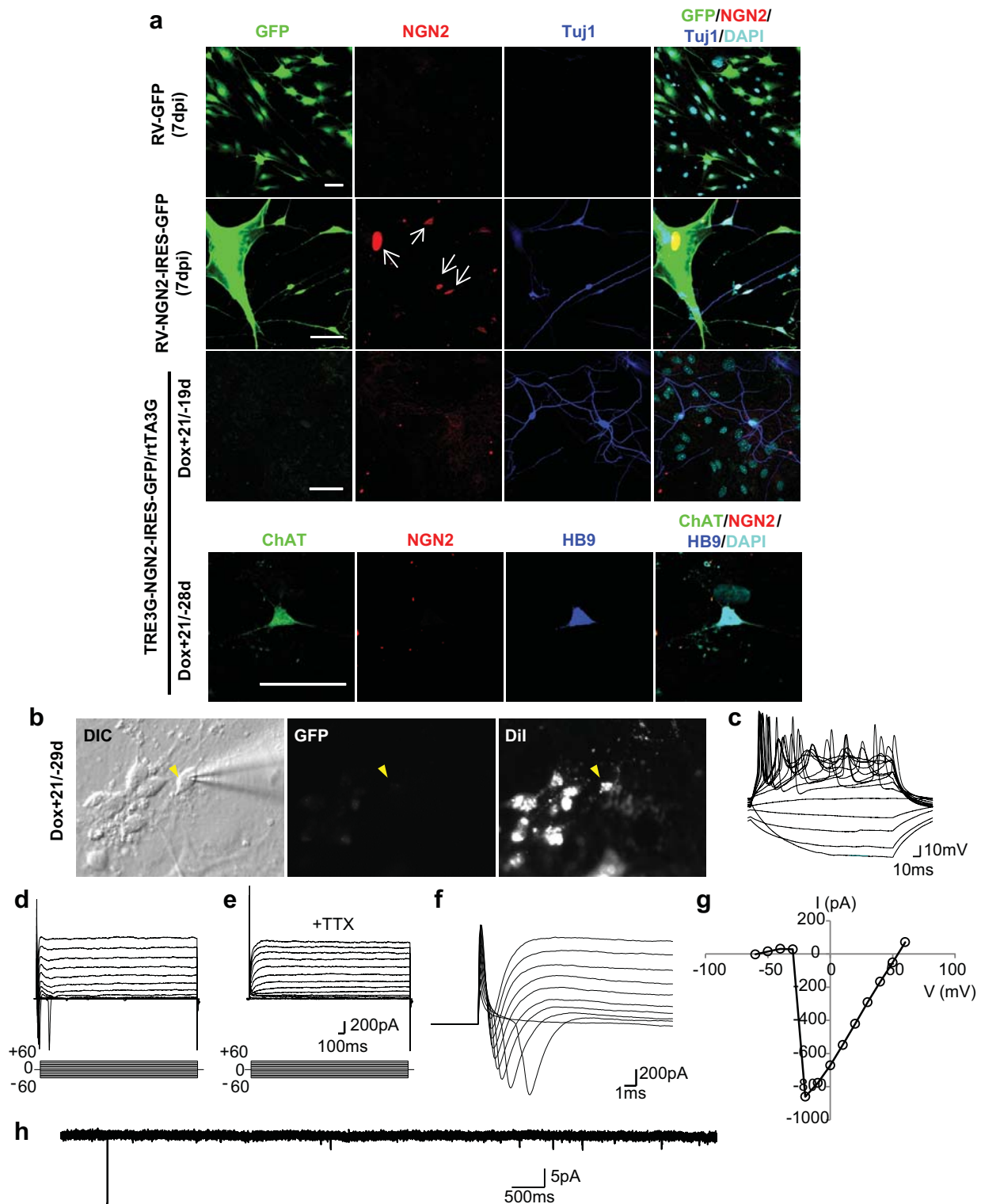
Supplementary Figure S8. Neuronal conversion from human fibroblasts does not pass through a progenitor state. **(a-c)** qRT-PCR analysis of markers for neural progenitors (means \pm s.e.m., $n=3$ independent samples at the indicated time points). Samples from adult human brain (Brn) or spinal cord (SC) were used as controls ($n=1$). **(d)** Markers for neural progenitors were not detected during the reprogramming process. **(e)** Antibody specificity was examined in human neural stem cells (hNSC). Scales, 50 μ m.



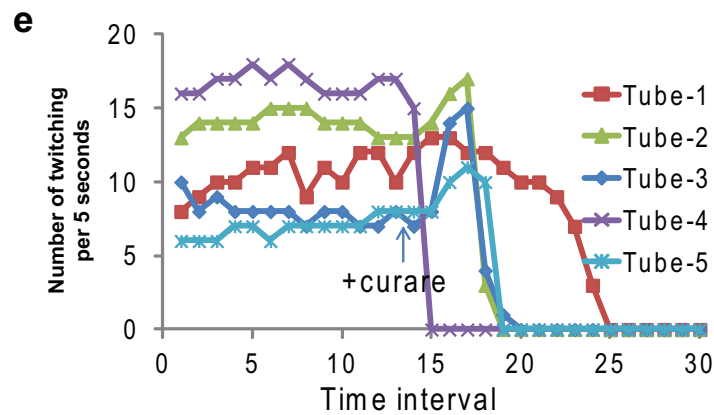
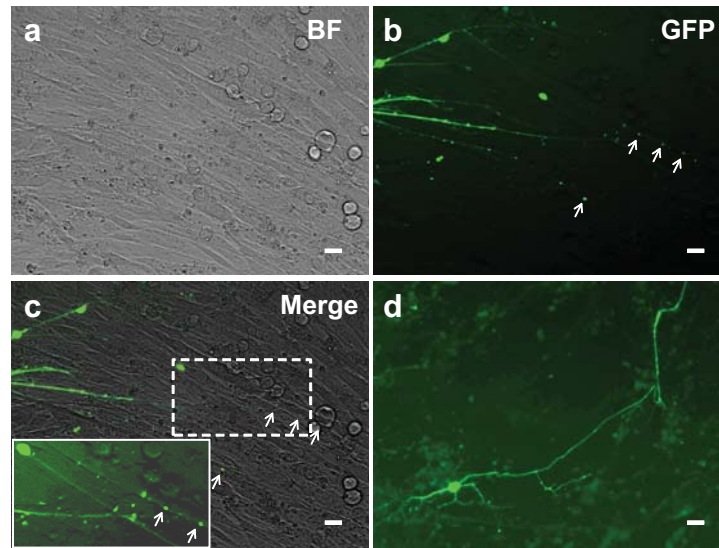
Supplementary Figure S9. Fezf2, a marker for upper motor neurons, was not detected in induced neurons by qRT-PCR (means \pm s.e.m., n=3). Samples from adult human brain (Brn) and spinal cord (SC) were used as controls (n=1).



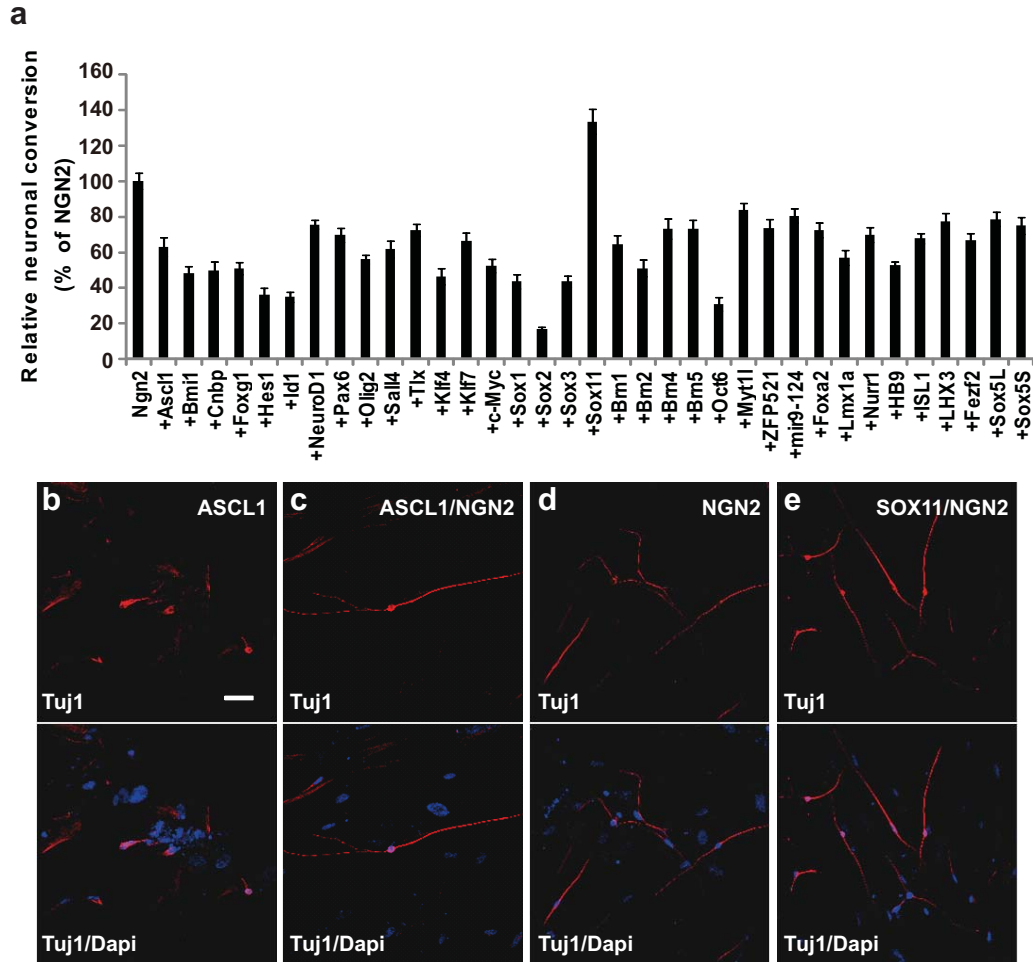
Supplementary Figure S10. NGN2-induced neurons depend on neurotrophic factors for long-term survival. (a-f) Withdrawing any of the indicated neurotrophic factors resulted in a loss of purified converted neurons. Surviving GFP⁺ cells were counted 3 days after removal of the indicated factors from triplicate samples (means \pm s.e.m.). Scale, 50 μ m.



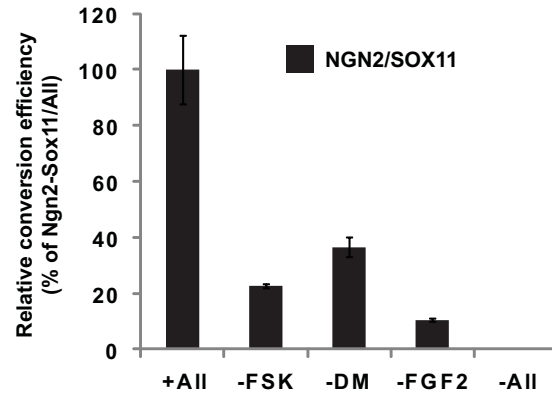
Supplementary Figure S11. Reprogrammed neurons are functionally mature after downregulation of exogenous NGN2. **(a)** Immunocytochemistry analysis showing hiCNs from MRC-5 fibroblasts express ChAT and HB9 at 49 dpi after turning off NGN2 expression by Dox withdrawal at 21 dpi (Dox+21/-28d). Lack of NGN2 expression was confirmed by staining at both 19 d and 28 d post Dox withdrawal. Antibody specificity was examined at 7 dpi using retrovirus (RV)-expressed NGN2-IRES-GFP (arrows). RV-GFP was used as a negative control. **(b)** Representative image of a patched hiCN 29 d post Dox withdrawal (Dox+21/-29d). These hiCNs were labeled at 21 dpi by Dil before plating on astrocytes-coated coverslips. **(c)** Action potentials were elicited by depolarizing current injection (-20 pA to +40 pA with 5pA interval) and resembled functionally mature neurons (n=8/8 recorded cells). **(d-e)** TTX-sensitive fast inward currents upon depolarization by voltage steps. **(f-g)** Sodium currents elicited above -30mV, with a peak current at -800 pA (n=5/7 recorded cells). **(h)** Spontaneous activity recorded from hiCNs co-cultured with mouse astrocytes (n=9/10 recorded cells). Scale, 50 μ m.



Supplementary Figure S12. hiCNs induce curare-sensitive contractions of co-cultured myotubes. (a-c) hiCNs extended long axons along the length of differentiated C2C12 myotubes and formed synaptic-like punctate structures (arrows). Live cell images were taken 7 days post co-culture. Scale, 50 μm . (d) hiCNs survived over 2 weeks in myotube-co-culture even in the absence of exogenous neurotrophic factors. (e) hiCNs stimulated rhythmic myotube contractions, which were sensitive to curare treatment.



Supplementary Figure S13. Enhancing NGN2 activity by SOX11. (a) IMR-90 fibroblasts were transduced with virus expressing NGN2 alone or in combination with the indicated factors. Tuj1⁺ cells were scored at 7 dpi (means \pm s.e.m., n=20 randomly selected 20x fields from triplicate samples). (b-e) Representative images of Tuj1⁺ cells at 7 dpi. The ASCL1-induced Tuj1⁺ cells showed very primitive unipolar morphology. Scale, 50 μ m.



Supplementary Figure S14. Highly efficient neuronal conversion from adult fibroblasts requires each of the identified extrinsic factors. Tuj1⁺ cells were counted at 21 dpi (means \pm s.e.m., n=3 wells).

Supplementary Table S1. List of small molecules

| | Name (Abbreviation) | Company (Catalog No.) | Function and Involved Signaling Pathway | Solvent (concentration) |
|---|------------------------------|------------------------------|---|--------------------------------|
| 1 | Forskolin (FSK) | Sigma (F6886) | cAMP activator, PKA pathway | DMSO (10 μ M) |
| 2 | Retinoic Acid (RA) | Sigma (R2625) | RA signaling pathways | DMSO (1 μ M) |
| 3 | Valproic Acid (VPA) | Sigma (P4543) | HDAC inhibitor, GSK3 β inhibitor, Notch-1 activator | DW (1 mM) |
| 4 | n-Butyric Acid (NaB) | Sigma (B-5887) | HDAC inhibitor | DW (1 mM) |
| 5 | Trichostatin A (TSA) | Sigma (T8552) | HDAC inhibitor | DW (0.1 μ M) |
| 6 | SB431542 (SB) | Sigma (S4317) | TGF- β R Inhibitor, Activin/TGF- β /SMAD signaling pathways | DMSO (10 μ M) |
| 7 | Dorsomorphin Compound C (DM) | EMD Chemicals (171260) | BMP inhibitor, BMP/SMAD signaling pathways | DMSO (1 μ M) |

Supplementary Table S2. List of genes examined for reprogramming human fibroblasts into neurons

| | Gene Name | Gene ID |
|----|------------------|----------------|
| 1 | Ngn2 | 63973 |
| 2 | Ascl1 | 429 |
| 3 | Bmi1 | 12151 |
| 4 | Cnbp | 12785 |
| 5 | Foxg1 | 2290 |
| 6 | Hes1 | 15205 |
| 7 | Id1 | 15901 |
| 8 | NeuroD1 | 4760 |
| 9 | Pax6 | 5080 |
| 10 | Olig2 | 10215 |
| 11 | Sall4 | 99377 |
| 12 | Tlx | 21907 |
| 13 | Klf4 | 9314 |
| 14 | Klf7 | 93691 |
| 15 | c-Myc | 4609 |
| 16 | Sox1 | 20664 |
| 17 | Sox2 | 6657 |
| 18 | Sox3 | 20675 |
| 19 | Sox11 | 20666 |
| 20 | Brn1 | 18993 |
| 21 | Brn2 | 5454 |
| 22 | Brn4 | 5456 |
| 23 | Brn5 | 19009 |
| 24 | Oct6 | 18991 |
| 25 | Myt1l | 17933 |

| | | |
|----|--------------|------------------------|
| 26 | ZNF521 | 25925 |
| 27 | mir9 cluster | 407046, 407047, 407051 |
| 28 | mir124 | 406907 |
| 29 | Foxa2 | 15376 |
| 30 | Lmx1a | 110648 |
| 31 | Nurr1 | 18227 |
| 32 | HB9 | 3110 |
| 33 | ISL1 | 3670 |
| 34 | LHX3 | 8022 |
| 35 | Fezf2 | 55079 |
| 36 | Sox5L | 6660 |
| 37 | Sox5S | 6660 |

Supplementary Table S3. Sources of examined human fibroblasts

| | Name | Repository (Catalog ID) | Origin | Age | Gender | Disease |
|---|-------------|------------------------------------|---------------|-----------------------|---------------|-------------------------------|
| 1 | IMR-90 | ATCC (CCL-186) | Fetal Lung | 16 Weeks Gestation | Female | Normal |
| 2 | MRC-5 | ATCC (CCL-171) | Fetal Lung | 14 Weeks Gestation | Male | Normal |
| 3 | GM05565 | Coriell | Skin | 3 Years | Male | Apparently Healthy |
| 4 | GM03813 | Coriell | Skin | 3 Years | Male | Spinal Muscular Atrophy |
| 5 | GM03814 | Coriell | Skin | NA ^a | Female | Apparently Healthy |
| 6 | GM07522 | Coriell | Skin | 19 Years | Female | Apparently Healthy |
| 7 | ND29563 | Coriell | Skin | 37 Years | Male | Amyotrophic Lateral Sclerosis |

Supplementary Table S4. qRT-PCR Primers used in this study for quantitative gene expression

| Gene Name | Primer Sequences | Sense/Antisense |
|------------------|--------------------------|------------------------|
| Cacna1h | TGACCTTCGGCAACTATGTG | Sense |
| | GGAGTTCTCTGAGCTTGTGG | Antisense |
| ChAT | GCACTCCAGCTCCTTCAC | Sense |
| | CACTGCACCAGGACGATG | Antisense |
| Fezf2 | GGTACTGAAGGAAACTCGGC | Sense |
| | CGGGTGAGATTATAGTGAGCG | Antisense |
| GFAP | GTGTCAGAAGGCCACCTCAAG | Sense |
| | TGGACTCCTTAATGACCTCTCCAT | Antisense |
| HB9 | GCACCAGTTCAAGCTCAAC | Sense |
| | GCTGCGTTTCCATTTTCATCC | Antisense |
| HOXA5 | GCAAGCTGCACATAAGTCATG | Sense |
| | AGGTAACGGTTGAAGTGG AAC | Antisense |
| HOXA6 | GTACACGAGCCCGGTTTAC | Sense |
| | CAGGTAGCGGTTGAAGTGG | Antisense |
| HOXA7 | AATTTCCGCATCTACCCCTG | Sense |
| | GTGGGCGATTTCAATGCG | Antisense |
| HOXA9 | AATGCTGAGAATGAGAGCGG | Sense |
| | GGGTCTGGTGTTTTGTATAGGG | Antisense |
| HOXB4 | TCGTCTACCCCTGGATGC | Sense |
| | GTGTCAGGTAGCGGTTGTAG | Antisense |
| HOXB6 | TGGATGCAGCGGATGAATTC | Sense |
| | CGTCAGGTAGCGATTGTAGTG | Antisense |
| HOXB7 | CCTGGATGCGAAGCTCAG | Sense |
| | CGTCAGGTAGCGATTGTAGTG | Antisense |
| HOXC6 | GACCAGAAAGCCAGTATCCAG | Sense |
| | AAATTCCTTCTCCAGTTCAGG | Antisense |
| HOXC8 | CTAACAGTAGCGAAGGACAAGG | Sense |
| | CTAGTTCCAAGGTCTGATACCG | Antisense |
| HOXC9 | AGCACAAAGAGGAGAAGGC | Sense |
| | CGTCTGGTACTTGGTGTAGG | Antisense |
| HOXC10 | AAAGGAGAGGGCCAAAGC | Sense |
| | GCGTCTGGTGTTTAGTATAGGG | Antisense |
| HOXC11 | ACCGGCTGCAGTATTTCTC | Sense |
| | ACAGTCCAGTTTTCCACCG | Antisense |
| HOXD9 | GGCTGTTGCTGAAGGAG | Sense |
| | CGTCTGGTATTTGGTGTAGGG | Antisense |
| ISL2 | CAACAGTATGGTGCCGAGTC | Sense |
| | AGAGCTTTCGGAAGTGAATG | Antisense |
| Kcnj6 | GAAGTGGAGACTGAAGAGGAAG | Sense |

| | | |
|----------|------------------------|-----------|
| | GACAAGGAAAGATTGTGTTGGG | Antisense |
| endoNGN2 | TCAGACATGGACTATTGGCAG | Sense |
| | GGGACAGGAAAGGGAACC | Antisense |
| vNGN2 | TCAGACATGGACTATTGGCAG | Sense |
| | ACACCGGCCTTATTCCAAG | Antisense |
| Olig2 | AGCTCCTCAAATCGCATCC | Sense |
| | AAAAGGTCATCGGGCTCTG | Antisense |
| Pax6 | GCCCTCACAACACCTACAG | Sense |
| | TCATAACTCCGCCATTAC | Antisense |
| Scn3b | GAAAGGTCTCAAAGCCGAAG | Sense |
| | CACTGCTCCTGTTCTATTCCTC | Antisense |
| SMN1 | CCACACCTAAAAGAAAACCTGC | Sense |
| | GCAATGGTAGCTGGGTAAATG | Antisense |
| Sox2 | CCGTTTCATCGACGAGGCTAA | Sense |
| | TAATCCGGGTGCTCCTTCAT | Antisense |

