

Figure S1: Simplified pathway of wax biosynthesis in Arabidopsis (redrawn from Greer *et al.* [16] and Samuels *et al.* [61]). The enzyme mid-chain alkane hydroxylase (MAH1) which, according to Greer *et al.* [16], catalyzes the enzymatic conversations of alkanes to secondary alcohols and ketones is highlighted in red.



Figure S2: Multiple Alignment of MLOC_15925.1, HO07G08 and re-sequenced *CYP96B22* construct used in VIGS experiments. The sequence of HO07G08 was taken from the IPK Crop EST database (http://pgrc.ipk-gatersleben.de/cr-est/) and used as BLASTN query against the transcript sequences published by the International Barley Sequencing Consortium [18]. The incomplete identity between MLOC_15925.1 and HO07G08 might be due to differences between barley cultivars or inaccuracies in the original sequencing of HO07G08 [27] or MLOC_15925.1. Identical parts of sequences are highlighted in yellow.

inoculation

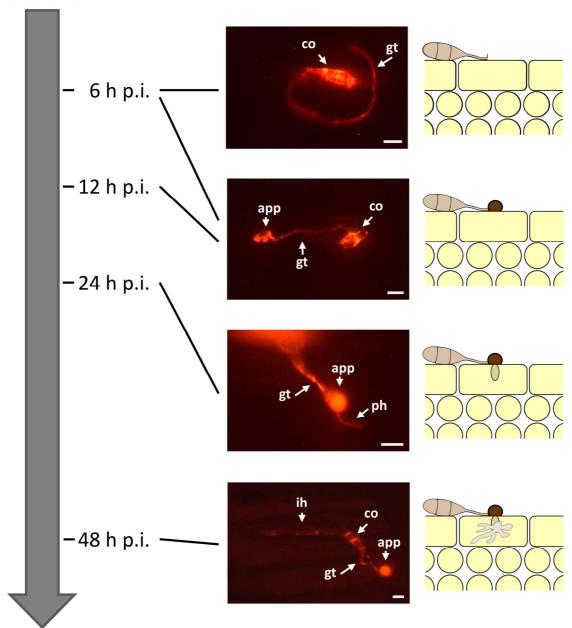


Figure S3: Time course study of the development of infection stages of *M. oryzae* on barley. Transgenic fungi labeled with DsRed were generated for the *M. oryzae* host isolate TH6772 and inoculated onto barley primary leaves. At 6 h after inoculation (h p.i.) germinated spores were found on the leaf surface some of which had already developed appressoria. At 12 h p.i. almost all germinated conidia had formed appressoria and at 24 h p.i. penetration hyphae got visible beneath appressoria which further developed into larger invasive hyphae detected around 48 h p.i. A similar kinetic was observed for the development of infection structures of a transgenic, GFP-expressing, nonhost *Magnaporthe* isolate (CD180), however in this case invasive hyphae were only rarely detected (data not shown). scale bar: 10 μm; co: conidium; gt: germ tube; app: appressorium; ph: penetration hypha; ih: bulbous invasive hypha

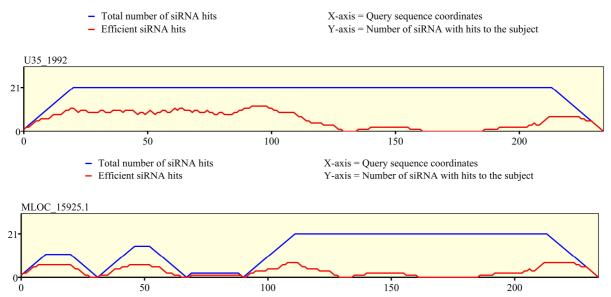


Figure S4: Prediction of targets in the transcriptome of barley by the siRNA used in this study. The target sequence for the *CYP96B22* silencing construct was predicted using the si-Fi software (http://labtools.ipk-gatersleben.de/). The 234 bp fragment of *CYP96B22* was used as a query to be scanned for potential generated siRNA and the predicted siRNA sequences were then checked for potential targets using the HarvEST database (assembly 35, http://harvest.ucr.edu/) or the high-confidence barley coding sequences [18]. In both databases a unique coding sequence was identified as potential target, both representing *CYP96B22* as deposited in HarvEST (U35_1992, http://harvest.ucr.edu/) or by Meyer *et al.* [18] (MLOC_15925.1).

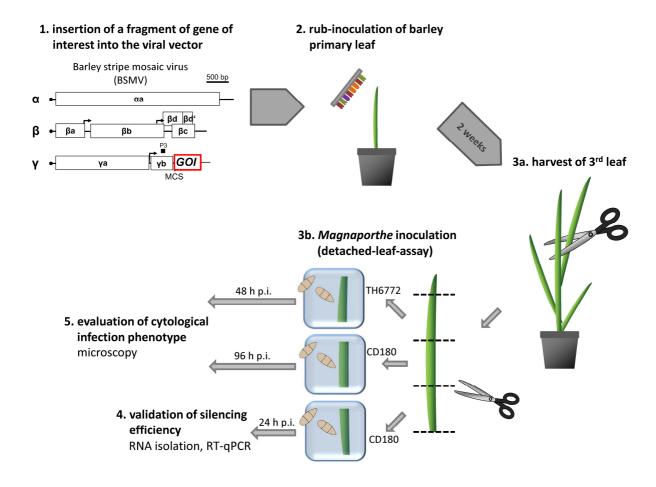


Figure S5: Experimental design of VIGS experiments.

A 234 bp fragment of *CYP96B22* was inserted into the γ -subunit of BSMV (1) (picture modified according to [59]). *In vitro* generated viral transcripts were rub-inoculated on barley primary leaves (2). Two weeks later the third leaf of plants showing viral disease symptoms was harvested and cut into three pieces, two of which were inoculated with the *Magnaporthe* nonhost isolate CD180 and one with the host isolate TH6772. Microscopic assessment of plant defense responses was done at time points indicated. Silencing efficiency was monitored by RT-qPCR using one leaf piece harvested at 24 h p.i. with CD180.