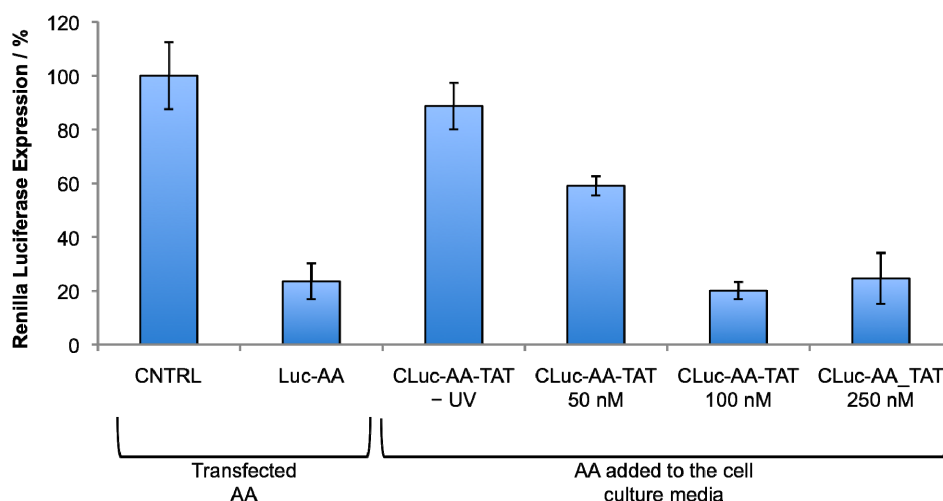


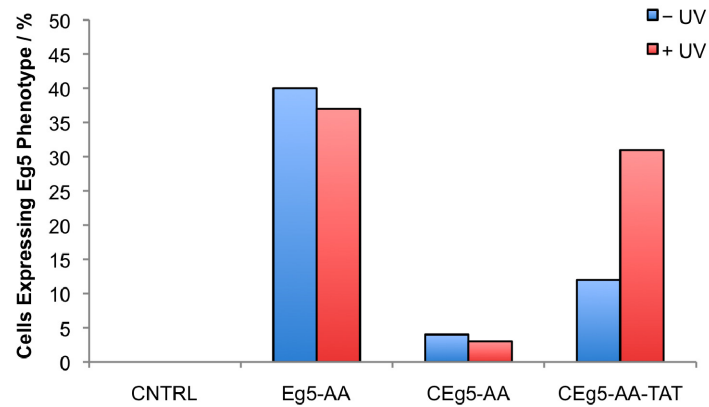
Cellular Delivery and Photochemical Activation of Antisense Agents through a Nucleobase Caging Strategy

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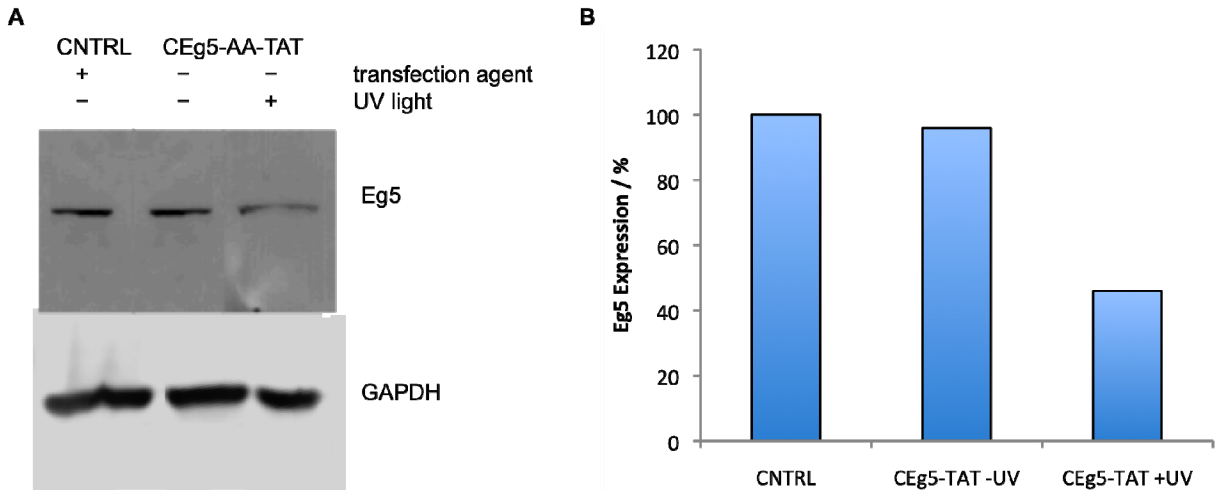
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Supporting Figure 1. Titration of HIV TAT conjugated antisense agents. Antisense agents (CNTRL and Luc-AA) were actively transfected into HEK293T cells. Increasing concentrations of CLuc-AA-TAT (0, 50, 100, 250 nM) were simply added to HEK293T cell culture media. Cells were either kept in the dark or irradiated (365 nm, 2 min, 25 W). The cells were expressing firefly and *Renilla* luciferase from pGL3 and pRL-TK plasmids, respectively, and a dual-luciferase assay was performed after 48 h. *Renilla* luciferase expression was normalized to firefly luciferase expression and the negative control antisense agent (CNTRL) was set to 100%. All experiments were performed in triplicate and error bars represent standard deviations.



Supporting Figure 2. HeLa cells were passaged into 4-well chamber slide (1 mL per well, $\sim 4 \times 10^4$ cells per well) and grown to $\sim 70\%$ confluence within 24 h. Conjugated antisense agents (100 nM) were added directly to the cell culture media and incubated overnight at 37 °C, 5% CO₂. The media was removed and the cells were irradiated for 2 min on a UV transilluminator (365 nm, 25 W). Control antisense agents were transfected into the cells with X-tremeGENE siRNA reagent (Roche). DMEM media was added and the cells were incubated at 37 °C, 5% CO₂ for 48 h. The cells were fixed with formaldehyde (3.75%) and permeabilized with Triton-X100, stained with Alexa Fluor 488 Phalloidin (ex/em 488 nm/495-630 nm, Invitrogen) and DAPI (ex/em 405 nm/410-495 nm, Invitrogen). 100 total cells were counted for the binucleated phenotype and plotted against the antisense agent treatment.

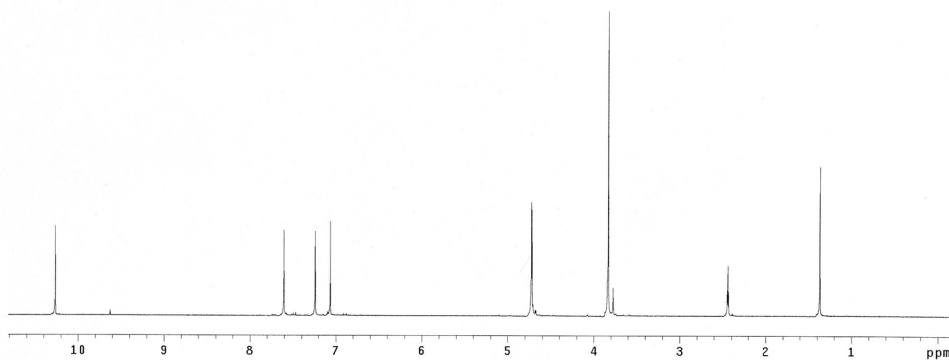
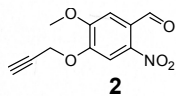


Supporting Figure 3. Western Blot of photochemical Eg5 inhibition. HeLa cells were passaged into 6-well plates (2 mL per well, $\sim 2 \times 10^5$ cells per well) and grown to $\sim 70\%$ confluence within 24 h. Conjugated antisense agents (100 nM) were added directly to the cell culture media and incubated overnight at 37 °C, 5% CO₂. The media was removed and the cells were irradiated for 2 min on a UV transilluminator (365 nm, 25 W). Control antisense agents were transfected into the cells with X-tremeGENE siRNA reagent (Roche). DMEM media was added and the cells were incubated at 37 °C, 5% CO₂ for 48 h. The cells were lysed with mammalian protein extraction buffer (GE Healthcare) and the lysate was analyzed by 12 % SDS-PAGE. A Western blot was performed using Eg5 (1:500 dilution, Santa Cruz Biotechnology) and GAPDH (1:1,000 dilution, Santa Cruz Biotechnology) primary antibodies, and a goat-anti-mouse-HRP (1:1,000 dilution, Santa Cruz Biotechnology) secondary antibody. The Western blot was developed by HRP colorimetric staining (Bio-Rad) and (A) imaged on a Typhoon 7000 phosphorimager (GE Healthcare). (B) The bands were quantified with *ImageQuant TL* software and the bands corresponding to Eg5 expression were normalized to GAPDH expression bands. For normalization purposes, the CNTRL antisense agent was set to 100% Eg5 expression.

¹H NMRs of synthesized compounds:

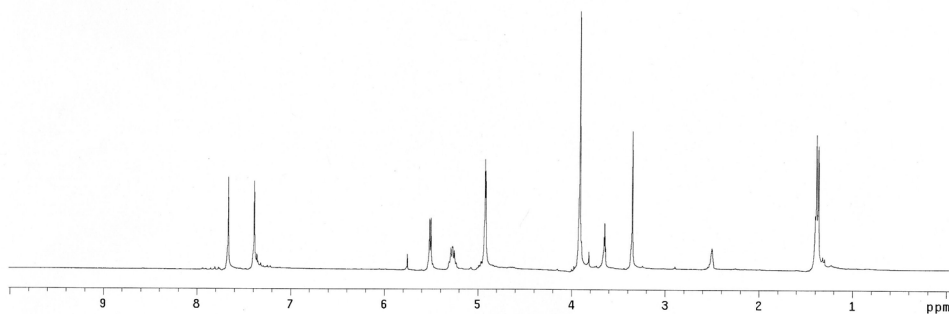
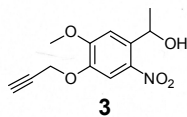
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DATA PROCESSING
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Total time 0 min, 49 sec



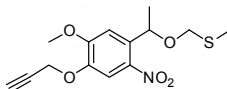
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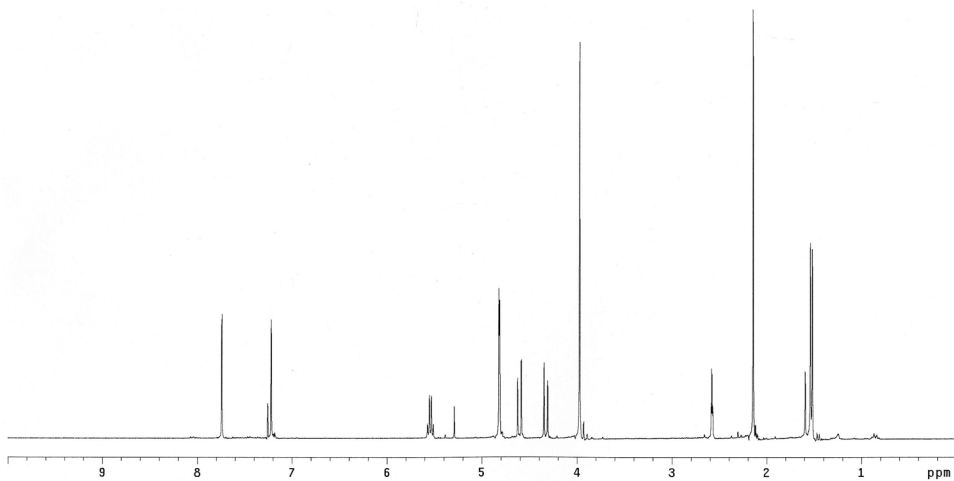


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DATA PROCESSING
FT size 32768
Total time 0 min, 49 sec

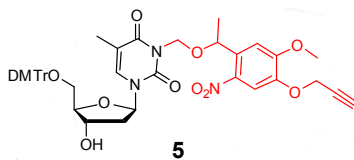


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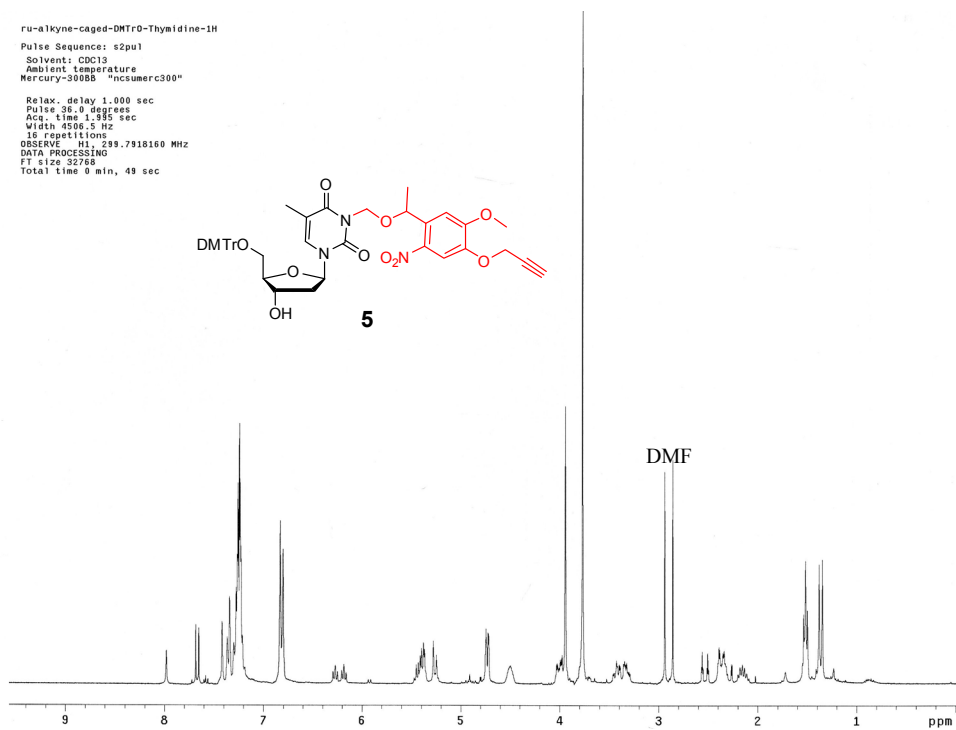


ru-alkyne-caged-DMTrO-Thymidine-1H

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18 repetitions
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DATA PROCESSING
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Total time 0 min, 49 sec



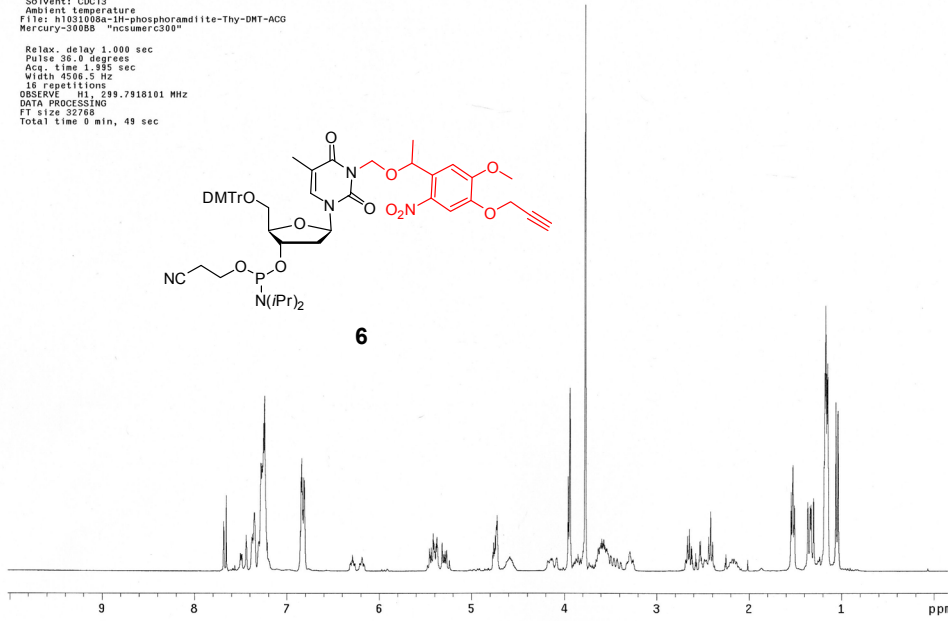
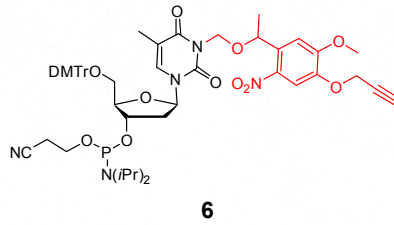
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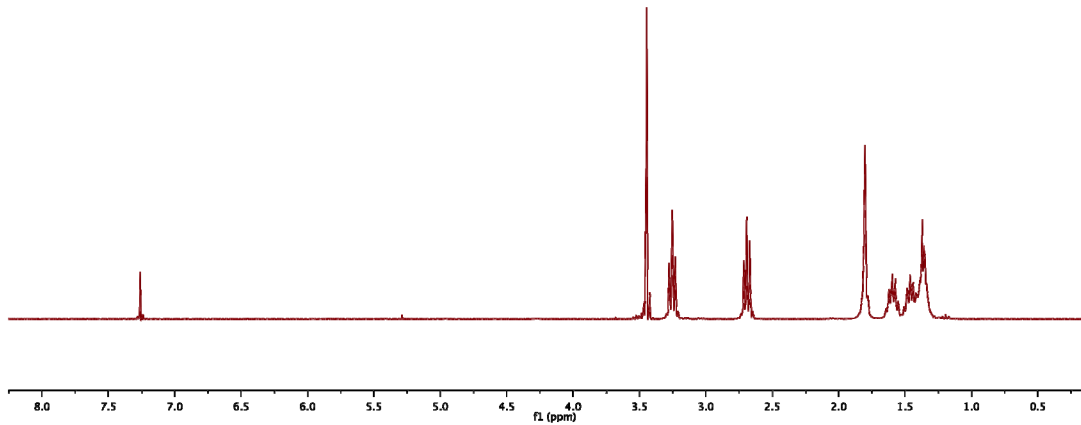
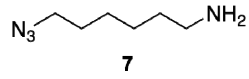
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Mercury-300BB "ncsummerc30B"

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Width 4506.5 Hz
18 repetitions
OBSERVE: H1, 299.7918101 MHz
DATA PROCESSING
F1 size 32768
Total time 0 min, 49 sec



MT03-169
MT03-169



MT04-110-ON
MT04-110-HPLC

