

Supplementary Table 1.**Primers Used for constructs.**

Primer Name	Sequence 5'-3'	Specification
IdeRRev524	GCTAGCTGAACGTGCTCGGTA	3' end of <i>ideR</i>
IdeRFw4	ATGCATAACGAGTTGGTTGATA	5' end of <i>ideR</i>
PptrIdeRFw7	ACACGAGGCCCTTTTCGTCTTTCAA	Forward primer for confirmation of <i>ideR</i> conditional mutant
PptrIdeRRev900	TCAGACTTTCTCGACCTTGACCGC	Reverse primer for the confirmation of <i>ideR</i> conditional mutant
BfrBpSM232Fw	AGACAATTGCGGATCCATGACAGAAT ACGAAGGGCCTAAGAC	Forward in fusion primer to amplify <i>bfrB</i> and BamHI site of pMV261.
BfrBpSM232Rev	TTCTGCAGCTGGATCCAACGGGACC ACTCGCTGAT	Reverse infusion primer to amplify <i>bfrB</i> and BamHI site of pMV261
BfrA Fw pMV261	GACAATTGCGGATCCGCCACGACAG	Forward infusion primer to amplify <i>bfrA</i> BamHI site of pMV261
BfrARev pMV261	TTCTGCAGCTGGATCCAGTGGTATCG	Reverse infusion primer to amplify <i>bfrA</i> gene and BamHI site of pMV261

Supplementary Table 2.

Primers used for qRT-PCR

Primer name	Sequence 5'-3'	Specification
16S UP#2	ATGACGGCCTTCGGGTTGTTA	Forward primer for 16S RNA
16S Rev#2	CGGCTGCTGGCACGTAGTTG	Reverse primer for 16S RNA
Rv2383cF	GCGACTTTCCCATCAGTGTT	Forward primer for <i>mbtB</i>
Rv2383cR	TAATAACGTCAACGCAAGTTCCG	Reverse primer for <i>mbtB</i>
Rv1348F	TCTATCTGTTCGTGGACTG	Forward primer for <i>irtA</i>
Rv1348R	CTTCGCCGTTTCATCTTCTCT	Reverse primer for <i>irtA</i>
Rv3841F244	CGAAACCAGTTCGACAGACC	Forward primer for <i>bfrB</i>
Rv3841R546	GCAAGAACCACTGCATGAAC	Reverse primer for <i>bfrB</i>

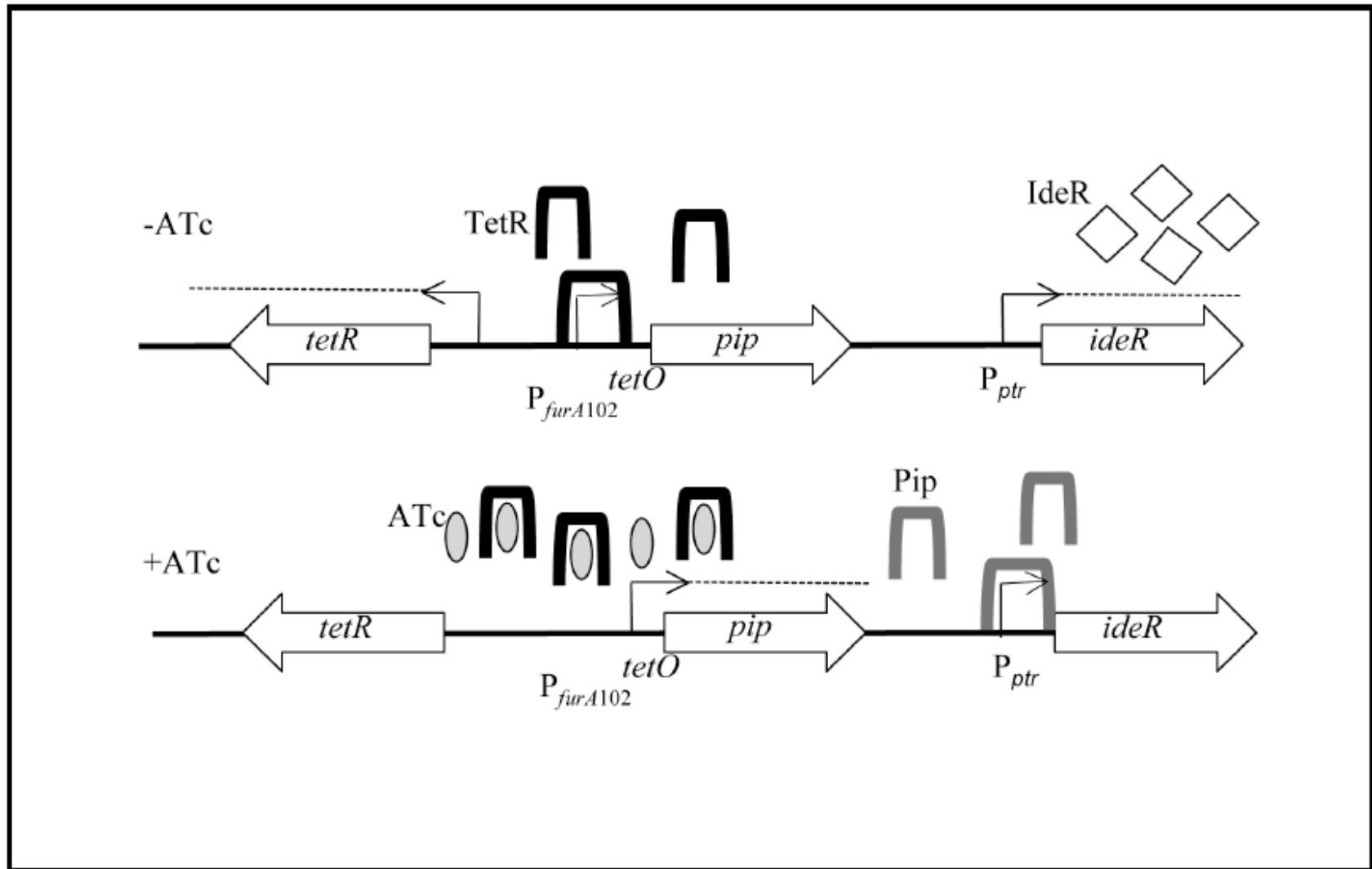


Figure S1. Control of *ideR* expression in ST217. The native *ideR* promoter was replaced by a Pip repressible promoter by homologous recombination as described in Materials and Methods in TB38.2 which has the *tetR* and *pip* integrated in the chromosome. In the absence of tetracycline (ATc), TetR binds to its operator repressing *pip* transcription and allowing *ideR* expression. In the presence of ATc, *pip* is transcribed and Pip represses *ideR* expression.

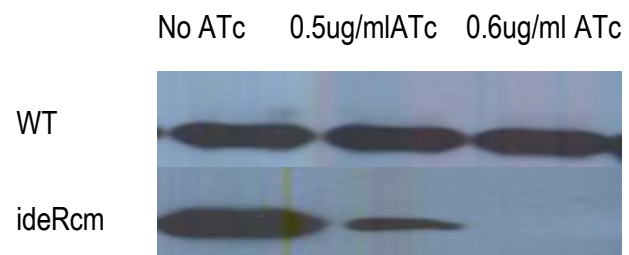
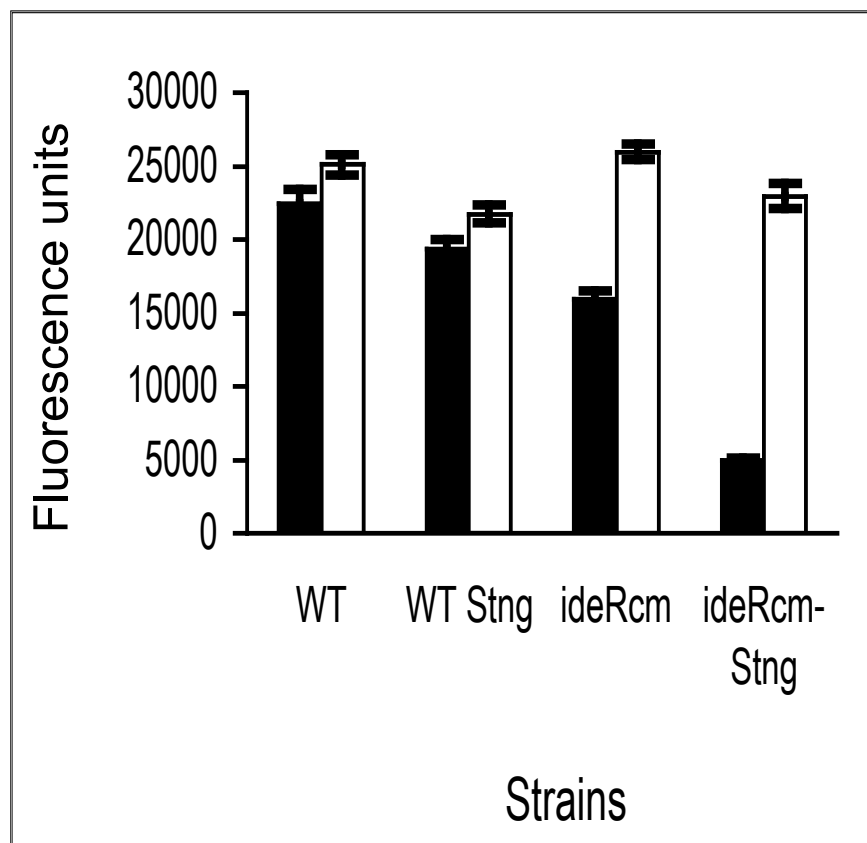


Figure S2. IdeR protein in the wild type and the *ideR* conditional. Wild type and the *ideR* conditional mutant were grown in 7H9. Protein extracts were obtained as described in Materials and Methods. Levels of IdeR were detected by Immunoblotting using anti-IdeR antibodies.

A



B

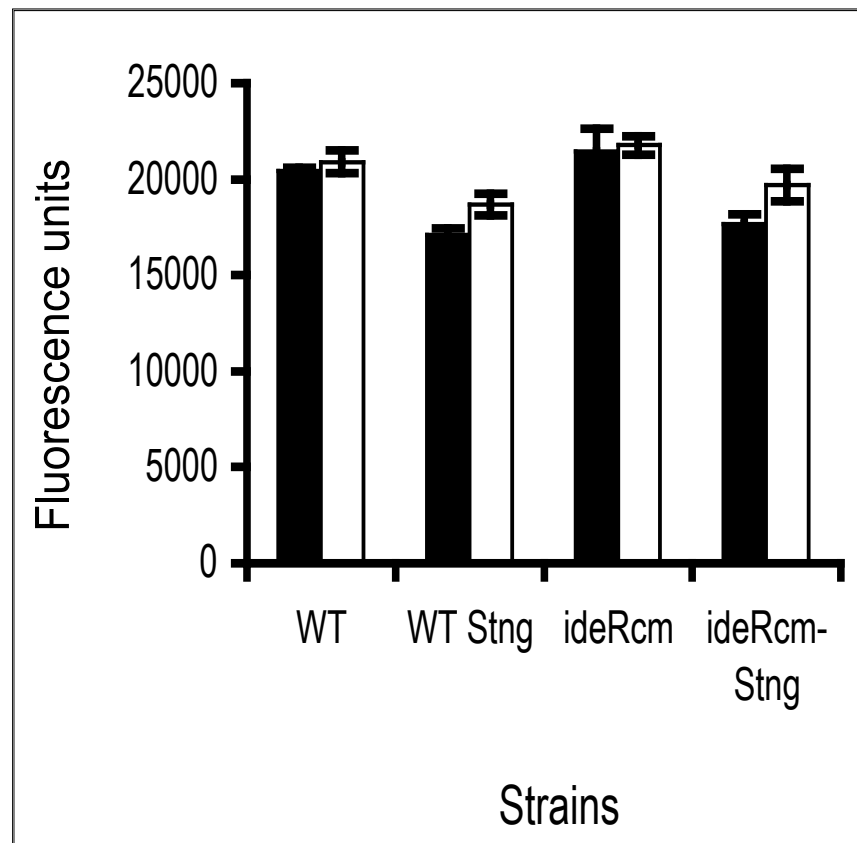


Figure S3. Sensitivity to Streptonigrin. The *ideR* conditional mutant (*ideRcm*) and the parental wild type strain (WT) were cultured in HIMM(A) or LMM (B) with ATc (dark bars) or without ATc (open bars) and with and without 0.6 $\mu\text{g}\cdot\text{ml}^{-1}$ streptonigrin (Stng). The data corresponds to the fluorescence resulting from reduction of Alamar Blue by each strain. Shown are mean values \pm standard deviations of three biological replicates.

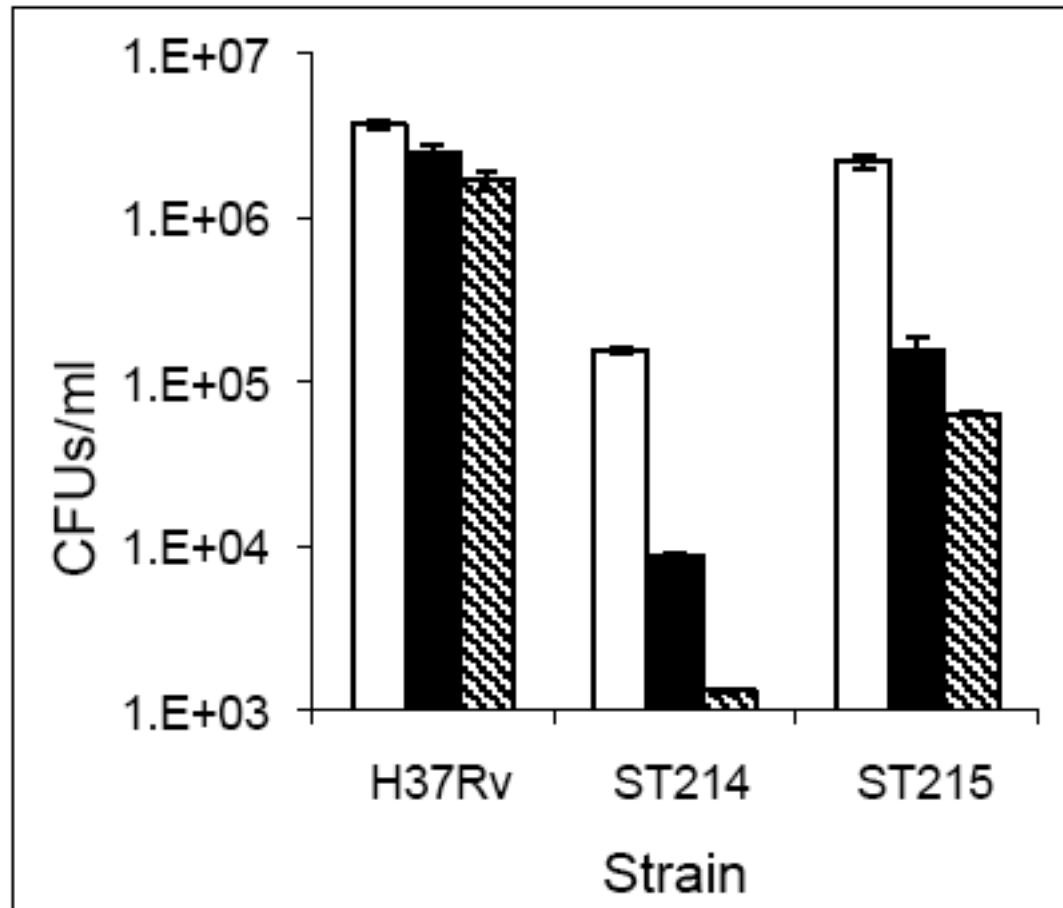


Figure S4. Sensitivity of the *bfrB* and *bfrA* mutants to nitrite. The *bfrB* (ST214) and *bfrA* (ST215) mutants and the parental strain H37Rv were exposed to 0 (open bars), 3 mM (dark bars) or 5mM (striped bars) of sodium nitrite at pH 5.5 Each day a sample was taken and dilutions plated onto 7H10. The data shows the mean \pm standard deviations of the number of CFUs recovered from biological triplicates of each strain at day three.