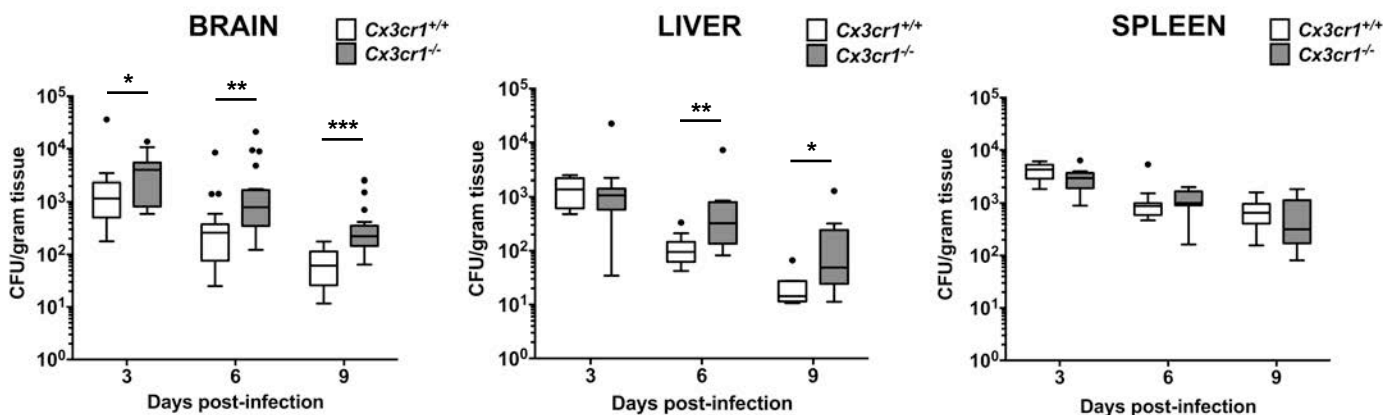


# Figure S1

## A

### Tissue Fungal Burden

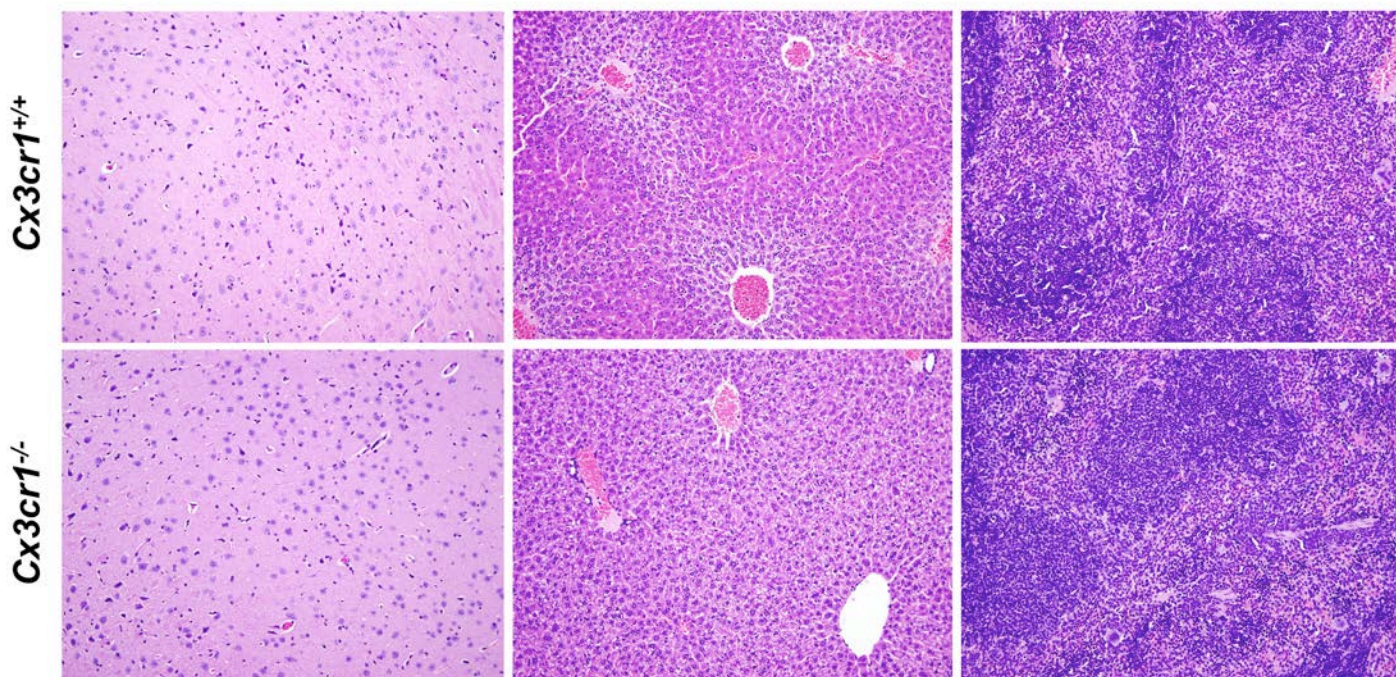


## B

### BRAIN

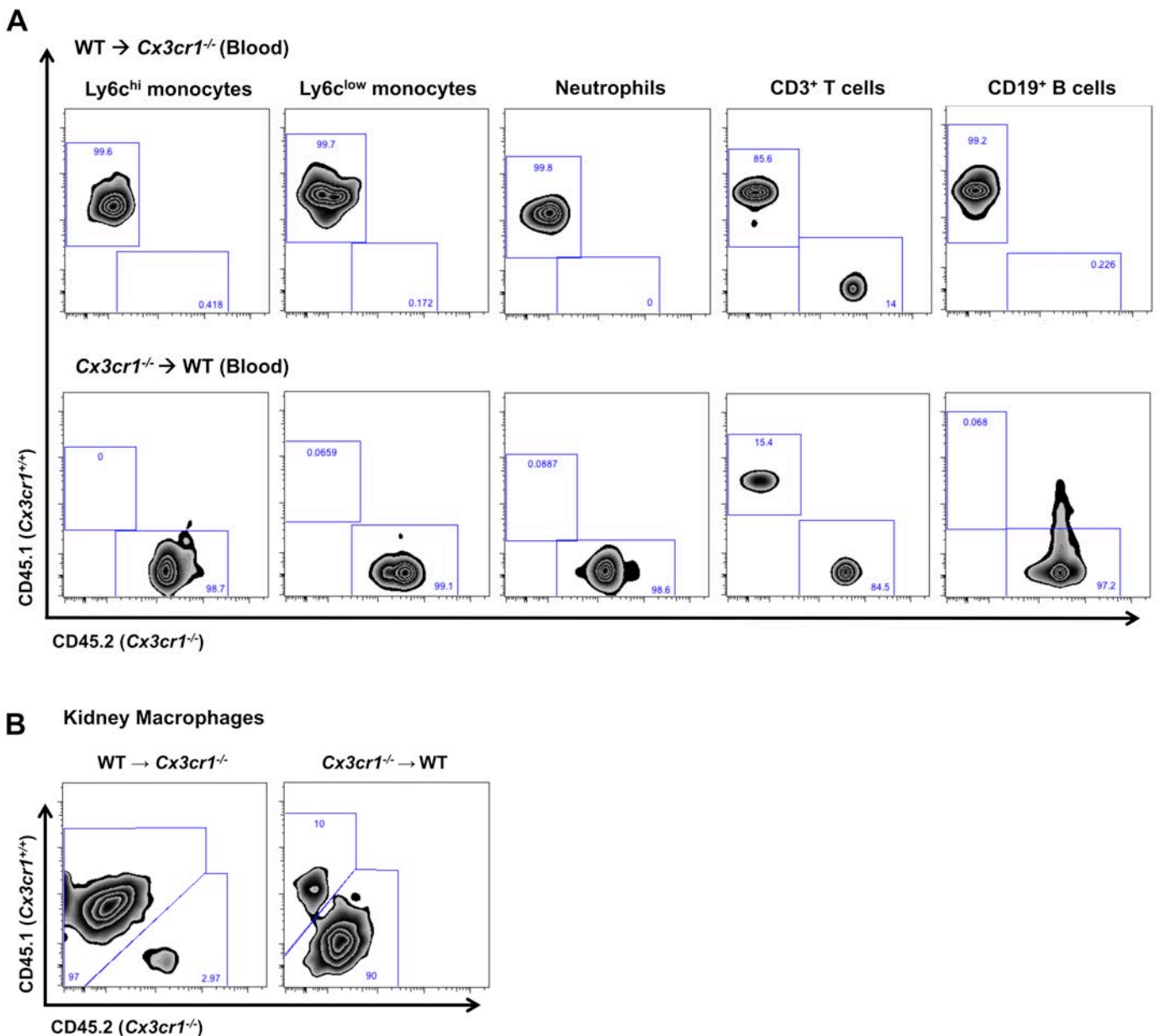
### LIVER

### SPLEEN



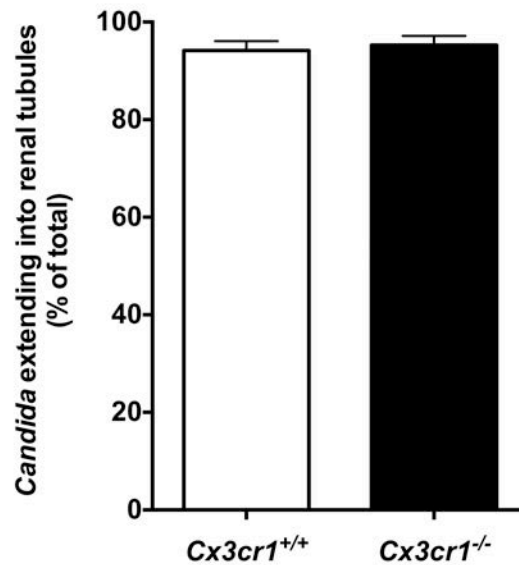
**Figure S1. Tissue fungal burden and histology in brain, liver and spleen of *Candida*-infected WT and *Cx3cr1*<sup>-/-</sup> mice. (A) Fungal burden. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.0001 (*n* = 10-21; two to three independent experiments). (B) Hematoxylin and Eosin staining (day 6 post-infection). Magnification, 200× (*n* = 8; two independent experiments).**

## Figure S2



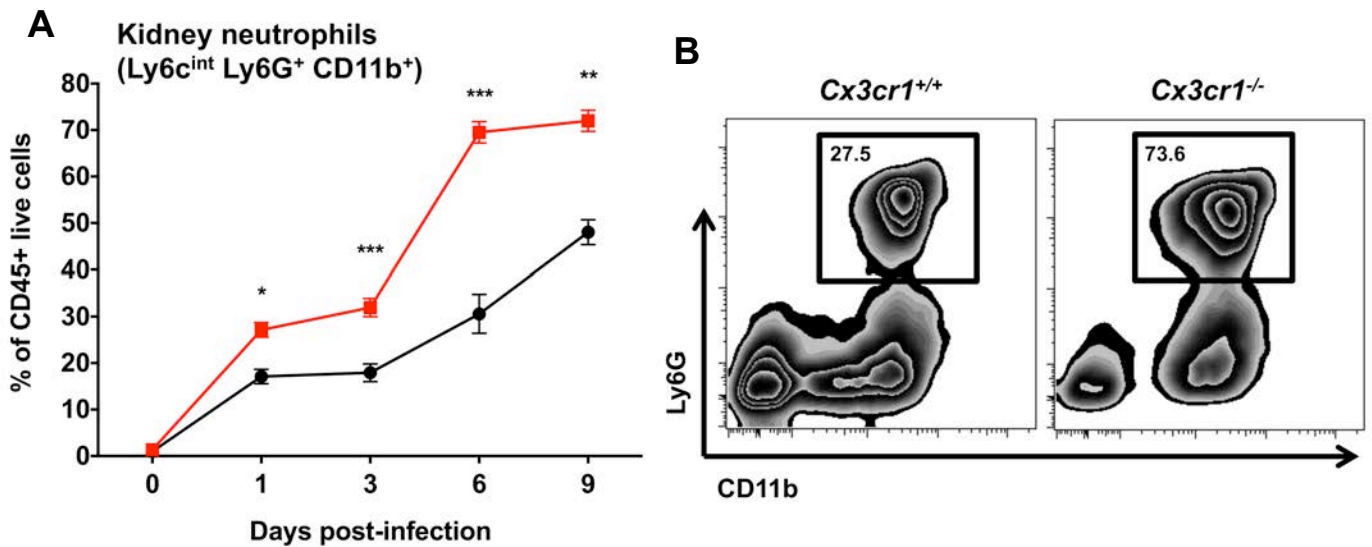
**Figure S2. Engraftment of bone-marrow radiation chimeras before *Candida* infection.** Whole bone marrow cells from WT (CD45.1<sup>+</sup>) and *Cx3cr1*<sup>-/-</sup> (CD45.2<sup>+</sup>) congenic mice were cross-transferred to irradiated recipients and evaluated for engraftment 8 weeks later. (A) Blood. Each column of FACS plots corresponds to the subpopulation labeled at the top, analyzed for CD45.1 and CD45.2 expression. WT donor→*Cx3cr1*<sup>-/-</sup> recipient mice (upper panel); *Cx3cr1*<sup>-/-</sup> donor→WT recipient mice (lower panel). Data are representative of 25 mice per group. (B) Kidney macrophages. FACS plots of kidney leukocytes purified from the indicated chimeric mice and gated on MHCII<sup>hi</sup>F4/80<sup>hi</sup>CD11b<sup>+</sup> cells. Data are representative of 4 mice.

**Figure S3**



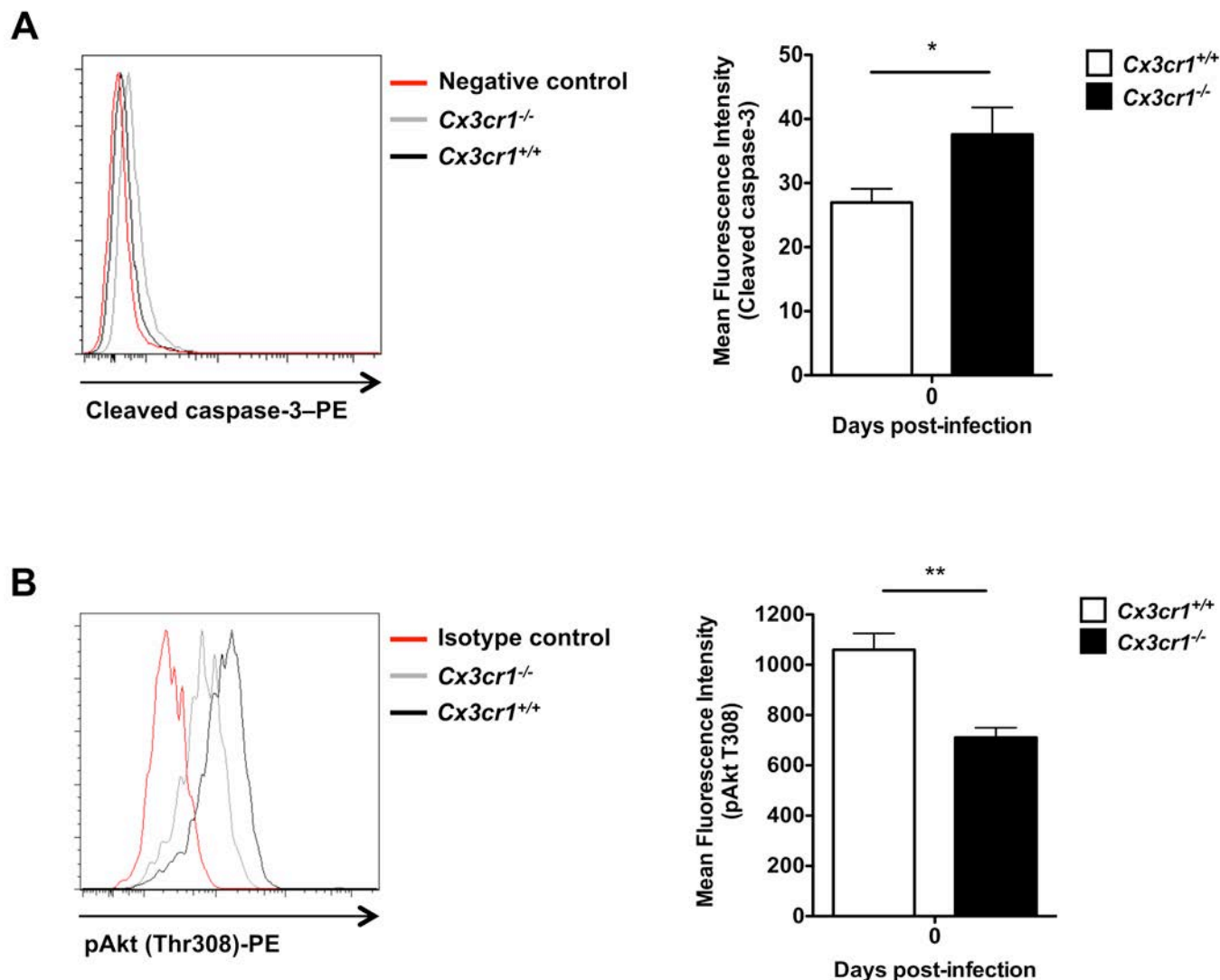
**Figure S3. *Candida* pseudohyphae rapidly extend into the renal collecting system by 24 hours after infection.** Percent of *Candida* pseudohyphal structures that have extended into renal tubules at day 1 post-infection in Cx3cr1<sup>+/+</sup> and Cx3cr1<sup>-/-</sup> mice ( $n = 11-13$ ; two independent experiments;  $P = 0.68$ ).

**Figure S4**



**Figure S4. *Cx3cr1* deficiency results in increased accumulation of neutrophils in the kidney after *Candida* infection.** (A) Percent of total CD45<sup>+</sup> leukocytes represented by Ly6c<sup>int</sup>Ly6G<sup>+</sup>CD11b<sup>+</sup> neutrophils that accumulated in *Cx3cr1*<sup>+/+</sup> and *Cx3cr1*<sup>-/-</sup> kidneys at different time-points after infection. \**P* < 0.01; \*\**P* < 0.001; \*\*\**P* < 0.0001 (*n* = 9-15; three to four independent experiments). (B) Representative FACS plots demonstrating increased frequency of Ly6c<sup>int</sup>Ly6G<sup>+</sup>CD11b<sup>+</sup> neutrophils in *Cx3cr1*<sup>-/-</sup> kidneys at day 6 post-infection (*n* = 9-15; three to four independent experiments).

Figure S5



**Figure S5. *Cx3cr1* deficiency is associated with increased caspase-3 cleavage and decreased Akt phosphorylation in kidney resident macrophages.** (A) Decreased expression of cleaved caspase-3 in *Cx3cr1*<sup>+/+</sup> kidney resident macrophages from uninfected mice. Left, representative FACS histogram. Right, summary mean fluorescence intensity data obtained by FACS. \**P*<0.05 (*n* = 9; three independent experiments). (B) Increased expression of pAKT (Thr308) in *Cx3cr1*<sup>+/+</sup> kidney resident macrophages from uninfected mice. Left, representative FACS histogram. Right, mean fluorescence intensity data obtained by FACS. \*\**P*<0.01 (*n* = 6). Data are shown from one of two independent experiments with similar pattern of results. All quantitative data are mean ± SEM.

**Table S1. Relative abundance of leukocyte subsets and their Cx3cr1 expression in the kidney at steady state and after *Candida* infection.**

Leukocyte subset*	% of total CD45 <sup>+</sup> leukocytes**					% of total Cx3cr1 <sup>+</sup> leukocytes***		
	Day post-infection							
	0	1	3	6	9	0	3	6
Neutrophils	0.5-1	12-21	14-28	17-61	25-60	N/A	N/A	N/A
Monocytes	3-5	35-39	12-39	4-11	4-10	6-15	21-30	11-17
Macrophages	53-67	16-21	25-47	26-47	15-33	77-87	55-65	62-69
Dendritic cells	9-12	7-14	4-9	3-11	2-5	3-7	9-13	10-14

N/A, not applicable

\* Neutrophils were defined as Ly6C<sup>int</sup>Ly6G<sup>+</sup>CD11b<sup>+</sup> cells, monocytes were defined as MHCII<sup>-</sup>F4/80<sup>int</sup>CD11c<sup>-</sup>CD11b<sup>+</sup> cells, macrophages were defined as MHCII<sup>hi</sup>F4/80<sup>hi</sup>CD11c<sup>low</sup>CD11b<sup>+</sup> cells, and dendritic cells were defined as MHCII<sup>hi</sup>CD11c<sup>hi</sup>F4/80<sup>low</sup> cells.

\*\* Range based on 9-15 animals tested (three to four independent experiments).

\*\*\* Range based on 4-6 animals tested (two independent experiments).

**Table S2. Number of candidemic subjects and non-infected controls from the two patient cohorts analyzed in the present study.**

	<b>Candidemia Cases n = 281</b>	<b>Non-infected Controls n = 384</b>	<b>Total</b>
<b>RUNMC</b>			
<b>Caucasian</b>	37	167	204
<b>DUMC</b>			
<b>Caucasian</b>	166	156	322
<b>African American</b>	78	61	139
			665

RUNMC, Radboud University Nijmegen Medical Center; DUMC, Duke University Hospital

**Table S3. Demographic and clinical characteristics of DUMC subjects**

	No Systemic Candidiasis n = 217	Systemic Candidiasis n = 244	p value
Male gender	112 (51.6%)	136 (55.7%)	0.376
Mean Age (years) adult (n = 443) pediatric (n = 18)	58.7 (56.4-60.9) --	52.5 (54.0-58.3) 6.7 (3.6-9.9)	0.11
Solid organ transplant	6 (2.8%)	35 (14.3%)	<0.0001
Malignancy	49 (22.6%)	69 (28.3%)	0.162
Chemotherapy	26 (12.0%)	37 (15.2%)	0.321
Neutropenia	6 (2.8%)	23 (9.4%)	0.0033
Surgery within last 30 days	121 (55.8%)	111 (45.7%)	0.0310
Receipt of total parenteral nutrition	7 (3.2%)	50 (21.1%)	<0.0001
HIV	0	3 (1.2%)	0.251*
Dialysis-dependent	16 (7.4%)	29 (12.1%)	0.089
Acute renal failure	49 (22.6%)	84 (34.6%)	0.0047
Immunocompromised state	95 (43.8%)	140 (57.6%)	0.0031
ICU admission within past 14 days	67 (30.9%)	118 (49.6%)	<0.0001
Liver disease	9 (4.3%)	58 (23.8%)	<0.0001
<i>Candida</i> spp.	N/A		
<i>albicans</i>		109 (44.7%)	
<i>glabrata</i>		64 (26.2%)	
<i>parapsilosis</i>		40 (16.4%)	
<i>tropicalis</i>		27 (11.1%)	
<i>krusei</i>		9 (3.7%)	
other		12 (4.9%)	
>1 <i>Candida</i> spp.		14 (5.8%)	

\*Fishers Exact p value

DUMC, Duke University Hospital; ICU, intensive care unit



**Table S4. Association of the mutant CX<sub>3</sub>CR1-M280 allele with susceptibility to systemic candidiasis in African-American subjects**

	<b>No Systemic Candidiasis n = 61</b>	<b>Systemic Candidiasis n = 78</b>	<b>p value</b>	<b>OR (95% CI)</b>
<i>CC</i>	47(77.1%)	65 (83.3%)	0.354	0.671 (0.289-1.560)
<i>CT + TT</i>	14 (23.9%)	13 (16.7%)		

**Table S5. Sequences of the primers used for qPCR with SYBR Green in the present study.**

<b>Gene product</b>	<b>Primer name<sup>*</sup></b>	<b>Primer sequence (5' → 3')</b>	<b>Amplicon length (bp)</b>
Gapdh	Gapdh F	aactttggcattgtggaagg	223
	Gapdh R	acacattgggggtaggaaca	
Cxcr2	Cxcr2 F	gggtggggagttcgtgtagaa	201
	Cxcr2 R	cgaggtgctaggattgagc	
Cxcl1	Cxcl1 F	gctgggattcacctcaagaa	180
	Cxcl1 R	tctccgttacttggggacac	
Cxcl2	Cxcl2 F	aagtttgccttgaccctgaa	180
	Cxcl2 R	aggcacatcaggtagatcc	
Il-4	Il-4 F	cctcacagcaacgaagaaca	155
	Il-4 R	atcgaaaagcccgaagagt	

\*F, forward; R, reverse.

Taqman primers and probes for Gapdh, Il-1 $\beta$ , Il-6, Ccr2, Cx3cr1 and Cx3cl1 were purchased from Applied Biosystems and their sequences are not available.