Supporting Information

Bedel et al. 10.1073/pnas.1320777110



Fig S1. Invariant NK T cells (iNKT) cells in 2A3-D Tg mice depends on CD1d for thymic development. (*A*) Purified lymphocytes from the thymus, spleen, liver, and peripheral lymph nodes (pLN) of C57BL/6 or 2A3-D Tg mice were stained for indicated markers to assess reactivity of iNKT cells to self-CD1d and PBS57-CD1d (data representative of n = 3). (*B*) Purified lymphocytes from the thymus, spleen, liver, and pLN of 2A3-D Tg CD1d1d2^{-/-} mice were stained for indicated markers to confirm that iNKT cell positive selection is still restricted to CD1d (data representative of n = 2). (C) Purified lymphocytes from the thymus, spleen, liver, and pLN of J α 18^{-/-} mice were stained for indicated markers to confirm the specificity of iNKT cell staining (data representative of n = 2).



Fig S2. Ly108 expression levels are similar on C57BL6 and 2A3-D Tg thymocytes. Purified lymphocytes from the thymus of C57BL/6 or 2A3-D Tg mice were stained for indicated markers to assess Ly108 expression level. The gating strategy to obtain total thymocytes is depicted and Ly108 gMFI (geometric mean flouorescent intensity) is indicated as a representative value of n = 3.



Fig S3. 2A3-D Tg NKT cells do not express more Annexin V compared with wild-type control. Total thymocytes were stained for CD24 and CD44 to define the different developmental stages. Here, the stages are defined as follows: stage 0 (CD24^{ligh}CD44^{low}), stage 1 (CD24^{low}CD44^{low}), stage 2/3 (CD24^{low}CD44^{ligh}). Total thymocytes were stained for Annexin V and 7-ADD to assess apoptosis and necrosis respectively (data representative of n = 2). ns, not significant.



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C	C57BL/6 2A														2A3	2A3-D															
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Fig 54. Complementary data for Fig. 5. (A) PBS57-CD1d tetramer-positive T-cell receptor $(TCR)-\beta^+$ cells were sorted from the spleen and peripheral lymph nodes of 2A3-D Tg mice. mRNA was extracted and transformed into cDNA. cDNAs were amplified by PCR with V α -specific forward primers and a C α -specific reverse primer. (*B*) Sequences of the V α 14-J α 18 rearrangements found in iNKT cells from the spleen of C57BL/6 and 2A3-D Tg mice. For each observed rearrangement, the number of sequences, contribution of the V α 14 and J α 18 chain and existence of p-addition (blue) or n-addiction is depicted (data representative of n = 2). (C) PCR analysis was performed to evaluate the use frequency of TCR- α joining (TRAJ) genes encoding productive, in-frame rearrangements with the TRAV11 family in iNKT cells from the liver or the pLN or C57BL/6 or 2A-D Tg mice. (*D*) Amino acid composition and size of the CDR3 of the α -chain in C57BL/6 or 2A3-D Tg in iNKT cells from the liver and pLN.



Fig S5. Specific loss of self-reactivity in V α 14 natural variants paired with 2A3-D Tg V β . (A) 5KC Hybridoma expressing 2A3-D Tg V β were transduced with indicated V α 14 natural variants and tested for reactivity with self-CD1d and PBS57-CD1d tetramers (data representative of n = 4). (B) For each 5KC hybridoma expressing a V α 14 natural variant, the gMFI of the PBS57-CD1d or self-CD1d tetramer staining was evaluated for a narrow slice of TCR expression. Relative percentage of this gMFI compared with the gMFI of the D94 variant with PBS57-CD1d and self-CD1d is shown (n = 3).



Fig S6. Hybridomas expressing the D94A V α 14 natural variant paired with 2A3-D Tg V β do not induce Egr-2 upon autoreactive response to CD1d-transfected A20 cells. The TCR⁻ 5KC hybridoma was transduced with the 2A3-D β -chain paired either with the D94 wild-type V α 14 iNKT chain or the A94 variant α -chain. Hybridoma were stimulated for 2 h with A20 lymphoma cells transfected or not with mouse CD1d in the presence or not of 200 ng/mL of the antigen PBS57. Following stimulation the levels of Egr-2 were measured by intracellular staining. The percentage of Egr-2⁺ cells in each condition is shown. Results are representative of two independent experiments.