

Figure S1 Methods used to identify conditional drug-sensitive suppressors of *nimA1*. Diagram of the genetic screen performed and the general steps involved to identify suppressors of *nimA1* temperature sensitivity.

			aa
А.	nidulans	MGRKPNQLILEFFIRGQKLEDASNRYQHTCKACGEKFPKGRIDSLTNHLVKKCTAIPLRDRQRVLLRLHELPDLADGDQNKDSNSAGQ	88
Ρ.	chrysogenum	MGRKPNQLILEFFHRGPKLEDASNRYQHTCKACGEVFPKGRIDSLTNHLVKKCQAIPLRDRQRVLLRLHELPDLTDGDSNKDSPSG	86
С.	immitis	MGRPNOLILEVERGPKLPDSSNTYEHTCKLGCENFFKGPDSLLGHLTKSCQAISLDKQRVIHLLHGLSSNTTGAGKAHGNSK5DRARANT	94
s.	sclerotiorum	MGRKPNPLILKYFERGAKLNDSSNRYHHTCRACSEKFPKGRLDTLTSHICKKCPALDRACKORLVLEFNNLPEIADRTNNQQMNIQTA	88
F.	oxysporum	MGRKPNPLILEYFVRGPKLNDNSNRYPHTCKQCGENFPKGRIDSLTTHITKKCPAISESDR#RACLELHGITNARAPVDRPPPEAQ	86
т.	reesei	MGRKPNPLILEYFVRGPKLNDNSNRYPHTCKQCGENFPKGRIDSLTTHITKKCPAISDSDRMRACLELHGITTARAAIDRPQGGAP	86
Ν.	crassa	MGRRPNALILQYFERGPKLQDQSNRYPHTCKSCGEHFPRGRLDSLTTHLTKKCPAIGEADRVNAIFTLSGMSHASHRLQQTQHAQGQVQQPLQVPTQVNG ****** !! !! ** ** * **** **** **** **	100
А.	nidulans	NKGKSGEVSFTTRQNFDGLNVLAEASRQVGASDQTKRGTPAFPQSVTVGGKTVVVDPALEAEGFQGHPT	157
Ρ.	chrysogenum	SKGKSADGGFPNRQNFDGLNVLAEASRQVGASDVTKRG-PGYTQSVTAGGKTVIVDPALEAEGFQGHSQ	154
С.	immitis mavdis	SPYSSRLANFSNPGGLOGLNVLAEASRRVGATNET	154
s.	sclerotiorum	INHOTAVRGRPF	170
F.	oxysporum	$PNG \Tilde{O}PVDVPNLP\Tilde{O}GWSALETLAEASR \tilde{O}VDLNE-NNRA \tilde{O}SVQAGVVDP \Tilde{O}PADPAN \tilde{O}TLDNPPVSYENRS \Tilde{O}QGNRGNGPSELDLQG$	180
Τ.	reesei	VRGOPTAQLALPOGWSALETLAEASROVDLNE-NSRGQKVSASDNTANFTANGGGFVPDRLELQEQFTLDNPPTTYDSP	164
N.	CIASSA	* *** ** : * *** ** :	102
А.	nidulans	OSDOMDEDVKIPVSPKSPTDATTIPSLP-PTGSLDPSSASPPLP-ETSLTPDPTATSROS-OLSMIAASASEMVPHGLPLDHDPSSGLSDG-	245
₽.	chrysogenum	Ğpdškaegtanaegtpqssnapsipalpshtpgehpasmspplg-dasmsppestsnarqs-qlsmiaasanemvpqgmsidsdsd	237
C.	immitis	VIDLNEHPTATSIQTSQSSFIQEEPHASACVSD-SMQLMPDSLSDLRHSSQLSLIAASASEIVPEDT	220
s.	sclerotiorum	PDG5DLAGL5HGRUPSNPTAPTSRIQTNNSLHHSPTTSPUNG-HMPL5-NSSSR5HMVPSLVRAASAANDLAFLTINGL5MD5DRALSGRG	240
F.	oxysporum	ALLTTNVLGAIOOPLPGTELSPEERLOALLPNSV-ASPDASNISVAVAATARLNPSLLDPOLVHETGTSTTPPSMDIPTPIEA	262
т.	reesei	$ {\tt FTATAAGKLP} \tilde{{\tt SPTP}} {\tt EL} \tilde{{\tt SPEERL}} \tilde{{\tt ALLPASE}} - {\tt ASPDTTNISVAVAATARLNPTFLDP} \tilde{{\tt C}} {\tt LVNTDIPPPTSSPPIVEAH} \tilde{{\tt QTTIDS}} $	245
Ν.	crassa	TVLGEQPKPEPVAPTDEERVLTAEERLHEILRVEENANADAANISMAAAAAARLHHGLLDPQLLGQEAAAAAEAASATAAAAAAVASSLSASEQA	277
А.	nidulans	LSKMSSAWNQQLSTQEQLLFDSLQEHDPTLTAATQRAASFPRPIAMNPNTOAKGFVNEFGNSTKPAKPKVRGRFSAARRREVOEVRKRG	334
Ρ.	chrysogenum	MKMN-QWNQQLSTHEQLLFDSLQEHDPSLTAATQRAASYPRPIAMNPNTQAKGFVNEFGNSTKPMKPKVRGRFTAERRREVQDVRKKG	324
с.	immitis	VEHLDSDQHITQFNHDAHDNAQCYVAYPRPIAINVNPQSHEPISVPKAISRITKHKSRSAFSEERRKEVQLVRKMG	296
в.	maydis	LHIMSDCAFFHAQCAGHMYNIQYNSLDFHLQDISAEHNQMNDNHAGNGUQXLAMAGNQYH-FTDFSMQAPAAPAVRGAFTDSKAEVQEVKAG DSGASAMQVDONAHMCONTYSFUNDSENTGYDCADDIOSPKSUBTSANDMTTFFSSPYCACGASHTKEVGEVENTGPBKFVSDFKTC	343
5. F.	oxvsporum	SPG-AAGA AVALAND	353
т.	reesei	VPVSLPTSLSDTSVSQPWGEMTYLTTTP-IPLLTENSPALPLMSRGGVRMDTSDGLINGRARHARSRFTPARRKEVQEVRKIG	328
Ν.	crassa	HQSHQSPPEVSPSIPATTPSQPWGEITYTADAFQTMPAGDSVPHTTASLTKGGFRLETTSKARHSRARFNATRRKQVQEVRKIG : *, *, **::*, :** *	361
А.	nidulans	ACIRCRMLKKPCSGDSPCTTCASVESARLWKHPCIRTRIADEFELYHANLHATLAFHDVSSIRNQVKFEIFAGRIEVTHFE-ESMVFVTLAGQQGHKPSA	433
Ρ.	chrysogenum	ACLRCRNLKKPCSGDTPCTTCASVESARLWKHPCLRTRLSDEFELYNANLNSTLAYHDTNSIKNQIKFEHYSGRIEVTHPE-ESNSFMTFSGLQGHRTSV	423
С.	mavdis	ACIDERALIARTOSPETCOSCENDERVWERDCOVERTENTREATOVALASVDIIDINNENQVEEDSTERTEVIEBUCSIEVIEBUCA	442
s.	sclerotiorum	SCIRCRNLKKFCTAGDPCDTCKGVESARLWKTPCIRTRVAHELCLYSAKLHTVLAHHQVLAAKGSANFONSPYQIEVSHFP-ETTIYASFPCLEAHDADT	452
F.	oxysporum	ACIRCRILRKNCGKGTPCDTCRKVLAPRVWRTGCVRTRLQEQLDLYSAGVQVVLSQNRINLLKNQLNMTHDGTMIEVSHPP-ETGKVILLEALVALLE	440
Τ.	reesei	ACIRCRILRKWCGKGTPCDTCRKVLAPRVWRTGCVRTRLHEQLDLYAAGVQVLSQWRINLLKDQLKLTNDGSLIEVSHPP-EAGKKIVFGALVY-PLD	425
N.	crassa	INTERCEPTION CONTRACTOR CONT	460
Α.	nidulans	STLDPQLQGLGDETQFQGPLQEIYLLDNDMDDVPGKIEMYIKKAGPFFYEREASDVMRPTLMLAAELAQQKKDLLLERVLELWVATHILVDADLHWKTFS	533
P.	chrysogenum	SALDPQLQALGDD-QFAGPSQEVYLLDADSDD1FSKLEMYIKNASYFYERTSPFMRPTLLLAAELNQGKKDILLERVLELWVATHILDDTELTWKTYY SNEDD01 pt troupter caupanity ti doever for the day of the second state of a sold to accur to wards ti voo to wards	496
в.	maydis	TOTTDIDPAIFSGLASPDLELISD-DDVSGKVDHYVKKMAGFADTEDSHFMKATVHSAKSIASLSPDGLISKSLELWALTRILTSRFEWHLFS	537
s.	sclerotiorum	$\label{eq:generalized_constraint} G_{LTSYILDTDHDDLSNKLEAYAKRMMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMPIFIDNEPSQFIRTTLQTALNLSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMPIFIDNEPSQFIRTTLQTALNSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMPIFIDNEPSQFIRTTLQTALNSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMPIFIDNEPSQFIRTTLQTALNSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMPIFIDNEPSQFIRTTLQTALNSIQKQDNFLSRALEFWTIVHIFVDSEIRWEMSAKRMPIFITANGKAKRAKRAKRATAKRATAKRAKRAKRAKRAKRAKRAKRAKRAKRAKRAKRAKRAKRA$	533
F.	oxysporum	PSESLAESRG-SKDSFFQVIMIDQDKEDVPGKVEAYMRDVFQLFIDREFSKPMKVTLSLALQQLQESEDDLLRKSLELWGLVESID-RERQWNVLE	534
T.	reesel	LSSSQAEIKEBURKFFRUVINI DUSSDIFARASSINKVVLTFFIERESKFRUVISAAGDDLLKASLELWGLVSDID-RERUWTILE	555
	crassa		
А.	nidulans	$\label{eq:linear} NPTLPPASYNSFAQPTDDGRIPIDEITNAESYALLCGQLRSATEKRAAALSKSVMNDLERRLLQRQQSGWFETFLVSLILLNCVERTCWLFRSWDDESALLCGQLRSATEKRAAALSKSVMNDLERRLLQRQQSGWFETFLVSLILLNCVERTCWLFRSWDDESALLCGQLRSATEKRAAALSKSVMNDLERRLLQRQQSGWFETFLVSLILLNCVERTCWLFRSWDDESALLCGQLRSATEKRAAALSKSVMNDLERRLLQRQQSGWFETFLVSLILLNCVERTCWLFRSWDDESALLCGQLRSATEKRAAALSKSVMNDLERRLLQRQQSGWFETFLVSLILLNCVERTCWLFRSWDDESALLGRGAAALSKSVMNDLERRLLQRQQSGWFETFLVSLILLNCVERTCWLFRSWDDESALLGRGAAALSKSVMNDLERRLLQRQQSGWFETFLVSLILLNCVERTCWLFRSWDDESALLGRGAAALSKSVMNDLERRLLQRQQSGWFETFLVSLILLNCVERTCWLFRSWDDESALLGRGAAALSKSVMNDLERRLLQRQQSGWFETFLVSLILLNCVERTCWLFRSWDDESALLGRGAAAASAAASAAAAAAAAAAAAAAAAAAAAAAAA$	630
Ρ.	chrysogenum	NPTLPPNSLHSLSQPSDEGRLP1EEVSDPESYGLLCSQLRAAMEKRASQLSKFVINDLERRLLQRQKCGWPDTfVAIILLLCCDRTCWLFRSWDRE	619
в.	mavdis	NFILFFALQFLISSSNGMIFFIGODGLESISLICSQUARABARABISAADINALBURUKUKUKUKUKUKUKUKUKUKUKUKUKUKUKUKUKUKU	593
s.	sclerotiorum	KAISGSSDEGGGQTKPILSTERSFAIIHTQLNSAAENKASQMCKGILNELERRLLNKÄÄŠKSSFELFLVAVITLNCVEKSTWLFKCWEGE	622
F.	oxysporum	$\label{eq:real} RPANKNEEPRRIQEAKSENDADIYTTLCMQLNAAAERKANNTSKALLSGMHRVLQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCLEKSTWAFKAWEQDSKVKVNFKMYLTAIIFLNCHAKAKAKAKAKAKAKAKAKAKAKAKAKAKAKAKAKAKAK$	623
T.	reesei	RPSAEGEEPKWIKEAQMENDADIYTMICMQLMAAASKKANSTSKSLLNHMDRLLTDSKTKVGPKITLATIFLDCVEKSTWAFKAWSQD	609
14.	ciassa		641
А.	nidulans	NFSQRWPLDKRPQYYAAQGDRFSDILHMLLKMRSLPPKANPLPDSGILKAV-EGSDENAIRWFDMIKITPLYLEQRQA-AGFDPTDSRSFDLRYGAKLLP	728
₽.	chrysogenum	NFSQRWPLDKRPPYYFNQSDRFADILEMLLRMRHIPPKATFRPDNGILKAV-DGSDEHAIRWFDMIQITPYFIEERKN-AVFDPSDSRSLDCHHSASLLE	717
С.	1mm1t1s mavdis	ELAINTELERREDET VAUGURF DELLATIELERREDET AUGUSTUTELERREDEGIGEVIRWERAIN DHUPLESKUT-VGEDESNMRSEDELKYCSTELL STTPREMPIERSPECTREDET ANGELKEREUPPEVISTENTELERLIGENERGET. UNDER DELKAVERWISCH ZUTTELEREN LED AUGUNDEVENTET ET EL	691 736
s.	sclerotiorum	DFHGQWPLDKPPONFANGGDDLINVLQMILRWREVPPKTFVRQDNGVLAVD-DTSEQVVREFFEQVQLSFTEVHDKOVRYFFDPADSRSYELRYISRLL	721
F.	oxysporum	HIRPGWPLERDPGVFTQQGGNLAGLLKMLLAIRKALPQTLRSEA-GKLITSEQDPVIGTYFQSIDLDYDTILARQDGSPFSPADSRSLEMILCSHLLI	720
T.	reesei	HLRPGWPLERDPSVFTQQGGNLAGLLKMLLSTRKALPQTMRGED-GKLTVQDKOPETTEYFRTLDLDVDTIEARQKSSQFSPADSRSLELAFCSHLL	706
Ν.	crassa	*** I * * * I II I I I I I I I I I I I	723
А	nidulans	PANUVA 734	
P .	chrysogenum	INNAT 722	
c.	immitis	SADNLG 697	
в.	maydis	GD 738	
5. F.	oxysporum	F_VOR / 20 TMTP_ 724	
T.	reesei	PNSP 710	
N.	crassa	LG 725	

Figure S2 ClustalW alignment (<u>http://workbench.sdsc.edu</u>) of SonC and its orthologs. Identical (*), conserved strong groups (:), and conserved weak groups (.) are indicated. Accession numbers for the sequences are: *A. nidulans* (AN1232; *Aspergillus* Genome Database, <u>http://www.aspgd.org/07-16-13</u>), *Penicillium chrysogenum* (Pc16g03990), *Coccidioides immitis* (CIMG_10322), *Bipolaris maydis* (COCC4DRAFT_161203, Δ641-688), *Sclerotinia sclerotiorum* (SS1G_09964), *Fusarium oxysporum* (FOXB_15136, Δ1-67), *Trichoderma reesei* (hypothetical protein, locus EGR51802), *Neurospora crassa* (NCU07692). Sequences were modified as noted in Fig. 5C before performing the alignment.

Time-lapse imaging of Mag1-GFP

Video shows time-lapse imaging of Mag1-GFP during mitosis (Fig. 2B). Images were collected every 1 min and are

displayed at 2 frames per second (fps). File S1 is available for download as an AVI at

Time-lapse imaging of SonC-GFP

Video shows time-lapse imaging of SonC-GFP during mitosis (Fig. 6C). Images were collected every 1 min and are

displayed at 2 fps. File S2 is available for download as an AVI at

3D rotation of SonC-GFP during mitosis

Video shows a 3D rotation over time of SonC-GFP during mitosis (Fig. 6C). Images were collected every 1 min, were

rotated in 10° increments, and are displayed at 25 fps. File S3 is available for download as an AVI file at

Time-lapse imaging of SonC-GFP during SAC arrest

Video shows time-lapse imaging of SonC-GFP in cells treated with 2.4 μ g/ml benomyl during SAC arrest (Fig. 6D). Images were collected every 1 min and are displayed at 2 fps. File S4 is available for download as an AVI at

3D rotation of SonC-GFP during SAC arrest

Video shows a 3D rotation over time of SonC-GFP during SAC arrest in cells treated with 2.4 µg/ml benomyl (Fig. 6D). Images were collected every 1 min, were rotated in 10^o increments, and are displayed at 25 fps. File S5 is available for download as an AVI at http://www.genetics.org/lookup/suppl/doi:10.1534/genetics.113.156745/-/DC1.

Time-lapse imaging of SonC-GFP and histone H1-mCherry during mitosis

Video shows time-lapse imaging of SonC-GFP together with histone H1-mCherry during mitosis (Fig. 7A). Images were

collected every 1 min and are displayed at 2 fps. File S6 is available for download as an AVI at

3D rotation of SonC-GFP and histone H1-mCherry during mitosis

Video shows a 3D rotation over time of SonC-GFP and histone H1–mCherry during mitosis (Fig. 7A). Images were collected every 1 min, were rotated in 10^o increments, and are displayed at 25 fps. File S7 is available for download as an AVI at http://www.genetics.org/lookup/suppl/doi:10.1534/genetics.113.156745/-/DC1.

SonC-GFP and histone H1-mCherry in a hooked projection

Video shows a 3D rotation over time of SonC-GFP together with histone H1-mCherry in nuclei that display a hooked

structure (Fig. 7B). Images were collected every 1 min, were rotated in 10⁰ increments, and are displayed at 25 fps.

File S8 is available for download as an AVI at

Time-lapse imaging of SonC-GFP and histone H1-mCherry in lagging chromosome arms

Video shows time-lapse imaging of SonC-GFP together with histone H1-mCherry can sometimes be seen as lagging chromosome arms following mitosis (Fig. 7C). Images were collected every 1 min and are displayed at 2 fps. File S9 is available for download as an AVI at http://www.genetics.org/lookup/suppl/doi:10.1534/genetics.113.156745/-/DC1.

Time-lapse imaging of SonC-GFP and Bop1-mCherry during mitosis

Video shows time-lapse imaging of SonC-GFP together with the nucleolar marker Bop1-mCherry during mitosis (Fig. 7D). Images were collected every 1 min and are displayed at 2 frames per second (fps). File S10 is available for download as an AVI at http://www.genetics.org/lookup/suppl/doi:10.1534/genetics.113.156745/-/DC1.

3D rotation of SonC-GFP and Bop1-mCherry during mitosis

Video shows a 3D rotation over time of SonC-GFP and Bop1–mCherry during mitosis (Fig. 7D). Images were collected every 1 min, were rotated in 10^o increments, and are displayed at 25 fps. File S11 is available for download as an AVI at http://www.genetics.org/lookup/suppl/doi:10.1534/genetics.113.156745/-/DC1.

Time-lapse imaging of SonC-GFP and Pol I-mCherry during mitosis

Video shows time-lapse imaging of SonC-GFP together with the NOR marker Pol I-mCherry during mitosis (Fig. 7E). Images were collected every 1 min and are displayed at 2 frames per second (fps). File S12 is available for download as an AVI at http://www.genetics.org/lookup/suppl/doi:10.1534/genetics.113.156745/-/DC1.

Table S1 Strains Used in This Study

NAME	GENOTYPE	SOURCE
C61	yA1 nicB8 pyroA4 niiA4 riboB2 veA1	(Dowzer and Kelly 1991)
CDS36	sonB1 nimA1 pyrG89 wA veA1	(De Souza <i>et al.</i> 2003)
CDS364	sonB1 pyroA4 wA3 veA1	(De Souza <i>et al.</i> 2006)
CDS509	sonA1 pyroA4 wA3 yA2 ¹ veA1	This study
JLA1	nimA1 pyrG89 wA2/wA31 yA21 veA1	This study
JLA73	nimA1 mutant D pyrG89 wA2/wA3? yA21 veA1	This study
JLA75	nimA1 mutant F pyrG89 wA2/wA31 yA21 veA1	This study
JLA77	nimA1 mutant C (sonC1) pyrG89 wA2/wA31 yA21 veA1	This study
JLA78	nimA1 mutant E pyrG89 wA2/wA31 yA21 veA1	This study
JLA184	nimA1 mutant 89 pyrG89 wA2/wA3? yA2 ¹ veA1	This study
JLA185	nimA1 sonA2 pyrG89 wA2/wA31 yA21 veA1	This study
JLA227	sonA2 riboB2 pyroA4 yA2 wA2/wA31 veA1	This study
JLA255	nimA1 sonA3 pyrG89 wA2/wA31 yA21 veA1	This study
JLA263	mag1-GFP::pyrG ^{Af} pyrG89 nkuA <i>1</i> ::argB argB2 pyroA4 SE15	This study
	nirA14 chaA1 wA3 fwA1 veA1	
JLA264	mag1 <i>∆</i> ::pyrG ^{Af} pyrG89 argB2 nirA14 ¹ wA3 chaA1 ¹ fwA1 ¹ yA2 ¹	This study
	veA1	
JLA265	sonC-GFP::pyrG ^{Af} pyrG89 argB2 wA3 nirA14 ¹ yA2 ¹ chaA1 ¹	This study
	fwA1 ¹ veA1	
JLA268	sonA3 wA3 yA2 ¹ veA1	This study
JLA319	sonC-GFP::pyrG ^{Af} H1-mCherry::pyro ^{Af} pyroA4 ¹ pyrG89 argB2	This study
	wA3 yA2 ¹ chaA1 ¹ fwA1 ¹ veA1	
JLA324	sonC-GFP::pyrG ^{Af} bop1-mCherry::pyrG ^{Af} pyrG89 argB2	This study
	nirA14 ¹ wA3 yA2 ¹ chaA1 ¹ fwA1 ¹ veA1	
JLA325	sonC-GFP::pyrG ^{Af} pol I-mCherry::pyrG ^{Af} pyrG89 argB2 nirA14 ¹	This study
	wA3 yA2 ¹ chaA1 ¹ fwA1 veA1	
KF018	nimA-GFP::pyrG ^{AF} H1-mCherry::pyro ^{AF} nirA14 ¹	This study
	sE15 ¹ wA3 veA1	

KF110	nimA-GFP::pyrG ^{AF} nup170-mCherry::pyroA ^{AF}	This study
	argB2 wA3 veA1	
KF120	nimA-GFP::pyrG ^{AF} pyrG89 fib-mCherry::pyrG ^{AF} wA3 veA1	This study
KF122	nimA-GFP::pyrG ^{AF} pyrG89 topo1-mCherry::pyrG ^{AF} pyroA4	This study
	argB2 wA3 veA1	
LPW29	nimA1 sonA1 riboA1/riboB2 ¹ pyrG89 wA2 veA1	(Wu <i>et al.</i> 1998)
LU178	cgrA-GFP::pyrG ^{AF} H1-mCherry::pyrG ^{AF} pyrG89 argB2 wA3	(Ukil <i>et al.</i> 2009)
	nirA14 ¹ fwaA1 ¹ chaA1 ¹ veA1	
LU193	bop1-GFP::pyrG ^{AF} H1-mCherry::pyrG ^{AF} pyrG89 argB2 wA3	(Ukil <i>et al.</i> 2009)
	SE15 pabaA1 pyroA4 nirA14¹ fwaA1¹ chaA1¹ veA1	
R153	pyroA4 wA3 veA1	C. F. Roberts
SO369	sonA1 pyrG89 wA2 nicA2 veA1	This study

¹ In some strains we have not confirmed some markers that could be covered by or are recessive to other markers in

the strain.

Table S2 Primers Used in This Study

Name	Purpose	Sequence
3'sonBcDNA	sonB amplification	GCTCGAGTCAGATCTCAGTA
5'UTRsonB	sonB amplification	ACGGATGGACGTTGATAACATACTG
AF-2	nimA sequencing	GGTGTCACTATGGTACG
AF-4	nimA sequencing	GTCATTGCGAGCTGCCT
AF-6	nimA sequencing	CTTGAGTCGCCGACGAA
AF-7	nimA sequencing	AAGATCCCATCGTCCGC
AR-2	nimA amplification	GCAGCAGCGCAAGAAAT
HP15	nimA GFP-tagging	GAACACCATTCTACATGTCTCCAG
HP16	nimA GFP-tagging	CCTCAAGTTGCGAATCACCTTTC
HP17	nimA GFP-tagging	TGAAAACGCAAGATCGTCCTAACC
HP18	nimA GFP-tagging	TGGCGTACGAGCGCTTCAAAACTG
HP19	nimA GFP-tagging	GGGGAAGAAAGGTGATTCGCAACTTGAG GGGAGCTGGTGCAGGCGCTGGAGCCAA AG
HP20	nimA GFP-tagging	ACCGCCGGTTAGGACGATCTTGCGTTTT CACTGTCTGAGAGGAGGCACTGATGCGT G
M13 Forward	pRG3-AMA1-NotI plasmid library	CTGGCCGTCGTTTTAC
	sequencing	
NucP1	sonB sequencing	TATTGTAAGGCTGCATGAAC
NucP2	sonB sequencing	AACACTGGCAGCTCACTCTT
NucP4	sonB sequencing	GGAGTGGACGACACATT
NucP8	sonB sequencing	AACAACCGCGCAAACAAGGA
NucP11	sonB sequencing	GTCCGCAAGCAAGGTCAGAA
NucP12	sonB sequencing	CCTCAGATCGTGAAAGGGCT
NucP15	sonB sequencing	GCAGTGGCTCAGGCTTTGGT
NucP16	sonB sequencing	GGCATCGTTCCAGTTCTTCC
NucP17	sonB sequencing	TTGGTGGATGCCTTGGTTAC
NucP20	sonB sequencing	ATGGAAGTGGGCAGGCAGGA

NucP21	sonB sequencing	ATTGCTGAGTGTTGCTGCCA
NucP23	sonB sequencing	GGCACAGCTGCTACTTCGTC
NucP24	sonB sequencing	ACGTGCTCCTCTGCCAATCT
NucP26	sonB sequencing	CGGCCAGTGTACCCACACAA
NucP27	sonB sequencing	GGCATTACCGCTGATGAACC
NucP28	sonB sequencing	GGCCACCTTGGACGGATTCT
NucP32	sonB sequencing	CCTGCGAAGGTCTTTGGCTG
NucP34	sonB sequencing	ACTCGCTACGACCGATCCTC
oMN33	pRG3-AMA1-NotI plasmid library	AAATAAGCTTGCATGCGC
	sequencing	
SP204-F	AN4389 amplification	CAAGCATTGACCGATGACACT
SP205-R	AN4389 amplification	GCTCTCATTAAGTCAATCCAGCAT
SP206-F	AN4389 sequencing	CCGGATTCTGATTGCTAAGGC
SP207-F	AN4389 sequencing	GTGGACAACCGAGAATCTCA
SP209-F	AN4389 sequencing	CGACAACTGGCACGCTTC
SP233-F	nimA sequencing	CGACTACTAACCTGCAGA
SP234-F	nimA amplification	CCCGTGCATTGATATTGACTCC
SP251	sonC amplification and deletion	CACCAGAGCACCGAAATTGC
SP252	sonC deletion and GFP-tagging	CCAGGTCGAGAATTGTCCTTG
SP253-F	sonC sequencing	CGGAGCGATAAATCTACGTCT
SP254-F	sonC sequencing	GGACTAAATGTGCTGGCCG
SP255-F	sonC amplification, GFP-tagging, and	CTGCATGGAACCAACAGCT
	sequencing	
SP256-F	sonC sequencing	GATCTGCATGCGACGTTAGC
SP257-F	sonC sequencing	CGCAGCCTACAGATGACGG
SP258-F	sonC deletion and GFP-tagging	GCATCAGTGCCTCCTCTCAGACAGATGGCAACAGTTGTGT GAAGTATGC
SP259-R	sonC deletion	GAAGAGCATTGTTTGAGGCGGACGAGATCGCGTGGCAG AATATAT
SP260-R	sonC GFP-tagging	GGCTCCAGCGCCTGCACCAGCTCCCGCATAGACATTTGCA GGCGGCAGC

SPF1	sonA amplification and sequencing	GCTCTTGATACCCGTCTCTC
SPF2	sonA sequencing	GTACATCAATATAGTCGACCT
SPF5	sonA sequencing	CTGCTGCTGGTCACCTGTAAG
SPR5	sonA amplification and sequencing	CGTGACTGTCTATACGTGCG
SPR8	sonA sequencing	CGAGATCGAGCATACGAGCC