

# CHEMBIOCHEM

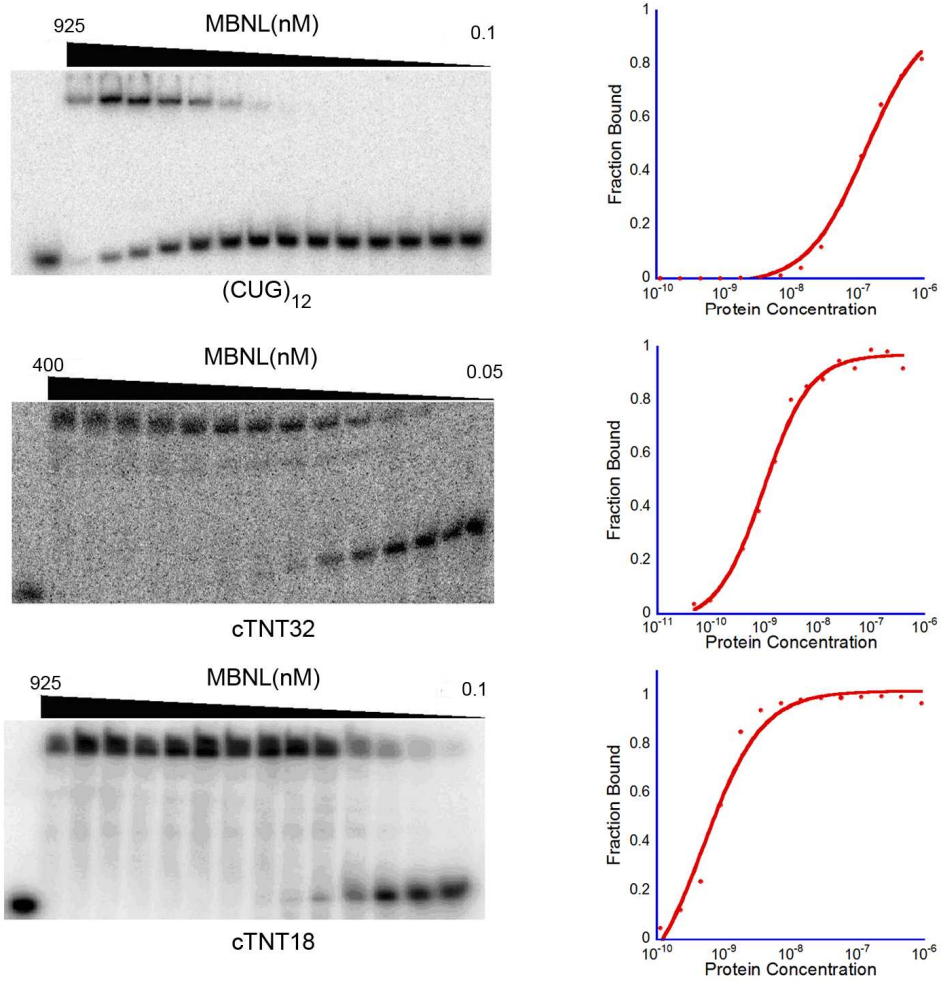
## Supporting Information

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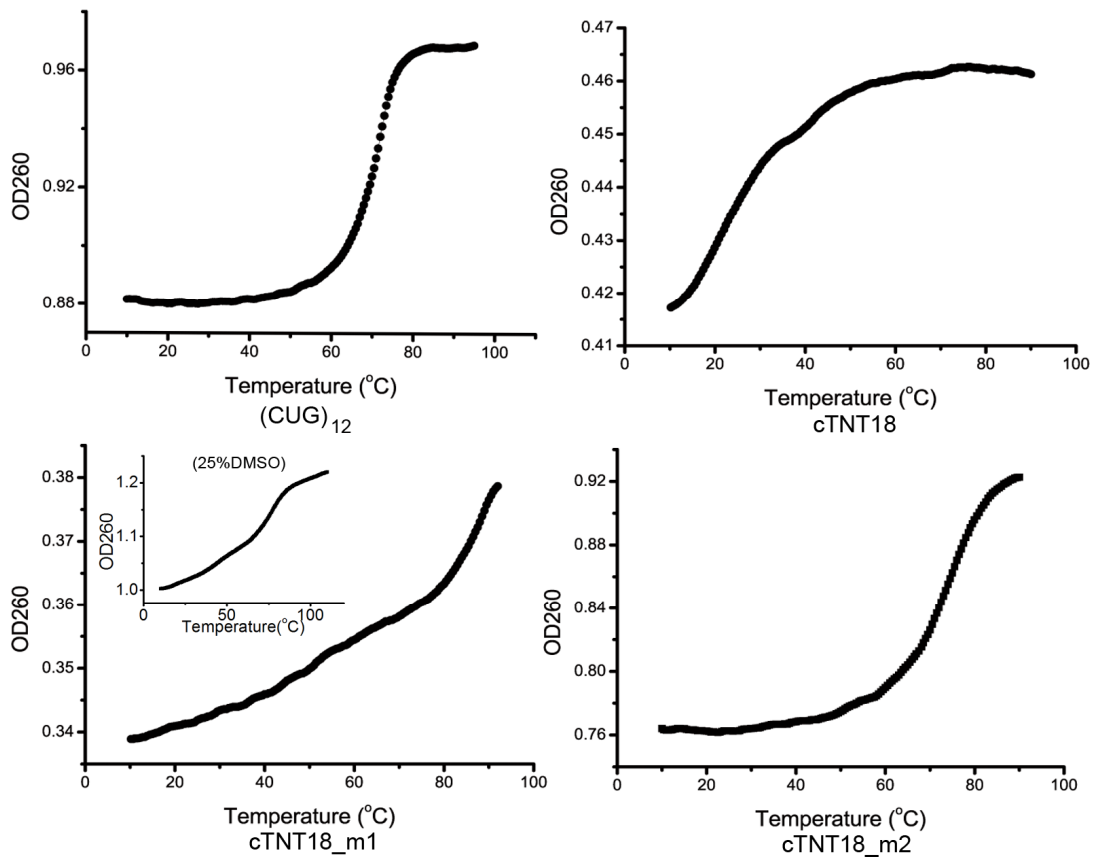
### **MBNL1–RNA Recognition: Contributions of MBNL1 Sequence and RNA Conformation**

Yuan Fu, Sreenivasa Rao Ramisetty, Nejmun Hussain, and Anne M. Baranger<sup>\*[a]</sup>

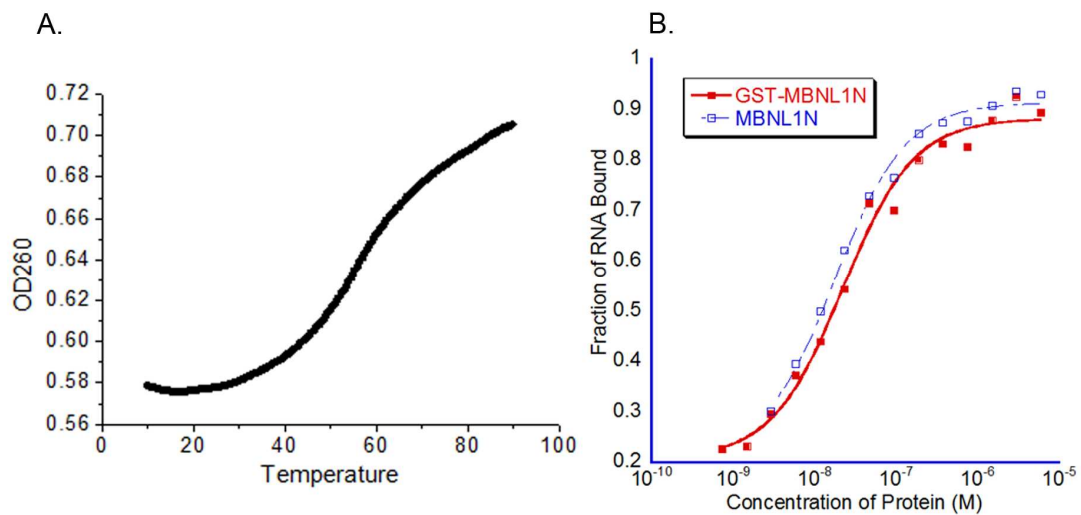
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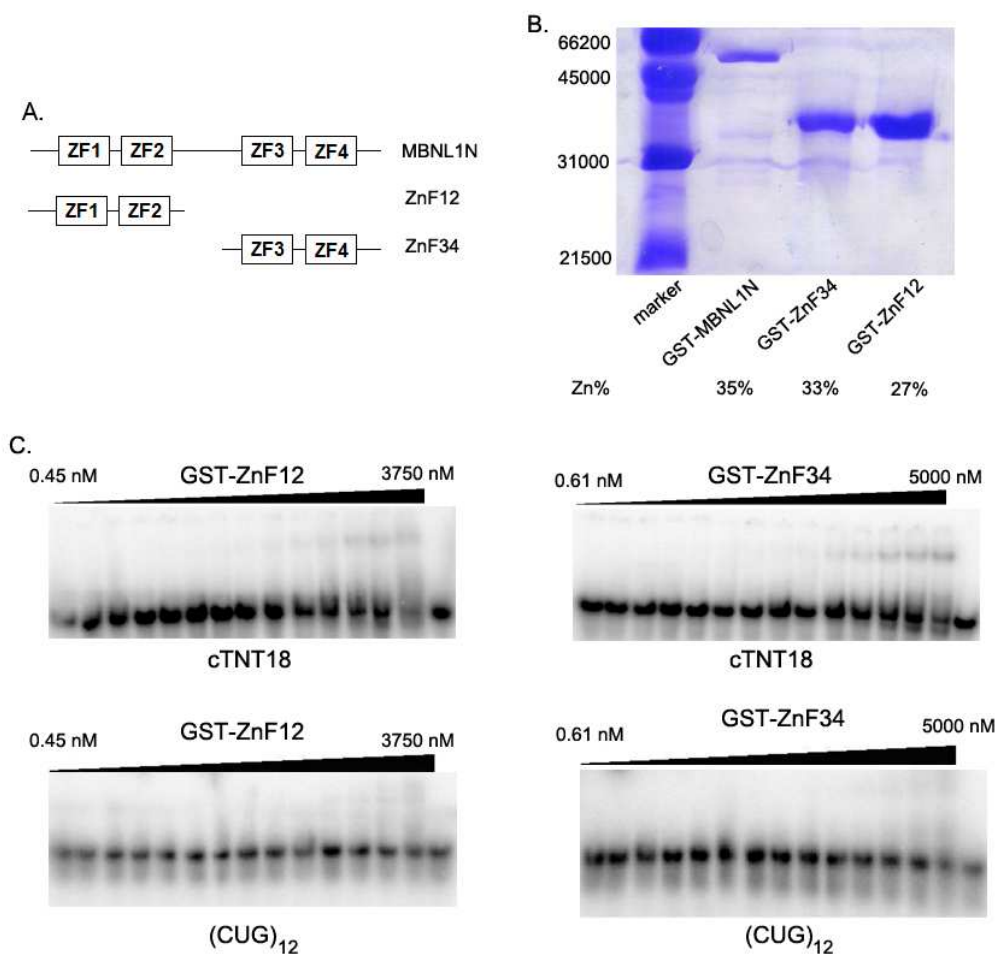
**Figure S1.** Representative results of gel mobility shift assays for MBNL1N with (CUG)<sub>12</sub> and cTNT RNAs. The protein concentration is noted on the top of each gel. The RNA concentration is 0.1 nM. The gels were run at 360V in 0.5X TB buffer.



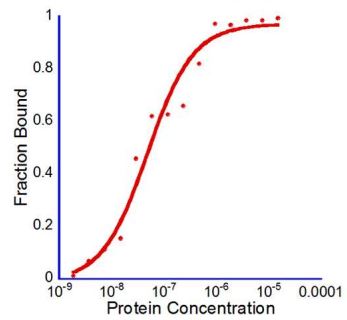
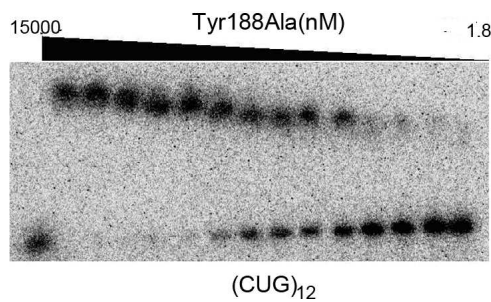
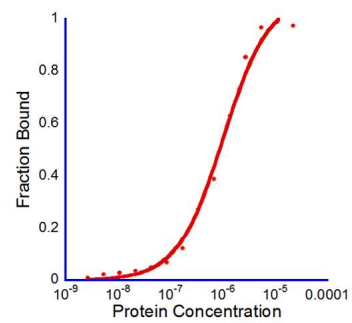
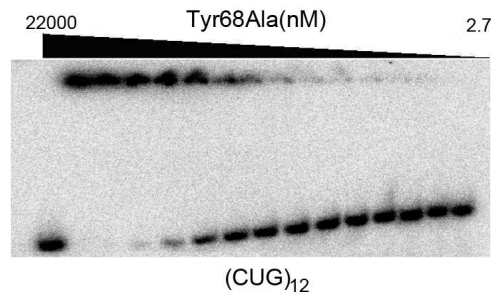
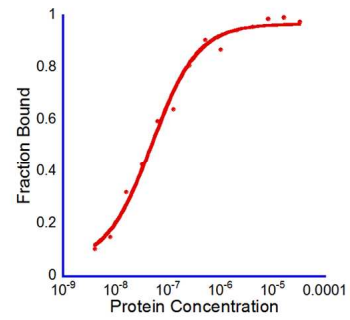
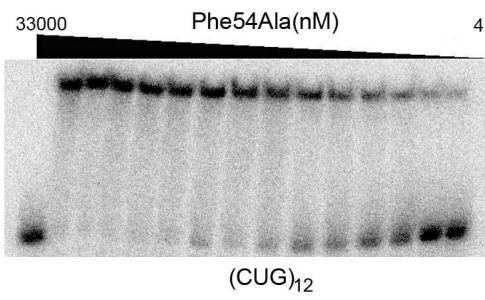
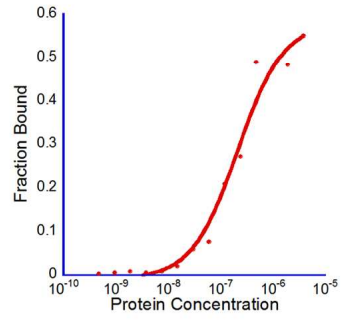
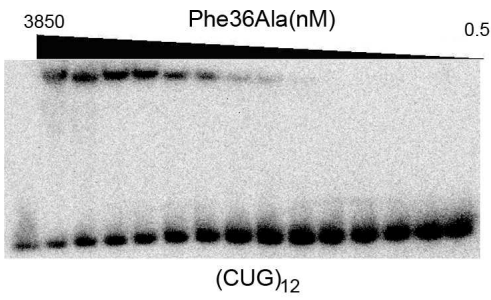
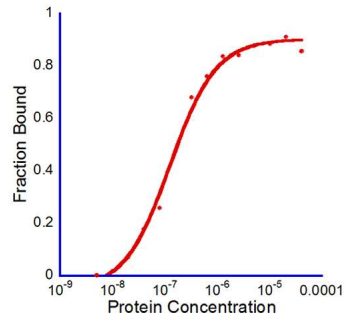
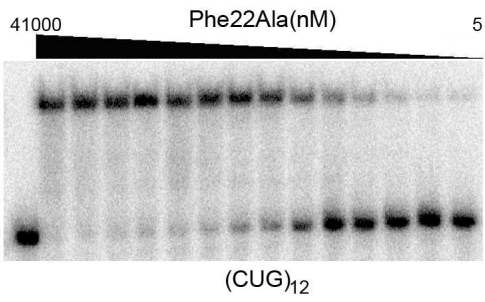
**Figure S2.** Representative melting curves of RNAs. The absorbance of each sample was monitored at 260 nm from 10 °C to 90 °C.

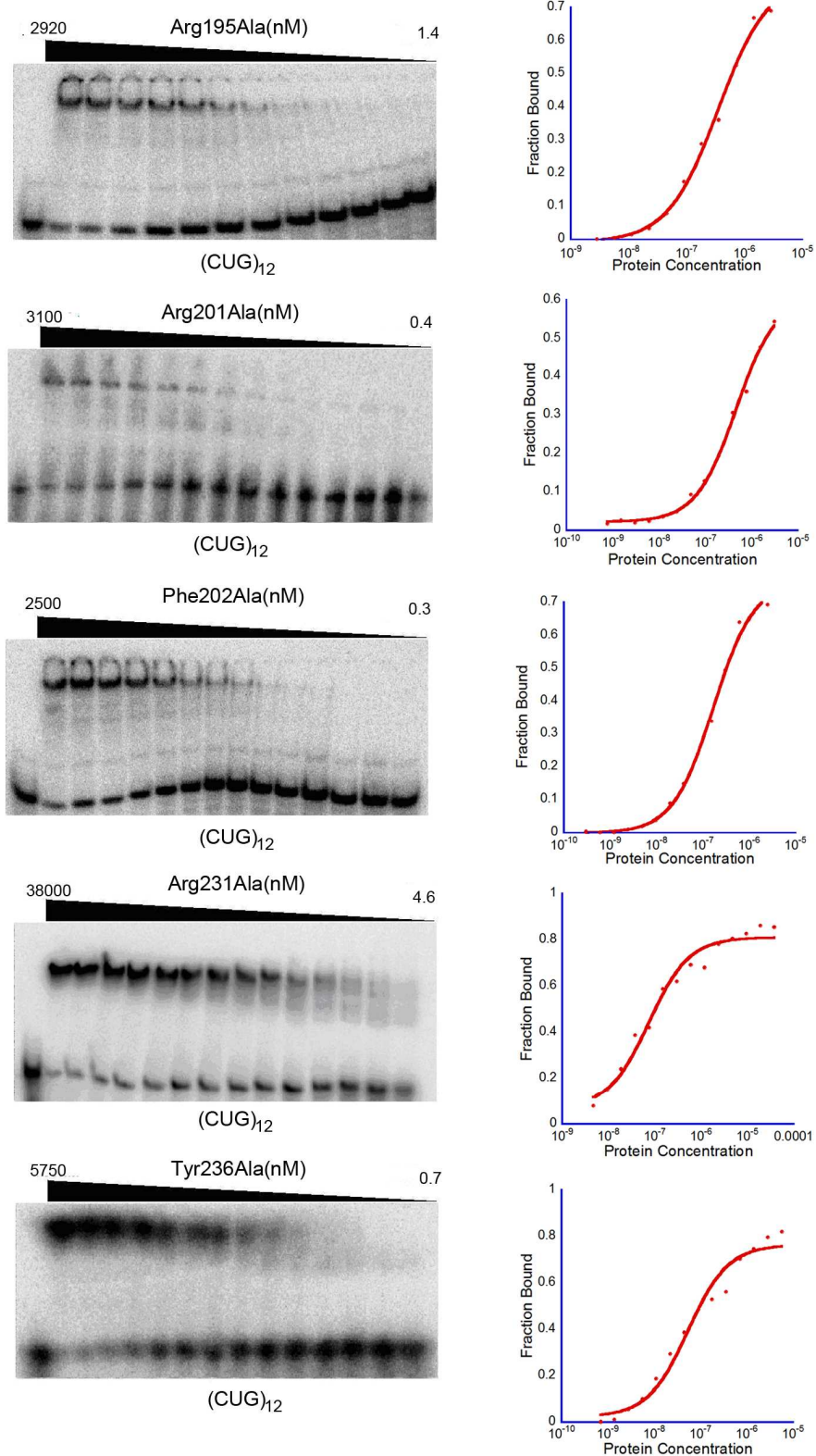


**Figure S3.** A) Melting temperature analysis of the quencher and fluorophore dual labeled RNA cTNT21\*. B) Curve fitting for gel mobility shift assays of MBNL1N and GST-MBNL1N interacting with cTNT21 RNA.



**Figure S4.** A) Schematic illustration of the relationship between truncated proteins and full-length MBNL1N. B) SDS-PAGE analysis of the purified proteins and analysis of zinc inclusion percentage by ICP-MS. C) Representative results of gel mobility shifts of truncated proteins with cTNT18 and (CUG)<sub>12</sub> RNAs.





**Figure S5.** Representative results of gel mobility shift assays of mutant proteins and MBNL1N with  $(CUG)_{12}$  RNA. The protein concentration is noted on top of each gel. The RNA concentration is 0.1 nM. The gels were run at 360V in 0.5X TB buffer.