

# Supplementary Material

to

## A Novel C5a-neutralizing Mirror-image (L-)Aptamer Prevents Organ Failure and Improves Survival in Experimental Sepsis

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Short title: C5a-neutralizing L-aptamer improves sepsis

**Figure S1.** Enrichment of bio-D-mC5a binding aptamers from a random RNA library.

**Figure S2.** C5a sequence alignment.

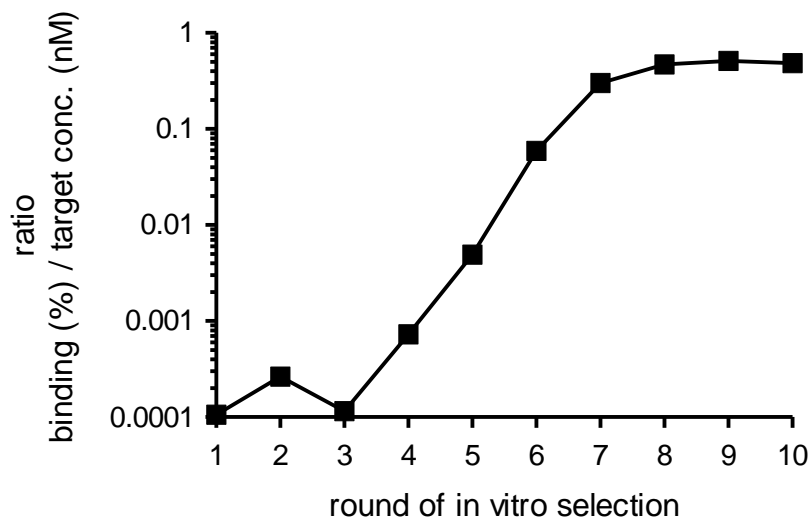
**Figure S3.** Reverse control Spiegelmers do not inhibit huC5a-induced chemotaxis.

**Figure S4.** NOX-D20 binds to mouse C5a.

**Figure S5.** NOX-D19 and NOX-D20 inhibit mouse C5a-induced chemotaxis.

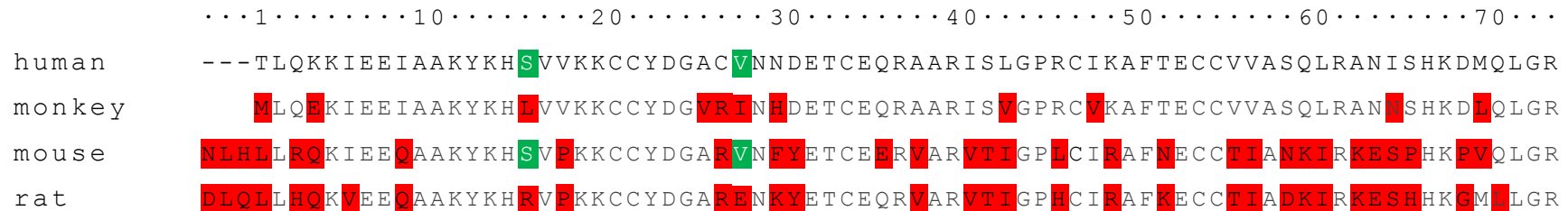
**Table S1.** Sequences and frequencies of RNA aptamers identified after 10 rounds of in vitro selection.

**Table S2.** Truncation of NOX-D19001-6xDNA.



**Figure S1. Enrichment of bio-D-mC5a binding aptamers from a random RNA library.** The ratio of the percentage of target-bound aptamers and the concentration of target applied to the respective selection round was used as a measure for the frequency and affinity of target-binding aptamers in the library.

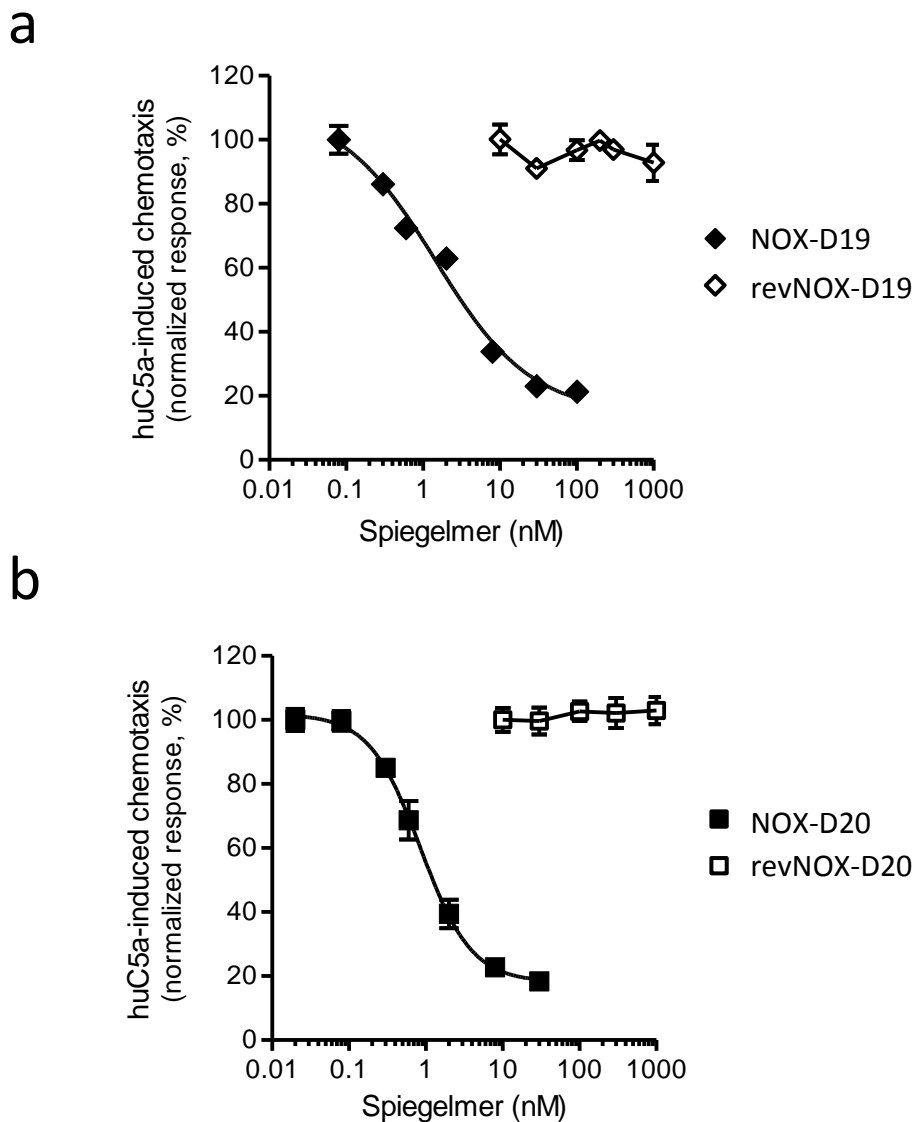
*Hoehlig K et al., A Novel C5a-neutralizing Mirror-image (L-)Aptamer Prevents Organ Failure and Improves Survival in Experimental Sepsis*



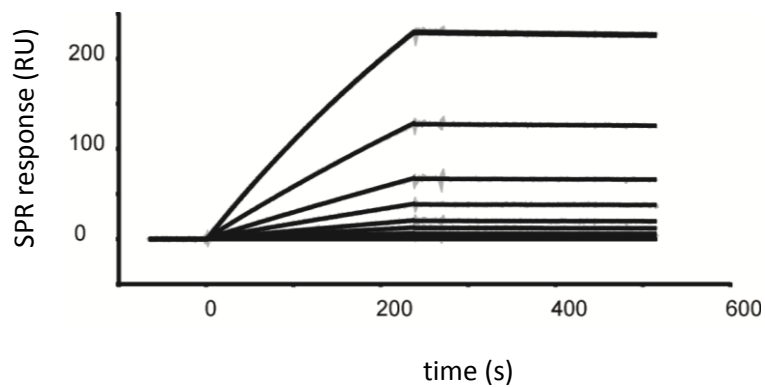
■ residues different from human C5a

■ residues shared by human and mouse C5a but different in monkey and rat C5a

**Figure S2. C5a sequence alignment.** Alignment of C5a protein sequences from human (UniProt ID: P01031), rhesus/cynomolgus monkey, mouse (UniProt ID: P06684), and rat (UniProt ID: P08650). Protein sequence of C5a from rhesus monkey was predicted from the genomic sequences (NCBI reference sequence: XP\_001095750.2). Identity of cynomolgus monkey C5a was confirmed by RT-PCR and sequencing.

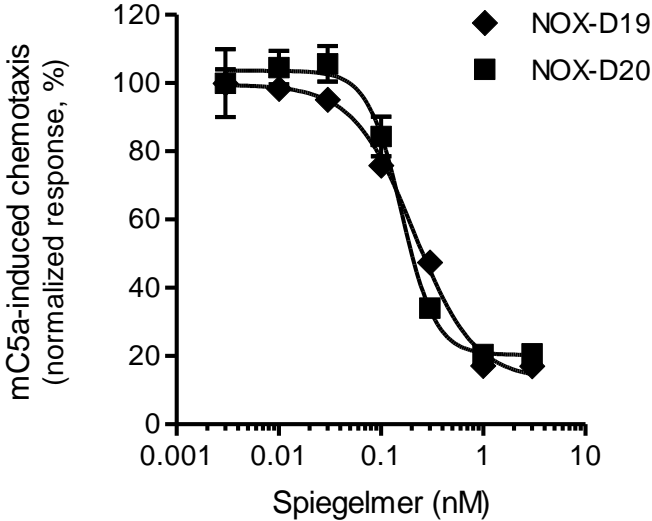


**Figure S3. Reverse control Spiegelmers do not inhibit huC5a-induced chemotaxis.** The oligonucleotide part of the PEGylated control Spiegelmers revNOX-D19 and revNOX-D20 is of the reverse sequence of NOX-D19 and NOX-D20, respectively. Chemotaxis of CD88<sup>+</sup> BA/F3 cell was stimulated with 0.1 nM human C5a pre-incubated with (a) NOX-D19 (closed diamonds), revNOX-D19 (open diamonds) and (b) NOX-D20 (closed squares), revNOX-D20 (open squares) at indicated concentrations. Mean  $\pm$  SD of triplicate measurement are shown. Data is representative for three independent experiments.



$k_a$ ( $10^6 M^{-1} s^{-1}$ )	$k_d$ ( $10^{-5} s^{-1}$ )	$K_d (= k_d/k_a)$
$2.44 \pm 0.001$	$4.65 \pm 0.09$	19 pM

**Figure S4. NOX-D20 binds to mouse C5a.** SPR measurement of NOX-D20 binding to mouse C5a. Kinetic rate constants are shown as mean  $\pm$  SEM. Data is representative for at least 3 individual measurements.



**Figure S5. NOX-D19 and NOX-D20 inhibit mouse C5a-induced chemotaxis.** Chemotaxis of CD88<sup>+</sup> BA/F3 cell was stimulated with 0.3 nM mouse C5a pre-incubated with NOX-D19 (closed diamonds) and NOX-D20 (black squares) at indicated concentrations. Stimulation with 0.3 nM mouse C5a stoichiometrically limits the sensitivity of the assay to IC<sub>50</sub> = 0.15 nM. Mean ± SD of triplicate measurement are shown. Data is representative for five independent experiments.

**Table S1.** Sequences and frequencies of RNA aptamers identified after 10 rounds of in vitro selection.

name	sequence <sup>a</sup>										frequency																																																							
	1	10	20	30	40	50	60	70	80																																																									
274-D5	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	G	A	U	G	U	G	G	G	U	U	G	A	G	G	G	G	U	U	G	U	G	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	16.7 %	
274-H6	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	G	A	U	G	U	G	G	G	G	U	U	G	A	G	G	G	G	U	U	G	U	G	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	16.7 %
274-C8	g	g	a	g	c	u	c	a	g	a	c	c	U	A	U	G	C	C	U	G	A	U	G	U	G	U	G	A	A	G	G	U	U	U	G	G	G	U	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %
274-C5	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	G	A	U	G	U	G	G	U	G	A	G	G	G	U	U	G	U	G	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %				
274-B5	g	g	a	g	c	u	c	a	g	a	c	c	U	A	U	G	C	C	U	G	A	U	G	U	G	U	G	A	A	G	G	U	U	U	G	U	G	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %				
274-F5	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	G	A	U	G	U	G	A	U	G	U	A	G	G	G	U	U	G	U	G	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %			
274-A5	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	G	A	U	G	U	G	U	A	G	G	G	G	U	U	G	U	G	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %					
274-B6	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	G	A	U	G	U	G	A	U	G	U	A	G	G	G	U	U	U	G	U	G	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %		
274-G8	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	G	A	U	G	U	G	G	U	A	G	G	G	G	U	U	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %				
274-G7	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	U	G	U	G	G	U	G	A	G	G	G	G	U	U	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %					
274-G6	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	G	A	U	G	U	G	G	A	U	G	G	G	G	U	U	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %				
274-H7	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	A	U	G	U	G	U	G	U	A	G	G	G	G	U	U	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %				
274-F6	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	A	U	G	U	G	U	A	G	G	G	G	U	U	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %						
274-H5	g	g	a	g	c	u	c	a	g	a	c	c	U	G	U	G	C	C	U	G	A	U	G	U	G	U	A	G	G	G	G	U	U	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %					
274-F8	g	g	a	g	c	u	c	a	g	a	c	c	C	G	U	G	C	C	U	G	A	U	G	U	G	U	A	A	G	G	G	A	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %						
274-A6	g	g	a	g	c	u	c	a	g	a	c	c	U	A	U	G	C	C	U	G	A	U	G	U	G	G	U	A	G	G	G	G	U	U	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %				
274-C6	g	g	a	g	c	u	c	a	g	a	c	c	U	U	U	G	C	C	U	G	A	U	G	U	G	U	A	G	G	G	G	U	U	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %					
274-D7	g	g	a	g	c	u	c	a	g	a	c	c	U	A	U	G	C	C	U	A	U	G	U	G	U	A	G	G	G	G	U	U	U	G	U	C	G	A	C	A	A	g	c	u	g	c	a	g	u	g	u	c	g	g	u	c	c	a	g	4.2 %						

<sup>a</sup> Fixed primer-binding sites are in lower case. Point mutations are highlighted.

**Table S2.** Truncation of NOX-D19001-6xDNA.

name	sequence	length	affinity ( $K_d$ ) <sup>a</sup>
NOX-D19001-6xDNA	<b>GCCUG</b> AUG (dU) GGUGGU (dG) (dA) AGGGUUGUUGGG (dU) G (dU) CGACGCA <b>(dC) AGGC</b>	44 nt	0.4 nM
NOX-D19001-6xDNA-011	<b>GC-UG</b> AUG (dU) GGUGGU (dG) (dA) AGGGUUGUUGGG (dU) G (dU) CGACGCA <b>(dC) A-GC</b>	42 nt	0.3 nM
NOX-D19001-6xDNA-018	<b>GCC-G</b> AUG (dU) GGUGGU (dG) (dA) AGGGUUGUUGGG (dU) G (dU) CGACGCA <b>(dC) -GGC</b>	42 nt	0.3 nM
NOX-D19001-6xDNA-012	<b>G--UG</b> AUG (dU) GGUGGU (dG) (dA) AGGGUUGUUGGG (dU) G (dU) CGACGCA <b>(dC) A--C</b>	40 nt	0.8 nM
NOX-D19001-6xDNA-020 = <b>NOX-D20001</b>	<b>G-C-G</b> AUG (dU) GGUGGU (dG) (dA) AGGGUUGUUGGG (dU) G (dU) CGACGCA <b>(dC) -G-C</b>	40 nt	0.4 nM
NOX-D19001-6xDNA-033	<b>G---G</b> AUG (dU) GGUGGU (dG) (dA) AGGGUUGUUGGG (dU) G (dU) CGACGCA <b>(dC) ---C</b>	38 nt	5.5 nM

<sup>a</sup> Equilibrium dissociation constants ( $K_d$ ) for Spiegelmers binding to huC5a measured by SPR.