

B Cell-Specific Loss of Lyn Kinase Leads to Autoimmunity

Chrystelle Lamagna^{*}, Yongmei Hu^{*}, Anthony L. DeFranco[†] and Clifford A. Lowell^{*, 2}

^{*} Departments of Laboratory Medicine and [†] Microbiology and Immunology, University of California, San Francisco, CA, 94143 USA

¹ This work was supported by the National Institutes of Health (AI65495 and AI68150 to C.A.L.; AI078869 to A.L.D).

² Address correspondence to Dr Clifford A. Lowell, University of California at San Francisco, Department of Laboratory Medicine, 513 Parnassus Avenue, Box 0451, San Francisco, CA 94143, USA. E-mail: clifford.lowell@ucsf.edu
Tel: 415-4762963; Fax: 415-5025462

³ Abbreviations used in this paper: SFK, Src family kinase; $[Ca^{2+}]_i$, intracellular Ca^{2+} concentration; T1, transitional stage 1; T2, transitional stage 2; T3, transitional stage 3; Fol., Follicular; MZ, marginal zone; GC, germinal center; Ig, immunoglobulins

Running title: B cell-specific Lyn deletion causes autoimmunity

Supplemental Materials

Figure S1: Generation of B cell-specific Lyn-deficient mice.

(A) Representative dot plots showing Lyn protein expression levels determined by intracellular flow cytometry analysis of peripheral blood leukocytes of the indicated lineages (2 month-old mice).

(B) Percentage of Lyn⁺ cells in the indicated peripheral blood populations of 2 month-old mice. Data represent mean \pm SEM of independent experiments from 5-8 mice per group. ** $P \leq 0.01$, *** $P \leq 0.001$ (One-way ANOVA).

Figure S2: Representative labeling and gating strategies used to analyze FACS data and define the different B cell subpopulations in the spleen.

Representative dot plots from spleen of 2 month-old WT mice. The numbers in gates represent the frequency of gated cells. Cells were first defined as DAPI⁻ and PerCP-Cy5.5⁻ population, PerCP-Cy5.5 being used to exclude CD11b⁺ and TCR β ⁺ cells. Subsequent gating allowed the identification of the following strategies B cell populations.

(A) Plasma cells were identified as B220^{lo/-} CD138^{hi} cells.

(B) Gating on the B220⁺ IgD⁻ population, followed by subsequent gating on Fas⁺ GL7⁺ cells allowed the identification of GC B cells.

(C) Gating strategies used to identify transitional T1, T2 and T3 cells, as well as mature follicular (I and II) and MZ B cells. The different subsets were defined as follow:

T1: CD19⁺ IgD^{lo} IgM^{hi} CD93⁺ CD23⁻ CD21⁻

T2: CD19⁺ IgD^{lo} IgM^{hi} CD93⁺ CD23⁺ CD21⁻

T3: CD19⁺ IgD^{hi} IgM^{lo} CD93⁺ CD23⁺ CD21^{lo}

MZ: CD19⁺ IgD^{lo} IgM^{hi} CD93⁻ CD23⁻ CD21⁺

Fol. I: CD19⁺ IgD^{hi} IgM^{lo} CD93⁻ CD21^{lo}

Fol. II: CD19⁺ IgD^{hi} IgM^{hi} CD93^{lo/-} CD21^{lo}

Figure S3: *Lyn*^{-/-} and *B-lyn*^{-/-} display altered architecture of the lymphoid tissues

H&E staining of spleen (upper panel) and lymph node (lower panel) sections from 8 month-old control, *lyn*^{-/-} and *B-lyn*^{-/-} mice. Representative pictures are shown. Scale bars represent 200 μm.

Figure S1 - Lamagna C. *et al.*

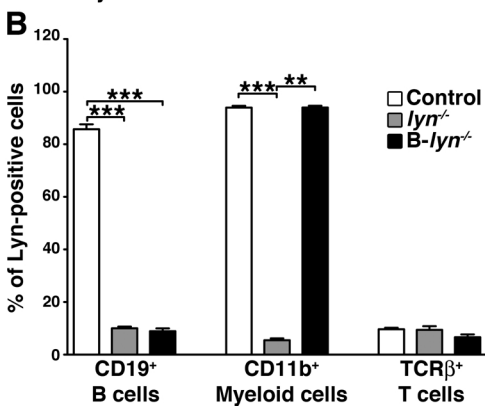
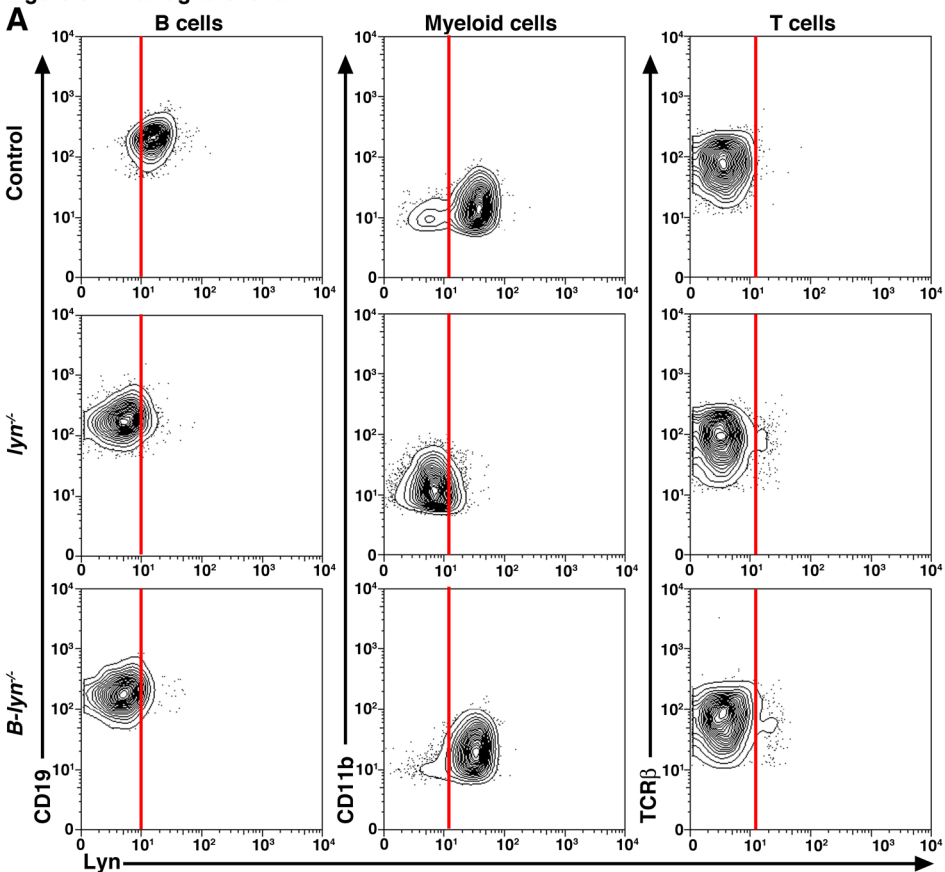


Figure S2 - Lamagna C. *et al.*

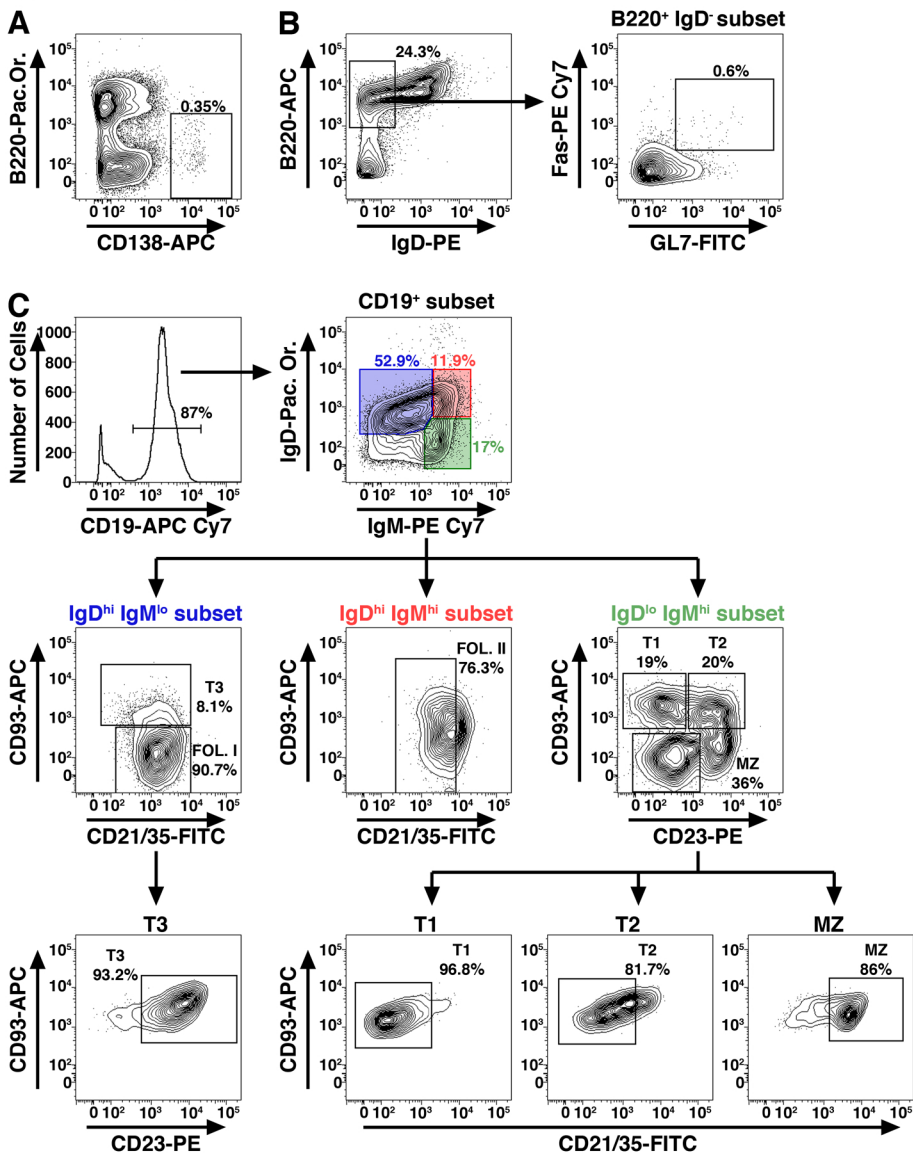


Figure S3 - Lamagna C. *et al.*

