

table S1: strains and plasmids

Strain or plasmid	Description	Reference
Strains		
<i>E. coli</i>		
TOP10	Competent <i>E. coli</i> for plasmid transformation	Invitrogen
BL21	BL21(DE3)pLysS strain for protein expression	Promega
BL21-121	BL21 with <i>relP</i> gene under P _{IPTG} control, amp	This work
BL21-122	BL21 with <i>relQ</i> gene under P _{IPTG} control, amp	This work
<i>S. aureus</i>		
RN4220	Restriction deficient <i>S. aureus</i> strain, r ⁻	(1)
RN4220-55	RN4220 with <i>rsh</i> gene under P _{spac} control in the chromosome, <i>ermC</i>	(2)
HG001	<i>rsbU</i> restored RN1 (8325), previously named RN1HG	(3, 4)
HG001-55	HG001 with <i>rsh</i> gene under P _{spac} control in the chromosome, <i>ermC</i>	(2)
HG001-86	HG001 <i>rsh</i> _{Syn} (Δ942-950nt)	(2)
HG001-229	HG001 <i>relP</i> (Δ450-536nt)	This work
HG001-230	HG001 <i>relQ</i> (Δ343-429nt)	This work
HG001-229-230	HG001 <i>relP/relQ</i> double mutant	This work
HG001-229-230-263	HG001 <i>rsh/relP/relQ</i> triple mutant referred as (p)ppGpp ⁰	This work
USA300JE2 NE554	<i>vraR</i> ::Tn bursa aurealis mutant	(5)
HG001 NE554	<i>vraR</i> ::Tn bursa aurealis mutant	This work
Plasmids		
pET15b	Protein expression vector, P _{IPTG} , amp	Novagen
pMUTIN4	Integrative vector including the IPTG-inducible promoter P _{spac} , (Ap ^r , Em ^r)	(6)
pKOR1	AHT inducible suicide mutagenesis vector	(7)
pCG86	pKOR1 with mutated <i>rsh</i> synthase domain (Δ942-950nt)	(2)
pCG263	pKOR1 with mutated <i>rsh</i> (Δ249-951nt)	This work
pCG229	pKOR1 with mutated <i>relP</i> synthase (Δ450-536nt)	This work
pCG230	pKOR1 with mutated <i>relQ</i> synthase (Δ343-429nt)	This work
pCG121	<i>relP</i> gene in protein expression vector pET15b	This work
pCG122	<i>relQ</i> gene in protein expression vector pET15b	This work
pCG248	anhydrotetracycline (ATc) inducible vector	(8, 9)
pCG258	ATc inducible vector with <i>relP</i>	(8, 9)
pCG259	ATc inducible vector with <i>relQ</i>	(8, 9)
pCG328	ATc inducible vector with <i>rsh</i> _{Hyd}	(8, 9)

References:

1. **Kreiswirth BN, Lofdahl S, Betley MJ, O'Reilly M, Schlievert PM, Bergdoll MS, Novick RP.** 1983. The toxic shock syndrome exotoxin structural gene is not detectably transmitted by a prophage. *Nature* **305**:709-712.
2. **Geiger T, Goerke C, Fritz M, Schafer T, Ohlsen K, Liebeke M, Lalk M, Wolz C.** 2010. Role of the (p)ppGpp synthase RSH, a RelA/SpoT homolog, in stringent response and virulence of *Staphylococcus aureus*. *Infect. Immun.* **78**:1873-1883.
3. **Pohl K, Francois P, Stenz L, Schlink F, Geiger T, Herbert S, Goerke C, Schrenzel J, Wolz C.** 2009. CodY in *Staphylococcus aureus*: a regulatory link between metabolism and virulence gene expression. *J. Bacteriol.* **191**:2953-2963.
4. **Herbert S, Ziebandt AK, Ohlsen K, Schafer T, Hecker M, Albrecht D, Novick R, Gotz F.** 2010. Repair of global regulators in *Staphylococcus aureus* 8325 and comparative analysis with other clinical isolates. *Infect. Immun.* **78**:2877-2889.
5. **Fey PD, Endres JL, Yajjala VK, Widhelm TJ, Boissy RJ, Bose JL, Bayles KW.** 2013. A genetic resource for rapid and comprehensive phenotype screening of nonessential *Staphylococcus aureus* genes. *MBio* **4**:e00537-00512.
6. **Vagner V, Dervyn E, Ehrlich SD.** 1998. A vector for systematic gene inactivation in *Bacillus subtilis*. *Microbiology* **144 (Pt 11)**:3097-3104.
7. **Bae T, Schneewind O.** 2006. Allelic replacement in *Staphylococcus aureus* with inducible counter-selection. *Plasmid* **55**:58-63.
8. **Schroder W, Goerke C, Wolz C.** 2013. Opposing effects of aminocoumarins and fluoroquinolones on the SOS response and adaptability in *Staphylococcus aureus*. *The Journal of antimicrobial chemotherapy* **68**:529-538.
9. **Helle L, Kull M, Mayer S, Marincola G, Zelder ME, Goerke C, Wolz C, Bertram R.** 2011. Vectors for improved Tet repressor-dependent gradual gene induction or silencing in *Staphylococcus aureus*. *Microbiology* **157**:3314-3323.

table S2: oligonucleotides

Purpose and Description	Template	Name	Sequence
Mutagenesis <i>relP</i> mutant	H G001	attB1relP2-for	GGGGACAAGTTTGTACAAAAA GCAGGCTTGGTCACTATCACA CGAA
		relPmut-rev	ACTTGCCACATATCCATACCT ATCCTAGGACCATTTCTTTAG GGTGCTGAAT
		relPmut-for	ATAGGTATGGATATGTGGGCA GGGGACCACTTTGTACAAGAA AGCTGGGTTGGGCGAGTATTT AGCAA
		attB2relP2-rev	GGGGACAAGTTTGTACAAAAA GCAGGCTTAAAGGTTGATGAT GTTCCG
<i>relQ</i> mutant	HG001	attB1relQ-for	CGTTGCCAGAAATTCATTGCT AACCTAGGACCACTTTCTTTAG TGTTACGAAT
		relQmut-rev	TTAGCAATGAATTTCTGGGCA GGGGACCACTTTGTACAAGAA AGCTGGGTCGCGGTAAATGAA TTTCTAA
		relQmut-for	GGGGACAAGTTTGTACAAAAA GCAGGCTATTAGGCGGTATCG TAGT
		attB2relQ-rev	GCCTACTACTGTAGTATGCAAC AAAAACCTGCGACAAT TTGCATACTACAGTAGTAGGC GGGGACCACTTTGTACAAGAA AGCTGGGTGCTGCGAATAAAT CATCTT
Construction of hybridization probes <i>vraR</i> <i>relP</i> <i>relQ</i>	HG001 HG001 HG001 HG001 HG001 HG001	vraRDIG-for	GGAATTTCAAGTTATCTATCAA
		vraRDIG-rev	TGAGTAACCTTTTCGCAATCA
		relPDIG-for	GTCGCACATTCTTTCAGT
		relPDIG-rev	CGTTATTAGGTTTCGTAGAGTT
		relQDIG-for	AAGCGGTTGATGAGTTGA
		relQDIG-rev	AAATACGCTGCTTCTGCC

<i>rsh</i>	HG001 HG001	rshDIG-for rshDIG-rev	CGGCTCTTCGTTATATTGATAA GGCAACTCAATAACATCAC
Protein purification			
RelP	HG001 HG001	XhoI pETrelP- for BamHI pETrel P-rev	CCCCCTCGAGATGTATGTAGAT CGAAAACCAT GGGGGGATCCCTACTCTGTTA TTTCAGAAATGAA
RelQ	HG001 HG001	XhoI pETrelQ- for BamHI pETrel Q-rev	GGGGCTCGAGATGAATCAATG GGATCAGTT GGGGGGATCCCTTAATCATTTTC ATGTTTTTT
ATc inducible plasmids			
pRelP	HG001 HG001	SallrelPfor SallrelPprev	CCGGGTCGACAAAATGTTAGGTA CAATAAAGA CCGGGTCGACGAACACACCTACT AACAT
pRelQ	HG001 HG001	SallrelQfor SallrelQrev	AATTGTCGACTGCAACCGTTGAA GAAACAT TTAAGTCGACCGCATGATTTATAA CACCTCG
pRSH _{Hyd}	HG001 HG001 HG001 HG001	EcoRIrel115 1 HDreplaceA Srev HDreplaceA Sfor EcoRIrelrev	AAAAGAATTCGTACCTAAATCATT GTTTAAGGCG ATACGGTGTATCTTCAATTACGGA GGCCAAAAAACCTGC GTAATTGAAGATACACCGTAT CCCCGAATTCCTCGATACCGACTA ATAACAATA

Underlined: artificial restriction sites

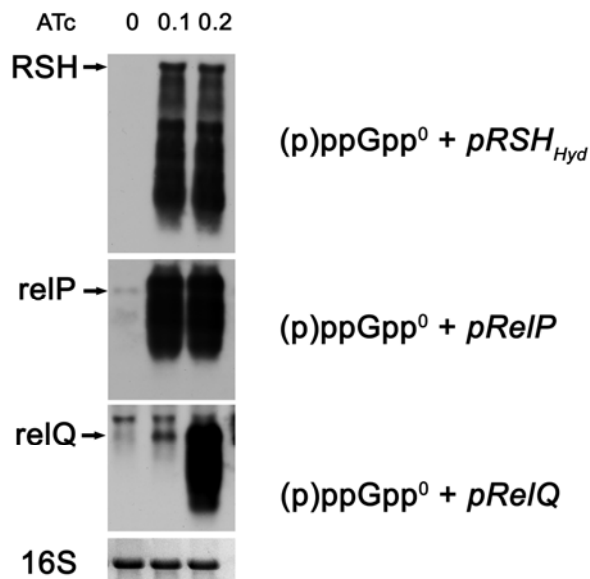


Figure S1: Overexpression of the three (p)ppGpp Synthases.

(p)ppGpp⁰ mutants complemented with *pRSH_{Hyd}*, *pRelP*, or *pRelQ* synthases were grown in CYPG to the exponential growth phase followed by further incubation with and without ATc ($\mu\text{g/ml}$) for one hour. RNA was hybridized with digoxigenin-labeled PCR fragments. The 16S rRNA detected in the ethidium bromide-stained gels is indicated as loading control in the lower lane.