SUPPLEMENTARY FIGURE LEGENDS

Supplementary Fig. S1: The *cdipt^{lop}* (*lop*) and *cdipt^{hi559}* (*hi559*) mutants exhibit identical GI phenotype and fail to rescue each other in complementation assay. (A) Lateral view of ISH of intestine-specific markers *vil1* and *anxa2b* showing reduced expression in the *hi559* intestine at 5-dpf. (B-F) Morphology of intestine shown by bright-field images (*left panel*) and fluorescent images (*right panel*, Nile-Red staining shown by *white outline*). (B) GI morphology of *hi559* larvae at 6-dpf shows smaller intestine and lumen. (C) GI morphology of *lop* larvae shows smaller intestine and lumen at 8-dpf comparable to the 6-dpf *hi559* larvae. (D) *Cdipt^{hi559/lop}* (*hi559* X *lop*) transheterozygote mutant larvae show smaller intestinal lumen at 7.5-dpf comparable to the 6dpf *hi559* larvae. (E) Wild-type embryos were microinjected with either control morpholino or splice-blocking morpholino against *cdipt* at 1-cell stage and larvae were analyzed at 6-dpf (Thakur et al., 2011). *Cdipt* morphants show smaller intestinal structures replicating *hi559* phenotype. (F) Wild-type larvae were treated with PI synthase inhibitor δ -HCH shows smaller intestinal structures compared to DMSO treated larvae.

Supplementary Fig. S2: Abnormal goblet cells and mucosecretion in *hi559* intestinal mucosa. (A) Wild-type intestinal luminal area appears clear with mucinous granules and microvilli projections from IECs. The *hi559* lumen is often filled with detached cells (*black arrow*) and bacterial overgrowth at 5.5-dpf (*red arrow*). Nuclear condensation (*white arrow*) and cytoplasmic vacuoles (*arrowhead*) are apparent in the detached IEC, suggesting apoptosis. (B) At 5-dpf, wild-type mucinous GCs are mature with largely dense mucinous theca, whereas the *hi559* GCs have poorly developed theca with abnormal vesicles, cytoplasmic necrosis and bacterial invasion (*red arrow*). (C-D) PAS-staining (*pink*) showing the mucin-secreting mid-intestinal GCs (*arrow*) and secreted mucinous layer (*arrowhead*) of the mid-intestinal epithelium at (C) 5-dpf and (D) 6-dpf. The thick secreted mucinous layer, which is seen at the apical border of wild-type intestinal mucosa, is greatly diminished in *hi559* and the mucinous cells are disarranged and often detached into the lumen. (E) Bar-chart showing the number of mid-intestinal (Mi) GCs in wild-type and *hi559* intestine at 5 and 6-dpf (*n*=7, **P*<0.05). Goblet cell

population is significantly depleted in *hi559* intestine at 6-dpf. Wild-type (WT) on *left* and *hi559* on *right panels*. Lu, intestinal lumen; Th, mucinous theca. Scale bar: A-B, 500 nm; C-D, 20 μ M.

Supplementary Fig. S3: PI deficiency causes necro-inflammattion. (A) H&E-staining shows nicely arranged mature cylindrical IECs in wild-type with thick apical border, whereas *hi559* epithelium shows necro-inflammatory injury with vacuolated IECs, thinner apical border and necrotic foci (*arrow*). (B) Confocal projection of intestines of DMSO and δ -HCH-treated *Tg(lyzc:egfp)* larvae, showing leukocyte infiltration (*arrow*). Right panel shows bar-chart of leukocytes in DMSO and δ -HCH-treated wild-type intestines at 6-dpf (*n*=15, **P*<0.05). Scale bar: A, 20 µM; B, 100 µM.

Supplementary Fig. S4: Activation of pro-inflammatory cascade in *hi559 larvae*. (A) Differential upregulation and the corresponding *P*-values of a set of pro-inflammatory gene features and APR complement cascade in *hi559* larvae. (B) GSEA enrichment plots and expression profile (shown by heat-map) of genes involved in the NF- κ B pathway, suggesting their enrichment in *hi559* larvae (*P*=0.006).

Supplementary Fig. S5: Unresolved ER stress pathology of *hi559* GI tissues. (A-B) ISH shows elevated expression of the crucial ER stress-UPR markers *hspa5* (A) and *xbp1* (B) in the *hi559* GI tissue. (C) RT-PCR showing activation of *xbp1* by increased transcription of unspliced (*xbp1-u*) and spliced *xbp1* (*xbp1-s*) in 5-dpf *hi559* GI tissues. β -actin was used as RT-PCR control. (D-E) TEM images of 6-dpf wild-type (*left panels*) and *hi559* pancreatic cells (*right panels*). (D) Ultrastructure of the secretory granules-containing endocrine pancreatic cells shows increased ER luminal expansion and vacuolization, multi-lamellar autophagic bodies (*red outline*), and increased mitophagy (*red arrow*) in *hi559*. (E) ER luminal expansion is also apparent in the *hi559* pancreatic acinar cells containing zymogen granules. Er: endoplasmic reticulum; Sg, secretory granules; Zm; zymogen granules; Mt, mitochondria. Scale bars: D-E, 500 nm.

Supplementary Fig. S6: Tunicamycin-induced ER stress causes abnormal GCs and necro-inflammation in wild-type GI tissues. (A) Bright-field image of DMSO and tunicamycin-treated larvae showing intestinal morphology (*red outline*). (B) H&E-

stained sections of DMSO and tunicamycin-treated larval esophagus showing GC morphology (*arrow*). Note the condensed nuclei of *hi559* GCs, suggesting apoptosis, in tunicamycin-treated larvae. (C) H&E-stained sections of DMSO and tunicamycin-treated larval intestine showing necro-inflammatory injury (*pink outline*) and increased cytoplasmic vesicles (*arrow*) in the IECs of the tunicamycin-treated intestine. (D) Confocal projection of Tg(mpx:gfp) larval intestine shows increased neutrophil infiltration in tunicamycin-treated larvae. Corresponding bar-chart is shown in the *right panel* (***P*<0.01). TN, tunicamycin. Scale bars: B, C, 20 μ M; D, 100 μ M.

Supplementary Fig. S7: Suppression of inflammation by antibiotics or antiinflammatory drugs. Larvae were treated with antibiotics (abx) or anti-inflammatory (ai) drugs from 3.5-dpf to 6-dpf. (A) ISH with *fabp2* shows no improvement of intestinal architecture in *hi559* larvae treated with abx or ai drugs. (B) ISH with *mpo* showing partial suppression of intestinal neutrophil infiltration (*arrows*) in abx-treated *hi559* larvae and complete suppression in ai-treated larvae. (C) H&E-staining shows clearing of luminal bacteria in abx-treated *hi559* larvae. Luminal bacteria (*asterisk*) are seen in aitreated larvae. IEC necrosis and shedding (*arrow*) and vacuolation (*arrowhead*) are still apparent in both abx- and ai-treated larvae. (E) Bar-charts showing significant reduction of intestinal bacterial density (gut CFU) in abx-treated *hi559* larvae (***P*<0.01). Scale bars: C, 20 μ M.

REFERENCES

Thakur, P. C., Stuckenholz, C., Rivera, M. R., Davison, J. M., Yao, J. K., Amsterdam, A., Sadler, K. C. and Bahary, N. (2011) 'Lack of de novo phosphatidylinositol synthesis leads to endoplasmic reticulum stress and hepatic steatosis in cdipt-deficient zebrafish', *Hepatology* 54(2): 452-62.















Symbol C4B CFB SOCS3 C3 HPX CEBPB FGA CP C9 FGB FGG CRP JUN STAT3 A2M HMOX2 PLG	Entrez Gene Name complement component 4B (Chido blood group) complement factor B suppressor of cytokine signaling 3 complement component 3 hemopexin CCAAT/enhancer binding protein (C/EBP), beta fibrinogen alpha chain ceruloplasmin (ferroxidase) complement component 9 fibrinogen beta chain fibrinogen gamma chain C-reactive protein, pentraxin-related jun proto-oncogene signal transducer and activator of transcription 3 alpha-2-macroglobulin heme oxygenase (decycling) 2	<i>P</i> -value 3.39E-05 4.14E-05 6.62E-04 4.88E-04 4.00E-02 2.37E-03 2.77E-03 3.28E-02 1.33E-03 2.68E-04 3.45E-04 2.32E-02 6.24E-04 2.47E-03 1.53E-02 4.84E-02 1.15E-02
HMOX2 PLG NRAS SERPING1	heme oxygenase (decycling) 2 plasminogen neuroblastoma RAS viral (v-ras) oncogene homolog serpin peptidase inhibitor, clade G (C1 inhibitor)	4.84E-02 1.15E-02 2.74E-02 4.01E-02
PLG NRAS SERPING1	plasminogen neuroblastoma RAS viral (v-ras) oncogene homolog serpin peptidase inhibitor, clade G (C1 inhibitor)	4.04E-02 1.15E-02 2.74E-02 4.01E-02

Α



























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