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Supporting Information

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Rapid and Specific Post-Synthesis Modification of DNA through a Biocompatible Condensation of 1,2-Aminothiols with 2-Cyanobenzothiazole

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Chaofeng Dai,* and Binghe Wang*^[a]**

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A Novel Method for the Rapid and Specific Post-synthesis Modification of DNA via a Bio-orthogonal Condensation of 1,2-Aminothiols with 2-Cyanobenzothiazole †

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Supporting Information

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1. General Information

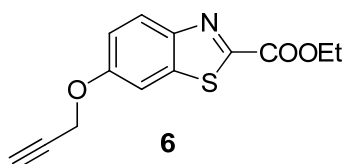
Solvents and reagents were purchased from VWR International, Oakwood Product Inc., or Sigma-Aldrich Co. and used without purification unless specified otherwise. When necessary, solid reagents were dried under high vacuum. Reactions with compounds sensitive to air or moisture were performed under argon. Solvent mixtures are indicated as volume/volume ratios. Thin layer chromatography (TLC) was run on Sorbtech

W/UV254 plates (0.25 mm thick), and visualized under UV-light or by a Ce-Mo staining solution (phosphomolybdate, 25 g; Ce(SO₄)₂·4H₂O, 10 g; conc. H₂SO₄, 60 mL; H₂O, 940 mL) with heating. Flash chromatography was performed using Fluka silica gel 60 (mesh size: 0.040-0.063 mm) using a weight ratio of ca. 30:1 for silica gel over crude compound. ¹H, ¹³C, and ³¹P-NMR spectra were recorded on a Bruker 400 spectrometer (400, 100, and 166 MHz, respectively) in deuterated chloroform (CDCl₃), methanol-*d*₄ (CD₃OD), and DMSO-*d*₆ with either tetramethylsilane (TMS) (0.00 ppm) or the NMR solvent as the internal reference. HPLC purification for CBT-TTP was carried out using a Shimadzu LC-10AT VP system with a Zobax C18 reversed-phase column (9.4 mm × 25 cm). The sample was eluted (6 mL/min) with buffer A (20 mM triethylammonium acetate, pH 6.9-7.1) and buffer B (50% acetonitrile, 20 mM triethyl ammonium acetate) with the following: 0 min 0% B; 20 min 20% B; 30 min 100% B; 38 min 100% B; 40 min 0% B; and 45 min 0% B.

DNA primers and templates were purchased from Integrated DNA Technologies. Klenow fragment (3'-5' exo), *Taq*. DNA polymerase, and Deep Vent_R (exo-) DNA polymerase were purchased from New England Biolabs. KOD-XL DNA polymerase was purchased from Novagen, Darmstadt. Reaction buffers were used as provided.

2. Experimental Procedure for the Synthesis of CBT-TTP

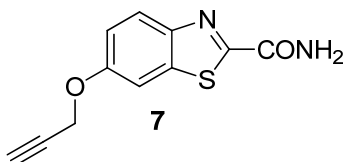
Ethyl 6-(prop-2-ynoxy)benzo[d]thiazole-2-carboxylate **6**



To a solution of **5** (2.23 g, 10 mmol) in DMF (30 mL) was added K₂CO₃ (2.70 g, 20 mmol) and the mixture was stirred at RT for 30 min. After addition of propargyl bromide (80% in toluene, 3.7 mL, 20 mmol), the reaction mixture was stirred at RT for 12 h before diluting with EtOAc (300 mL). Then the mixture was washed with water (50 mL × 3) and brine (50 mL), and dried over Na₂SO₄. Solvent evaporation followed by flash chromatography (Hex: EA, 4:1) gave a yellow solid (2.35 g, 90% yield). ¹H NMR (CDCl₃): 8.14 (d, *J* = 9.1 Hz, 1H), 7.49 (d, *J* = 2.3 Hz, 1H), 7.24 (dd, *J* = 2.3, 9.1 Hz, 1H), 4.80 (s, 2H), 4.55 (q, *J* = 8.7 Hz, 2H), 2.58 (s, 1H), 1.48 (t, *J* = 8.7 Hz); ¹³C NMR (CDCl₃):

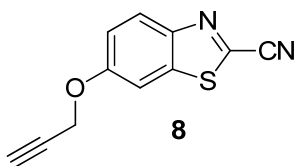
160.7, 157.4, 156.4, 148.3, 138.4, 126.3, 117.9, 105.1, 63.0, 56.4, 14.3. HRMS (ESI) for $C_{13}H_{12}NO_3S$ $[M+H]^+$ Calcd. 262.0545, Found 262.0544.

6-(Prop-2-ynyloxy)benzo[d]thiazole-2-carboxamide **7**



To a solution of ester **6** (2.35 g, 9 mmol) in EtOH 100 mL was added ammonium hydroxide 40 mL. The reaction mixture was heated to 75 °C for 5 h and then cooled to RT. Solvent was removed using a rota-evaporator, and the resulting precipitate was filtered, washed with H₂O, EtOH and EtOAc to give a white solid (1.54 g, yield 73%). ¹H NMR (CDCl₃): 8.41 (s, 1H), 8.03 (d, *J* = 9.1 Hz, 1H), 8.01 (s, 1H), 7.82 (d, *J* = 2.3 Hz, 1H), 7.26 (dd, *J* = 2.3, 9.1 Hz, 1H), 4.92 (d, *J* = 2.2 Hz, 2H,), 3.64 (t, *J* = 2.2 Hz, 1H); ¹³C NMR (CDCl₃): 163.7, 162.4, 157.3, 148.7, 138.9, 125.7, 118.3, 107.3, 79.8, 79.6, 57.1. IR film 3424.7, 3301.3, 1696.0, 1600.8, 1502.0, 1399.5, 1256.1, 1230.5; HRMS (ESI) for $C_{11}H_9N_2O_2S$ $[M+H]^+$ Calcd 233.0380; found 233.0387.

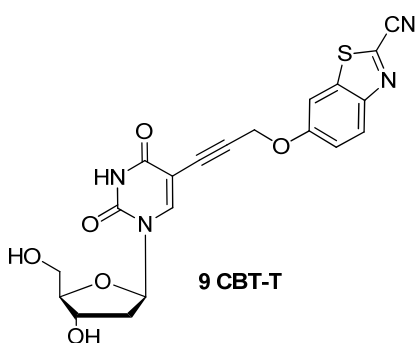
6-(Prop-2-ynyloxy)benzo[d]thiazole-2-carbonitrile **8**



To a suspension of amide **7** (1.1 g, 4.7 mmol) in anhydrous pyridine (25 mL) was added a solution of phosphorous oxychloride (0.85 mL, 9.4 mmol) in CH₂Cl₂ (5 mL) dropwise at 0 °C. After stirring at 0 °C for 1 h, the reaction was slowly warmed to ambient temperature and the resulting solution was stirred for an additional 12 h. CH₂Cl₂ (200 mL) was added, and the reaction was quenched by addition of H₂O 20 mL. The organic phase was separated, washed with H₂O (20 mL × 2), brine (20 mL), and dried over Na₂SO₄. Solvent evaporation followed by flash chromatography (Hex: EtOAc, 6:1) afforded

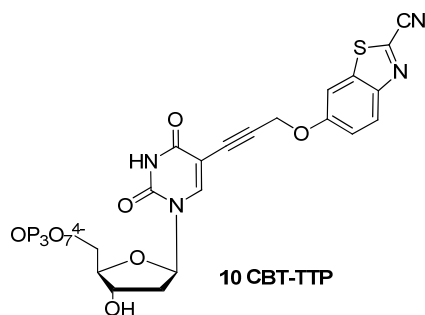
compound **8** as a light yellow solid (900 mg, yield 82%). ^1H NMR (CDCl_3): 8.14 (d, $J = 9.1$ Hz, 2H), 7.51 (d, $J = 2.5$ Hz, 1H), 7.25 (dd, $J = 2.5, 9.1$ Hz, 1H), 4.83 (d, $J = 2.4$ Hz, 2H), 2.61 (t, $J = 2.4$ Hz, 1H); ^{13}C NMR (CDCl_3): 158.2, 147.4, 137.2, 134.0, 126.0, 118.8, 113.1, 104.7, 56.5; IR film 3255.3, 2232.2, 2126.5, 1606.5, 1551.5, 1470.1, 1256.8, 1230.3, 1132.0, 1049.9, 127.7. HRMS (ESI) for $\text{C}_{11}\text{H}_7\text{N}_2\text{OS}$ $[\text{M}+\text{H}]^+$ Calcd. 215.0279, Found 215.0289

CBT-T **9**



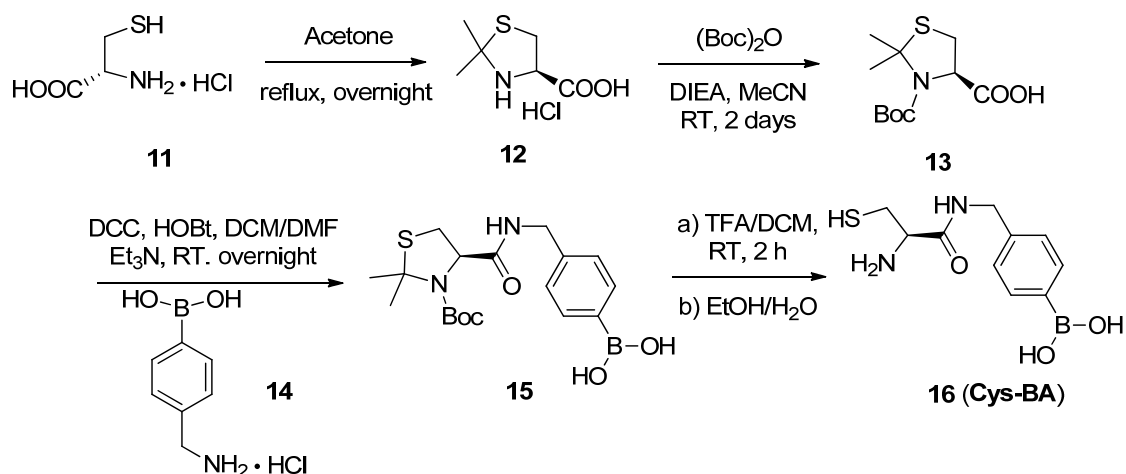
To a mixture of 5-iodo-deoxyuridine (708 mg, 2 mmol), $\text{Pd}(\text{PPh}_3)_4$ (230 mg, 0.2 mmol), and CuI (76 mg, 0.4 mmol) in anhydrous DMF (12 mL) under argon was added DIPEA (0.62 mL, 4 mmol) followed by a solution of compound **8** (700 mg, 3.0 mmol) in DMF (4 mL). After stirring at RT overnight, DMF was evaporated, and the remaining dark gum was purified by flash chromatography (DCM: MeOH, 20 : 1) to afford compound **9** as a pale yellow solid (630 mg, yield 72%). ^1H NMR (CD_3OD): 8.39 (s, 1H), 8.08 (d, $J = 9.2$ Hz, 2H), 7.81 (d, $J = 2.0$ Hz, 1H), 7.34 (dd, $J = 2.0, 9.2$ Hz, 1H), 6.22 (t, $J = 6.4$ Hz), 5.07 (s, 2H), 4.39 (m, 1H), 3.94 (d, $J = 3.2$ Hz), 3.83 (m, 1H), 3.23 (s, 1H), 2.30 (m, 1H), 2.20 (m, 1H); ^{13}C NMR (CD_3OD): 164.3, 159.9, 151.1, 148.4, 146.4, 138.8, 135.6, 126.5, 120.3, 114.2, 106.4, 99.2, 89.2, 88.4, 87.2, 80.6, 71.8, 62.5, 58.1, 55.8, 41.8; IR film 3413.7, 2229.5, 1725.8, 1667.6, 1616.6, 1600.8, 1475.2, 1372.7, 1278.4, 1229.3, 1129.9, 1091.8, 1001.6; HRMS (ESI) for $\text{C}_{20}\text{H}_{16}\text{N}_4\text{O}_6\text{SNa}$ $[\text{M}+\text{Na}]^+$ Calcd 463.0683; Found 463.0674.

CBT-TTP **10**



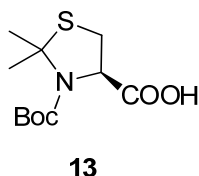
To a solution of CBT-T **9** (70 mg, 0.16 mmol) and proton sponge (42 mg, 0.19 mmol, pre-dried in vacuum over P_2O_5 overnight) in anhydrous trimethylphosphate (0.6 mL) was added freshly distilled $POCl_3$ (18 μ L, 0.19 mmol) dissolved in anhydrous trimethylphosphate (0.2 mL) dropwise via a syringe with stirring under argon at 0 °C. The reaction mixture was further stirred in an ice-bath for 2 h and then a solution of tributylammonium pyrophosphate (226 mg, 0.48 mmol) and tri-*n*-butylamine (0.4 mL,) in 1.0 mL of anhydrous DMF was added in one portion. The mixture was stirred at RT for 10 min and then triethylammonium bicarbonate solution (0.1 M, pH 8, 11 mL) was added. After stirring at RT for an additional hour, the resulting reaction mixture was transferred to a 50-mL centrifuge tube. Then EtOH (33 mL) was added followed by 3 M NaCl solution (1.0 mL). After vortexing for 10 sec, the centrifuge tube was placed at – 80 °C for 1 h, and then centrifuged at 5000 rpm for 20 min. After removing the supernatant, the resulting pellet was purified by HPLC and lyophilized to give a pale yellow powder (9 mg, 10%). 1H NMR (D_2O): 8.57 (s, 1H), 8.05 (d, J = 6.8 Hz, 2H), 7.65 (s, 1H), 7.27 (d, J = 9.2 Hz, 1H), 6.11 (t, J = 6.4 Hz), 5.00 (s, 2H), 4.11-4.13(m, 3H), 2.27-2.31 (m, 2H); ^{13}C NMR (D_2O): 158.5, 151.5, 147.1, 146.0, 138.1, 135.7, 125.7, 119.9, 113.9, 106.2, 99.2, 88.9, 86.5, 86.4, 86.3, 79.5, 70.8, 65.8, 58.1, 58.0, 39.3, 17.5; ^{31}P NMR (D_2O) -10.9, -11.5, -23.4. HRMS (ESI) for $C_{20}H_{18}N_4O_{15}P_3S$ [M-H] $^-$ Calcd. 678.9702; Found 678.9683.

3. Experimental Procedure for the Synthesis of Cys-BA



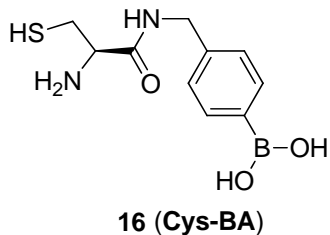
Scheme S1. Synthesis of Compound **16 (Cys-BA)**

L-(-)-(tert-Butoxycarbonyl)-2,2-dimethylthiazolidine-4-carboxylic acid **13**



Compound **13** was prepared starting from L-cysteine hydrochloride (**11**) in two steps following literature procedures.^[1]

Cys-BA (**16**)

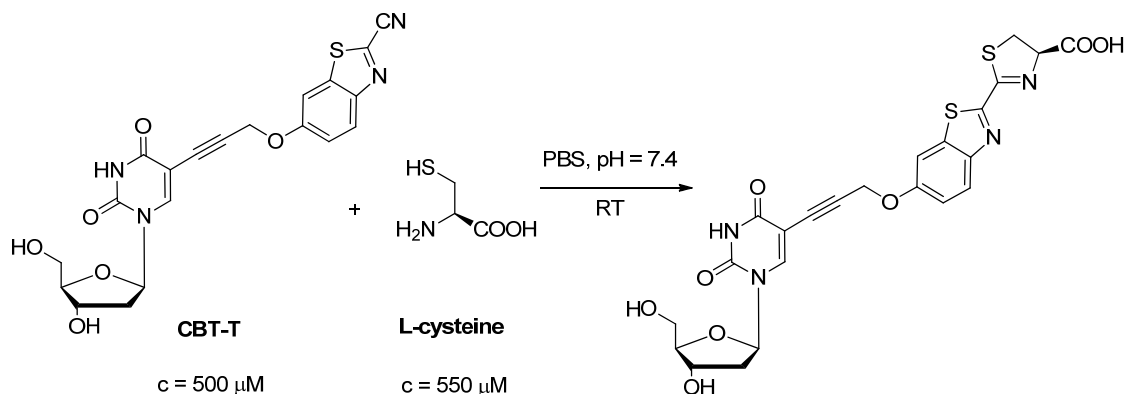


To a mixture of compound **13** (836 mg, 3.2 mmol), **14** (500 mg, 2.7 mmol), and HOBT (397 mg, 2.9 mmol) in anhydrous DCM (20 mL) and DMF (20 mL) under ice-bath, was added Et₃N (0.7 mL, 5.3 mmol) followed DCC (607 mg, 2.9 mmol) in one portion. The mixture was stirred at RT overnight and the resulting solid was filtered off. The filtrate was concentrated under vacuum and the residue was purified by flash chromatography (DCM: MeOH, 30:1) to afford intermediate **15**. ¹H NMR (CDCl₃): 7.83 (d, *J* = 6.8 Hz,

2H), 7.25 (d, $J = 6.8$ Hz, 2H), 4.82 (brs, 1H), 4.48 (s, 2H), 3.25 (m, 2H), 1.82 (s, 3H), 1.77 (s, 3H), 1.50 (s, 6H).

Compound **15** was stirred in DCM/TFA (20 mL/20 mL) at RT for another 2 h. Solvent was evaporated under vacuum and the residue was treated with diethyl ether (100 mL). The resulting precipitate was collected through filtration. After washing three times with diethyl ether, the collected precipitate was dissolved in EtOH/H₂O (300 mL/300 mL) with stirring for 30 mins. Evaporation of the solvent yielded compound **16** as a white solid (497 mg, 73% in two steps). ¹H NMR (CD₃OD): 7.67 (d, $J = 4.0$ Hz, 2H), 7.25 (d, $J = 4.0$ Hz, 2H), 4.39 (m, 1H), 4.08 (m, 1H), 3.00 (m, 2H); ¹³C NMR (CD₃OD): 168.2, 140.7, 134.9, 127.7, 55.8, 44.2, 26.1; HRMS (ESI) for C₁₀H₁₅BN₂O₃S [M+H]⁺ Calcd 255.0969; Found 255.0965.

4. Kinetic studies of the reaction between CBT-T and 1, 2-aminothiols (L-cysteine)



Scheme S2. Reaction scheme of CBT-T with L-cysteine

To a solution of CBT-T **9** in acetonitrile was added a solution of L-cysteine in PBS buffer (pH = 7.4) to afford the final concentration of CBT-T (500 μ M), L-cysteine (550 μ M) in a mixture of acetonitrile/PBS buffer (v/v: 1/4). The resulting mixture was stirred at room temperature and monitored by HPLC at different time points in a similar way as literature reported.^[2] Applying similar kinetic analysis reported in the references, $1/[\text{CBT-T}]$ vs reaction time was plotted and the linear regression analysis of $1/[\text{CBT-T}]$ vs. time gave the second-order rate constant as 22 M⁻¹ s⁻¹.

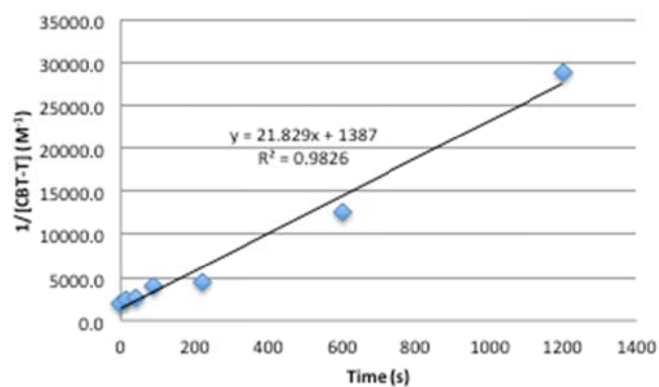


Figure S1. Linear regression analysis of $1/[CBT-T]$ vs. time of the CBT-T condensation reaction

5. MALDI-TOF spectra of DNA products

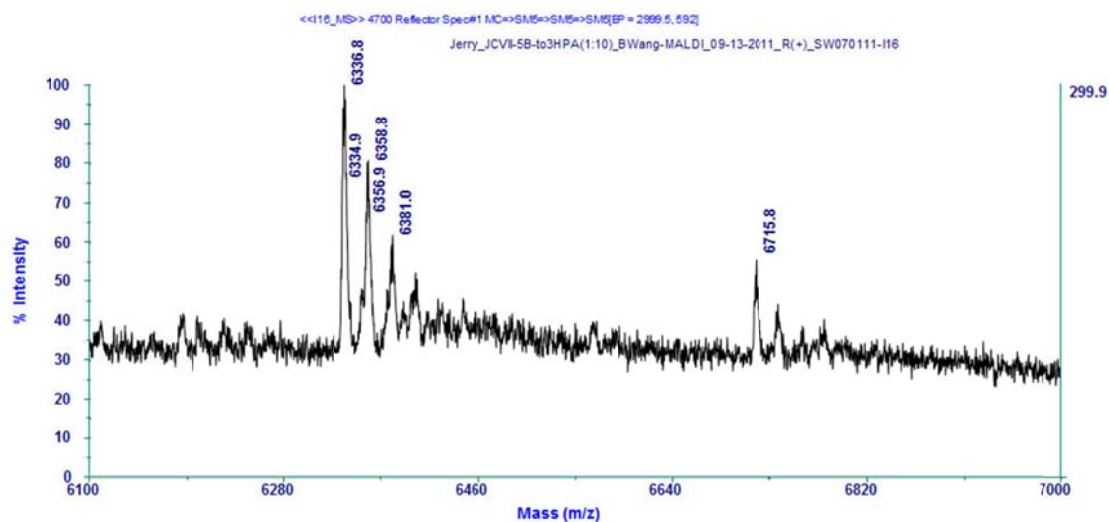


Figure S2. MALDI-TOF spectra of CBT-DNA₂₁ (Template-1)

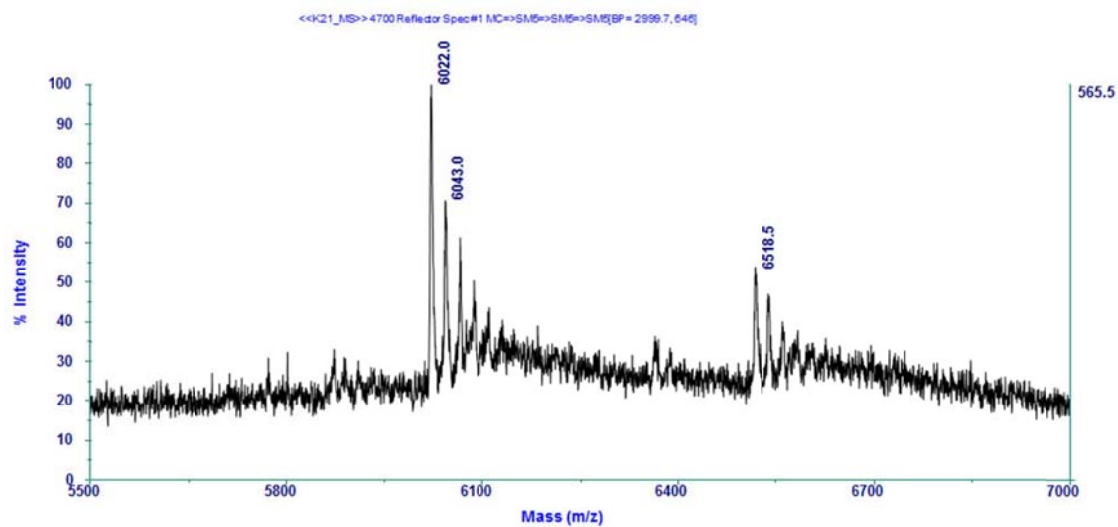


Figure S3. MALDI-TOF spectra of dNTP-DNA₂₁ (Template-1)

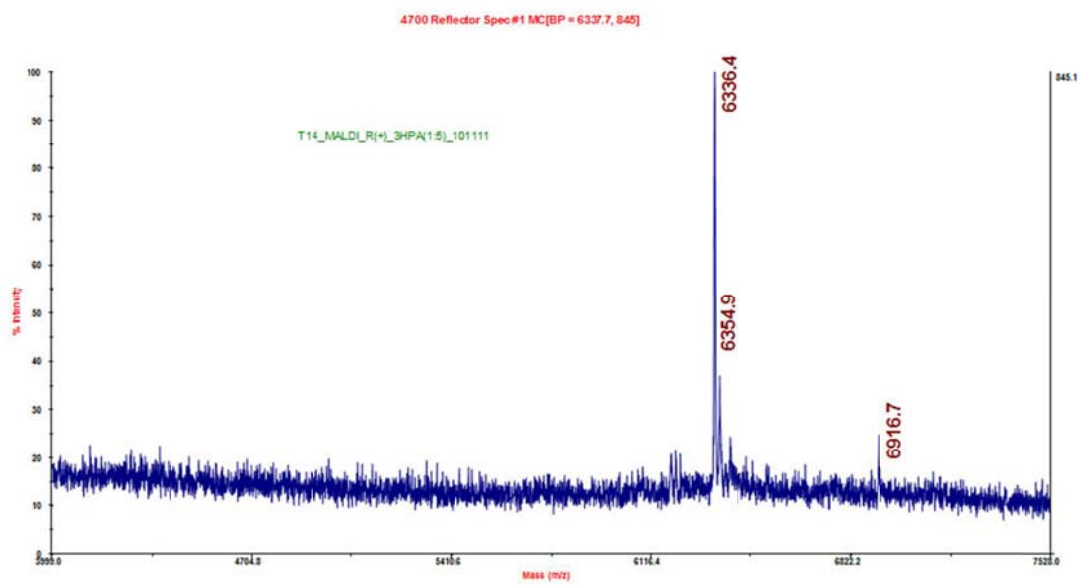


Figure S4. MALDI-TOF spectra of click product of CBT-DNA₂₁ and Cys-BA (Template-1)

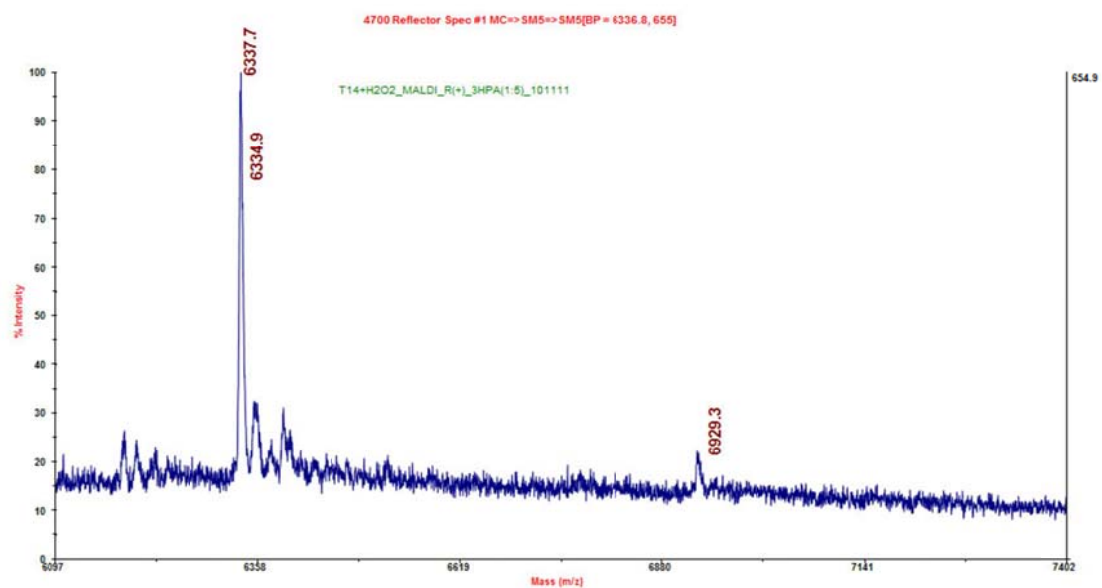


Figure S5. MALDI-TOF spectra of H₂O₂ treated click product of CBT-DNA₂₁ (Template-1)

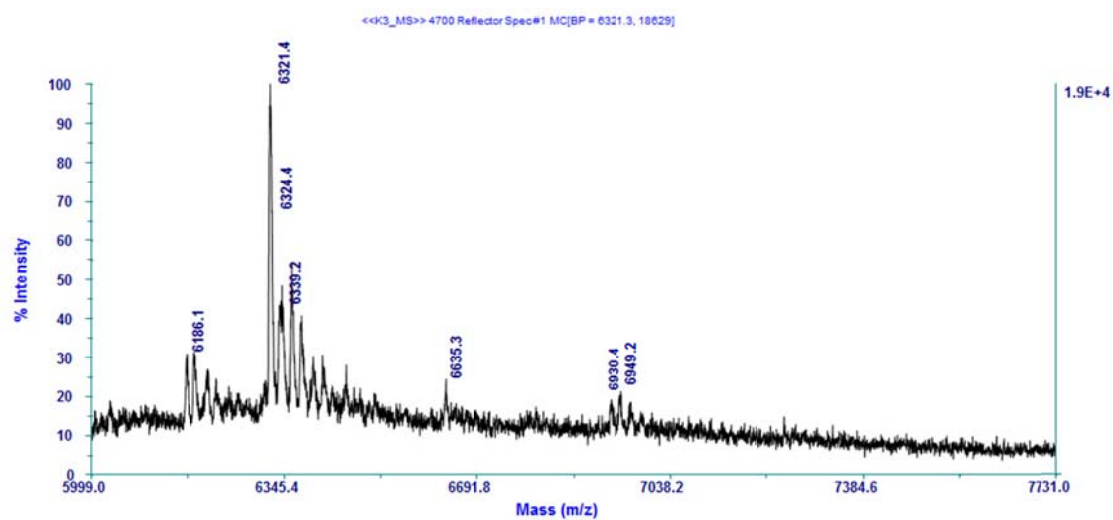


Figure S6. MALDI-TOF spectra of CBT-DNA₂₁ (Template-2)

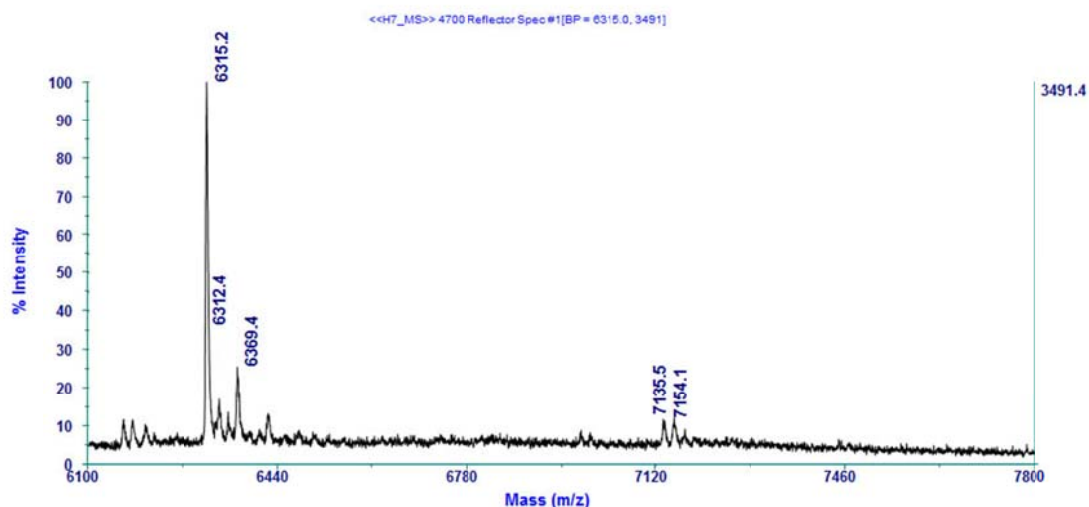


Figure S7. MALDI-TOF spectra of CBT-DNA₂₁ (Template-3)

6. Thermodenaturation of duplex DNAs

The UV-melting temperature studies were carried out by Cary 300 UV-Vis Spectrophotometer and a temperature control module. Duplex DNA samples were prepared through Klenow fragment primer extension reactions with dTTP or CBT-TTP. Post-synthetic sample was prepared by click reaction of CBT-DNA₂₁ with Cys-BA. All samples were cleaned using Millipore Amicon 10 kDa spin column. The samples (1 μ M DNA duplexes) were dissolved in PBS buffer (pH 7.4). The samples were heated to 80 °C and cooled to 6 °C at a rate of 0.5 °C per min. 4 Rounds of melting were performed for each sample. The melting temperature data and curves are presented in Table S1 and Figure S7.

Table S1. Melting temperatures (T_m) of DNA dueplexes

DNA/Rounds	1	2	3	4	Average
dNTPs-DNA ₂₁	72.97	73.02	73.02	72.97	73.00 \pm 0.03
CBT-DNA ₂₁	70.94	70.97	70.97	70.97	70.96 \pm 0.02
CBT-DNA ₂₁ + Cys-BA	69.87	70.92	69.92	70.92	70.24 \pm 0.59

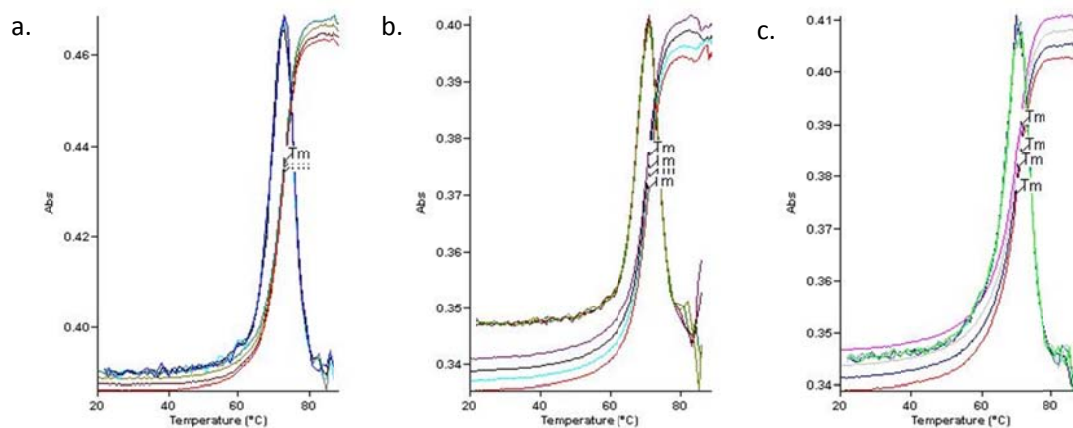


Figure S8. UV-melting curves of DNA duplexes, a. dNTPs-DNA₂₁, b. CBT-DNA₂₁, c. dNTPs-DNA₂₁ + Cys-BA.

References:

- [1] D. S. Kemp and R. I. Carey, *J. Org. Chem* **1989**, *54*, 3640-3646.
 [2] a) G. L. Liang, H. J. Ren and J. H. Rao, *Nat. Chem.* **2010**, *2*, 54-60; b) J. M. Baskin, J. A. Prescher, S. T. Laughlin, N. J. Agard, P. V. Chang, I. A. Miller, A. Lo, J. A. Codelli and C. R. Bertozzi, *P. Natl. Acad. Sci. USA.* **2007**, *104*, 16793-16797.

7. NMR spectrum of synthesized compounds (Please see next page)

PHJ-8-125
CDC13
04-17-2011

8.153
8.131
7.490
7.484
7.269
7.254
7.247
7.231
7.225

4.804
4.798
4.570
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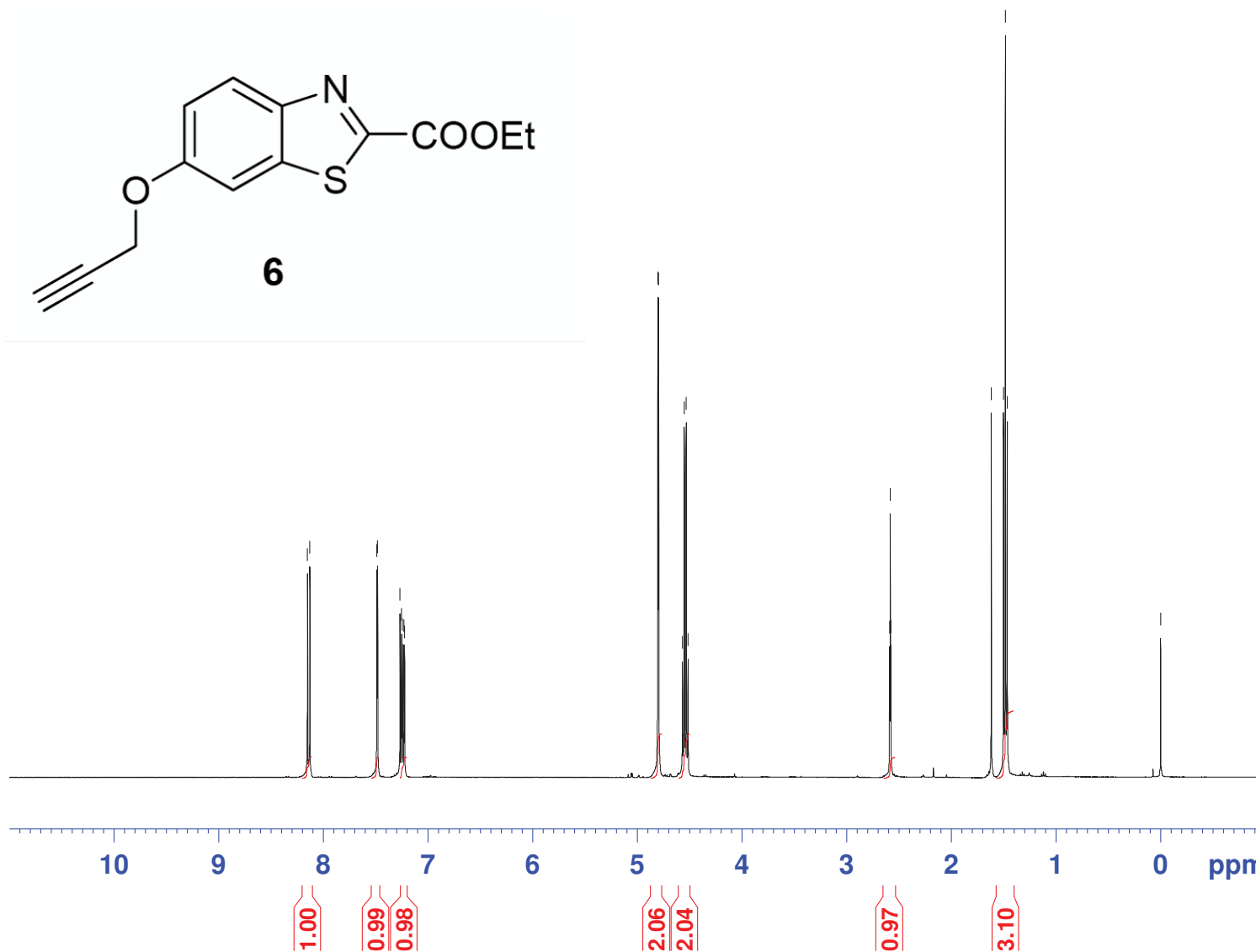
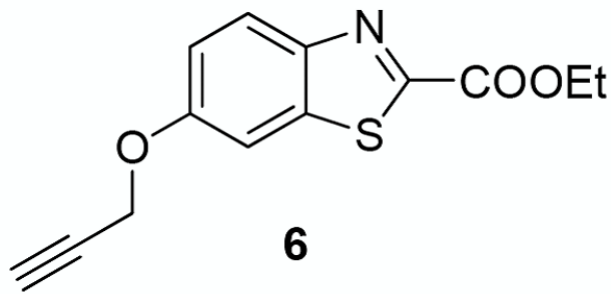


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FIDRES 0.125483 Hz
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RG 161
DW 60.800 usec
DE 6.50 usec
TE 296.6 K
D1 1.00000000 sec

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P1 13.50 usec
PLW1 16.00000000 W
SFO1 400.1424008 MHz

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DCF-IV-14-C13



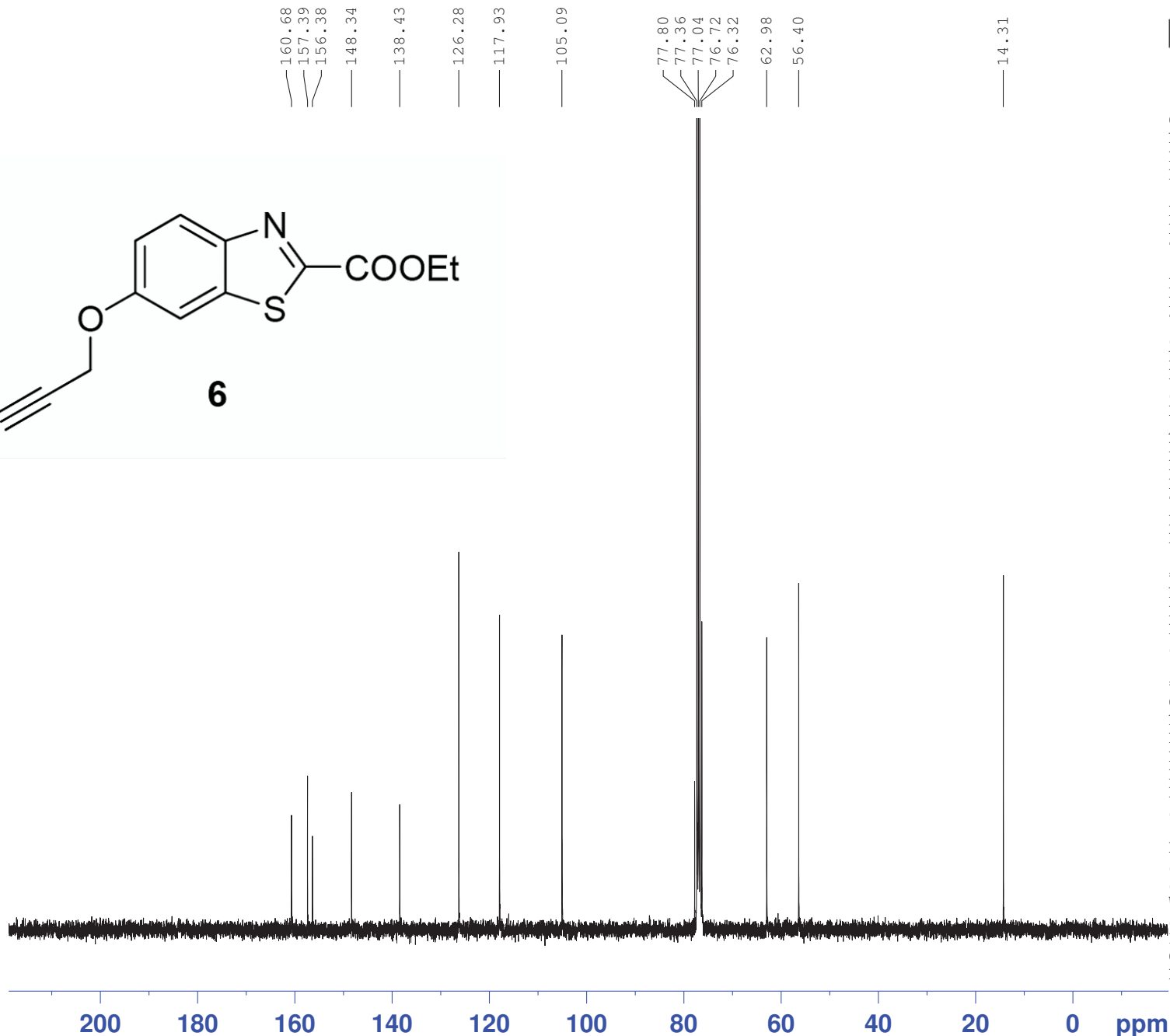
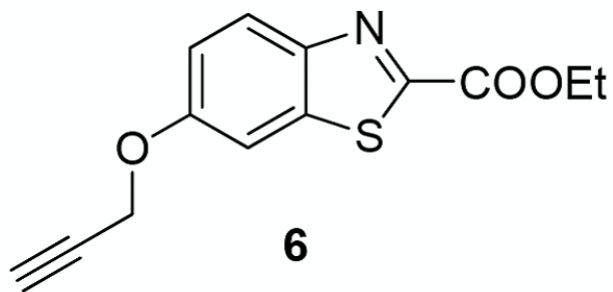
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D1 2.00000000 sec
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SFO1 100.6253441 MHz

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PLW12 0.28864691 W
PLW13 0.14466980 W
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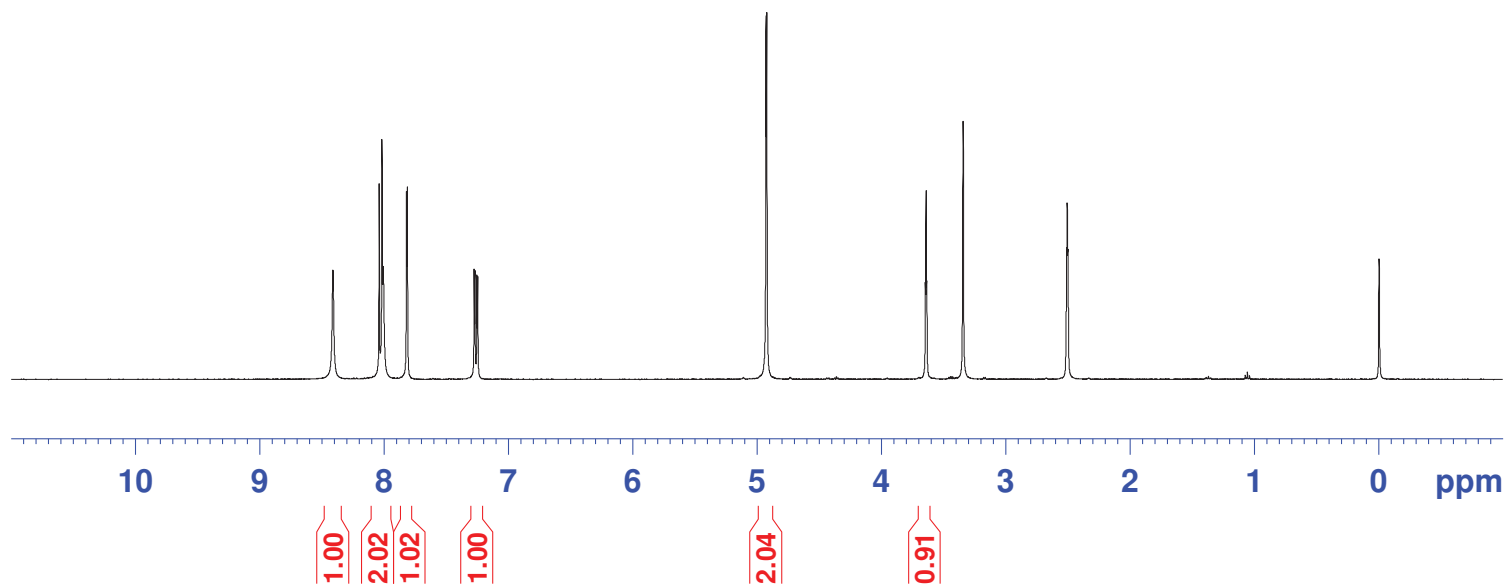
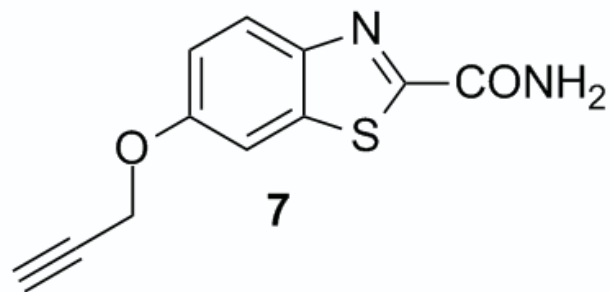
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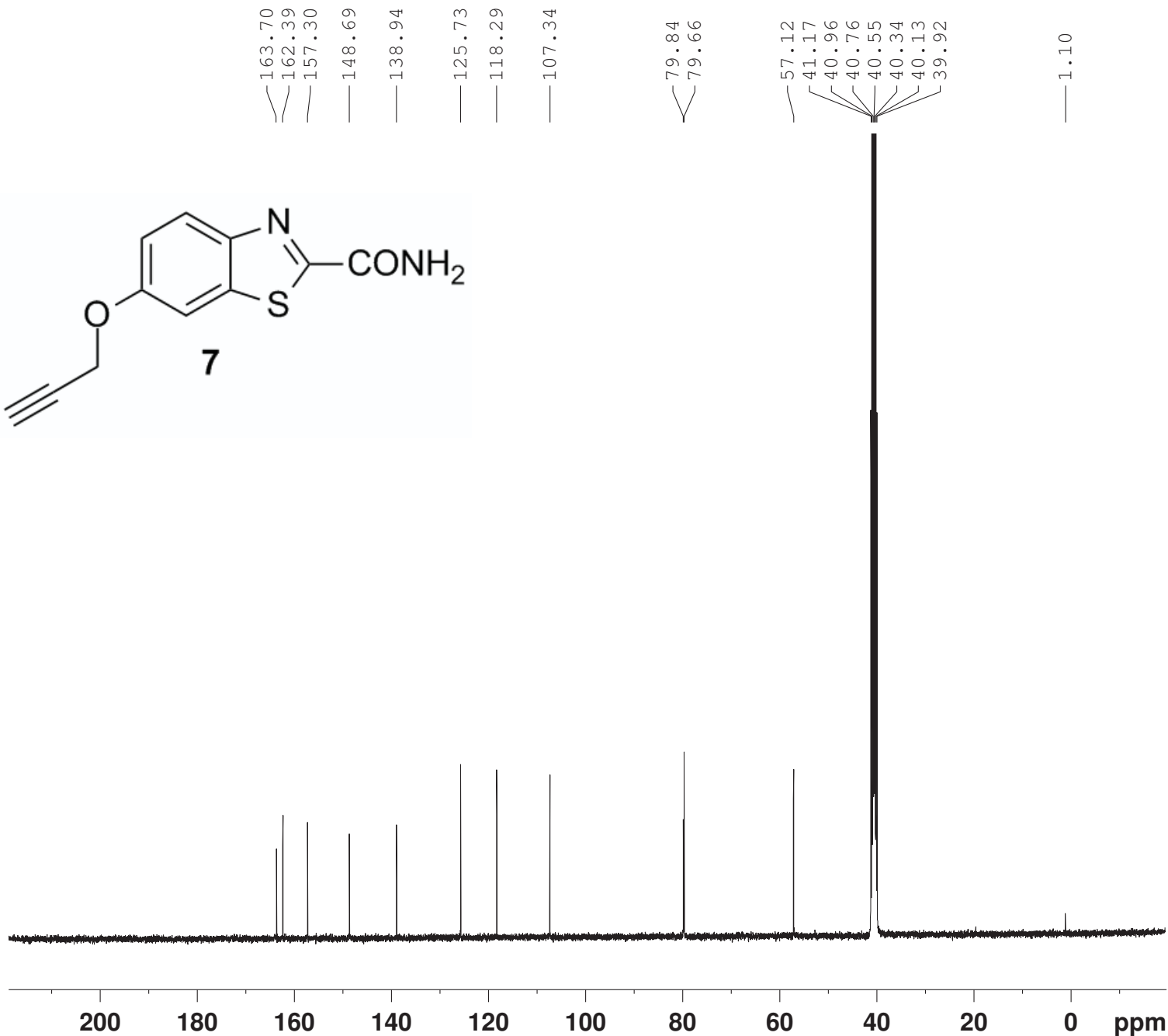
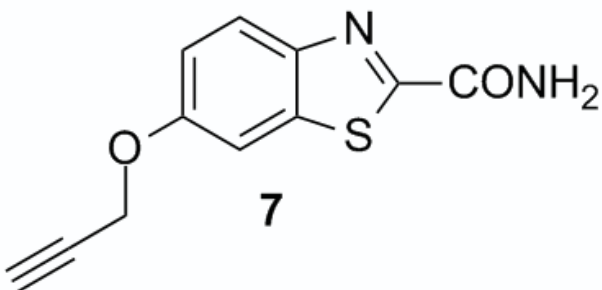
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DE 6.50 usec
TE 295.1 K
D1 1.00000000 sec

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F2 - Processing parameters
SI 65536
SF 400.1403249 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

DCF-IV-15-C13



Current Data Parameters
NAME DCF-IV-15
EXPNO 4
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110214
Time 2.02
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 10000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 297.3 K
D1 2.00000000 sec
D11 0.03000000 sec

==== CHANNEL f1 =====
NUC1 13C
P1 9.00 usec
PLW1 53.48799896 W
SFO1 100.6253441 MHz

==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PLW2 12.22599983 W
PLW12 0.28864691 W
PLW13 0.14466980 W
SFO2 400.1416006 MHz

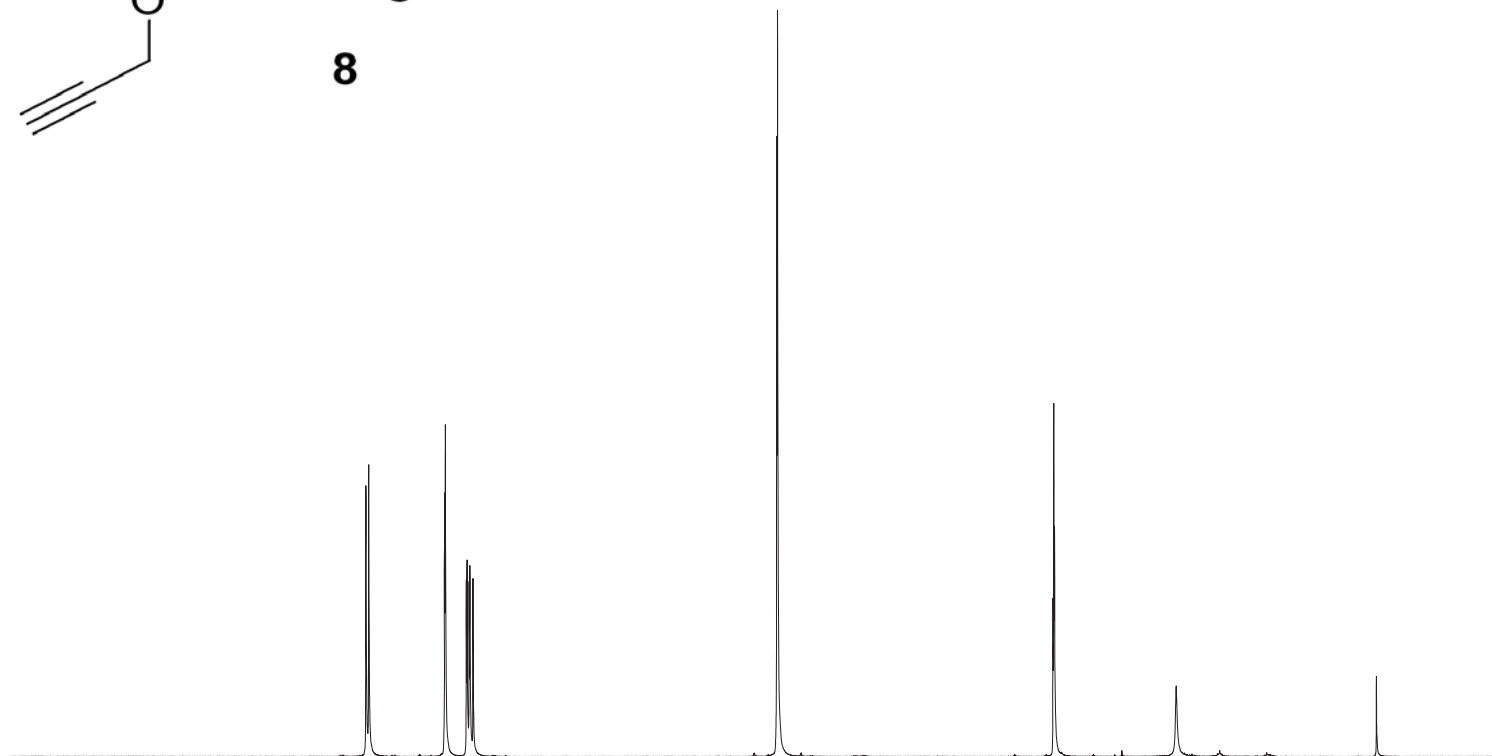
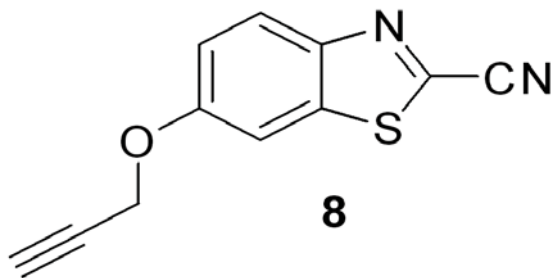
F2 - Processing parameters
SI 32768
SF 100.6152848 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

DCF-IV-17-H1

8.146
8.123
7.513
7.507
7.339
7.333
7.316
7.310
7.285

4.842
4.836

2.621
2.615
2.609



10 9 8 7 6 5 4 3 2 1 0 ppm

1.00
0.99
0.93
0.34

2.00

0.92



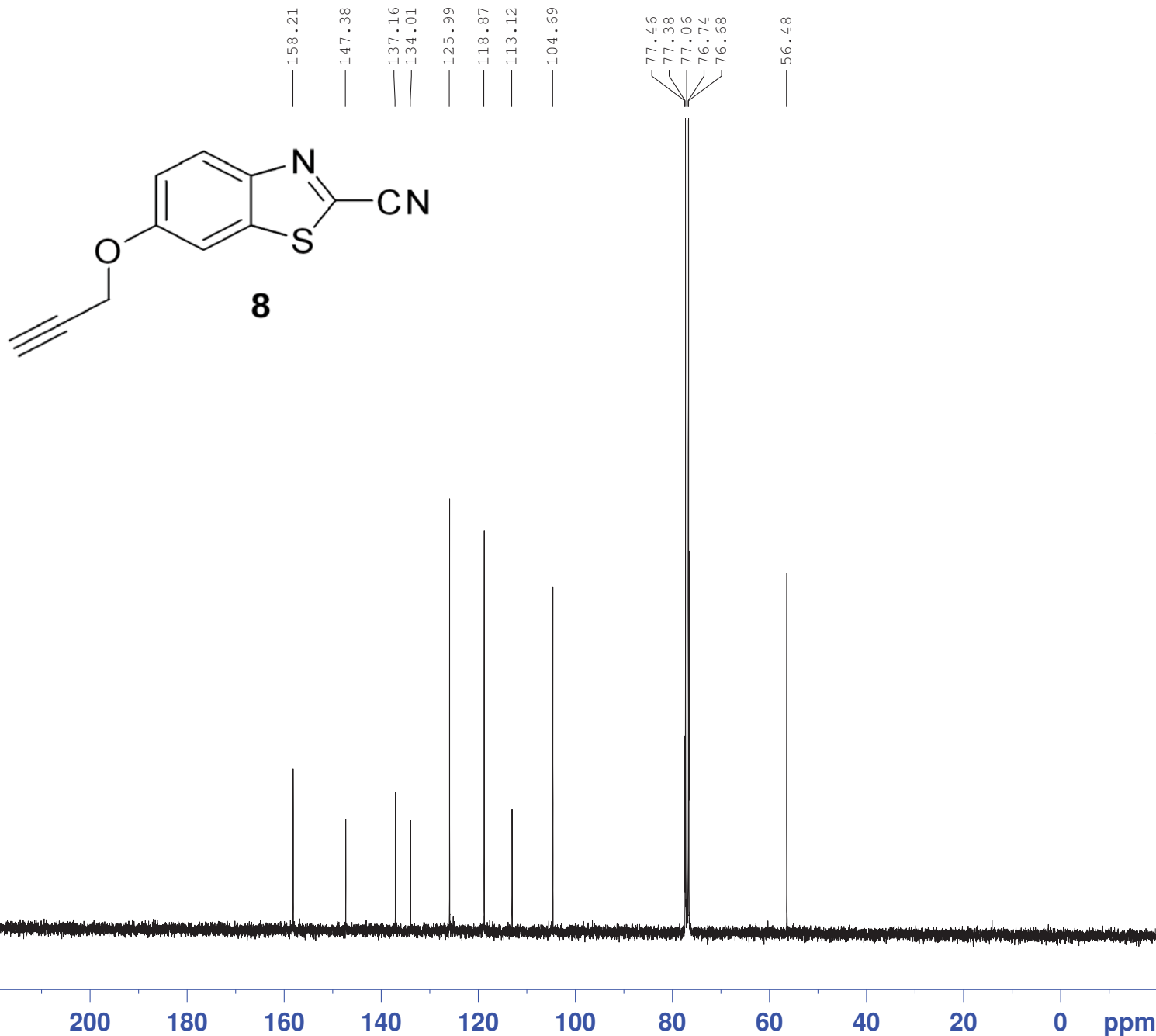
Current Data Parameters
NAME DCF-IV-17
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110210
Time 18.41
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 181
DW 60.800 usec
DE 6.50 usec
TE 295.3 K
D1 1.00000000 sec

==== CHANNEL f1 =====
NUC1 1H
P1 14.00 usec
PLW1 12.22599983 W
SFO1 400.1424710 MHz

F2 - Processing parameters
SI 65536
SF 400.1400000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

DCF-IV-17b-C13



Current Data Parameters
NAME DCF-IV-17b
EXPNO 2
PROCNO 1

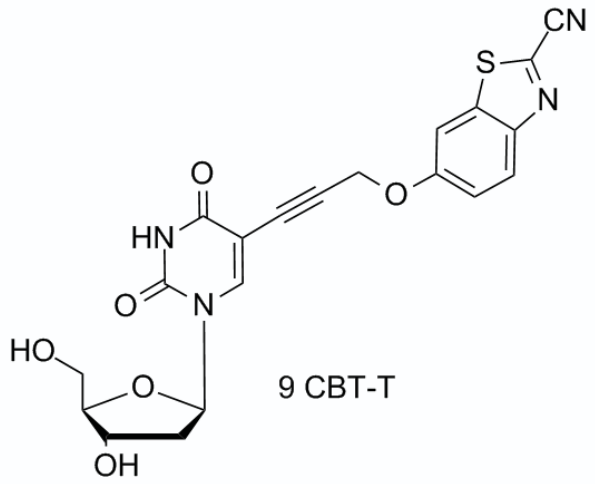
F2 - Acquisition Parameters
Date_ 20110213
Time 15.36
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 388
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 295.9 K
D1 2.00000000 sec
D11 0.03000000 sec

==== CHANNEL f1 =====
NUC1 13C
P1 9.00 usec
PLW1 53.48799896 W
SFO1 100.6253441 MHz

==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PLW2 12.22599983 W
PLW12 0.28864691 W
PLW13 0.14466980 W
SFO2 400.1416006 MHz

F2 - Processing parameters
SI 32768
SF 100.6152890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

8.403
8.109
8.086
7.835
7.829
7.372
7.366
7.350
7.343
6.237
6.221
6.205
5.079
4.896
4.622
4.405
4.398
4.390
3.949
3.941
3.812
3.804
3.763
3.754
3.724
3.367
3.337
3.333
3.329
3.325
3.321
3.274
3.255
3.237
2.309
2.303
2.235
2.218
1.405
1.399
1.386
1.382
1.368
0.021

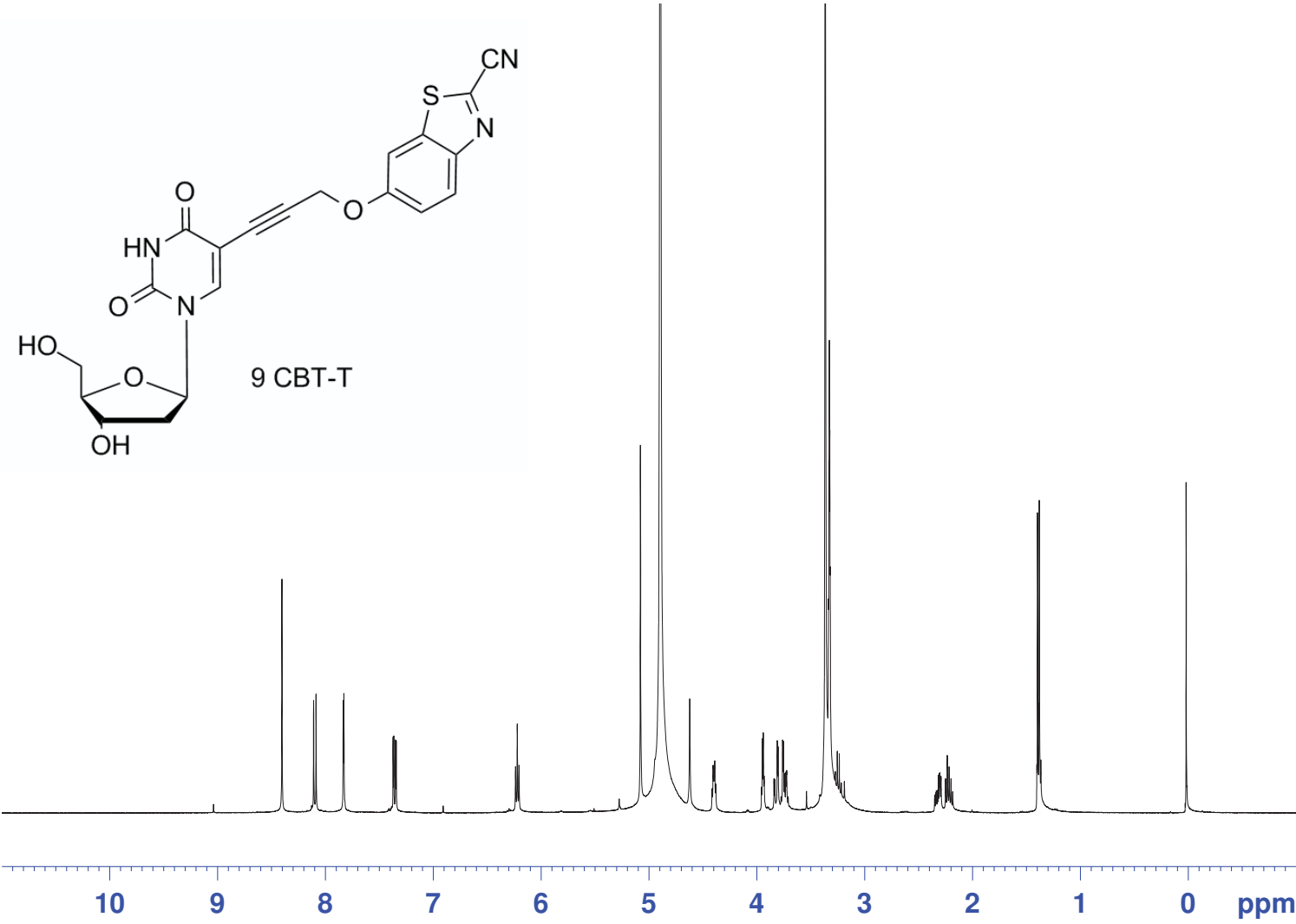


Current Data Parameters
NAME DCF-IV-21
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110219
Time 15.14
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT MeOD
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 32
DW 60.800 usec
DE 6.50 usec
TE 296.1 K
D1 1.00000000 sec

==== CHANNEL f1 =====
NUC1 1H
P1 14.00 usec
PLW1 12.22599983 W
SFO1 400.1424710 MHz

F2 - Processing parameters
SI 65536
SF 400.1400000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

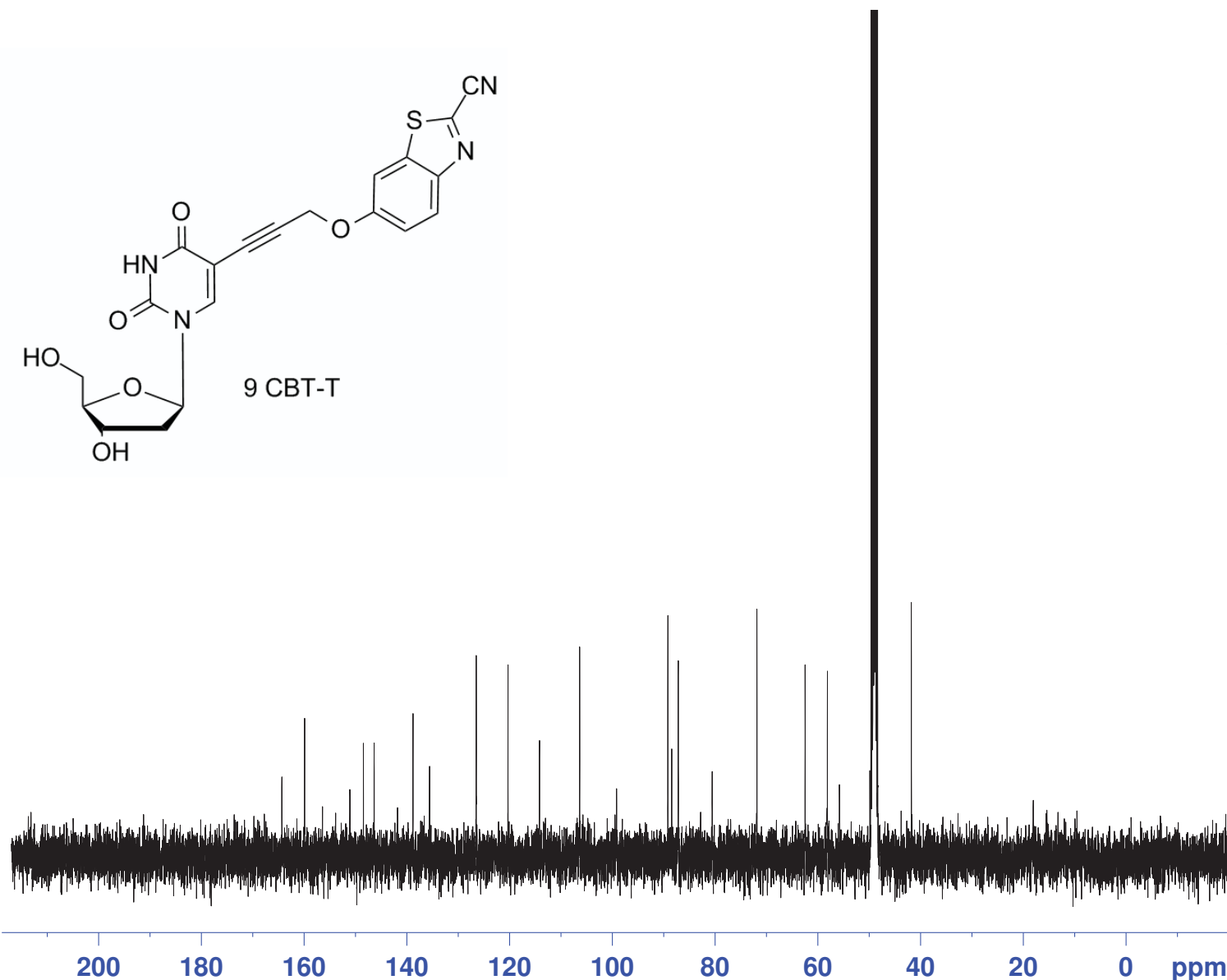
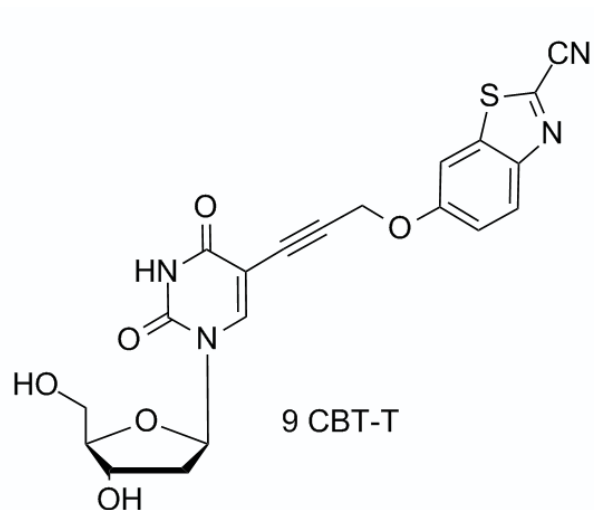


1.00
1.03
0.96
1.01
1.02
1.99
1.12
1.06
0.96
1.06
1.30
13.66
5.32
1.00
1.24
2.60

dcf-IV-21-C13



164.32
159.89
151.08
148.44
146.36
138.80
135.57
126.48
120.30
114.17
106.35
99.17
89.18
88.43
87.15
80.55
71.84
62.46
58.12
55.78
49.85
49.64
49.43
49.21
49.00
48.79
48.57
48.36
41.79



Current Data Parameters
NAME dcf-IV-21-C13
EXPNO 2
PROCNO 1

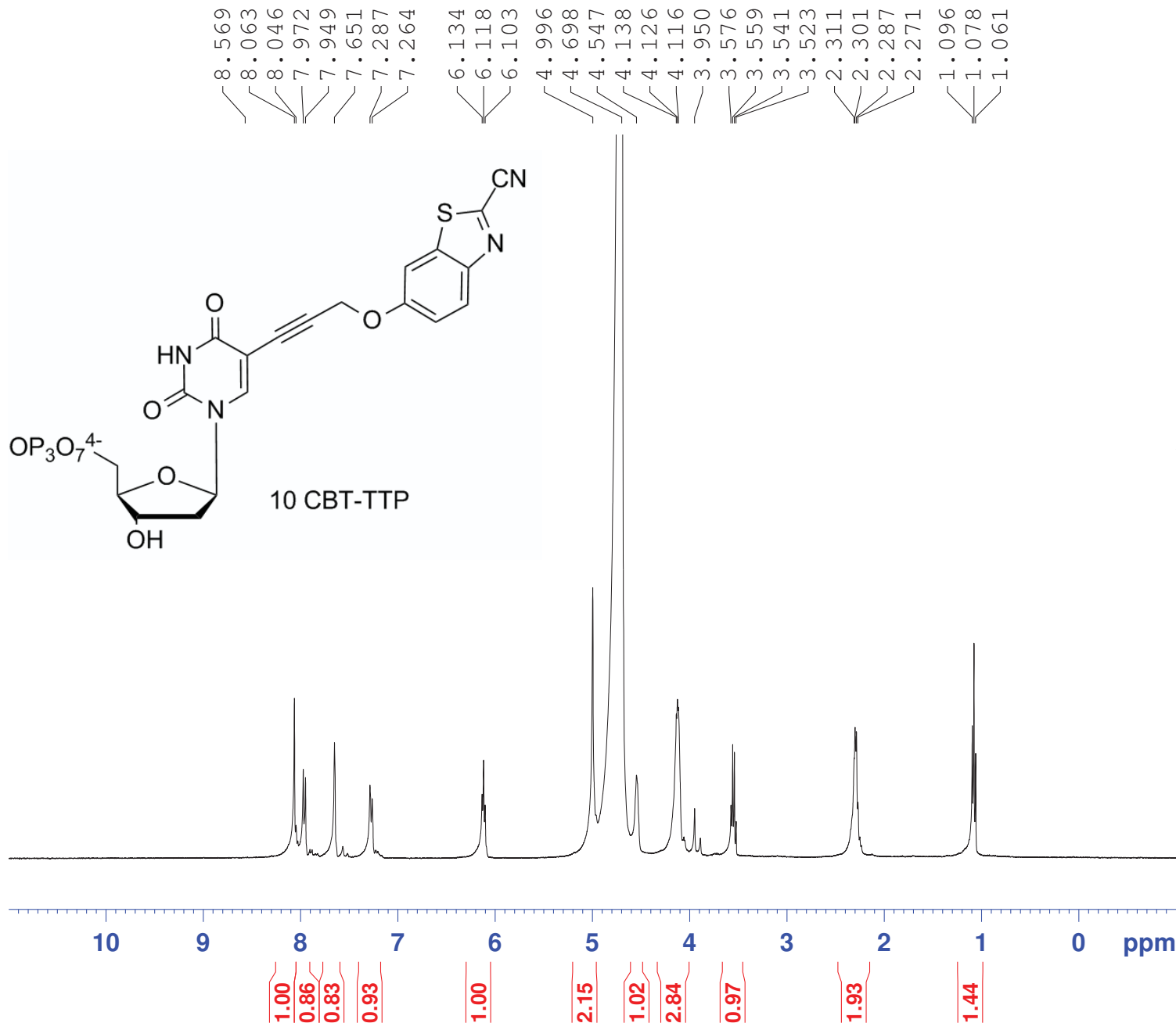
F2 - Acquisition Parameters
Date_ 20120124
Time 10.05
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 423
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 298.3 K
D1 2.00000000 sec
D11 0.03000000 sec

==== CHANNEL f1 =====
NUC1 13C
P1 9.00 usec
PLW1 62.00000000 W
SFO1 100.6253441 MHz

==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PLW2 16.00000000 W
PLW12 0.36000001 W
PLW13 0.29159999 W
SFO2 400.1416006 MHz

F2 - Processing parameters
SI 32768
SF 100.6155389 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

dcf-IV-45-pure-H1



Current Data Parameters
NAME dcf-IV-45-pure
EXPNO 3
PROCNO 1

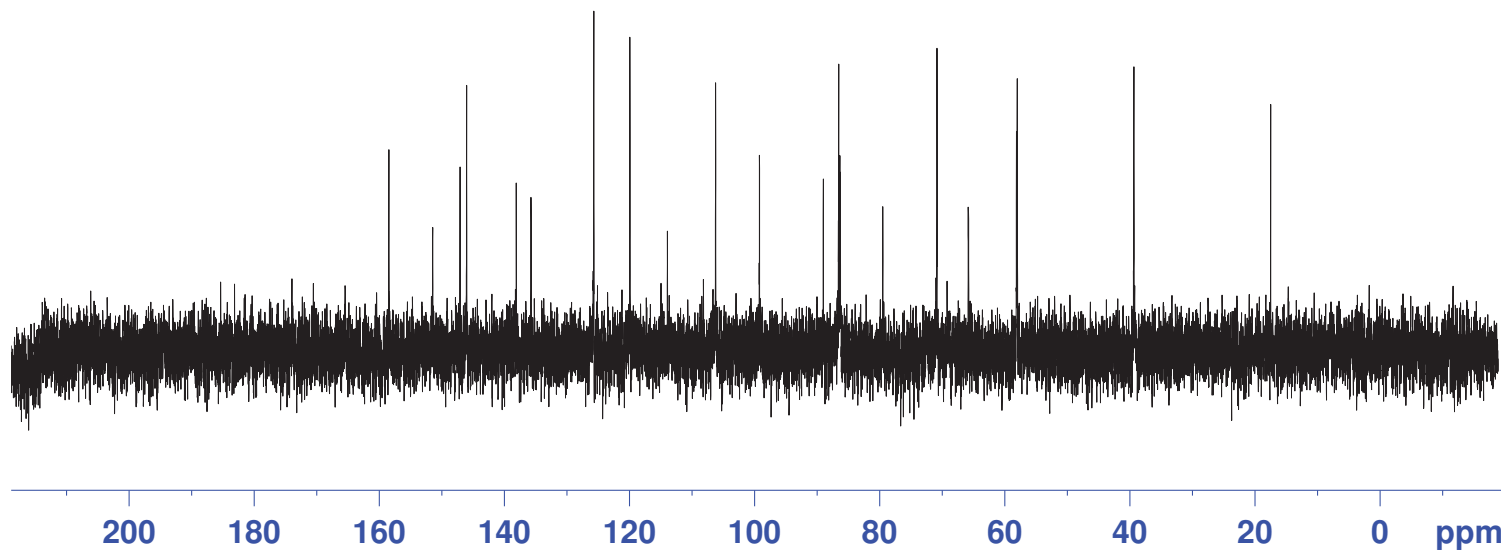
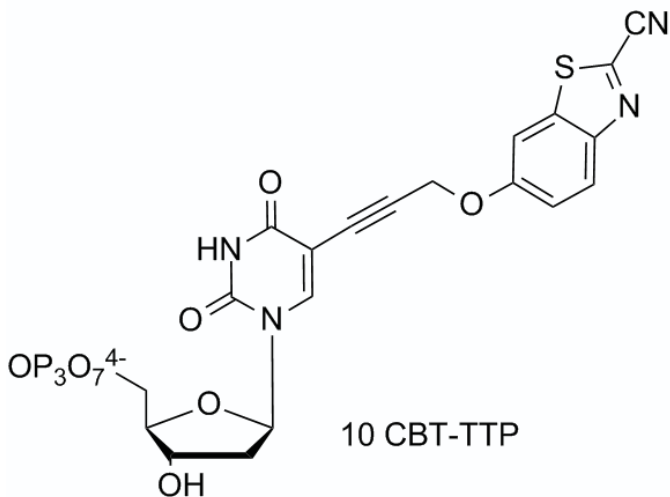
F2 - Acquisition Parameters
Date_ 20110523
Time 18.59
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT D2O
NS 100
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 128
DW 60.800 usec
DE 6.50 usec
TE 296.8 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PLW1 16.00000000 W
SFO1 400.1424710 MHz

F2 - Processing parameters
SI 65536
SF 400.1400000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

dcf-IV-45-C13

158.46
151.46
147.07
146.03
138.09
135.73
125.70
119.92
113.92
106.22
99.20
88.99
86.53
86.35
86.27
79.49
70.82
65.82
58.12
57.99
39.34
17.47



Current Data Parameters
NAME dcf-IV-45-C13
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110523
Time 21.33
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT D2O
NS 11000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 297.3 K
D1 2.00000000 sec
D11 0.03000000 sec

==== CHANNEL f1 =====
NUC1 13C
P1 9.00 usec
PLW1 62.00000000 W
SFO1 100.6253445 MHz

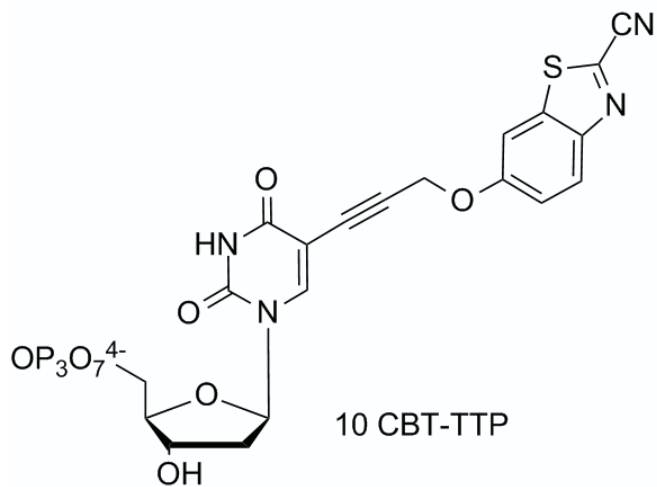
==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PLW2 16.00000000 W
PLW12 0.36000001 W
PLW13 0.29159999 W
SFO2 400.1416006 MHz

F2 - Processing parameters
SI 32768
SF 100.6152207 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

dcf-IV-45-P31

-10.85
-10.98
-11.49
-11.60

-23.35

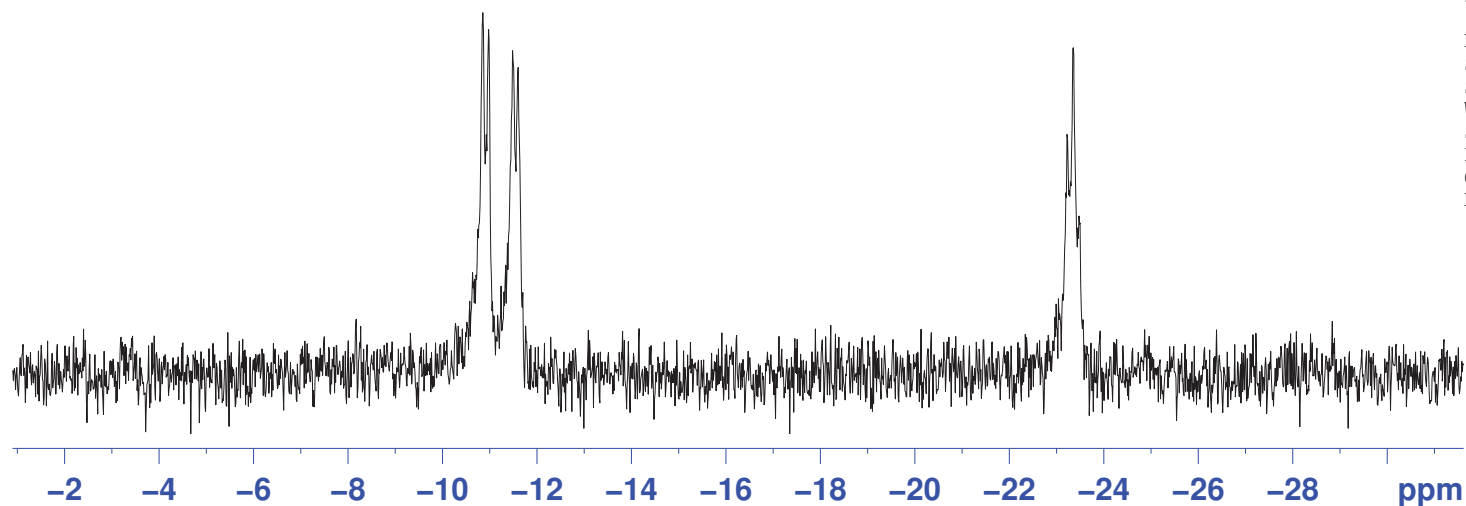


Current Data Parameters
NAME dcf-IV-45
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110522
Time 14.47
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT D2O
NS 32
DS 4
SWH 64102.563 Hz
FIDRES 0.978127 Hz
AQ 0.5112308 sec
RG 203
DW 7.800 usec
DE 6.50 usec
TE 296.5 K
D1 2.00000000 sec

==== CHANNEL f1 =====
NUC1 31P
P1 14.00 usec
PLW1 12.5000000 W
SFO1 161.9816417 MHz

F2 - Processing parameters
SI 32768
SF 161.9796410 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



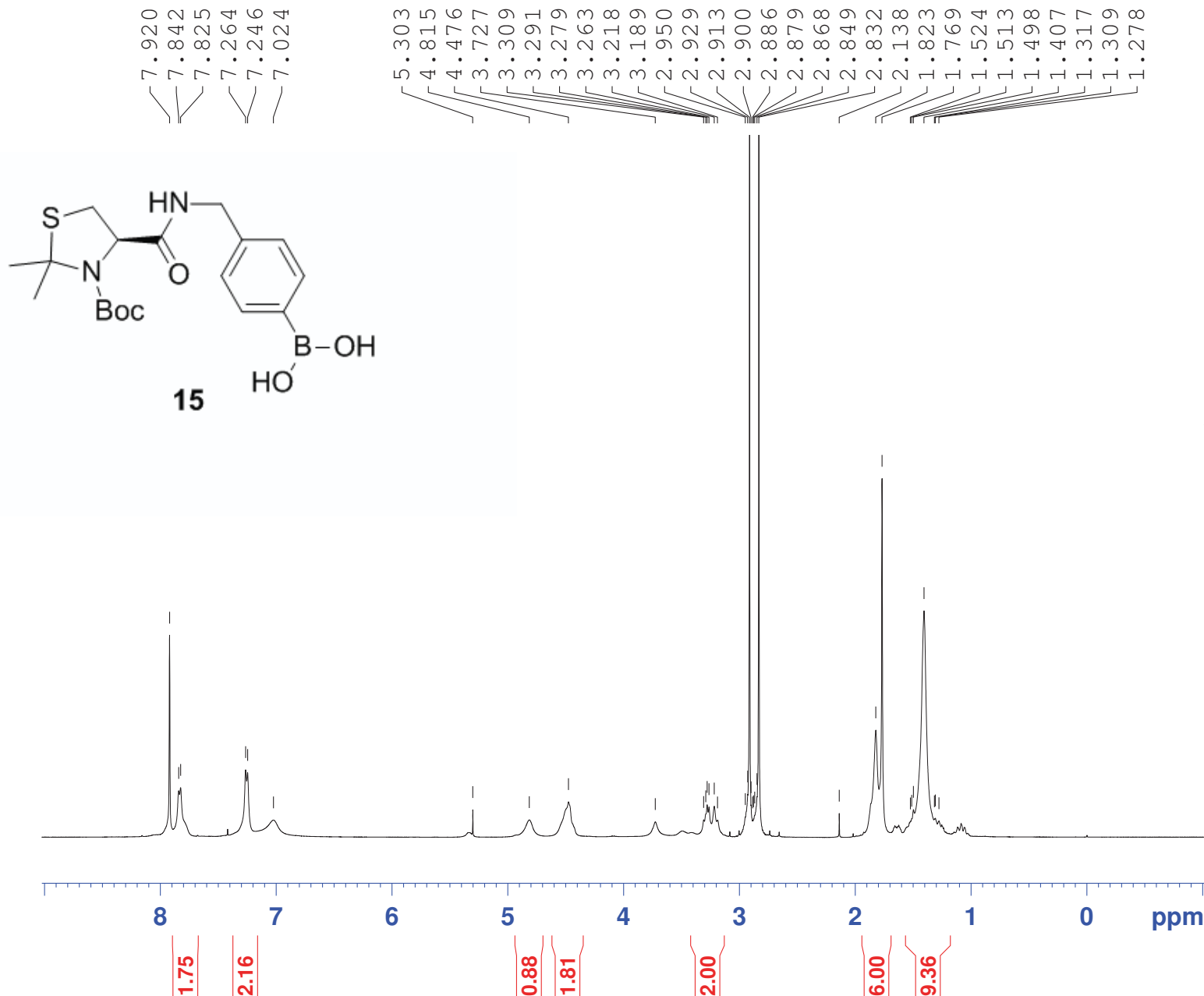


Current Data Parameters
NAME JC-VI-83
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110413
Time 16.18
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 2
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 10
DW 60.800 usec
DE 6.50 usec
TE 296.3 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PLW1 16.0000000 W
SFO1 400.1420007 MHz

F2 - Processing parameters
SI 65536
SF 400.1399452 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00





Current Data Parameters
NAME JC-VI-83-3
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110418
Time 17.03
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT MeOD
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 16
DW 60.800 usec
DE 6.50 usec
TE 297.0 K
D1 1.00000000 sec

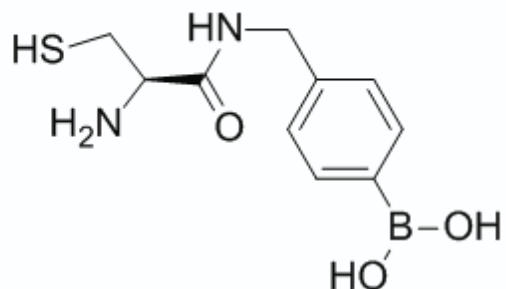
===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PLW1 16.00000000 W
SFO1 400.1424008 MHz

F2 - Processing parameters
SI 65536
SF 400.1400000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

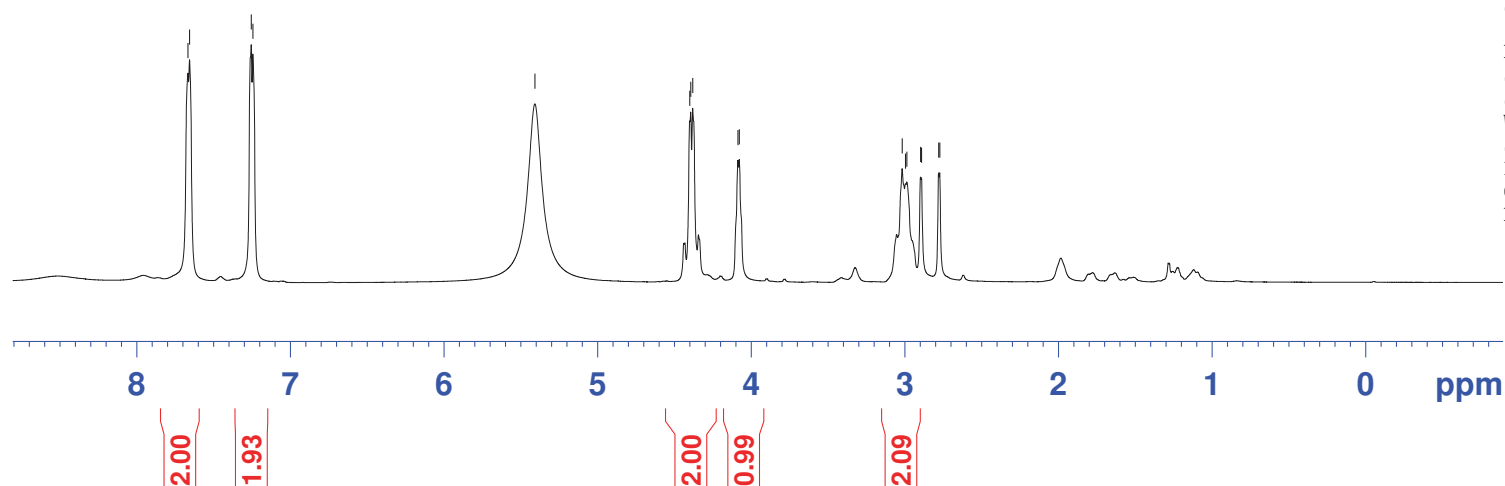
7.667
7.657
7.255
7.244

5.409

4.401
4.394
4.381
4.087
4.078
3.018
2.996
2.987
2.898
2.892
2.780
2.773



16 (Cys-BA)





Current Data Parameters
NAME JC-VI-83-3
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110418
Time 17.57
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG jmod
TD 65536
SOLVENT MeOD
NS 33
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 297.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
D20 0.00689655 sec

=====
CHANNEL f1
NUC1 13C
P1 9.00 usec
P2 18.00 usec
PLW1 62.00000000 W
SFO1 100.6238353 MHz

=====
CHANNEL f2
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PLW2 16.00000000 W
PLW12 0.36000001 W
SFO2 400.1416006 MHz

F2 - Processing parameters
SI 32768
SF 100.6151715 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

168.23

140.69

134.93

127.60

55.81

44.19

26.12

