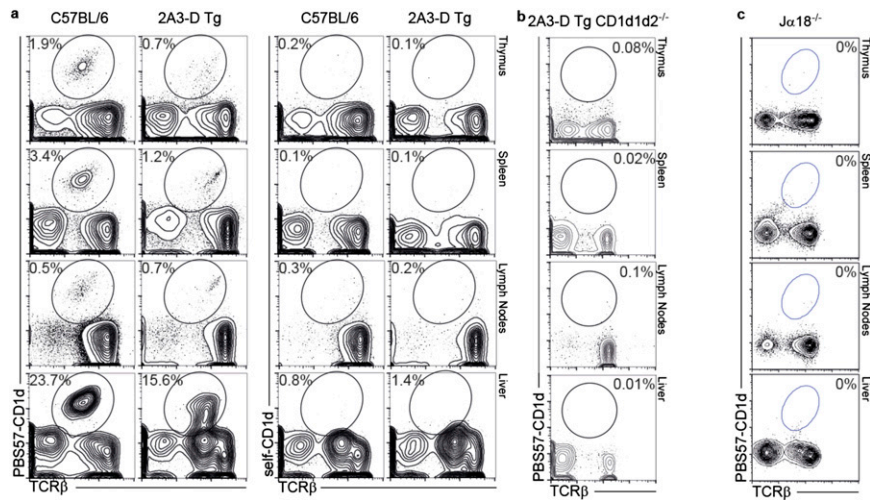
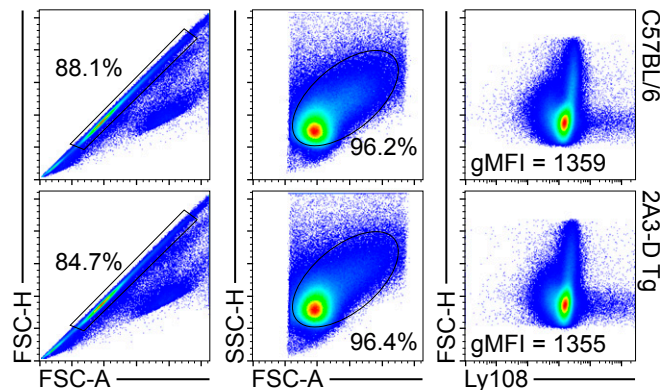


# Supporting Information

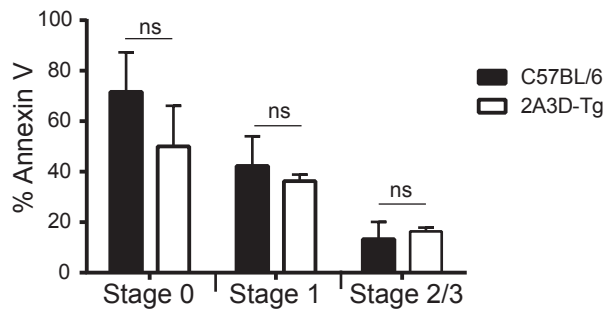
Bedel et al. 10.1073/pnas.1320777110



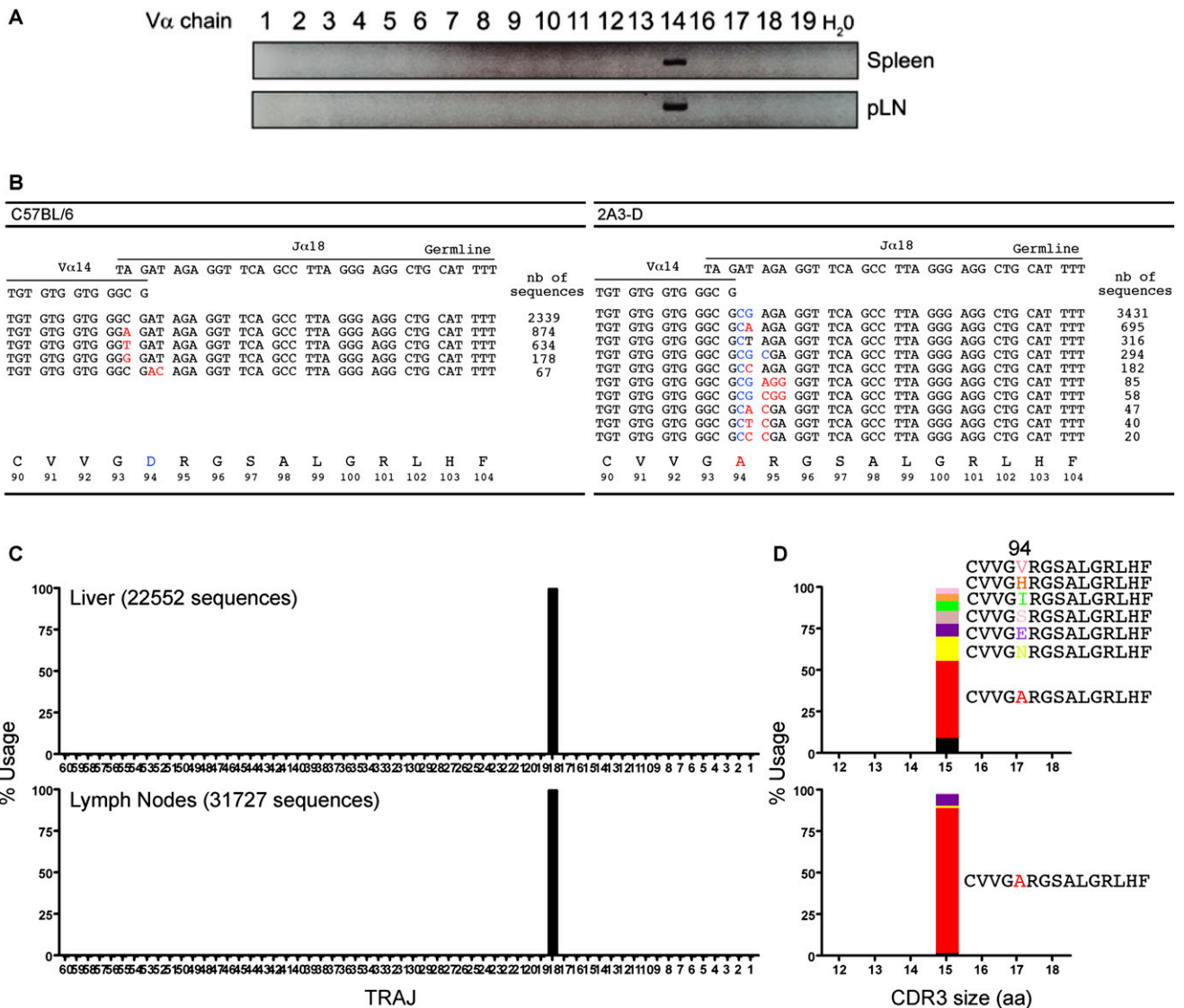
**Fig S1.** Invariant NK T cells (iNKT) cells in 2A3-D Tg mice depends on CD1d for thymic development. (A) Purified lymphocytes from the thymus, spleen, liver, and peripheral lymph nodes (pLN) of C57BL/6 or 2A3-D Tg mice were stained for indicated markers to assess reactivity of iNKT cells to self-CD1d and PBS57-CD1d (data representative of  $n = 3$ ). (B) Purified lymphocytes from the thymus, spleen, liver, and pLN of 2A3-D Tg CD1d1d2<sup>-/-</sup> mice were stained for indicated markers to confirm that iNKT cell positive selection is still restricted to CD1d (data representative of  $n = 2$ ). (C) Purified lymphocytes from the thymus, spleen, liver, and pLN of Jα18<sup>-/-</sup> mice were stained for indicated markers to confirm the specificity of iNKT cell staining (data representative of  $n = 2$ ).



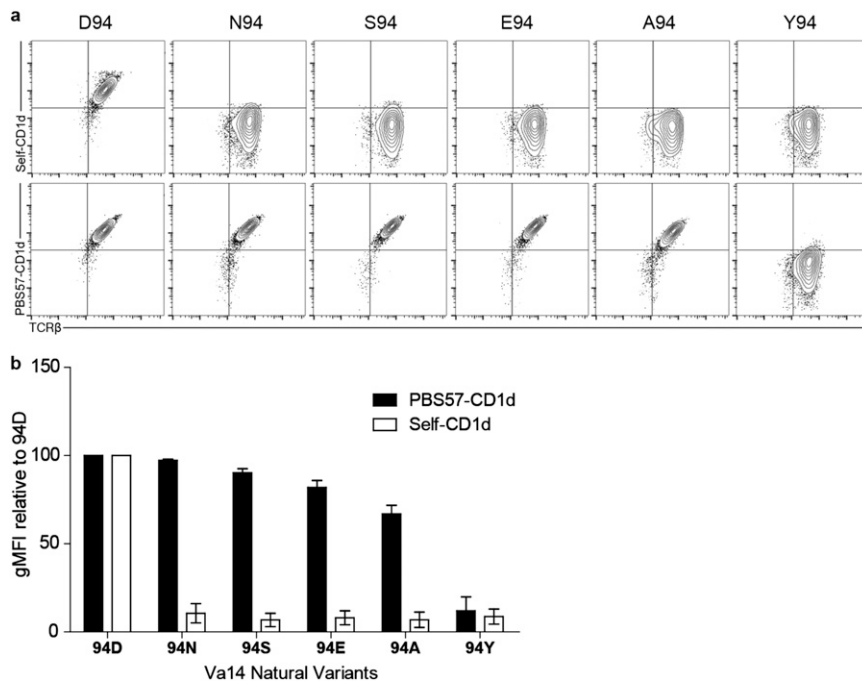
**Fig S2.** Ly108 expression levels are similar on C57BL/6 and 2A3-D Tg thymocytes. Purified lymphocytes from the thymus of C57BL/6 or 2A3-D Tg mice were stained for indicated markers to assess Ly108 expression level. The gating strategy to obtain total thymocytes is depicted and Ly108 gMFI (geometric mean fluorescence intensity) is indicated as a representative value of  $n = 3$ .



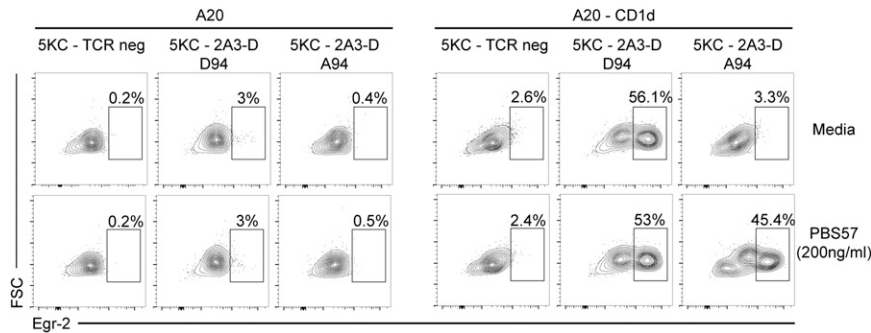
**Fig S3.** 2A3-D Tg NKT cells do not express more Annexin V compared with wild-type control. Total thymocytes were stained for CD24 and CD44 to define the different developmental stages. Here, the stages are defined as follows: stage 0 ( $CD24^{high}CD44^{low}$ ), stage 1 ( $CD24^{low}CD44^{low}$ ), stage 2/3 ( $CD24^{low}CD44^{high}$ ). Total thymocytes were stained for Annexin V and 7-ADD to assess apoptosis and necrosis respectively (data representative of  $n = 2$ ). ns, not significant.



**Fig S4.** Complementary data for Fig. 5. (A) PB557-CD1d tetramer-positive T-cell receptor (TCR)- $\beta^+$  cells were sorted from the spleen and peripheral lymph nodes of 2A3-D Tg mice. mRNA was extracted and transformed into cDNA. cDNAs were amplified by PCR with V $\alpha$ -specific forward primers and a C $\alpha$ -specific reverse primer. (B) Sequences of the V $\alpha$ 14-J $\alpha$ 18 rearrangements found in iNKT cells from the spleen of C57BL/6 and 2A3-D Tg mice. For each observed rearrangement, the number of sequences, contribution of the V $\alpha$ 14 and J $\alpha$ 18 chain and existence of p-addition (blue) or n-addition is depicted (data representative of  $n = 2$ ). (C) PCR analysis was performed to evaluate the use frequency of TCR- $\alpha$  joining (TRAJ) genes encoding productive, in-frame rearrangements with the TRAV11 family in iNKT cells from the liver or the pLN or C57BL/6 or 2A-D Tg mice. (D) Amino acid composition and size of the CDR3 of the  $\alpha$ -chain in C57BL/6 or 2A3-D Tg in iNKT cells from the liver and pLN.



**Fig 55.** Specific loss of self-reactivity in Vα14 natural variants paired with 2A3-D Tg Vβ. (A) 5KC Hybridoma expressing 2A3-D Tg Vβ were transduced with indicated Vα14 natural variants and tested for reactivity with self-CD1d and PBS57-CD1d tetramers (data representative of  $n = 4$ ). (B) For each 5KC hybridoma expressing a Vα14 natural variant, the gMFI of the PBS57-CD1d or self-CD1d tetramer staining was evaluated for a narrow slice of TCR expression. Relative percentage of this gMFI compared with the gMFI of the D94 variant with PBS57-CD1d and self-CD1d is shown ( $n = 3$ ).



**Fig 56.** Hybridomas expressing the D94A Vα14 natural variant paired with 2A3-D Tg Vβ do not induce Egr-2 upon autoreactive response to CD1d-transfected A20 cells. The TCR<sup>-</sup> 5KC hybridoma was transduced with the 2A3-D β-chain paired either with the D94 wild-type Vα14 iNKT chain or the A94 variant α-chain. Hybridomas were stimulated for 2 h with A20 lymphoma cells transfected or not with mouse CD1d in the presence or not of 200 ng/mL of the antigen PBS57. Following stimulation the levels of Egr-2 were measured by intracellular staining. The percentage of Egr-2<sup>+</sup> cells in each condition is shown. Results are representative of two independent experiments.