

Supporting Information

Supplementary Table 1. List of primers used in this study

Supplementary Fig. 1. Expression of *AtPrx1*, *AtPrx22*, *AtPrx39* and *AtPrx69* in *bri1-9* and *BRI1-GFP* plants compared with that of the wild type. ANOVA statistical analyses were performed and significant differences are indicated by P values (*: $P \leq 0.004$, **: $P \leq 0.002$). The unpaired t-test was performed (***: P value = 0.5).

Supplementary Fig. 2. Gross morphologies and H_2O_2 concentrations of T-DNA insertional *AtPrx* mutants. (A) Phenotypes of T-DNA insertional *AtPrx* mutants compared with those of wild type. Pictures were taken of 4-week-old plants grown under long-day conditions. (B) Hydrogen peroxide concentrations were determined from 10-day-old seedlings.

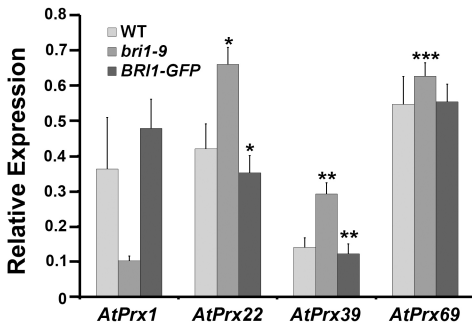
Supplementary Fig. 3. DAB-staining pattern of the *BRI1-GFP* plants transformed with *AtPrx1*, *AtPrx22*, and *AtPrx69* under normal conditions.

Supplementary Table 1. Primer Sequences

Primer Name	Sequence (5' to 3')	Use
<i>AtPrx1 RT F</i>	GGGGGTCACACCATTGGAATCTC	Confirmation of null <i>AtPrx1</i> expression in the <i>atprx1</i> mutant
<i>AtPrx1 RT R</i>	GGAGGGAGAATGGCCTGAGTCTG	
<i>AtPrx22 RT F</i>	GACATGTCCGTGTTCTTTTCGAGCATTCA	Confirmation of null <i>AtPrx22</i> expression in the <i>atprx22</i> mutant
<i>AtPrx22 RT R</i>	TATTAGTCGAAATTGTTCTCACGTGCG	
<i>AtPrx39 RT F</i>	CACCGGCGTTGGAGACCAAGATCC	Confirmation of null <i>AtPrx39</i> expression in the <i>atprx39</i> mutant
<i>AtPrx39 RT R</i>	CTCCGATCCTCCGGCGAAGCGTTTG	
<i>AtPrx69 RT F</i>	CTGCTGGCTGCGGTCTAGTAAGGGG	Confirmation of null <i>AtPrx69</i> expression in the <i>atprx69</i> mutant
<i>AtPrx69 RT R</i>	GGCTCGGGTCTCGGGATCCTTCC	
<i>Tubulin F</i>	ACCATGAGCAGCTTTCTGTGCCTG	Amplification of Arabidopsis <i>tubulin</i> for normalization
<i>Tubulin R</i>	CGCCGACTTCCTCATAGTCCTTC	
<i>AtrbohD F</i>	GGAAGGATGGACTGGCATTGTG	Monitoring the <i>AtrbohD</i> expression in response to cold
<i>AtrbohD R</i>	GTACGCTCAGTAATCGTCTCCG	
<i>AtrbohF F</i>	GAACGATCGGCGACGGTGGTCATTTG	Monitoring the <i>AtrbohF</i> expression in response to cold
<i>AtrbohF R</i>	CGAAATCGGAGCGATAGATGTAACCATT	
<i>AtPrx1 F</i>	GGGGTACCATGGCGATCAAGAACATTCTCGC	Amplification of full length <i>AtPrx1</i> for overexpression construct
<i>AtPrx1 R</i>	ACGCGTCGACTTAGTTAGGGAAGGCGCATCTC	
<i>AtPrx22 F</i>	GGGGTACCATGGGGTTTTCTCCTTCATTTTC	Amplification of full length <i>AtPrx22</i> for overexpression construct
<i>AtPrx22 R</i>	ACGCGTCGACTCAGATAGAACTCACAACACCAT	
<i>AtPrx69 F</i>	TCCCCCGGGATGGGTCGTGGTTACAATTTGC	Amplification of full length <i>AtPrx69</i> for overexpression construct
<i>AtPrx69 R</i>	GCGTCGACTTAGTTGATGGCGGAACAAACC	

RNA was purified from the seedlings grown for 10 days in normal conditions and then exposed to cold for 24 hours. The RNA samples were treated with RNase-free RQ1 DNase (Promega) and used for first-strand cDNA synthesis with the Superscript^{III}-MMLV reverse transcriptase (Invitrogen) using oligo d(T₁₅) as the primer. Second-strand synthesis was performed using the same aliquot of first-strand cDNA as the template. PCR conditions were as follows: pre-denaturation at 94°C for 4 min., denaturation at 94°C for 30 sec., primer-annealing at 52°C for 30 sec., elongation at 72°C for 30 sec. for 25 cycles or 28 cycles depending on the experiment, and post-elongation at 72°C for 7 min. The expression of each gene was normalized to β -*Tubulin*.

Supplementary Fig.1 Kim *et al.*

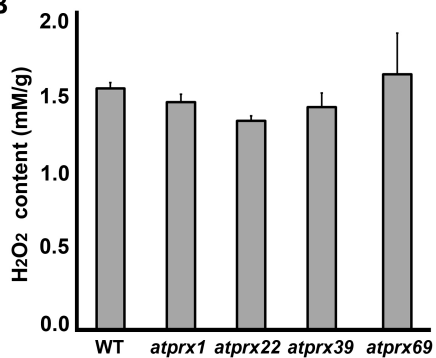


Supplementary Fig. 2 Kim *et al.*

A



B



Supplementary Fig. 3 Kim *et al.*



BRI1-GFP

***AtPrx1OE/
BRI1-GFP***

***AtPrx22OE/
BRI1-GFP***

***AtPrx69OE/
BRI1-GFP***