

Supplementary Materials and Methods

Cell Lines and Cell Culture Conditions: KYSE150 and Colo824 cell lines were obtained from DSMZ (Braunschweig, Germany). The KYSE150 cell line was established from the poorly differentiated esophageal squamous cell carcinoma resected from upper (cervical) esophagus of a 49-year-old Japanese woman after receiving radiotherapy (the tumor was invading contiguous structures) (Shimada *et al.*, 1992). KYSE 150 cells were cultured in 49% RPMI-1640 + 49% Ham's F12 + 2% fetal bovine serum (FBS). Colo824 cell line was established from the pleural fluid of a 52-year-old patient with breast carcinoma. Colo824 cells were cultured in 15% FBS RPMI-1640. HCC70 and HCC1954 cell lines were obtained from ATCC (Manassas, VA, USA). HCC70 and HCC1954 cells were cultured in 10% FBS RPMI-1640. MCF10A is a spontaneously immortalized, but nontransformed human mammary epithelial cell line derived from the breast tissue of a 36-year-old patient with fibrocystic changes (Soule *et al.*, 1990). MCF10A cells were cultured in Ham's F-12 medium supplemented with 0.1% bovine serum albumin, fungizone (0.5 µg/mL), gentamicin (5 µg/mL), ethanolamine (5 mmol/L), HEPES (10 mmol/L), transferrin (5 µg/mL), 3,3',5'-Triiodo-L-Thyronine (T3) (10 µmol/L), selenium (50 µmol/L), hydrocortisone (1 µg/mL), insulin (5 µg/mL) and 10 ng/ml epidermal growth factor (EGF). The isolation and culture of the SUM series of HBC cell lines have been described in detail previously (Forozan *et al.*, 2000; Forozan *et al.*, 1999). SUM-44 and SUM-190 cells were cultured in Ham's F-12 medium supplemented with 0.1% bovine serum albumin, fungizone (0.5 µg/mL), gentamicin (5 µg/mL), ethanolamine (5 mmol/L), HEPES (10 mmol/L), transferrin (5 µg/mL), 3,3',5'-Triiodo-L-Thyronine (T3) (10 µmol/L), selenium (50 µmol/L), hydrocortisone (1 µg/mL), and insulin. SUM-52, SUM-149 and SUM-225 cells were cultured with 5% fetal bovine serum, fungizone (0.5 µg/mL), gentamicin (5 µg/mL), hydrocortisone (1 µg/mL) and insulin (5 µg/mL).

References

- Forozan F, Mahlamaki EH, Monni O, Chen Y, Veldman R, Jiang Y *et al* (2000). CGH analysis of 38 breast cancer cell lines: A basis for interpreting cDNA microarray data. *Cancer Res* **60**: 4519-4525.
- Forozan F, Veldman R, Ammerman CA, Parsa NZ, Kallioniemi A, Kallioniemi O *et al* (1999). Molecular cytogenetic analysis of 11 new human breast cancer cell lines. *Br J Cancer* **81**: 1328-1334.
- Shimada Y, Imamura M, Wagata T, Yamaguchi N, Tobe T (1992). Characterization of 21 newly established esophageal cancer cell lines. *Cancer* **69**: 277-84.
- Soule HD, Maloney TM, Wolman SR, Peterson WD, Jr., Brenz R, McGrath CM *et al* (1990). Isolation and characterization of a spontaneously immortalized human breast epithelial cell line, MCF-10. *Cancer Res* **50**: 6075-86.

Supplementary Table 1: Array CGH data on chromosome 9p using an Agilent 244K chip (A) or 44K chip (B) in 9 cancer cell lines.

Supplementary Figure Legend

Figure S1. A summary of GASC1 amplification across multiple GISTIC analyses performed on the entire dataset (copy-number profiles of 3131 cancer samples) or subsets representing different cancer types.

Figure S2. Genome view of the PTPRD locus analyzed on the Agilent oligonucleotide array (Agilent Technology) in KYSE150 cells.

Figure S3. Schematic representation of PTPRD primers for genomic real-time PCR.

Figure S4. Relative copy number of the PTPRD gene in KYSE150, HCC1954 and control cells determined by genomic real time-PCR. Genomic DNA was used as a template and PCR amplification of PTPRD fragments of intron 7-exon 8 and intron 8-exon 9 were normalized against the PCR amplification of β -actin. Copy numbers in control cells are set as 0 arbitrarily. Relative copy numbers were shown as \log_2 values.

Figure S5: Summary of regions at chromosome 9 with significant copy number alterations in basal-like primary breast tumor, brain metastasis and xenograft samples that were adopted from Ding et al's published Supplementary Information .

Figure S6: Protein levels of p53 in HCC1954 cells stably expressing control shRNA, UHRF2 shRNA#1 or shRNA#2 were analyzed by Western blot.

Figure S7: Detection of RB protein in HCC1954 cells stably expressing non-silencing shRNA (lane 1), UHRF2 shRNA#1 (lane 2), or UHRF2 shRNA#2 (lane 3). Whole cell extracts of MCF10A cells were resolved on the same gel to serve as a migration control for the hypophosphorylated (p) form and the

hyperphosphorylated (pp) form of RB protein. MCF10A cells were either EGF-starved (-EGF) for 24 hours or EGF-starved for 24 hours followed by stimulation with EGF (+EGF) for 24 hours.

Figure S1

Summary

Amplifications

Deletions

JMJD2C ([chr9:6747653-7165648](#))

Cancer Subset	In Peak?	Nearest Peak	#Genes in Peak	Q-value	Frequency of Amplification		
					Overall	Focal	High-level
all_cancers	No	chr9:137916695-140207187	76	1.0	0.115	0.0287	0.0105
Lung_SC	Yes	chr9:235706-8362053	41	1.0	0.4	0.1	0.05
Breast	Yes	chr9:235706-18592403	56	1.0	0.1564	0.0453	0.0453
all_lung	Yes	chr9:235706-8548331	41	1.0	0.1344	0.0478	0.0065
Esophageal_squamous	No	No peak on chromosome	0	1.0	0.2045	0.0227	0.0
Myeloproliferative_disorder	No	No peak on chromosome	0	1.0	0.0558	0.0	0.0
Ovarian	No	No peak on chromosome	0	1.0	0.1942	0.068	0.0194
all_neural	No	No peak on chromosome	0	1.0	0.1198	0.0046	0.0046
Acute_lymphoblastic_leukemia	No	chr9:36988416-36998984	0	1.0	0.0307	0.0	0.0
Colorectal	No	chr9:135358506-140207187	101	1.0	0.1925	0.0062	0.0124
Glioma	No	No peak on chromosome	0	1.0	0.122	0.0	0.0
Hepatocellular	No	No peak on chromosome	0	1.0	0.0826	0.0165	0.0
Lung_NSC	No	No peak on chromosome	0	1.0	0.1201	0.045	0.0041
Medulloblastoma	No	No peak on chromosome	0	1.0	0.1328	0.0078	0.0078
Melanoma	No	chr9:26573226-29470065	11	1.0	0.0811	0.045	0.0090
Prostate	No	No peak on chromosome	0	1.0	0.1087	0.0217	0.0
Renal	No	No peak on chromosome	0	1.0	0.0238	0.0159	0.0
all_epithelial	No	chr9:34233377-36127464	49	1.0	0.133	0.0388	0.0119
all_hematologic	No	chr9:4648449-5698176	13	1.0	0.0515	0.0043	0.0043

Figure S2

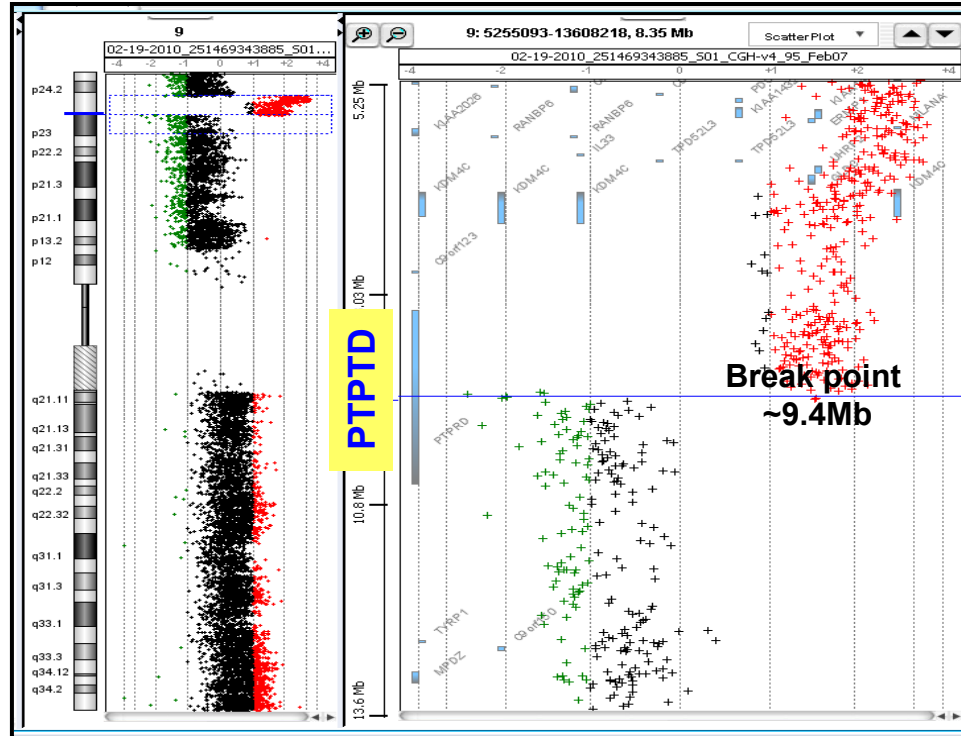


Figure S3

Schematic Representation of PTPRD PCR primers

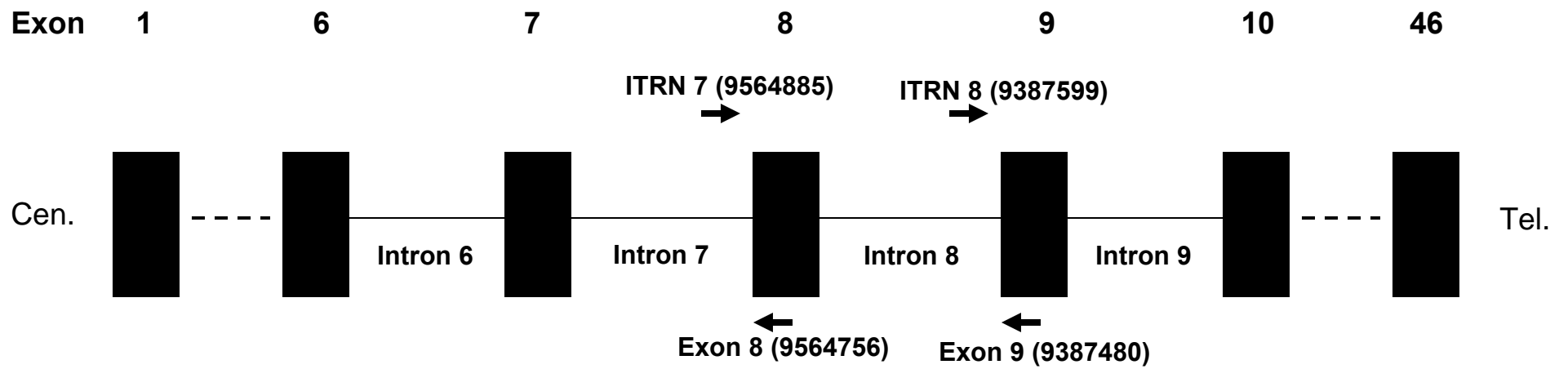


Figure S4

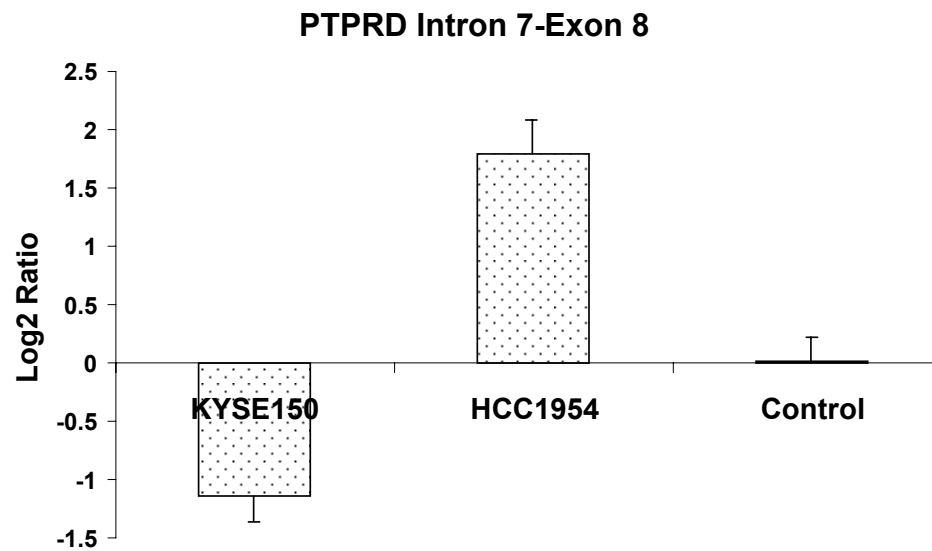
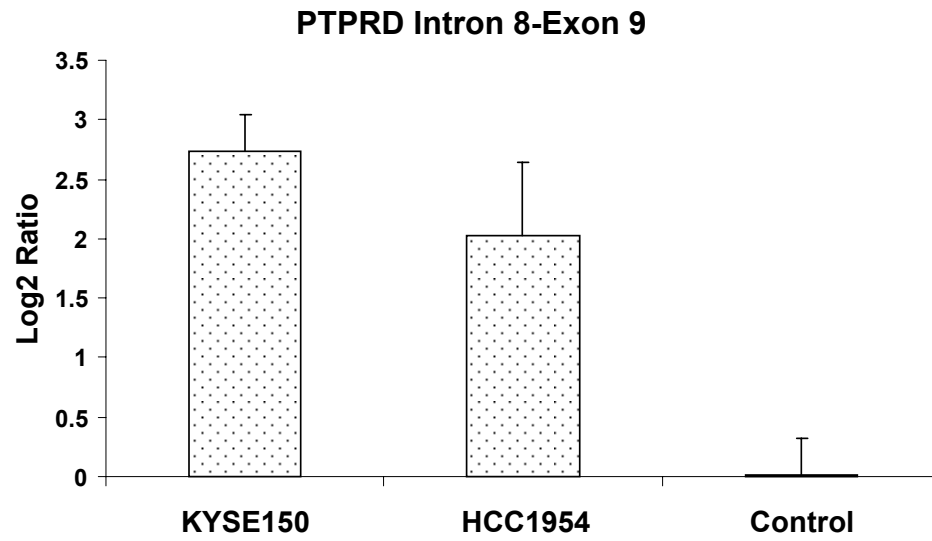



Figure S5


Supplementary Information for Genome Remodeling in a Basal-like Breast Cancer Metastasis and Xenograft (Ding et al. 2010)

Supplementary Table 5. Summary of regions with significant copy number alterations in primary tumor.




Chromosome	Start position	End position	Size	Number of markers	HMM state	Copy Number	LLR score
9	200000	8280000	8080000	809	3	2.83	42.04
9	17560000	20390000	2830000	284	3	2.8	12.37
9	139030000	140130000	1100000	111	3	3.1	12.88

Supplementary Table 6. Summary of regions with significant copy number alterations in brain metastasis.



Chromosome	Start position	End position	Size	Number of markers	HMM state	Copy Number	LLR score
9	200000	8880000	8680000	869	3	2.67	36.97
9	13350000	15370000	2020000	203	3	2.73	10.38
9	16000000	20580000	4580000	459	3	2.64	16.29
9	139020000	140130000	1110000	112	3	3.44	21.78

Supplementary Table 7. Summary of regions with significant copy number alterations in xenograft.



Chromosome	Start position	End position	Size	Number of markers	HMM state	Copy Number	LLR score
9	200000	7780000	7580000	759	3	2.84	48.75
9	12990000	19670000	6680000	669	3	2.74	30.2
9	138340000	140130000	1790000	180	3	4.9	80.77

Figure S6

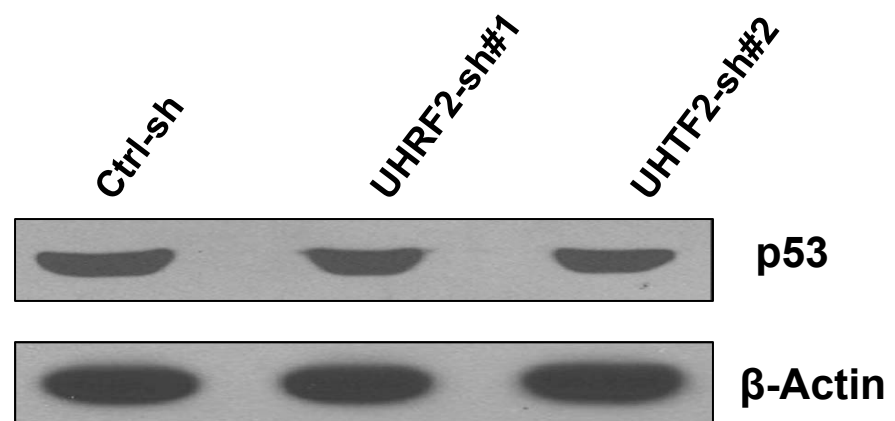


Figure S7

