Supplementary Figure 1. Reduced RB1 protein expression in osteosarcomas derived in WT, $Rb1^{+/-}$ and $II-6^{-/-}$ mice. Protein (25 μ g) from wild-type normal bone, $Rb1^{+/-}$ tissue and tumors was resolved on a 7% SDS-PAGE gel and probed for the presence of RB1.



Supplementary Figure 2. Markers of senescence 10 days after exposure to 4Gy radiation. A) SA- β -Gal staining of hOB shEV (left panel), and shRB1 (right panel) 10X magnification. B) Percentage of flattened cells shEV and shRB1 10 days after 4Gy IR (4 panels were counted for each cell line with 53-301 cells per field), STDEV is shown. C) Western blot analysis of cyclin dependent inhibitors: p16 and p21 (CDKN2A and CDKN1A), anti-Histone H3 (tri-methyl K9) (H3K9) and ACTIN.



Supplementary Figure 3. Changes in cytokine/chemokine protein expression by bone cells following IR using BD bead arrays and FACS analysis. Condition media was collected from control cells (CM) and cells irradiated at 4Gy (CM ⁴⁵Ca) after 7 days. Of the 20 proteins analyzed (IL-1α, IL-1β, IL-2, IL-4, IL-5, IL-6, IL-10, IL-12p70, IL-13, IL-17A, IL-21, G-CSF, GM-CSF, RANTES, MIG, MCP1, MIP1α, MIP1β, TNFα and IFN_γ), only 5 proteins were detectable following irradiation from condition media: IL-6, MCP1, RANTES, MIP1α and MIP1β.



Supplementary Figure 4. Recombination in *Osx-Cre+ Rb1*^{*fl/fl*} mice. Genomic DNA was extracted from calvaria, tibia (flushed of bone marrow cells) an osteosarcoma (derived from *Osx-Cre p53*^{*fl/fl*} *pRb*^{*fl/fl*}) and muscle. PCR was performed with control (*Recql4* WT allele PCR) and *Rb1* deletion PCR (% loxP and RbCre3-2 primers) to detect the exon 3-deleted allele. For quantification the unexcised allele shows up as a band at approximately 1200-1500 bp. In non-tumor bone approximately 50% excision was observed. In tumor 100% excision was observed.



gDNA 1. Calvaria 2. Tibia 3. Osteosarcoma 4. Muscle

5. Blank lane

Supplementary Figure 5. Growth of *II-6^{-/-}* osteosarcoma cell lines is suppressed when implanted into a wild-type background. A) Shows a repeat cross transplantation experiment: cell lines wild-type #18 and *II-6^{-/-}* #12. B) Shows an independently derived set of cell lines WT#5 and *II-6^{-/-}*#13. Cells (1 x 10⁶) were implanted subcutaneously into flanks of mice. Tumor volumes plotted for each cohort (WT/WT, WT/*II-6^{-/-}*, *II-6^{-/-}*/*II-6^{-/-}* and *II-6^{-/-}*/WT) over 60 days.



Days post cell line implantation

Supplementary Figure 6. Increased *II-6* transcript levels in *II-6^{-/-}* osteosarcomas grown in WT mice. qRT-PCR was carried out on RNA derived from WT tumors implanted in WT mice and in *II-6^{-/-}* tumors implanted in WT mice. Values are shown as expression relative to WT cell line \pm SEM and are representative of three independent experiments.



Supplementary Figure 7. *II-6^{-/-}* tumor cells had reduced capacity to undergo senescence compared to WT tumor cells following IR. Cell lines derived from WT and *II-6^{-/- 45}*Ca induced tumors were cultured ex vivo and irradiated at 4 Gy. Seven days after irradiation, cells were stained for (A) SA- β -Gal and (B) the number of flattened cells were quantitated. Values show % of cells that are SA- β -Gal positive and/or flattened ± SEM and are representative of two independent experiments.



Supplementary Figure 8. Dot plot showing differential infiltration of immune cells post-IR in spines of $Rb^{+/+}$ compared to $Rb^{t/t}$ mice. Dot plots showing representative images for CD8, NKT cells and macrophages.



Supplementary Table 1. Enrichment of expression of SASP genes among genes regulated by RB1 following ionizing radiation. The SASP signature derived from Coppe *et al* (Coppe et al., 2008) was significantly enriched in hOB compared to RB1 knockdown cells following IR (FDR q-value 0.0054). RANK = rank in gene list; ES = enrichment score, which reflects the degree to which a gene set is overrepresented at the top or bottom of a ranked list of genes.

PROBE	SYMBOL	GENE TITLE	RANK	ES	ENRICHED
8131803	IL6	interleukin 6 (interferon, beta 2)	13	0.0903	Yes
8006433	CCL2	chemokine (C-C motif) ligand 2	19	0.1771	Yes
8112139	IL6ST	interleukin 6 signal transducer	52	0.2474	Yes
8152512	TNFRSF11B	TNF receptor superfamily, member 11b	65	0.3165	Yes
8054722	IL1B	interleukin 1, beta	108	0.3746	Yes
7965322	KITLG	KIT ligand	568	0.3955	Yes
8095680	IL8	interleukin 8	634	0.4287	Yes
7929032	FAS	Fas (TNF receptor superfamily, member 6)	671	0.4623	Yes
7960518	TNFRSF1A	TNF receptor superfamily, member 1A	713	0.4949	Yes
8018972	TIMP2	TIMP metallopeptidase inhibitor 2	829	0.5232	Yes
8018966	TIMP2	TIMP metallopeptidase inhibitor 2	882	0.5531	Yes
8138542	IL6	interleukin 6 (interferon, beta 2)	949	0.5817	Yes
8048864	CCL20	chemokine (C-C motif) ligand 20	1885	0.5725	Yes
8107887	CSF2	colony stimulating factor 2	2234	0.5818	Yes
8135909	LEP	leptin	2517	0.5920	Yes
8033987	ICAM3	intercellular adhesion molecule 3	3569	0.5717	No
8097256	FGF2	fibroblast growth factor 2 (basic)	3941	0.5740	No
8095697	CXCL1	chemokine (C-X-C motif) ligand 1	4031	0.5859	No
8014342	CCL16	chemokine (C-C motif) ligand 16	4938	0.5674	No
8032899	TICAM1	toll-like receptor adaptor molecule 1	5147	0.5727	No
7955694	IGFBP6	insulin-like growth factor binding protein 6	6317	0.5425	No
8039484	IL11	interleukin 11	7104	0.5244	No
8075316	OSM	oncostatin M	10770	0.4015	No
8007100	IGFBP4	insulin-like growth factor binding protein 4	10773	0.4058	No
8145244	TNFRSF10C	TNF receptor superfamily, member 10c	10980	0.4028	No
8097553	IL15	interleukin 15	11456	0.3899	No
8014369	CCL3	chemokine (C-C motif) ligand 3	11643	0.3869	No
7983630	FGF7	fibroblast growth factor 7	12252	0.3685	No
8107970	IL13	interleukin 13	15173	0.2672	No
8006453	CCL8	chemokine (C-C motif) ligand 8	15918	0.2423	No
8140556	HGF	hepatocyte growth factor	18146	0.1682	No
8132694	IGFBP1	insulin-like growth factor binding protein 1	19023	0.1421	No
7973084	ANG	angiogenin	20531	0.0959	No
8006459	CCL13	chemokine (C-C motif) ligand 13	20739	0.0952	No
7897877	TNFRSF1B	TNF receptor superfamily, member 1B	22164	0.0544	No
7980233	PGF	placental growth factor	23313	0.0254	No
8151447	IL7	interleukin 7	23643	0.0255	No
8029006	AXL	AXL receptor tyrosine kinase	26729	-0.0594	No
8037374	PLAUR	plasminogen activator, urokinase receptor	26976	-0.0441	No
8048205	IGFBP2	insulin-like growth factor binding protein 2	27094	-0.0235	No
8140358	CCL26	chemokine (C-C motif) ligand 26	27619	-0.0123	No
8071737	MIF	macrophage migration inhibitory factor	28619	0.0073	No

Supplementary Table 2. Differential infiltration of immune cells into mouse tumors. Tumors from cross-transplantation experiments (3-4/genotype) (tumor genotype/mouse genotype: WT/WT, *II-6^{-/-}*, WT/*II-6^{-/-}* and *II-6^{-/-}*/WT) were excised and tumors digested and immune cell subsets residing in spine analyzed using FACS analysis at days 14 and day 27. Mean and SEM are shown. Significance was determined by 2-tailed student's *t*-test.

				P-value
	WT/WT	II-6 ^{-/-} /II-6 ^{-/-}	II-6 ^{-/} /WT	WT/WT compared to II-6 ^{-/-} /WT
CD4	0.09 ± 0.01	0.19 ± 0.03	0.75 ± 0.07	0.01
CD8	0.06 ± 0.02	0.14 ± 0.04	0.41 ± 0.07	0.04
NK	24.9 ± 1.02	8.74 ± 1.31	18.5 ± 1.0	0.02
NKT	0.04 ± 0.05	0.09 ± 0.01	0.18 ± 0.04	0.04
B-cell	2.65 ± 1.79	2.06 ± 0.22	2.04 ± 0.39	NS
Macrophage	4.55 ± 2.51	5.70 ± 0.33	7.27 ± 2.13	NS
Dendritic cell	5.10 ± 2.90	5.80 ± 0.29	7.70 ± 2.41	NS
Neutrophil	1.28 ± 0.50	2.39 ± 0.30	4.60 ± 0.20	0.002

Supplementary Table 3. Radiation induced changes in immune cells numbers in

spines of C57/BI6 mice. Mice were injected with saline or 4 μ Ci/g of ⁴⁵Ca and culled 2 weeks post-injection, spines were digested into single cell suspension and FACs analysis carried out to determine immune cell numbers (7 mice/cohort). Mean and SEM are shown. *P*-values were determined by unpaired 2-tailed student's *t*-test.

	Saline	⁴⁵ Ca	Fold change	P-value
CD4	25.6 ± 4.80	16.2 ± 3.00	0.6	NS
CD8	18.5 ± 6.60	10.9 ± 3.60	0.6	NS
NK	3.00 ± 0.30	6.50 ± 1.80	2.2	0.04
NKT	0.03 ± 0.01	0.23 ± 0.15	8.6	0.04
B cell	0.26 ± 0.01	0.29 ± 0.09	1.1	NS
Macrophage	11.5 ± 3.00	15.4 ± 5.70	1.3	NS
Dendritic cell	0.70 ± 0.10	0.69 ± 0.06	1.0	NS
Neutrophil	5.25 ± 0.62	7.35 ± 0.55	1.4	0.04

Supplementary Table 4. Osteoblast specific knockout of RB1 results in differential immune cell infiltration following exposure to ⁴⁵Ca. $Rb1^{+/+}$ and $Rb1^{fl/fl}$ (6 mice/cohort) were injected with 4 μ Ci/g ⁴⁵Ca and mice at 2 weeks post-injection. FACs analysis of immune cell subsets in spines was carried out. This experiment was repeated independently three times. The mean ± SEM are shown. *P*-values were determined by unpaired 2-tailed student's *t*-test.

	Rb1 ^{+/+}	Rb1 ^{fl/fl}	Fold change	P-value
CD4	0.01 ± 0.02	0.01 ± 0.00	1	NS
CD8	0.15 ± 0.27	0.09 ± 0.01	0.6	NS
NK	0.82 ± 0.21	0.48 ± 0.05	0.58	NS
NKT	0.09 ± 0.02	0.05 ± 0.01	0.55	0.04*
B-cell	0.71 ± 0.06	0.73 ± 0.07	1.03	NS
Macrophage	0.36 ± 0.04	0.35 ± 0.03	0.97	NS
Dendritic cell	1.08 ± 0.07	1.07 ± 0.13	0.98	NS
Neutrophil	1.71 ± 0.23	1.15 ± 0.15	0.67	NS

Supplementary Table 5. Differential immune cell infiltration in spines of *II-6^{-/-}* mice compared to *wildtype* mice following exposure to radiation. Cohorts of mice C57/BI6 *II-* $6^{-/-}$ or WT (n=6 cohort) were injected with ⁴⁵Ca at 28 days of age. At 14 days post IR spines were excised, digested to a single cell suspension and analyzed by FACS to determine numbers of resident immune cells. *NKT cells assayed in this experiment by NK1.1 and TCR β . Mean and SEM are shown. Significance was determined by 2-tailed student's *t*-test. Cell numbers were normalized to differences in baseline differences between immune cell subsets in control compared to *II-6^{-/-}* spleen.

	WT	II-6 ^{-/-}	Fold change	P-value	
CD4	0.01 ± 0.00	0.01 ± 0.01	1	NS	
CD8	0.65 ± 0.10	0.89 ± 0.20	1.38	NS	
NK	0.80 ± 0.09	0.60 ± 0.20	0.75	NS	
NKT*	0.24 ± 0.01	0.13 ± 0.02	0.53	<0.01	
B cell	2.27 ± 0.58	1.68 ± 0.60	0.75	0.03	
Macrophage	3.60 ± 0.54	2.40 ± 0.30	0.66	NS	
Dendritic cell	0.63 ± 0.20	0.59 ± 0.30	0.93	NS	
Neutrophil	0.17 ± 0.06	0.09 ± 0.01	0.52	NS	