

Y. Zhang<sup>1</sup>, Y. Song<sup>1</sup>, S. Ravindran<sup>1</sup>, Q. Gao<sup>1</sup>,  
C.C. Huang<sup>1</sup>, A. Ramachandran<sup>1</sup>, A. Kulkarni<sup>2</sup>,  
and A. George<sup>1\*</sup>

<sup>1</sup>Brodie Tooth Development Genetics & Regenerative Medicine Research Laboratory, Department of Oral Biology, University of Illinois at Chicago, Chicago, IL 60612, USA; and <sup>2</sup>NIDCR, NIH, Bethesda, MD, USA; \*corresponding author, anneg@uic.edu

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## APPENDIX

### MATERIALS & METHODS

#### Exosome Isolation from Culture Medium

Supernatant was collected from confluent T4-4 cells and centrifuged at 400 *g* at 4°C for 5 min to remove debris. The cleared supernatants were processed with an Exosome extraction kit (Life Technologies, Grand Island, NY, USA) according to the manufacturer's protocol. The final pellet was re-suspended in 200  $\mu$ L of PBS for immune transmission electron microscopy (TEM) and Western blot analysis.

#### Exosome Staining

Exosomes were mounted on carbon-/formvar-coated nickel grids (Electron Microscopy Sciences, Hartfield, PA, USA) fixed with 4% neutral formalin in PBS for 5 min and washed. They were permeabilized with 0.1% Triton X 100 in PBS and blocked with 5% BSA. Exosomes were labeled with rabbit anti-dentin phosphophoryn (DPP) or anti-CD-63 (Santa Cruz Biotechnology, Santa Cruz, CA, USA). They were washed and incubated with 20 nm gold-conjugated goat anti-rabbit antibody at a 1:100 dilution. The grids were washed and negatively stained with 1% aqueous uranyl acetate and viewed with a JEOL TEM at 80 kV.

**Appendix Table.** Mouse DSPP Primers for Subcloning into pDsRED-GFP and pRF

Primer	Sequence
Region 1 72 bp of DSP and 24 bp of DGP	-forward ATACTAGTGACACCCGAGATGCAGAG -reverse ACGAATTCGTTGGGACCTTCAGTTTC
Region 2 126 bp of DSP and 90 bp of DGP	-forward GCACTAGTGAAGAAGGCGACAGTACC -reverse ACGAATTCCTCCATTGCTATCTTTACT
Region 3 126 bp of DSP	-forward GCACTAGTGAAGAAGGCGACAGTACC -reverse ACGAATTCCTGATCTTGGCTCTTCCCA
Region 4 267 bp of DGP	-forward CACTAGTGAATAGAACTGAAGGTCCCAAC -reverse ACGAATTCCTTGCATGGACTCGTC
DSP+DGP (without signal peptide)	-forward ACTAGTATCCGGTCCCCAGTTAGTACCA -reverse ACGAATTCCTTGCATGGACTCGTC
DSPP (without signal peptide and stop codon)	-forward ACTAGTATCCGGTCCCCAGTTAGTACCA -reverse ACTAGTATCATCACTGGTTGAGTGGTTACTGTC

DSSP=dentin sialophosphoprotein; DGP= dentin glycoprotein; DSP=dentin sialoprotein.

# DSPP Contains an IRES Element Responsible for the Translation of Dentin Phosphophoryn

## Kidney Capsule Protocol

Mandibular molars were isolated from E13.5 dentin sialophosphoprotein (DSPP) null mouse embryos (a kind gift from Dr. Ashok Kulkarni). They were then enzyme-digested and dissociated into single-cell suspensions. Isolated cells were cultured as single layers in dishes with 10% FBS DMEM media (Song *et al.*, 2006). These cells were transfected with replication-incompetent lentivirus media containing DPP. The transfected cells were pelleted and recombined with the dental epithelium isolated from DSPP knock-out E10.5 mouse embryos.

## Immunohistochemistry

Extracellular matrix (ECM) was isolated as previously published, and immunohistochemical analysis was performed for the localization of DSP and DPP (Ravindran *et al.*, 2012). The experimental protocol used was according to published protocols with anti-DPP (1:200) (rabbit polyclonal) and anti-dentin sialoprotein (DSP), (1:1,000) (monoclonal antibody, a kind gift from Dr. Chunlin Qin).

## APPENDIX REFERENCES

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