



A cross-sectional study of Mycoplasma genitalium prevalence and correlates in women in the general population and attending sexually transmitted infection clinics in London

Journal:	<i>BMJ Open</i>
Manuscript ID:	bmjopen-2013-003947
Article Type:	Research
Date Submitted by the Author:	03-Sep-2013
Complete List of Authors:	Svenstrup, Helle; University College London, Institute for Women's Health Dave, Sangeeta; University College London, Institute for Women's Health Carder, Caroline; UCLH Microbiology Lab, Chlamydia Laboratory Grant, Paul; UCLH Microbiology Lab, Chlamydia Laboratory Morris-Jones, Stephen; UCLH Microbiology Lab, Chlamydia Laboratory Kidd, Ian; UCLH Microbiology Lab, Chlamydia Laboratory Stephenson, Judith; University College London, Institute for Women's Health
Primary Subject Heading:	Sexual health
Secondary Subject Heading:	Epidemiology
Keywords:	EPIDEMIOLOGY, GENITOURINARY MEDICINE, MICROBIOLOGY

SCHOLARONE™
Manuscripts

Only

1
2
3 **A cross-sectional study of *Mycoplasma genitalium* prevalence and correlates in women attending**
4 **a national chlamydia screening programme or sexually transmitted infection clinics in London**
5
6

7 Svenstrup HF¹, Dave SS¹, Carder C², Grant P², Morris-Jones S², Kidd M² and Stephenson JM¹

8
9 ¹Research Department of Reproductive Medicine, Institute for Women's Health, University College
10 London, 65 Whitfield Street, W1T 4EU, UK

11 ²Clinical Microbiology and Virology, University College London Hospitals NHS Foundation Trust,
12 65 Whitfield Street, W1T 4EU, UK
13
14

15
16
17 **Corresponding author:**

18 Dr. Helle F Svenstrup

19 Rigensgade 11, DK-1316 København K, Denmark

20 Fax +45 43 99 99 11

21 Direct phone +45 43 31 33 77,

22 Mobile +45 20 65 12 64

23 hfs@awapatent.com
24
25
26
27
28

29 **Co-authors:**

30 Sangeeta Shashikant Dave, Institute for Women's Health, University College London, London,
31 United Kingdom, sangeeta.dave@ucl.ac.uk
32
33

34 Caroline Carder, Clinical Microbiology and Virology, University College London Hospitals NHS
35 Foundation Trust, London, United Kingdom, caroline.carder@uclh.nhs.uk
36
37

38 Paul Grant, Clinical Microbiology and Virology, University College London Hospitals NHS
39 Foundation Trust London, United Kingdom paul.grant@ucl.ac.uk
40
41

42 Stephen Morris-Jones, Clinical Microbiology and Virology, University College London Hospitals
43 NHS Foundation Trust London, United Kingdom, stephen.morris-jones@uclh.nhs.uk
44
45

46 Ian Kidd, Clinical Microbiology and Virology, University College London Hospitals NHS Foundation
47 Trust, London, United Kingdom, i.kidd@ucl.ac.uk
48
49

50 Judith Stephenson, Institute for Women's Health, University College London, London, United
51 Kingdom, judith.stephenson@ucl.ac.uk
52

53 **Key words:** Genotyping / *Mycoplasma genitalium* / National Chlamydia Screening Programme /
54 Real-time PCR
55
56

57 **Word count:** 3461
58
59
60

ABSTRACT

Objective:

To determine *Mycoplasma genitalium* prevalence and correlates among young women undergoing population based screening or clinic based testing for chlamydia infection.

Design:

Cross-sectional study

Setting:

National Chlamydia Screening Programme (NCSP) and two London STI clinics

Participants:

2441 women aged 15 to 64 years who participated in the NCSP and 2172 women who attended two London STI clinics over a four month period in 2009.

Outcome measures:

1. *M.genitalium* prevalence (%)
2. Age-adjusted odds ratios (aORs) for correlates of *M.genitalium* infection

Results

The overall prevalence of *M. genitalium* and *C.trachomatis* was 3.0% and 5.4%, respectively. Co-infection was relatively uncommon (0.5% of all women); however 9% of women with *C.trachomatis* also had *M.genitalium* infection. *M.genitalium* was more frequently detected in swab than urine samples (3.9% vs. 1.3%, $p<0.001$) with a significantly higher mean bacterial load ($p=<0.001$). Among NCSP participants, *M.genitalium* was significantly more likely to be diagnosed in women of black/black British ethnicity (aOR 2.3, 95% CI 1.2-4.5, $p=0.01$). *M.genitalium* and *C. trachomatis* and were both significantly associated with multiple sexual partners in the past year (aOR 2.4, 95% CI 1.3-4.4, $p=0.01$ and aOR 2.0, 95% CI 1.4-2.8, $p<0.01$). Among STI clinic attendees, *M.genitalium* was more prevalent in women who were less than 25 years in age.

Conclusions

M.genitalium is a relatively common infection among young women in London. It is significantly more likely to be detected in vulvo-vaginal swabs than in urine samples. Co-infection with chlamydia is uncommon. The clinical effectiveness of testing and treatment strategies for *M.genitalium* needs further investigation.

Article summary

Article Focus

- How common is *Mycoplasma genitalium* in women in the general population and those attending sexually transmitted infection (STI) clinics in London?
- How are *M.genitalium* prevalence and bacterial load associated with sample type?
- How much co-infection is there between *M.genitalium* and *Chlamydia trachomatis* in the study sample?

Key messages

- At 3% prevalence *Mycoplasma genitalium* is a relatively common infection amongst women participating in the National Chlamydia Screening Programme and attending STI clinics in London.
- *M.genitalium* is more prevalent and has a higher mean bacterial load in cervical and self-taken vaginal swabs than in first catch urine samples.
- Only 0.5% of women had both chlamydia and *M.genitalium* infection and 9% of women with chlamydia had *M.genitalium* infection. Chlamydia treatment is therefore likely to have little impact on treating *M.genitalium* infection overall. In women with both infections antimicrobial treatment for chlamydia is likely

1
2
3 to be sub-optimal treatment for *M.genitalium* with the risk of increasing
4
5 macrolide resistance.
6
7
8

9
10 **Strengths and limitations**

11 **Strengths**

- 12
13
14
15 • This is the largest UK based *M. genitalium* prevalence study to date to provide
16 estimates for both community and STI clinic based populations.
17
18
19
20 • *M.genitalium* PCR results were confirmed positive by genotype sequencing.
21
22

23 **Limitations**

- 24
25
26 • Our analysis of potential correlates for *M.genitalium* and *C.trachomatis* is
27 limited by availability of data.
28
29
30

31 **Competing interests**

32 None
33

34 **Funding**

35
36
37 This work was supported by UCLH/UCL Comprehensive Biomedical Research Centre grant
38 no. 59.
39
40
41

42 **Data sharing statement**

43
44 There is no additional data available
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

INTRODUCTION

Pelvic inflammatory disease (PID) and its sequelae (chronic pelvic pain, ectopic pregnancy and tubal infertility) are major causes of morbidity in women in developed and developing countries.¹ In the USA more than \$10 billion is spent annually in treating these conditions.² *Chlamydia trachomatis* and *Neisseria gonorrhoeae*, two sexually transmitted infections (STIs) are known causes of PID. However in up to 70% of PID cases no cause is found³ and there is increasing evidence that *Mycoplasma genitalium* might be a cause of PID.⁴⁻⁸

There is also strong evidence that it is sexually transmitted.^{5;6} It is significantly associated with endometritis and⁹ tubal factor infertility¹⁰ although the association with cervicitis is complex.^{11;12} As with *C.trachomatis* it can be asymptomatic, acting as a reservoir for further spread.¹³ It may also be associated with human immunodeficiency virus acquisition.¹⁴

Although at present *M.genitalium* is not routinely tested for in most countries, there is interest in introducing testing and treatment. However, before this is done there is a need to gain a better understanding of the infection to avoid repeating the problems encountered with *C. trachomatis* screening.¹⁵ In the United Kingdom (UK) there are few data on the prevalence of *M.genitalium* infection in different population groups of women. Oakeshott *et al.* found that *M.genitalium* prevalence was 3.3% among young women in a community based sample who took part in a *C. trachomatis* screening trial in the UK.¹⁶ Estimates from studies in other countries indicate that the prevalence of *M. genitalium* is 40% to 60% lower than the prevalence of *C. trachomatis*, with little co-infection.^{17;18} The recommended treatment for uncomplicated chlamydia infection is a single dose of azithromycin 1g stat. There is growing evidence of considerably lower *M.genitalium* cure rates with this dose of azithromycin

1
2
3 compared with *C. trachomatis* (79-87% vs. 92-97%, respectively).¹⁹⁻²¹ This may be due to
4
5 genotypic variations in *M.genitalium* resistance to antibiotic treatment and an extended course
6
7 of azithromycin or moxifloxacin has been shown to have superior cure rates.^{6;11}
8
9

10 We investigated the prevalence of *M.genitalium* by real-time PCR and determined its
11
12 correlates in the largest *M. genitalium* prevalence study among women screened for *C.*
13
14 *trachomatis* in the National Chlamydia Screening Programme (NCSP) and STI clinics in the
15
16 United Kingdom (UK).
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

METHODS

Patients and specimens

We used an unlinked anonymised method to test routinely collected and stored cervical swabs, self-taken vaginal swabs and first catch urine samples for *M.genitalium*. The samples were from 2180 women aged 15 to 64 years who had *C. trachomatis* screening when they attended two STI clinics in central and North London and 2455 women aged 15 to 24 years who participated in the NCSP in London in a four month period in 2009. Each clinic offers comprehensive STI screening, treatment and partner notification services to symptomatic and asymptomatic women and men, irrespective of age. Samples from all female clinic attendees were eligible for the study. The NCSP is a national screening programme for chlamydia in the UK among women and men who are under 25 years old in age. The NCSP samples were from a variety of low and high STI risk settings within two London boroughs. In 2009 the majority of participating sites from which the samples were tested were family planning clinics (47%), universities (17%) and general practices (16%). Other testing sites included pharmacies, abortion services, outreach, young persons' services, schools and postal testing (Tina Sharp, NCSP Chlamydia Co-ordinator, personal communication).

The samples were originally collected from the NCSP and clinics and transported to the microbiology laboratory at University College London Hospital in 3 mL (self-taken vaginal and cervical swabs) or 4 mL (urine samples diluted 1:1) of APTIMA transport medium (Gen-Probe Inc., San Diego, USA) for routine *C. trachomatis* testing. After *C. trachomatis* testing the negative samples were stored for 6 weeks at -20°C and positive samples were stored for 3 months at -20°C before they were released for testing as part of this study. Available demographic, sexual behaviour, clinical PID diagnosis and sexually

1
2
3 transmitted infections data were recorded before samples were unlinked from all personal
4
5 identifiers prior to *M.genitalium* testing.
6
7

8 ***M. genitalium* testing**

9
10 Samples were thawed and DNA from 200 µL of the APTIMA transport medium was purified
11
12 by BioRobot 9604 automated workstation using the QIAamp® Virus BioRobot® 9604 Kit
13
14 (QIAGEN, Hilden, Germany). Before freezing and storing the eluate at -20°C it was tested by
15
16 quantitative PCR (qPCR) adapted from a method by Jensen *et al.*^{17;22} The qPCR targeted the
17
18 MgPa adhesion gene (MG191) using MgPa-355FW and MgPa-432R primers and MgPa-380
19
20 MGB probe (primers and probes were provided by Applied Biosystems, Warrington, UK).
21
22 Pilot laboratory work showed no difference in Aptima transport medium and PBS spiked
23
24 with *M.genitalium* DNA in different concentrations.
25
26
27
28

29 We introduced a degenerate oligonucleotide ('wobble') in the forward primer to
30
31 account for a frequent detected base substitution that has previously been shown to be
32
33 successful in another study by Chalker *et al.*²³ As an internal control for PCR inhibition we
34
35 used murine CMV (mCMV) and primers mCMVTAQ1 (forward primer) and mCMVTAQ2
36
37 (reverse primer) and mCMVTAQPR probe labelled with JOE (Primers and probe were
38
39 provided by Eurofins MWG Operon) designed by Garson *et al.*²⁴ The qPCR assays were
40
41 performed in 25 µL volumes; comprising 1x EXPRESS qPCR Supermix (Universal,
42
43 Invitrogen™, Life technologies Ltd. Paisley, UK), 0.4 µM forward and reverse primers, 0.2
44
45 µM probes and 7.5 µL of samples, and nuclease-free water (Promega UK Ltd., Southampton,
46
47 UK).
48
49
50
51

52 Thermal cycling was performed on an ABI 7500 Real-time PCR instrument using the
53
54 following conditions: hotstart at 95°C for 2 min and 1 cycle, denaturation at 95°C for 15 sec,
55
56 annealing and extension at 60°C for 1 min and 45 cycles. The data was analysed using
57
58
59
60

1
2
3 Sequence Detection Software (SDS) version 1.4 with manual baseline/threshold settings to
4
5 estimate quantification cycle.
6
7

8 Positive samples were re-extracted and retested by qPCR. If these tested negative the
9
10 samples was re-extracted and tested by qPCR a third time. If negative again the sample was
11
12 considered equivocal and was excluded from the analysis.
13
14

15 ***M. genitalium* genotyping**

16
17 *M. genitalium* PCR positive samples were sequenced by MgPa1-3 typing assay
18 according to Hjort *et al.* 2006.²⁵ The assay was modified with respect to PCR reagents and
19
20 PCR conditions. In a total volume of 50 µL the following were mixed: 25 µL of Taq PCR
21
22 Master Mix kit (QIAGEN, Hilden, Germany), 0.4 µM of mgpa-1 and mgpa-3 primers, 5 µL
23
24 of template, and nuclease-free water. To increase the sensitivity of the assay 10 µL of the
25
26 template was used in cases where the bacterial load was less than 1 genome copy per µL.
27
28
29
30
31

32 The PCR was performed on an ABI9700 instrument and in 3- step cycling conditions:
33
34 denaturation at 94 °C for 1 min, annealing at 60 °C for 1 min and extension at 72 °C for 1 min
35
36 and 50 cycles.
37
38

39 The amplified product were purified manually by QIAquick PCR purification kit (QIAGEN,
40
41 Hilden, Germany) and sent to the UCL sequencing service for sequencing of both the forward
42
43 and reverse strand.
44
45
46
47

48 **Statistical analysis**

49
50 We have only included data from women who are at least 15 years old in the analysis.
51
52 Data were analysed using SPSS® 14.0 for Windows. Paired sample T-test was used to
53
54 compare the difference of mean values. Multiple logistic regression analysis was used to
55
56
57
58
59
60

investigate the relationship between *M. genitalium* or *C. trachomatis* infection and demographic and sexual behaviour characteristics in women attending NCSP or STI clinics.

Categorical variables in the NCSP model included participant age, specimen type, a new sexual partner within three months, more than one partner within 12 months and ethnicity. The categorical variables included in the STI model were participant age, specimen type, current STI infections and ethnicity. Prevalence, odds ratios adjusted for age (aOR) and 95% confidence intervals (CIs) were calculated and values of $p < 0.05$ were considered statistically significant.

Ethics approval

On the advice of the chair of the local ethics committee, ethical approval was not required since the study team received anonymised samples for testing in the study from the laboratory and no other identifiable data were available.

RESULTS

Of 4635 samples, we excluded 21 samples for which the *M. genitalium* test result was equivocal and included 4613 samples in our analysis (figure 1).

NCSP participants were aged 15 to 25 years whereas STI clinic attendees were aged 15 to 64 years. Women attending the two clinics had significantly different mean ages (26.3 years, SD 7.7 vs. 28.6 years, SD 7.4 years, $p < 0.0001$). The highest prevalence of *M. genitalium* and *C. trachomatis* was in age groups 15 to 24 years in NCSP and the STI clinics. As we only had ethnicity data for 39% (851/2172) of the STI clinic attendees, we did not compare ethnicity across the clinics.

M. genitalium and *C. trachomatis* prevalence

As shown in table 1, the overall prevalence of *M. genitalium* and *C. trachomatis* was 3.0% (138/4613, 95% CI 2.5-3.5%) and 5.4% (249/4613, 95% CI 4.8-6.1%), respectively.

The overall co-infection rate was 0.5% (23/4613, 95% CI 0.3-0.7%). Of 249 women with *C. trachomatis*, 23 (9%) women had *M. genitalium* infection.

Among NCSP participants, *M. genitalium* and *C. trachomatis* frequency were 2.3% (57/2441, 95% CI 1.7-2.9%) and 6.8% (166/2441), 95% CI 5.8-7.8%), respectively.

Table 1. *M. genitalium* and *C. trachomatis* prevalence among NCSP and STI clinic attendees

Infection	Clinic 2 N=716 N (%), 95% CI)	Clinic 1 N=1456 N (%), 95% CI)	NCSP N=2441 N (%), 95% CI)	Total N=4613 N (%), 95% CI)
<i>M. genitalium</i> and <i>C. trachomatis</i>	3 (0.4, 0-0.9)	4 (0.3, 0-0.6)	16 (0.7, 0.4-1.0)	23 (0.5, 0.3-0.7)
Total <i>M. genitalium</i>	38 (5.3, 3.7-7.0)	43 (3.0, 2.0-3.9)	57 (2.3, 1.7-2.9)	138 (3.0, 2.5-3.5)
<i>M. genitalium</i> only	35 (4.9, 3.3-6.5)	41 (2.8, 2.0-3.7)	39 (1.6, 1.1-2.1)	115 (2.5, 2.0-2.9)
Total <i>C. trachomatis</i>	23 (3.2, 1.9-4.5)	60 (4.1, 3.1-5.1)	166 (6.8, 5.8-7.8)	249 (5.4, 4.8-6.1)
<i>C. trachomatis</i> only	20 (2.8, 1.6-4.0)	56 (3.8, 2.9-4.8)	150 (6.1, 5.2-7.1)	226 (4.9, 4.3-5.5)

M. genitalium infection significantly differed between the two clinics (5.3%, 95% CI 3.7-7.0% and 3.0%, 95% CI 2.1-3.8%, $p<0.01$) but the difference was not significant after adjusting for age ($p=0.16$). *C. trachomatis* did not differ significantly between the two clinics (3.2%, 95% CI 1.9-4.5% and 4.1%, 95% CI 3.1-5.1%, $p=0.30$).

1
2
3 **Association of *M. genitalium* and *C. trachomatis* with sexual behaviour and demographic**
4 **characteristics of participants in the National Chlamydia Screening Programme (NCSP)**
5
6
7

8 Table 2 shows the association of *M. genitalium* and *C. trachomatis* with sexual
9 behaviour and demographic characteristics among NCSP participants. *M. genitalium* was less
10 frequently detected than *C. trachomatis* in both age groups (15-19 years old 2.8%, 29/1045 vs.
11 8.3%, 83/1045 and 20-24 years old 2.0%, 28/1396 vs. 5.7%, 79/1396, respectively). When
12 adjusted for age *M. genitalium* was significantly more common in black/black British women
13 compared with white women (aOR 2.3, 95% CI 1.2-4.5, p=0.01). Women who reported
14 multiple sexual partners in the past twelve months were twice as likely to have both
15 *M. genitalium* and *C. trachomatis* infections compared with women who reported only one
16 partner (aOR 2.4, 95% CI 1.3-4.4, p=0.01) and (aOR 2.0, 95% CI 1.4-2.8, p<0.01),
17 respectively. Women who reported new sexual partners in the previous three months were
18 also more likely to have *C. trachomatis* infection (aOR 1.6, 95% CI 1.1-2.3, p=0.01). Those
19 who did not self-identify as white, black/black British, Asian/Asian British or mixed ethnicity
20 were less likely to be infected with *C. trachomatis* compared with white women (aOR 0.6,
21 95% CI 0.4-0.9, p=0.01).
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Table 2. Association of characteristics with *M. genitalium* and *C. trachomatis* in NCSP attendees

Characteristic	(N=2441) % of women with characteristic	<i>M.genitalium</i> % (proportion of women)	aOR ^a (95% CI)	P-value	<i>C.trachomatis</i> % (proportion of women)	aOR ^a (95% CI)	P-value
Age:							
15-19	41.6	2.8 (29/1045)			8.3 (87/1045)		
20-24	56.5	2.0 (28/1396)			5.7 (79/1396)		
Ethnicity							
White	46.6	2.0 (23/1138)	1		7.4 (84/1138)	1	
Black or Black British	12.8	4.8 (15/314)	2.3 (1.2-4.5)	0.01	8.3 (26/314)	1.1 (0.7-1.7)	0.83
Asian or Asian British	4.4	1.9 (2/108)	0.9 (0.2-4.0)	0.93	6.2 (5/108)	0.6 (0.3-1.6)	0.33
Mixed	7.7	3.7 (7/187)	1.8 (0.8-4.3)	0.18	10.2 (19/187)	1.3 (0.8-2.3)	0.29
Other Ethnic Groups	28.4	1.4 (10/694)	0.7 (0.3-1.5)	0.35	4.6 (32/694)	0.6 (0.4-0.9)	0.01
New sexual partner in previous 3 months							
Yes	31.5	3.2(25/770)	1.5 (0.8-2.6)	0.20	9.2 (71/770)	1.6 (1.1-2.3)	0.01
No	39.3	2.2 (21/959)	1		5.8 (56/959)	1	
Don't want to answer	0.2	0.0 (0/6)	-	-	0.0 (0/6)	-	-
Not filled in	28.9	1.6 (11/706)	0.7 (0.3-1.4)	0.33	5.5 (39/706)	0.9 (0.6-1.4)	0.69
Sex with > 1 partner within 12 months							
Yes	30.8	3.9 (29/751)	2.4 (1.3-4.4)	0.01	10.0 (75/751)	2.0 (1.4-2.8)	<0.01
No	39.5	1.7 (16/963)	1		5.4 (52/963)	1	
Don't want to answer	0.3	0.0 (0/8)	-	-	0.0 (0/8)	-	-
Not filled in	29.5	1.7 (12/719)	1.0 (0.5-2.1)	0.99	5.4(39/719)	1.0 (0.6-1.5)	0.99
Specimen							
Cervical/	1.3	3.2 (1/31)	3.3 (0.4-25.8)	0.26	9.7 (3/31)	2.0 (0.6-7.4)	0.21
Self-taken vaginal	40.4	4.2 (41/986)	4.2 (2.3-7.6)	<0.001	9.3 (92/986)	2.0 (1.5-2.8)	<0.001
First catch urine	58.3	1.0 (15/1424)	1		5.0 (71/1424)	1	

aOR^a odds ratios adjusted for age only

Association of *M. genitalium* and *C. trachomatis* with sexual behaviour and demographic characteristics of STI clinic attendees

Table 3 shows the association of *M. genitalium* and *C. trachomatis* with sexual behaviour and demographic characteristics among STI clinic attendees. The age distribution for both *M. genitalium* and *C. trachomatis* was similar with infections more frequently detected in younger women (15 to 19 years 9.7%, 18/186 vs. 6.4%, 12/186, respectively and 20 to 24 years 6.2%, 41/665 vs. 6.0%, 40/665) than other age groups. *M. genitalium* was more frequently detected in 15 to 19 year old women than *C. trachomatis* although this was not statistically significant ($p=0.28$).

Table 3. Association of characteristics with *M. genitalium* and *C. trachomatis* in women attending two London STI clinics

Characteristic	(N=2172) % of women with characteristic	<i>M. genitalium</i> (%) (proportion of women)	aOR ^a (95% CI)	P-value	<i>C. trachomatis</i> proportion of women)	aOR ^a (95% CI)	P-value
Age:							
15-19	8.6	9.7 (18/186)			6.4 (12/186)		
20-24	30.6	6.2 (41/665)			6.0 (40/665)		
25-29	28.6	1.6 (10/621)			2.9 (18/621)		
30-34	15.6	2.3 (9/339)			3.2 (11/339)		
35- 64	16.6	0.8 (3/361)			0.6 (2/361)		
Ethnicity							
White	23.0	6.0 (30/499)	1		7.0 (35/499)	1	
Black or Black British	6.9	7.4 (11/149)	1.2 (0.6-2.5)	0.60	4.0 (6/149)	0.5 (0.2-1.3)	0.54
Asian or Asian British	1.7	17.6 (6/36)	3.1 (1.2-8.1)	0.19	5.6 (2/36)	0.8 (0.2-3.4)	0.73
Mixed	3.9	4.8 (4/84)	0.7 (0.2-2.1)	0.54	7.1 (6/84)	0.9 (0.4-2.3)	0.91
Other Ethnic groups	3.9	9.5 (8/83)	1.6 (0.7-3.7)	0.24	3.6 (3/83)	0.5 (0.1-1.6)	0.49
Unknown	60.8	1.7 (22/1321)	0.5 (0.2-1.1)	0.09	2.3 (31/1321)	0.7 (0.3-1.4)	0.66
Specimen							
Cervical/	90.3	3.8 (75/1961)	1.4 (0.6-3.2)	0.48	3.4 (38/1130)	0.7 (0.4-1.6)	0.44
Self-taken vaginal					4.3 (36/831)		
First catch urine	9.7	2.8 (6/211)	1		4.3 (9/211)	1	

aOR^a odds ratios adjusted for age only

Specimen type and bacterial load

Overall *M. genitalium* was detected in 3.7% (43/1161), 4.0% (74/1817) and 1.3% (21/1635) of cervical swabs, self-taken vulval swabs and first-void urine samples, respectively. Since *M. genitalium* frequency in cervical and self-taken swabs was similar ($p=0.86$), the results for the two groups of swabs were merged and tested against first-void urine samples in the statistical model. *M. genitalium* was significantly more likely to be detected in swabs compared with urine specimens (3.9% vs. 1.3%, $P<0.001$).

The overall prevalence of *C. trachomatis* in cervical swabs, self-taken vulval swabs and first-void urine samples was 3.5% (41/1161), 7.0% (128/1817) and 4.9% (80/1635), respectively. *C. trachomatis* significantly differed between cervical and self-taken swabs ($p<0.001$) and the two groups were separately tested against the urine samples in the statistical model.

The majority (58%, 1424/2441) of specimens provided by the women in NCSP were urine samples. However swab samples were almost four times more likely to test positive for *M. genitalium* compared with urine samples (aOR 3.6, 95% CI 1.9-6.7, $p<0.001$) and *C. trachomatis* prevalence was almost twice as high among swabs compared with urine samples (aOR 1.8, 95% CI 1.2-2.4 $p=0.001$). Conversely the majority (90.3%, 1961/2172) of clinic specimens were swabs. *M. genitalium* and *C. trachomatis* in the clinic swab and urine specimens also differed (*M. genitalium* 3.8%, 75/1961 vs. 2.8%, 6/211 and *C. trachomatis* 3.8%, 74/1961 vs. 4.3%, 9/211, respectively).

In quantitative analysis of *M. genitalium* positive specimens, mean *M. genitalium* bacterial load in swab and urine samples did not significantly differ between the clinics or NCSP. Clinic data were therefore combined for comparison of the mean bacterial load in different specimen types. There was no difference in overall cervical and self-taken vaginal

1
2
3 swab bacterial loads (3.72 (CI 3.39-4.05) vs. 3.91 (CI 3.66-4.17) \log_{10} genome copies/mL,
4
5 equivalent to geometric means of 5,218 (CI 2,438-11,171) and 8,192 (CI 4,575-14,669)
6
7 organisms/mL, respectively) ($p=0.349$). The overall mean bacterial load in swabs 3.84 (CI
8
9 3.52-4.11) equivalent to 6,705 (CI 3506-12,920) organisms/mL was significantly higher than
10
11 in first-void urine samples (3.14 (CI 2.87-3.41) equivalent to 1386 (CI 740-2,597)
12
13 organisms/mL) ($p<0.0001$, equal variances not assumed).
14
15

16 17 **Genetic diversity**

18
19
20 The absence of false positive results was confirmed by the presence of 57 different
21
22 genotypes by sequence analysis of 127 *M. genitalium* positive specimens and 13 sequences
23
24 from previously isolated strains (figure 2). The discriminatory index by Hunter and Gaston *et*
25
26 *al.* 1988²⁶ was calculated to be 0.94 both with and without inclusion of the previously
27
28 isolated strain sequences. None of the sequenced samples were identical with the type strain
29
30 G37 used as a PCR standard control. Genetic diversity data are available in FASTA format
31
32 for download in the supplementary material.
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

DISCUSSION

Overall *M.genitalium* was relatively common with a prevalence of 3.0% among NCSP participants and STI clinic attendees. *M.genitalium* was more likely to be found in swabs compared with urine samples (3.9% vs. 1.3%, respectively) and the mean bacterial load was also much higher (6,705 (CI 3,506-12,920) organisms/mL vs. 1386 (CI 740-2,597) organisms/mL, respectively).

Only 0.5% of all the women had both *C. trachomatis* and *M.genitalium* infections. Among women who had *C. trachomatis*, 9% were co-infected with *M.genitalium*. Among NCPS participants the age-adjusted odds of detecting *M.genitalium* were twice as high among women of black/black British ethnicity (aOR 2.3) and those reporting multiple sexual partners in the past year (aOR 2.4) compared with women of white ethnicity or those who reported only one partner, respectively. After adjusting for age, *C. trachomatis* was also significantly more likely to be diagnosed in women with multiple partners (aOR 2.0) and new sexual partners in the previous three months (aOR 1.6) but was less likely to be detected in women who did not give a self-identified ethnic group (aOR 0.6) compared with reporting only one partner, not reporting new partners or being of white ethnicity, respectively. No significant associations were observed for either infection among STI clinic attendees. However among STI clinic attendees *M.genitalium* was as, if not more likely, to be detected as *C. trachomatis* among women aged 15-24 years (15-19 years 9.7% vs.6.4% and 20-24 years 6.2% vs. 6.0%, respectively). It was also more likely to be detected among STI clinic attendees aged 15-24 years compared with NCSP participants (15-19 years 9.7% vs.2.8% and 20-24 years 6.2% vs. 2.0%, respectively).

This is the largest UK based *M. genitalium* study to date to provide prevalence estimates for both community and STI clinic based populations. Transport media may affect

1
2
3 the sensitivity of DNA based PCR tests. The study samples were originally collected in
4
5 Aptima medium. We therefore tested Aptima and PBS media with *M.genitalium* DNA and
6
7 did not find any differences. We confirmed positive *M.genitalium* PCR results by genotype
8
9 sequencing. Our analysis of *M.genitalium* and *C.trachomatis* correlates is limited by
10
11 availability of data: only age and ethnicity were available for both clinic and NCSP datasets
12
13 and ethnicity data was missing for 61% of STI clinic attendees. There is also a possibility that
14
15 some young women may have had chlamydia tests through both the NCSP and the STI
16
17 clinics during the sample collection period. It is not possible to quantify this although we
18
19 speculate that the numbers are likely to be low given the relatively short time frame.
20
21

22
23
24 Our STI clinic *M.genitalium* prevalence is similar to that found in several studies of
25
26 female STI clinic attendees (4.5% to 7%)^{27;28} although other studies have reported a much
27
28 higher prevalence (19.3% to 38.2%).^{29;30} In lower risk non-STI clinic attendees such as
29
30 college students the prevalence has been shown to range from <1% to 5%.^{5;31} In one clinic in
31
32 our study *M.genitalium* prevalence was higher than *C. trachomatis* and the lower *C.*
33
34 *trachomatis* prevalence may reflect variation during the short study period as may the higher
35
36 *M.genitalium* prevalence. We speculate that it may also be due to variations in chlamydia
37
38 screening uptake and therefore *C. trachomatis* prevalence in different parts of London.
39
40
41

42
43 The higher prevalence of *M.genitalium* in women attending clinics than the NCSP (3-
44
45 5.3% vs. 2.3%, respectively) may in part reflect the higher proportion of swabs taken in
46
47 clinics than in NCSP settings. Urine samples have been shown to be less sensitive for
48
49 *M.genitalium* diagnosis than swabs (61% to 65% compared with 74% to 91%).^{32;33} It is
50
51 therefore likely that our NCSP *M.genitalium* prevalence is an underestimation. Although urine
52
53 sample sensitivity may be increased by up-concentrating the samples by centrifugation this is
54
55 not a practical step for large scale testing. A higher bacterial load may be associated with
56
57 symptoms as has been shown for men.²² This may also explain the difference in prevalence
58
59
60

1
2
3 between the two populations since STI clinic attendees are more likely to be symptomatic
4
5 than NCSP participants. The association of *M.genitalium* with multiple sexual partners and
6
7 black ethnicity has been previously observed.^{16,34} Additional risk factors include younger age
8
9 as observed in our STI clinic attendees, bacterial vaginosis, being symptomatic, cervicitis,
10
11 douching, smoking, prior miscarriage, menstrual cycle, social class and marital
12
13 status.^{12;16;30;34-37}

14
15
16
17 *M.genitalium* appears to be a relatively common infection among women in London.
18
19 The low level of *M.genitalium* and *C. trachomatis* co-infection (0.5%) suggests that
20
21 diagnosing and treating chlamydia will have little impact on *M.genitalium*. However
22
23 Azithromycin 1g used to treat uncomplicated *C. trachomatis* infection appears to be sub-
24
25 optimal for *M.genitalium* treatment³⁸

26
27
28
29 To avoid the problems encountered with *C. trachomatis* screening and *M.genitalium*
30
31 antimicrobial resistance, prior to introducing routine testing for *M.genitalium*, further research
32
33 is needed to better understand its natural history, the role of asymptomatic and symptomatic
34
35 *M.genitalium* in PID and determine optimum treatment guidelines.
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Acknowledgements

The authors would like to thank Menelaos Pavlou (Centre for Sexual Health and HIV Research, Mortimer Market Centre, London, UK) and Tina Sharp (Mortimer Market Centre, London, UK, previously NCSP Chlamydia Co-ordinator, Camden Primary Care Trust) for extracting clinical data from the Sexually Transmitted Disease Clinics and the National Chlamydia Screening Programme, respectively. We thank Dr. Stephane Hue, Center for Virology, UCL for aligning the sequenced fragments and create the resulting phylogenetic tree.

Authors Contribution

All authors contributed to conception and design of the study and / or to acquisition of data. HS performed the experiments. HS SSD and JS drafted the paper and all authors contributed to critical revision of the paper.

Reference List

- 1
2
3
4
5
6
7 (1) Barrett S, Taylor C. A review on pelvic inflammatory disease. *Int J STD AIDS* 2005; 16(11):715-
8 720.
- 9
10 (2) Simms I, Stephenson JM. Pelvic inflammatory disease epidemiology: what do we know and
11 what do we need to know? *Sex Transm Infect* 2000; 76(2):80-87.
- 12
13 (3) Haggerty CL, Ness RB. Diagnosis and treatment of pelvic inflammatory disease. *Womens*
14 *Health (Lond Engl)* 2008; 4(4):383-397.
- 15
16 (4) Haggerty CL, Taylor BD. Mycoplasma genitalium: an emerging cause of pelvic inflammatory
17 disease. *Infect Dis Obstet Gynecol* 2011; 2011:959816.
- 18
19 (5) McGowin CL, Anderson-Smits C. Mycoplasma genitalium: an emerging cause of sexually
20 transmitted disease in women. *PLoS Pathog* 2011; 7(5):e1001324.
- 21
22 (6) Taylor-Robinson D, Jensen JS. Mycoplasma genitalium: from Chrysalis to multicolored
23 butterfly. *Clin Microbiol Rev* 2011; 24(3):498-514.
- 24
25 (7) Bjartling C, Osser S, Persson, K. The association between Mycoplasma genitalium and pelvic
26 inflammatory disease after termination of pregnancy. *BJOG* 2010; 117(3):361-364.
- 27
28 (8) Bjartling C, Osser S, Persson, K. Mycoplasma genitalium in cervicitis and pelvic inflammatory
29 disease among women at a gynecologic outpatient service. *Am J Obstet Gynecol*
30 2012;206(6):476-478.
- 31
32 (9) Cohen CR, Manhart LE, Bukusi EA, Astete S, Brunham RC, Holmes KK et al. Association
33 between Mycoplasma genitalium and acute endometritis. *Lancet* 2002; 359(9308):765-766.
- 34
35 (10) Svenstrup HF, Fedder J, Kristoffersen SE, Trolle B, Birkelund S, Christiansen G. Mycoplasma
36 genitalium, Chlamydia trachomatis, and tubal factor infertility - a prospective study. *Fertility*
37 *and Sterility* 2008; 90(3):513-520.
- 38
39 (11) Manhart LE, Broad JM, Golden MR. Mycoplasma genitalium: should we treat and how? *Clin*
40 *Infect Dis* 2011;53(s3):s129-s142.
- 41
42 (12) Mobley VL, Hobbs MM, Lau K, Weinbaum BS, Getman DK, Sena AC. Mycoplasma genitalium
43 infection in women attending a sexually transmitted infection clinic: diagnostic specimen
44 type, coinfections, and predictors. *Sex Transm Dis* 2012; 39(9):706-709.
- 45
46 (13) Jensen J S. Mycoplasma genitalium: the aetiological agent of urethritis and other sexually
47 transmitted diseases. *J Eur Acad Dermatol Venereol* 2004; 18(1):1-11.
- 48
49 (14) Mavedzenge SN, Van Der PB, Weiss HA, Kwok C, Mambo F, Chipato T et al. The association
50 between Mycoplasma genitalium and HIV-1 acquisition in African women. *AIDS* 2012;
51 26(5):617-624.
- 52
53 (15) Low N, Bender N, Nartey L, Shang A, Stephenson JM. Effectiveness of chlamydia screening:
54 systematic review. *International Journal of Epidemiology* 2009; 38(2):435-448.
- 55
56
57
58
59
60

- 1
2
3 (16) Oakeshott P, Aghaizu A, Hay P, Reid F, Kerry S, Atherton H et al. Is *Mycoplasma genitalium* in
4 women the "New Chlamydia?" A community-based prospective cohort study. *Clin Infect Dis*
5 2010; 51(10):1160-1166.
6
7 (17) Anagrus C, Lore B, Jensen JS. *Mycoplasma genitalium*: prevalence, clinical significance, and
8 transmission. *Sex Transm Infect* 2005; 81(6):458-462.
9
10 (18) Andersen B, Sokolowski I, Ostergaard L, Moller JK, Olesen F, Jensen JS. *Mycoplasma*
11 *genitalium*: prevalence and behavioural risk factors in the general population. *Sexually*
12 *Transmitted Infections* 2007; 83(3):237-241.
13
14 (19) Jensen JS, Bradshaw CS, Tabrizi SN, Fairley CK, Hamasuna R. Azithromycin Treatment Failure
15 in *Mycoplasma genitalium* Positive Patients with Nongonococcal Urethritis Is Associated
16 with Induced Macrolide Resistance. *Clinical Infectious Diseases* 2008; 47(12):1546-1553.
17
18 (20) Lau CY, Qureshi AK. Azithromycin Versus Doxycycline for Genital Chlamydial Infections: A
19 Meta-Analysis of Randomized Clinical Trials. *Sexually Transmitted Diseases* 2002; 29(9):497-
20 502.
21
22 (21) Horner PJ. Azithromycin antimicrobial resistance and genital Chlamydia trachomatis
23 infection: duration of therapy may be the key to improving efficacy. 2012; 88(3):154-156.
24
25 (22) Jensen JS, Bjornelius E, Dohn B, Lidbrink P. Use of TaqMan 5' nuclease real-time PCR for
26 quantitative detection of *Mycoplasma genitalium* DNA in males with and without urethritis
27 who were attendees at a sexually transmitted disease clinic. *J Clin Microbiol* 2004; 42(2):683-
28 692.
29
30 (23) Chalker VJ, Jordan K, Ali T, Ison C. Real-time PCR detection of the mg219 gene of unknown
31 function of *Mycoplasma genitalium* in men with and without non-gonococcal urethritis and
32 their female partners in England. *J Med Microbiol* 2009; 58(7):895-899.
33
34 (24) Garson JA, Grant PR, Ayliffe U, Ferns RB, Tedder RS. Real-time PCR quantitation of hepatitis B
35 virus DNA using automated sample preparation and murine cytomegalovirus internal
36 control. *J Virol Methods* 2005; 126(1-2):207-213.
37
38 (25) Hjorth SV, Bjornelius E, Lidbrink P, Falk L, Dohn B, Berthelsen L et al. Sequence-based typing
39 of *Mycoplasma genitalium* reveals sexual transmission. *J Clin Microbiol* 2006; 44(6):2078-
40 2083.
41
42 (26) Hunter PR, Gaston MA. Numerical index of the discriminatory ability of typing systems: an
43 application of Simpson's index of diversity. *J Clin Microbiol* 1988; 26(11):2465-2466.
44
45 (27) Manhart LE, Critchlow CW, Holmes KK, Dutro SM, Eschenbach DA, Stevens CE et al.
46 Mucopurulent cervicitis and *Mycoplasma genitalium*. *J Infect Dis* 2003; 187(4):650-657.
47
48 (28) Moi H, Reinton N, Moghaddam A. *Mycoplasma genitalium* in women with lower genital tract
49 inflammation. *Sex Transm Infect* 2009; 85(1):10-14.
50
51 (29) Casin I, Vexiau-Robert D, De La SP, Eche A, Grandry B, Janier M. High prevalence of
52 *Mycoplasma genitalium* in the lower genitourinary tract of women attending a sexually
53 transmitted disease clinic in Paris, France. *Sex Transm Dis* 2002; 29(6):353-359.
54
55
56
57
58
59
60

- 1
2
3 (30) Mobley VL, Hobbs MM, Lau K, Weinbaum BS, Getman DK. Mycoplasma genitalium infection
4 in women attending a sexually transmitted infection clinic: diagnostic specimen type,
5 coinfections, and predictors. *Sex Transm Dis* 2012; 39(9):706-709.
6
7 (31) Jensen AJ, Kleveland CR, Moghaddam A, Haaheim H, Hjelmevoll SO, Skogen V. Chlamydia
8 trachomatis, Mycoplasma genitalium and Ureaplasma urealyticum among students in
9 northern Norway. *J Eur Acad Dermatol Venereol* 2013; 27(1):e91-e96.
10
11 (32) Lillis RA, Nsuami MJ, Myers L, Martin DH. Utility of urine, vaginal, cervical, and rectal
12 specimens for detection of Mycoplasma genitalium in women. *J Clin Microbiol* 2011;
13 49(5):1990-1992.
14
15 (33) Wroblewski JK, Manhart LE, Dickey KA, Hudspeth MK, Totten PA. Comparison of
16 transcription-mediated amplification and PCR assay results for various genital specimen
17 types for detection of Mycoplasma genitalium. *J Clin Microbiol* 2006; 44(9):3306-3312.
18
19 (34) Walker J, Fairley CK, Bradshaw CS, Tabrizi SN, Chen MY, Twin J et al. 'The difference in
20 determinants of Chlamydia trachomatis and Mycoplasma genitalium in a sample of young
21 Australian women'. *BMC Infect Dis* 2011; 11:35.
22
23 (35) Short VL, Totten PA, Ness RB, Astete SG, Kelsey SF, Murray P et al. The demographic, sexual
24 health and behavioural correlates of Mycoplasma genitalium infection among women with
25 clinically suspected pelvic inflammatory disease. *Sex Transm Infect* 2010; 86(1):29-31.
26
27 (36) Oakeshott P, Hay P, Taylor-Robinson D, Hay S, Dohn B, Kerry S et al. Prevalence of
28 Mycoplasma genitalium in early pregnancy and relationship between its presence and
29 pregnancy outcome. *BJOG* 2004; 111(12):1464-1467.
30
31 (37) Vandepitte J, Muller E, Bukenya J, Nakubulwa S, Kyakuwa N, Buve A et al. Prevalence and
32 correlates of Mycoplasma genitalium infection among female sex workers in Kampala,
33 Uganda. *J Infect Dis* 2012; 205(2):289-296.
34
35 (38) Twin J, Jensen JS, Bradshaw CS, Garland SM, Fairley CK, Min LY et al. Transmission and
36 selection of macrolide resistant Mycoplasma genitalium infections detected by rapid high
37 resolution melt analysis. *PLoS One* 2012;7(4):e35593
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

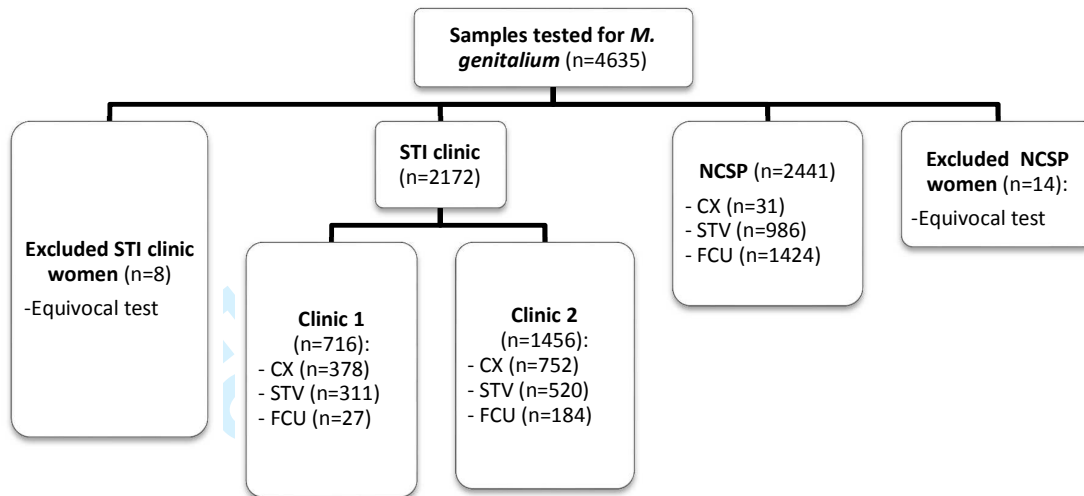


Figure 1. *M. genitalium* prevalence study sample source from the National Chlamydia Screening Programme (NCSP) and sexually transmitted infection clinics (STI) and sample types and numbers tested

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60



Figure 2: Phylogenetic tree showing clustering of 127 DNA sequences from the *M. genitalium* positive specimens of the study (marked with grey font) and 13 DNA sequences from *M. genitalium* strain from patients with no known sexual relationship (marked with black font) 139x198mm (300 x 300 DPI)



A cross-sectional study of Mycoplasma genitalium infection and correlates in women undergoing population based screening or clinic based testing for Chlamydia infection in London

Journal:	<i>BMJ Open</i>
Manuscript ID:	bmjopen-2013-003947.R1
Article Type:	Research
Date Submitted by the Author:	05-Dec-2013
Complete List of Authors:	Svenstrup, Helle; University College London, Institute for Women's Health Dave, Sangeeta; University College London, Institute for Women's Health Carder, Caroline; UCLH Microbiology Lab, Chlamydia Laboratory Grant, Paul; UCLH Microbiology Lab, Chlamydia Laboratory Morris-Jones, Stephen; UCLH Microbiology Lab, Chlamydia Laboratory Kidd, Ian; UCLH Microbiology Lab, Chlamydia Laboratory Stephenson, Judith; University College London, Institute for Women's Health
Primary Subject Heading:	Sexual health
Secondary Subject Heading:	Epidemiology
Keywords:	EPIDEMIOLOGY, GENITOURINARY MEDICINE, MICROBIOLOGY

SCHOLARONE™
Manuscripts

only

1
2
3 **A cross-sectional study of *Mycoplasma genitalium* infection and correlates in women undergoing**
4 **population based screening or clinic based testing for Chlamydia infection in London**
5
6

7 Svenstrup HF¹, Dave SS¹, Carder C², Grant P², Morris-Jones S², Kidd M² and Stephenson JM¹

8
9 ¹Research Department of Reproductive Medicine, Institute for Women's Health, University College
10 London, 65 Whitfield Street, W1T 4EU, UK

11
12 ²Clinical Microbiology and Virology, University College London Hospitals NHS Foundation Trust,
13 65 Whitfield Street, W1T 4EU, UK
14

15
16
17 **Corresponding author:**

18 Dr. Helle F Svenstrup

19 Rigensgade 11, DK-1316 København K, Denmark

20 Fax +45 43 99 99 11

21 Direct phone +45 43 31 33 77,

22 Mobile +45 20 65 12 64

23 hfs@awapatent.com
24
25
26
27
28

29 **Co-authors:**

30 Sangeeta Shashikant Dave, Institute for Women's Health, University College London, London,
31 United Kingdom, sangeeta.dave@ucl.ac.uk
32
33

34 Caroline Carder, Clinical Microbiology and Virology, University College London Hospitals NHS
35 Foundation Trust, London, United Kingdom, caroline.carder@uclh.nhs.uk
36
37

38 Paul Grant, Clinical Microbiology and Virology, University College London Hospitals NHS
39 Foundation Trust London, United Kingdom paul.grant@ucl.ac.uk
40
41

42 Stephen Morris-Jones, Clinical Microbiology and Virology, University College London Hospitals
43 NHS Foundation Trust London, United Kingdom, stephen.morris-jones@uclh.nhs.uk
44
45

46 Ian Kidd, Clinical Microbiology and Virology, University College London Hospitals NHS Foundation
47 Trust, London, United Kingdom, i.kidd@ucl.ac.uk
48
49

50 Judith Stephenson, Institute for Women's Health, University College London, London, United
51 Kingdom, judith.stephenson@ucl.ac.uk
52

53 **Key words:** Genotyping / *Mycoplasma genitalium* / National Chlamydia Screening Programme /
54 Real-time PCR
55
56

57 **Word count: 3049**
58
59
60

ABSTRACT

Objective:

To determine *Mycoplasma genitalium* infection and correlates among young women undergoing population based screening or clinic based testing for chlamydia infection.

Design:

Cross-sectional study

Setting:

National Chlamydia Screening Programme (NCSP) and two London STI clinics

Participants:

2441 women aged 15 to 64 years who participated in the NCSP and 2172 women who attended two London STI clinics over a four month period in 2009.

Outcome measures:

1. *M.genitalium* prevalence in defined populations (%)
2. Age-adjusted odds ratios (aORs) for correlates of *M.genitalium* infection

Results

The overall frequency of *M. genitalium* and *C.trachomatis* was 3.0% and 5.4%, respectively. Co-infection was relatively uncommon (0.5% of all women); however 9% of women with *C.trachomatis* also had *M.genitalium* infection. *M.genitalium* was more frequently detected in swab than urine samples (3.9% vs. 1.3%, $p<0.001$) with a significantly higher mean bacterial load ($p=<0.001$). Among NCSP participants, *M.genitalium* was significantly more likely to be diagnosed in women of black/black British ethnicity (aOR 2.3, 95% CI 1.2-4.5, $p=0.01$). *M.genitalium* and *C. trachomatis* and were both significantly associated with multiple sexual partners in the past year (aOR 2.4, 95% CI 1.3-4.4, $p=0.01$ and aOR 2.0, 95% CI 1.4-2.8, $p<0.01$). Among STI clinic attendees, *M.genitalium* was more common in women who were less than 25 years in age.

Conclusions

M.genitalium is a relatively common infection among young women in London. It is significantly more likely to be detected in vulvo-vaginal swabs than in urine samples. Co-infection with chlamydia is uncommon. The clinical effectiveness of testing and treatment strategies for *M.genitalium* needs further investigation.

Article summary

Article Focus

- How common is *Mycoplasma genitalium* in women in the general population and those attending sexually transmitted infection (STI) clinics in London?
- How are *M.genitalium* infection and bacterial load associated with sample type?
- How much co-infection is there between *M.genitalium* and *Chlamydia trachomatis* in the study sample?

Key messages

- At 3% *Mycoplasma genitalium* is a relatively common infection amongst women participating in the National Chlamydia Screening Programme and attending STI clinics in London.
- *M.genitalium* is more common and has a higher mean bacterial load in cervical and self-taken vaginal swabs than in first catch urine samples.
- Only 0.5% of women had both chlamydia and *M.genitalium* infection and 9% of women with chlamydia had *M.genitalium* infection. Chlamydia treatment is therefore likely to have little impact on treating *M.genitalium* infection overall. In women with both infections antimicrobial treatment for chlamydia is likely

1
2
3 to be sub-optimal treatment for *M.genitalium* with the risk of increasing
4
5 macrolide resistance.
6
7

8
9
10 Strengths and limitations

11 Strengths

- 12
13
14
15 • This is the largest UK based cross-sectional study to date to provide estimates
16
17 of *M. genitalium* prevalence in both community and STI clinic based
18
19 populations.
20
21
22 • *M.genitalium* PCR results were confirmed positive by genotype sequencing.
23
24

25 Limitations

- 26
27
28 • Our analysis of potential correlates for *M.genitalium* and *C.trachomatis* is
29
30 limited by availability of data.
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

INTRODUCTION

Pelvic inflammatory disease (PID) and its sequelae (chronic pelvic pain, ectopic pregnancy and tubal infertility) are major causes of morbidity in women in developed and developing countries.¹ In the USA more than \$10 billion is spent annually in treating these conditions.² *Chlamydia trachomatis* and *Neisseria gonorrhoeae*, two sexually transmitted infections (STIs) are known causes of PID. However in up to 70% of PID cases no cause is found³ and there is increasing evidence that *Mycoplasma genitalium* might be a cause of PID.⁴⁻⁸

There is also strong evidence that it is sexually transmitted.^{5;6} It is significantly associated with endometritis and⁹ tubal factor infertility¹⁰ although the association with cervicitis is complex.^{11;12} As with *C.trachomatis* it can be asymptomatic, acting as a reservoir for further spread.¹³ It may also be associated with human immunodeficiency virus acquisition.¹⁴

Although at present *M.genitalium* is not routinely tested for in most countries, there is interest in introducing testing and treatment. However, before this is done there is a need to gain a better understanding of the infection to avoid repeating the problems encountered with *C. trachomatis* screening.¹⁵ In the United Kingdom (UK) there are few data on the frequency of *M.genitalium* infection in different population groups of women. Oakeshott *et al.* found that *M.genitalium* prevalence was 3.3% among young women in a community based sample who took part in a *C. trachomatis* screening trial in the UK.¹⁶ Estimates from studies in other countries indicate that levels of *M. genitalium* are 40% to 60% lower than *C. trachomatis*, with little co-infection.^{17;18} The recommended treatment for uncomplicated chlamydia infection is a single dose of azithromycin 1g stat. There is growing evidence of considerably lower *M.genitalium* cure rates with this dose of azithromycin compared with *C. trachomatis*

1
2
3 (79-87% vs. 92-97%, respectively).¹⁹⁻²¹ Resistance has been shown to develop following 1g of
4
5 azithromycin and macrolide resistance is endemic in some populations.²²⁻²⁴
6
7

8 We investigated *M.genitalium* infection by real-time PCR and determined its
9
10 correlates in the largest cross-sectional study of *M. genitalium* among women screened for *C.*
11
12 *trachomatis* in the National Chlamydia Screening Programme (NCSP) and STI clinics in the
13
14 United Kingdom (UK).
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

METHODS

Patients and specimens

We used an unlinked anonymised method to test routinely collected and stored cervical swabs, self-taken vaginal swabs and first catch urine samples for *M.genitalium*. The samples were from 2180 women aged 15 to 64 years who had *C. trachomatis* screening when they attended two STI clinics in central and North London and 2455 women aged 15 to 24 years who participated in the NCSP in London in a four month period in 2009. Each clinic offers comprehensive STI screening, treatment and partner notification services to symptomatic and asymptomatic women and men, irrespective of age. Samples from all female clinic attendees were eligible for the study. The NCSP is a national screening programme for chlamydia in the UK among women and men who are under 25 years old in age. The NCSP samples were from a variety of low and high STI risk settings within two London boroughs. In 2009 the majority of participating sites from which the samples were tested were family planning clinics (47%), universities (17%) and general practices (16%). Other testing sites included pharmacies, abortion services, outreach, young persons' services, schools and postal testing (Tina Sharp, NCSP Chlamydia Co-ordinator, personal communication).

The samples were originally collected from the NCSP and clinics and transported to the microbiology laboratory at University College London Hospital in 3 mL (self-taken vaginal and cervical swabs) or 4 mL (urine samples diluted 1:1) of APTIMA transport medium (Gen-Probe Inc., San Diego, USA) for routine *C. trachomatis* testing. After *C. trachomatis* testing the negative samples were stored for 6 weeks at -20°C and positive samples were stored for 3 months at -20°C before they were released for testing as part of this study. Available demographic, sexual behaviour, clinical PID diagnosis and sexually

1
2
3 transmitted infections data were recorded before samples were unlinked from all personal
4
5 identifiers prior to *M.genitalium* testing.
6
7

8 ***M. genitalium* testing**

9
10 Samples were thawed and DNA from 200 µL of the APTIMA transport medium was purified
11
12 by BioRobot 9604 automated workstation using the QIAamp® Virus BioRobot® 9604 Kit
13
14 (QIAGEN, Hilden, Germany). Before freezing and storing the eluate at -20°C it was tested by
15
16 quantitative PCR (qPCR) adapted from a method by Jensen *et al.*^{17;25} The qPCR targeted the
17
18 MgPa adhesion gene (MG191) using MgPa-355FW and MgPa-432R primers and MgPa-380
19
20 MGB probe (primers and probes were provided by Applied Biosystems, Warrington, UK).
21
22 Pilot laboratory work showed no difference in Aptima transport medium and PBS spiked
23
24 with *M.genitalium* DNA in different concentrations.
25
26
27
28

29 We introduced a degenerate oligonucleotide ('wobble') in the forward primer to
30
31 account for a frequent detected base substitution that has previously been shown to be
32
33 successful in another study by Chalker *et al.*²⁶ As an internal control for PCR inhibition we
34
35 used murine CMV (mCMV) and primers mCMVTAQ1 (forward primer) and mCMVTAQ2
36
37 (reverse primer) and mCMVTAQPR probe labelled with JOE (Primers and probe were
38
39 provided by Eurofins MWG Operon) designed by Garson *et al.*²⁷. The qPCR assays were
40
41 performed in 25 µL volumes; comprising 1x EXPRESS qPCR Supermix (Universal,
42
43 InvitrogenTM, Life technologies Ltd. Paisley, UK), 0.4 µM forward and reverse primers, 0.2
44
45 µM probes and 7.5 µL of samples, and nuclease-free water (Promega UK Ltd., Southampton,
46
47 UK).
48
49
50
51

52 Thermal cycling was performed on an ABI 7500 Real-time PCR instrument using the
53
54 following conditions: hotstart at 95°C for 2 min and 1 cycle, denaturation at 95°C for 15 sec,
55
56 annealing and extension at 60°C for 1 min and 45 cycles. The data was analysed using
57
58
59
60

1
2
3 Sequence Detection Software (SDS) version 1.4 with manual baseline/threshold settings to
4
5 estimate quantification cycle.
6
7

8 Positive samples were re-extracted and retested by qPCR. If these tested negative the
9
10 samples was re-extracted and tested by qPCR a third time. If negative again the sample was
11
12 considered equivocal and was excluded from the analysis.
13
14

15 ***M. genitalium* genotyping**

16
17 *M. genitalium* PCR positive samples were sequenced by MgPa1-3 typing assay
18 according to Hjort *et al.* 2006.²⁸ The assay was modified with respect to PCR reagents and
19
20 PCR conditions. In a total volume of 50 µL the following were mixed: 25 µL of Taq PCR
21
22 Master Mix kit (QIAGEN, Hilden, Germany), 0.4 µM of mgpa-1 and mgpa-3 primers, 5 µL
23
24 of template, and nuclease-free water. To increase the sensitivity of the assay 10 µL of the
25
26 template was used in cases where the bacterial load was less than 1 genome copy per µL.
27
28
29
30
31

32 The PCR was performed on an ABI9700 instrument and in 3- step cycling conditions:
33
34 denaturation at 94 °C for 1 min, annealing at 60 °C for 1 min and extension at 72 °C for 1 min
35
36 and 50 cycles.
37
38

39 The amplified product were purified manually by QIAquick PCR purification kit (QIAGEN,
40
41 Hilden, Germany) and sent to the UCL sequencing service for sequencing of both the forward
42
43 and reverse strand.
44
45
46
47

48 **Statistical analysis**

49
50 We have only included data from women who are at least 15 years old in the analysis.
51
52 Data were analysed using SPSS[®] 14.0 for Windows. Paired sample T-test was used to
53
54 compare the difference of mean values. Multiple logistic regression analysis was used to
55
56
57
58
59
60

investigate the relationship between *M. genitalium* or *C. trachomatis* infection and demographic and sexual behaviour characteristics in women attending NCSP or STI clinics.

Categorical variables in the NCSP model included participant age, specimen type, a new sexual partner within three months, more than one partner within 12 months and ethnicity. The categorical variables included in the STI model were participant age, specimen type, current STI infections and ethnicity. Frequency, odds ratios adjusted for age (aOR) and 95% confidence intervals (CIs) were calculated and values of $p < 0.05$ were considered statistically significant.

Ethics approval

On the advice of the chair of the local ethics committee, ethical approval was not required since the study team received anonymised samples for testing in the study from the laboratory and no other identifiable data were available.

RESULTS

Of 4635 samples, we excluded 21 samples for which the *M. genitalium* test result was equivocal and included 4613 samples in our analysis (figure 1).

NCSP participants were aged 15 to 24 years whereas STI clinic attendees were aged 15 to 64 years. Women attending the two clinics had significantly different mean ages (20.1 years, SD 2.5 vs. 27.8 years, SD 7.6 years, $p < 0.0001$). The highest prevalence of *M. genitalium* and *C. trachomatis* was in age groups 15 to 24 years in NCSP and the STI clinics. As we only had ethnicity data for 39% (851/2172) of the STI clinic attendees, we did not compare ethnicity across the clinics.

***M. genitalium* and *C. trachomatis* infection**

As shown in table 1, the overall frequency of *M. genitalium* and *C. trachomatis* was 3.0% (138/4613, 95% CI 2.5-3.5%) and 5.4% (249/4613, 95% CI 4.8-6.1%), respectively. The overall co-infection rate was 0.5% (23/4613, 95% CI 0.3-0.7%). Of 249 women with *C. trachomatis*, 23 (9%) women had *M. genitalium* infection.

Among NCSP participants, *M. genitalium* and *C. trachomatis* frequency were 2.3% (57/2441, 95% CI 1.7-2.9%) and 6.8% (166/2441), 95% CI 5.8-7.8%), respectively.

Table 1. *M. genitalium* and *C. trachomatis* infection among NCSP and STI clinic attendees

Infection	Clinic 2 N=716 N (% , 95% CI)	Clinic 1 N=1456 N (% , 95% CI)	NCSP N=2441 N (% , 95% CI)	Total N=4613 N (% , 95% CI)
<i>M. genitalium</i> and <i>C. trachomatis</i>	3 (0.4, 0-0.9)	4 (0.3, 0-0.6)	16 (0.7, 0.4-1.0)	23 (0.5, 0.3-0.7)
Total <i>M. genitalium</i>	38 (5.3, 3.7-7.0)	43 (3.0 , 2.0-3.9)	57 (2.3, 1.7-2.9)	138 (3.0, 2.5-3.5)
<i>M. genitalium</i> only	35 (4.9, 3.3-6.5)	41 (2.8, 2.0-3.7)	39 (1.6, 1.1-2.1)	115 (2.5, 2.0-2.9)
Total <i>C. trachomatis</i>	23 (3.2, 1.9-4.5)	60 (4.1, 3.1-5.1)	166 (6.8, 5.8-7.8)	249 (5.4, 4.8-6.1)
<i>C. trachomatis</i> only	20 (2.8, 1.6-4.0)	56 (3.8, 2.9-4.8)	150 (6.1, 5.2-7.1)	226 (4.9, 4.3-5.5)

M. genitalium infection significantly differed between the two clinics (5.3%, 95% CI 3.7-7.0% and 3.0%, 95% CI 2.1-3.8%, $p<0.01$) but the difference was not significant after adjusting for age ($p=0.16$). *C. trachomatis* did not differ significantly between the two clinics (3.2%, 95% CI 1.9-4.5% and 4.1%, 95% CI 3.1-5.1%, $p=0.30$).

1
2
3 **Association of *M. genitalium* and *C. trachomatis* with sexual behaviour and demographic**
4 **characteristics of participants in the National Chlamydia Screening Programme (NCSP)**
5
6
7

8 Table 2 shows the association of *M. genitalium* and *C. trachomatis* with sexual
9 behaviour and demographic characteristics among NCSP participants. *M. genitalium* was less
10 frequently detected than *C. trachomatis* in both age groups (15-19 years old 2.8%, 29/1045 vs.
11 8.3%, 83/1045 and 20-24 years old 2.0%, 28/1396 vs. 5.7%, 79/1396, respectively). When
12 adjusted for age *M. genitalium* was significantly more common in black/black British women
13 compared with white women (aOR 2.3, 95% CI 1.2-4.5, p=0.01). Women who reported
14 multiple sexual partners in the past twelve months were twice as likely to have both
15 *M. genitalium* and *C. trachomatis* infections compared with women who reported only one
16 partner (aOR 2.4, 95% CI 1.3-4.4, p=0.01) and (aOR 2.0, 95% CI 1.4-2.8, p<0.01),
17 respectively. Women who reported new sexual partners in the previous three months were
18 also more likely to have *C. trachomatis* infection (aOR 1.6, 95% CI 1.1-2.3, p=0.01). Those
19 who did not self-identify as white, black/black British, Asian/Asian British or mixed ethnicity
20 were less likely to be infected with *C. trachomatis* compared with white women (aOR 0.6,
21 95% CI 0.4-0.9, p=0.01).
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Table 2. Association of characteristics with *M. genitalium* and *C. trachomatis* in NCSP attendees

Characteristic	(N=2441) % of women with characteristic	<i>M.genitalium</i> % (proportion of women)	aOR ^a (95% CI)	P-value	<i>C.trachomatis</i> % (proportion of women)	aOR ^a (95% CI)	P-value
Age:							
15-19	41.6	2.8 (29/1045)			8.3 (87/1045)		
20-24	56.5	2.0 (28/1396)			5.7 (79/1396)		
Ethnicity							
White	46.6	2.0 (23/1138)	1		7.4 (84/1138)	1	
Black or Black British	12.8	4.8 (15/314)	2.3 (1.2-4.5)	0.01	8.3 (26/314)	1.1 (0.7-1.7)	0.83
Asian or Asian British	4.4	1.9 (2/108)	0.9 (0.2-4.0)	0.93	6.2 (5/108)	0.6 (0.3-1.6)	0.33
Mixed	7.7	3.7 (7/187)	1.8 (0.8-4.3)	0.18	10.2 (19/187)	1.3 (0.8-2.3)	0.29
Other Ethnic Groups	28.4	1.4 (10/694)	0.7 (0.3-1.5)	0.35	4.6 (32/694)	0.6 (0.4-0.9)	0.01
New sexual partner in previous 3 months							
Yes	31.5	3.2(25/770)	1.5 (0.8-2.6)	0.20	9.2 (71/770)	1.6 (1.1-2.3)	0.01
No	39.3	2.2 (21/959)	1		5.8 (56/959)	1	
Don't want to answer	0.2	0.0 (0/6)	-	-	0.0 (0/6)	-	-
Not filled in	28.9	1.6 (11/706)	0.7 (0.3-1.4)	0.33	5.5 (39/706)	0.9 (0.6-1.4)	0.69
Sex with > 1 partner within 12 months							
Yes	30.8	3.9 (29/751)	2.4 (1.3-4.4)	0.01	10.0 (75/751)	2.0 (1.4-2.8)	<0.01
No	39.5	1.7 (16/963)	1		5.4 (52/963)	1	
Don't want to answer	0.3	0.0 (0/8)	-	-	0.0 (0/8)	-	-
Not filled in	29.5	1.7 (12/719)	1.0 (0.5-2.1)	0.99	5.4(39/719)	1.0 (0.6-1.5)	0.99
Specimen							
Cervical/	1.3	3.2 (1/31)	3.3 (0.4-25.8)	0.26	9.7 (3/31)	2.0 (0.6-7.4)	0.21
Self-taken vaginal	40.4	4.2 (41/986)	4.2 (2.3-7.6)	<0.001	9.3 (92/986)	2.0 (1.5-2.8)	<0.001
First catch urine	58.3	1.0 (15/1424)	1		5.0 (71/1424)	1	

aOR^a odds ratios adjusted for age only

Association of *M. genitalium* and *C. trachomatis* with sexual behaviour and demographic characteristics of STI clinic attendees

Table 3 shows the association of *M. genitalium* and *C. trachomatis* with sexual behaviour and demographic characteristics among STI clinic attendees. The age distribution for both *M. genitalium* and *C. trachomatis* was similar with infections more frequently detected in younger women (15 to 19 years 9.7%, 18/186 vs. 6.4%, 12/186, respectively and 20 to 24 years 6.2%, 41/665 vs. 6.0%, 40/665) than other age groups. *M. genitalium* was more frequently detected in 15 to 19 year old women than *C. trachomatis* although this was not statistically significant ($p=0.28$).

Table 3. Association of characteristics with *M. genitalium* and *C. trachomatis* in women attending two London STI clinics

Characteristic	(N=2172) % of women with characteristic	<i>M. genitalium</i> (%) (proportion of women)	aOR ^a (95% CI)	P-value	<i>C. trachomatis</i> proportion of women)	aOR ^a (95% CI)	P-value
Age:							
15-19	8.6	9.7 (18/186)			6.4 (12/186)		
20-24	30.6	6.2 (41/665)			6.0 (40/665)		
25-29	28.6	1.6 (10/621)			2.9 (18/621)		
30-34	15.6	2.3 (9/339)			3.2 (11/339)		
35- 64	16.6	0.8 (3/361)			0.6 (2/361)		
Ethnicity							
White	23.0	6.0 (30/499)	1		7.0 (35/499)	1	
Black or Black British	6.9	7.4 (11/149)	1.2 (0.6-2.5)	0.60	4.0 (6/149)	0.5 (0.2-1.3)	0.54
Asian or Asian British	1.7	17.6 (6/36)	3.1 (1.2-8.1)	0.19	5.6 (2/36)	0.8 (0.2-3.4)	0.73
Mixed	3.9	4.8 (4/84)	0.7 (0.2-2.1)	0.54	7.1 (6/84)	0.9 (0.4-2.3)	0.91
Other Ethnic groups	3.9	9.5 (8/83)	1.6 (0.7-3.7)	0.24	3.6 (3/83)	0.5 (0.1-1.6)	0.49
Unknown	60.8	1.7 (22/1321)	0.5 (0.2-1.1)	0.09	2.3 (31/1321)	0.7 (0.3-1.4)	0.66
Specimen							
Cervical/	90.3	3.8 (75/1961)	1.4 (0.6-3.2)	0.48	3.4 (38/1130)	0.7 (0.4-1.6)	0.44
Self-taken vaginal					4.3 (36/831)		
First catch urine	9.7	2.8 (6/211)	1		4.3 (9/211)	1	

aOR^a odds ratios adjusted for age only

Specimen type and bacterial load

Overall *M. genitalium* was detected in 3.7% (43/1161), 4.0% (74/1817) and 1.3% (21/1635) of cervical swabs, self-taken vulval swabs and first-void urine samples, respectively. Since *M. genitalium* frequency in cervical and self-taken swabs was similar ($p=0.86$), the results for the two groups of swabs were merged and tested against first-void urine samples in the statistical model. *M. genitalium* was significantly more likely to be detected in swabs compared with urine specimens (3.9% vs. 1.3%, $P<0.001$).

The overall frequency of *C. trachomatis* in cervical swabs, self-taken vulval swabs and first-void urine samples was 3.5% (41/1161), 7.0% (128/1817) and 4.9% (80/1635), respectively. *C. trachomatis* significantly differed between cervical and self-taken swabs ($p<0.001$) and the two groups were separately tested against the urine samples in the statistical model.

The majority (58%, 1424/2441) of specimens provided by the women in NCSP were urine samples. However swab samples were almost four times more likely to test positive for *M. genitalium* compared with urine samples (aOR 3.6, 95% CI 1.9-6.7, $p<0.001$) and *C. trachomatis* infection was almost twice as high among swabs compared with urine samples (aOR 1.8, 95% CI 1.2-2.4 $p=0.001$). Conversely the majority (90.3%, 1961/2172) of clinic specimens were swabs. *M. genitalium* and *C. trachomatis* in the clinic swab and urine specimens also differed (*M. genitalium* 3.8%, 75/1961 vs. 2.8%, 6/211 and *C. trachomatis* 3.8%, 74/1961 vs. 4.3%, 9/211, respectively).

In quantitative analysis of *M. genitalium* positive specimens, mean *M. genitalium* bacterial load in swab and urine samples did not significantly differ between the clinics or NCSP. Clinic data were therefore combined for comparison of the mean bacterial load in different specimen types. There was no difference in overall cervical and self-taken vaginal

1
2
3 swab bacterial loads (3.72 (CI 3.39-4.05) vs. 3.91 (CI 3.66-4.17) \log_{10} genome copies/mL,
4
5 equivalent to geometric means of 5,218 (CI 2,438-11,171) and 8,192 (CI 4,575-14,669)
6
7 organisms/mL, respectively) ($p=0.349$). The overall mean bacterial load in swabs 3.84 (CI
8
9 3.52-4.11) equivalent to 6,705 (CI 3506-12,920) organisms/mL was significantly higher than
10
11 in first-void urine samples (3.14 (CI 2.87-3.41) equivalent to 1386 (CI 740-2,597)
12
13 organisms/mL) ($p<0.0001$, equal variances not assumed).
14
15

16 17 **Genetic diversity**

18
19
20 The absence of false positive results was confirmed by the presence of 57 different
21
22 genotypes by sequence analysis of 127 *M. genitalium* positive specimens and 13 sequences
23
24 from previously isolated strains (figure 2). The discriminatory index by Hunter and Gaston *et*
25
26 *al.* 1988²⁹ was calculated to be 0.94 both with and without inclusion of the previously
27
28 isolated strain sequences. None of the sequenced samples were identical with the type strain
29
30 G37 used as a PCR standard control. Genetic diversity data are available in FASTA format
31
32 for download in the supplementary material.
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

DISCUSSION

Overall *M.genitalium* was relatively common at 3.0% among NCSP participants and STI clinic attendees. *M.genitalium* was more likely to be found in swabs compared with urine samples (3.9% vs. 1.3%, respectively) and the mean bacterial load was also much higher (6,705 (CI 3,506-12,920) organisms/mL vs. 1386 (CI 740-2,597) organisms/mL, respectively).

Only 0.5% of all the women had both *C. trachomatis* and *M.genitalium* infections. Among women who had *C. trachomatis*, 9% were co-infected with *M.genitalium* compared with <5% in population based studies.^{16;18;30;31} Among NCSP participants the age-adjusted odds of detecting *M.genitalium* were twice as high among women of black/black British ethnicity (aOR 2.3) and those reporting multiple sexual partners in the past year (aOR 2.4) compared with women of white ethnicity or those who reported only one partner, respectively. After adjusting for age, *C. trachomatis* was also significantly more likely to be diagnosed in women with multiple partners (aOR 2.0) and new sexual partners in the previous three months (aOR 1.6) but was less likely to be detected in women who did not give a self-identified ethnic group (aOR 0.6) compared with reporting only one partner, not reporting new partners or being of white ethnicity, respectively. No significant associations were observed for either infection among STI clinic attendees.

This is the largest UK based *M. genitalium* study to date to provide estimates of infection among both community and STI clinic based populations. Transport media may affect the sensitivity of DNA based PCR tests. The study samples were originally collected in Aptima medium. We therefore tested Aptima and PBS media with *M.genitalium* DNA and did not find any differences. We confirmed positive *M.genitalium* PCR results by genotype sequencing. Our analysis of *M.genitalium* and *C.trachomatis* correlates is limited by

1
2
3 availability of data: only age and ethnicity were available for both clinic and NCSP datasets
4
5 and ethnicity data was missing for 61% of STI clinic attendees. There is also a possibility that
6
7 some young women may have had chlamydia tests through both the NCSP and the STI
8
9 clinics during the sample collection period. It is not possible to quantify this although we
10
11 speculate that the numbers are likely to be low given the relatively short time frame.
12
13

14
15 Our STI clinic *M.genitalium* frequency is similar to that found in several studies of
16
17 female STI clinic attendees (4.5% to 7%)^{32;33} although other studies have reported a much
18
19 higher frequencies (19.3% to 38.2%).^{34;35} In lower risk non-STI clinic attendees such as
20
21 college students infection has been shown to range from <1% to 5%^{5;36} which is in keeping
22
23 with our estimate in the chlamydia screening population. The higher frequency of
24
25 *M.genitalium* in women attending clinics than the NCSP (3-5.3% vs. 2.3%, respectively) may
26
27 in part reflect the higher proportion of swabs taken in clinics than in NCSP settings. Urine
28
29 samples have been shown to be less sensitive for *M.genitalium* diagnosis than swabs (61% to
30
31 65% compared with 74% to 91%).^{37;38} It is therefore likely that our NCSP *M.genitalium*
32
33 frequency is an underestimation. Although urine sample sensitivity may be increased by up-
34
35 concentrating the samples by centrifugation this is not a practical step for large scale testing.
36
37 A higher bacterial load may be associated with symptoms as has been shown for men.²⁵ This
38
39 may also explain the difference in infection between the two populations since STI clinic
40
41 attendees are more likely to be symptomatic than NCSP participants. The association of
42
43 *M.genitalium* with multiple sexual partners and black ethnicity has been previously
44
45 observed.^{16;31} Additional risk factors include younger age as observed in our STI clinic
46
47 attendees, bacterial vaginosis, being symptomatic, cervicitis, douching, smoking, prior
48
49 miscarriage, menstrual cycle, social class and marital status.^{12;16;31;35;39-41}
50
51
52
53
54

55
56 *M.genitalium* appears to be a relatively common infection among women in London.
57
58 The low level of *M.genitalium* and *C. trachomatis* co-infection (0.5%) suggests that
59
60

1
2
3 diagnosing and treating chlamydia will have little impact on *M.genitalium*. However
4
5 Azithromycin 1g used to treat uncomplicated *C. trachomatis* infection appears to be sub-
6
7 optimal for *M.genitalium* treatment²⁴ This treatment dose has also been associated with the
8
9 development of *M.genitalium* macrolide resistance in some studies of predominantly
10
11 symptomatic men.^{22 24} The risk of inadvertent *M.genitalium* antibiotic resistance in co-
12
13 infected women who are treated for chlamydia with 1g of azithromycin is therefore
14
15 potentially a cause for concern although further research is required to confirm this.
16
17
18

19 To avoid the problems encountered with *C. trachomatis* screening and *M.genitalium*
20
21 antimicrobial resistance, prior to introducing routine testing for *M.genitalium*, further research
22
23 is needed to better understand its natural history, the role of asymptomatic and symptomatic
24
25 *M.genitalium* in PID and determine optimum treatment guidelines.
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Acknowledgements

The authors would like to thank Menelaos Pavlou (Centre for Sexual Health and HIV Research, Mortimer Market Centre, London, UK) and Tina Sharp (Mortimer Market Centre, London, UK, previously NCSP Chlamydia Co-ordinator, Camden Primary Care Trust) for extracting clinical data from the Sexually Transmitted Disease Clinics and the National Chlamydia Screening Programme, respectively. We thank Dr. Stephane Hue, Center for Virology, UCL for aligning the sequenced fragments and create the resulting phylogenetic tree.

Authors Contribution

All authors contributed to conception and design of the study and / or to acquisition of data. HS performed the experiments. HS SSD and JS drafted the paper and all authors contributed to critical revision of the paper.

Competing interests

None

Funding

This work was supported by UCLH/UCL Comprehensive Biomedical Research Centre grant no. 59.

Data sharing statement

There is no additional data available

Reference List

- (1) Barrett S, Taylor C. A review on pelvic inflammatory disease. *Int J STD AIDS* 2005; 16(11):715-720.
- (2) Simms I, Stephenson JM. Pelvic inflammatory disease epidemiology: what do we know and what do we need to know? *Sex Transm Infect* 2000; 76(2):80-87.
- (3) Haggerty CL, Ness RB. Diagnosis and treatment of pelvic inflammatory disease. *Womens Health (Lond Engl)* 2008; 4(4):383-397.
- (4) Haggerty CL, Taylor BD. Mycoplasma genitalium: an emerging cause of pelvic inflammatory disease. *Infect Dis Obstet Gynecol* 2011; 2011:959816.
- (5) McGowin CL, Anderson-Smits C. Mycoplasma genitalium: an emerging cause of sexually transmitted disease in women. *PLoS Pathog* 2011; 7(5):e1001324.
- (6) Taylor-Robinson D, Jensen JS. Mycoplasma genitalium: from Chrysalis to multicolored butterfly. *Clin Microbiol Rev* 2011; 24(3):498-514.
- (7) Bjartling C. The association between Mycoplasma genitalium and pelvic inflammatory disease after termination of pregnancy. 2010.
- (8) Bjartling C. Mycoplasma genitalium in cervicitis and pelvic inflammatory disease among women at a gynecologic outpatient service. 2012.
- (9) Cohen CR, Manhart LE, Bukusi EA, et al. Association between Mycoplasma genitalium and acute endometritis. *Lancet* 2002; 359(9308):765-766.
- (10) Svenstrup HF, Fedder J, Kristoffersen SE, et al. Mycoplasma genitalium, Chlamydia trachomatis, and tubal factor infertility - a prospective study. *Fertility and Sterility* 2008; 90(3):513-520.
- (11) Manhart LE, Broad JM, Golden MR. Mycoplasma genitalium: should we treat and how? *Clin Infect Dis* 2011; 53 (s3): s129-42.
- (12) Mobley VL, Hobbs MM, Lau K, Weinbaum BS, et al. Mycoplasma genitalium infection in women attending a sexually transmitted infection clinic: diagnostic specimen type, coinfections, and predictors. *Sex Transm Dis* 2012; 39(9):706-709.
- (13) Jensen JS. *Mycoplasma genitalium* - the etiological agent of urethritis and other Sexually Transmitted Diseases. *Sex Transm Dis* 2003.
- (14) Mavedzenge SN, Van Der PB, Weiss HA, et al. The association between Mycoplasma genitalium and HIV-1 acquisition in African women. *AIDS* 2012; 26(5):617-624.

- 1
2
3 (15) Low N, Bender N, Nartey L, et al. Effectiveness of chlamydia screening: systematic review.
4 *International Journal of Epidemiology* 2009; 38(2):435-448.
5
6 (16) Oakeshott P, Aghaizu A, Hay P, et al. Is *Mycoplasma genitalium* in women the "New
7 Chlamydia?" A community-based prospective cohort study. *Clin Infect Dis* 2010;
8 51(10):1160-1166.
9
10 (17) Anagrus C, Lore B, Jensen JS. *Mycoplasma genitalium*: prevalence, clinical significance, and
11 transmission. *Sex Transm Infect* 2005; 81(6):458-462.
12
13 (18) Andersen B, Sokolowski I, Ostergaard L, et al. *Mycoplasma genitalium*: prevalence and
14 behavioural risk factors in the general population. *Sexually Transmitted Infections* 2007;
15 83(3):237-241.
16
17 (19) Jensen JS, Bradshaw CS, Tabrizi SN, et al. Azithromycin Treatment Failure in *Mycoplasma*
18 *genitalium* Positive Patients with Nongonococcal Urethritis Is Associated with Induced
19 Macrolide Resistance. *Clinical Infectious Diseases* 2008; 47(12):1546-1553.
20
21 (20) Lau CY, Qureshi AK. Azithromycin Versus Doxycycline for Genital Chlamydial Infections: A
22 Meta-Analysis of Randomized Clinical Trials. *Sexually Transmitted Diseases* 2002; 29(9).
23
24 (21) Horner PJ. Azithromycin antimicrobial resistance and genital Chlamydia trachomatis
25 infection: duration of therapy may be the key to improving efficacy. 2012.
26
27 (22) Anagrus C, Lore B, Jensen JS. Treatment of *Mycoplasma genitalium*. Observations from a
28 Swedish STD clinic. *PLoS One* 2013; 8(4):e61481.
29
30 (23) Tagg KA, Jeoffreys NJ, Couldwell DL, et al. Fluoroquinolone and macrolide resistance-
31 associated mutations in *Mycoplasma genitalium*. *J Clin Microbiol* 2013; 51(7):2245-2249.
32
33 (24) Twin J. Transmission and selection of macrolide resistant *Mycoplasma genitalium* infections
34 detected by rapid high resolution melt analysis. 2012.
35
36 (25) Jensen JS, Bjornelius E, Dohn B, Lidbrink P. Use of TaqMan 5' nuclease real-time PCR for
37 quantitative detection of *Mycoplasma genitalium* DNA in males with and without urethritis
38 who were attendees at a sexually transmitted disease clinic. *J Clin Microbiol* 2004; 42(2):683-
39 692.
40
41 (26) Chalker VJ, Jordan K, Ali T, Ison C. Real-time PCR detection of the mg219 gene of unknown
42 function of *Mycoplasma genitalium* in men with and without non-gonococcal urethritis and
43 their female partners in England. *J Med Microbiol* 2009; 58(Pt 7):895-899.
44
45 (27) Garson JA, Grant PR, Ayliffe U, et al Real-time PCR quantitation of hepatitis B virus DNA using
46 automated sample preparation and murine cytomegalovirus internal control. *J Virol*
47 *Methods* 2005; 126(1-2):207-213.
48
49 (28) Hjorth SV, Bjornelius E, Lidbrink P, et al. Sequence-based typing of *Mycoplasma genitalium*
50 reveals sexual transmission. *J Clin Microbiol* 2006; 44(6):2078-2083.
51
52 (29) Hunter PR, Gaston MA. Numerical index of the discriminatory ability of typing systems: an
53 application of Simpson's index of diversity. *J Clin Microbiol* 1988; 26(11):2465-2466.
54
55
56
57
58
59
60

- 1
2
3 (30) Manhart LE, Holmes KK, Hughes JP, et al. Mycoplasma genitalium among young adults in the
4 United States: an emerging sexually transmitted infection. *Am J Public Health* 2007; 97(6):
5 1118-1125.
6
7 (31) Walker J, Fairley CK, Bradshaw CS, et al. 'The difference in determinants of Chlamydia
8 trachomatis and Mycoplasma genitalium in a sample of young Australian women'. *BMC*
9 *Infect Dis* 2011; 11:35.
10
11 (32) Manhart LE, Critchlow CW, Holmes KK, et al. Mucopurulent cervicitis and Mycoplasma
12 genitalium. *J Infect Dis* 2003; 187(4):650-657.
13
14 (33) Moi H, Reinton N, Moghaddam A. Mycoplasma genitalium in women with lower genital tract
15 inflammation. *Sex Transm Infect* 2009; 85(1):10-14.
16
17 (34) Casin I, Vexiau-Robert D, De La SP, et al. High prevalence of Mycoplasma genitalium in the
18 lower genitourinary tract of women attending a sexually transmitted disease clinic in Paris,
19 France. *Sex Transm Dis* 2002; 29(6):353-359.
20
21 (35) Mobley VL, Hobbs MM, Lau K, et al. Mycoplasma genitalium infection in women attending a
22 sexually transmitted infection clinic: diagnostic specimen type, coinfections, and predictors.
23 *Sex Transm Dis* 2012; 39(9):706-709.
24
25 (36) Jensen AJ, Kleveland CR, Moghaddam A, et al. Chlamydia trachomatis, Mycoplasma
26 genitalium and Ureaplasma urealyticum among students in northern Norway. *J Eur Acad*
27 *Dermatol Venereol* 2013; 27(1):e91-e96.
28
29 (37) Lillis RA, Nsuami MJ, Myers L, et al. Utility of urine, vaginal, cervical, and rectal specimens for
30 detection of Mycoplasma genitalium in women. *J Clin Microbiol* 2011; 49(5):1990-1992.
31
32 (38) Wroblewski JK, Manhart LE, Dickey KA, et al. Comparison of transcription-mediated
33 amplification and PCR assay results for various genital specimen types for detection of
34 Mycoplasma genitalium. *J Clin Microbiol* 2006; 44(9):3306-3312.
35
36 (39) Short VL, Totten PA, Ness RB, et al. The demographic, sexual health and behavioural
37 correlates of Mycoplasma genitalium infection among women with clinically suspected
38 pelvic inflammatory disease. *Sex Transm Infect* 2010; 86(1):29-31.
39
40 (40) Oakeshott P, Hay P, Taylor-Robinson D, et al. Prevalence of Mycoplasma genitalium in early
41 pregnancy and relationship between its presence and pregnancy outcome. *BJOG* 2004;
42 111(12):1464-1467.
43
44 (41) Vandepitte J, Muller E, Bukonya J, et al. Prevalence and correlates of Mycoplasma genitalium
45 infection among female sex workers in Kampala, Uganda. *J Infect Dis* 2012; 205(2):289-296.
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 **A cross-sectional study of *Mycoplasma genitalium* ~~prevalence-infection~~ and correlates in women**
4 **~~attending a national chlamydia screening programme or sexually transmitted infection~~**
5 **~~clinics undergoing population based screening or clinic based testing for Chlamydia infection in~~**
6 **London**
7
8
9

10 Svenstrup HF¹, Dave SS¹, Carder C², Grant P², Morris-Jones S², Kidd M² and Stephenson JM¹

11 ¹Research Department of Reproductive Medicine, Institute for Women's Health, University College
12 London, 65 Whitfield Street, W1T 4EU, UK

13 ²Clinical Microbiology and Virology, University College London Hospitals NHS Foundation Trust,
14 65 Whitfield Street, W1T 4EU, UK

15
16
17
18
19
20 **Corresponding author:**

21 Dr. Helle F Svenstrup

22 Rigensgade 11, DK-1316 København K, Denmark

23 Fax +45 43 99 99 11

24 Direct phone +45 43 31 33 77,

25 Mobile +45 20 65 12 64

26 hfs@awapatent.com
27
28
29
30
31

32 **Co-authors:**

33 Sangeeta Shashikant Dave, Institute for Women's Health, University College London, London,
34 United Kingdom, sangeeta.dave@ucl.ac.uk

35
36
37 Caroline Carder, Clinical Microbiology and Virology, University College London Hospitals NHS
38 Foundation Trust, London, United Kingdom, caroline.carder@uclh.nhs.uk

39
40 Paul Grant, Clinical Microbiology and Virology, University College London Hospitals NHS
41 Foundation Trust London, United Kingdom paul.grant@ucl.ac.uk

42
43 Stephen Morris-Jones, Clinical Microbiology and Virology, University College London Hospitals
44 NHS Foundation Trust London, United Kingdom, stephen.morris-jones@uclh.nhs.uk

45
46 Ian Kidd, Clinical Microbiology and Virology, University College London Hospitals NHS Foundation
47 Trust, London, United Kingdom, i.kidd@ucl.ac.uk

48
49 Judith Stephenson, Institute for Women's Health, University College London, London, United
50 Kingdom, judith.stephenson@ucl.ac.uk
51
52
53
54
55
56
57
58
59
60

Key words: Genotyping / *Mycoplasma genitalium* / National Chlamydia Screening Programme / Real-time PCR

Word count: [3049](#)

ABSTRACT

Objective:

To determine *Mycoplasma genitalium* ~~prevalence~~infection and correlates among young women undergoing population based screening or clinic based testing for chlamydia infection.

Design:

Cross-sectional study

Setting:

National Chlamydia Screening Programme (NCSP) and two London STI clinics

Participants:

2441 women aged 15 to 64 years who participated in the NCSP and 2172 women who attended two London STI clinics over a four month period in 2009.

Outcome measures:

1. *M.genitalium* ~~prevalence in defined populations~~prevalence (%)
2. Age-adjusted odds ratios (aORs) for correlates of *M.genitalium* infection

Results

The overall ~~prevalence~~frequency of *M. genitalium* and *C.trachomatis* was 3.0% and 5.4%, respectively. Co-infection was relatively uncommon (0.5% of all women); however 9% of women with *C.trachomatis* also had *M.genitalium* infection. *M.genitalium* was more frequently detected in swab than urine samples (3.9% vs. 1.3%, $p<0.001$) with a significantly higher mean bacterial load ($p<0.001$). Among NCSP participants, *M.genitalium* was significantly more likely to be diagnosed in women of black/black British ethnicity (aOR 2.3, 95% CI 1.2-4.5, $p=0.01$). *M.genitalium* and *C. trachomatis* and were both significantly associated with multiple sexual partners in the past year (aOR 2.4, 95% CI 1.3-4.4, $p=0.01$)

1
2
3 and aOR 2.0, 95% CI 1.4-2.8, $p < 0.01$). Among STI clinic attendees, *M.genitalium* was more
4 ~~prevalent~~ common in women who were less than 25 years in age.
5
6
7

8 9 10 **Conclusions**

11 *M.genitalium* is a relatively common infection among young women in London. It is
12 significantly more likely to be detected in vulvo-vaginal swabs than in urine samples. Co-
13 infection with chlamydia is uncommon. The clinical effectiveness of testing and treatment
14 strategies for *M.genitalium* needs further investigation.
15
16
17

18 19 **Article summary**

20 21 22 Article Focus

- 23
24 • How common is *Mycoplasma genitalium* in women in the general population
25 and those attending sexually transmitted infection (STI) clinics in London?
26
27
- 28
29 • How are *M.genitalium* ~~prevalence~~ infection and bacterial load associated with
30 sample type?
31
32
- 33
34 • How much co-infection is there between *M.genitalium* and *Chlamydia*
35 *trachomatis* in the study sample?
36
37
38
39

40 41 Key messages

- 42
43 • At 3% ~~prevalence~~ *Mycoplasma genitalium* is a relatively common infection
44 amongst women participating in the National Chlamydia Screening
45 Programme and attending STI clinics in London.
46
47
- 48
49 • *M.genitalium* is more ~~prevalent~~ common and has a higher mean bacterial load
50 in cervical and self-taken vaginal swabs than in first catch urine samples.
51
52
- 53
54 • Only 0.5% of women had both chlamydia and *M.genitalium* infection and 9%
55 of women with chlamydia had *M.genitalium* infection. Chlamydia treatment is
56
57
58
59
60

1
2
3 therefore likely to have little impact on treating *M.genitalium* infection overall.

4
5 In women with both infections antimicrobial treatment for chlamydia is likely

6
7 to be sub-optimal treatment for *M.genitalium* with the risk of increasing

8
9 macrolide resistance.
10

11 12 13 14 Strengths and limitations

15 16 Strengths

- 17
18
19
20
21
22
23
24
25
26
27
28
29
- This is the largest UK based *M. genitalium* prevalence cross-sectional study to date to provide estimates of *M. genitalium* prevalence in ~~for~~ both community and STI clinic based populations.
 - *M.genitalium* PCR results were confirmed positive by genotype sequencing.

30 31 32 Limitations

- Our analysis of potential correlates for *M.genitalium* and *C.trachomatis* is limited by availability of data.

33 34 35 36 37 38 39 **Competing interests**

40
41 None

42 43 44 **Funding**

45
46 This work was supported by UCLH/UCL Comprehensive Biomedical Research Centre grant
47 no. 59.

48 49 50 **Data sharing statement**

51
52 There is no additional data available
53
54
55
56
57
58
59
60

INTRODUCTION

Pelvic inflammatory disease (PID) and its sequelae (chronic pelvic pain, ectopic pregnancy and tubal infertility) are major causes of morbidity in women in developed and developing countries.¹ In the USA more than \$10 billion is spent annually in treating these conditions.² *Chlamydia trachomatis* and *Neisseria gonorrhoeae*, two sexually transmitted infections (STIs) are known causes of PID. However in up to 70% of PID cases no cause is found³ and there is increasing evidence that *Mycoplasma genitalium* might be a cause of PID.⁴⁻⁸

There is also strong evidence that it is sexually transmitted.^{5;6} It is significantly associated with endometritis and⁹ tubal factor infertility¹⁰ although the association with cervicitis is complex.^{11;12} As with *C.trachomatis* it can be asymptomatic, acting as a reservoir for further spread.¹³ It may also be associated with human immunodeficiency virus acquisition.¹⁴

Although at present *M.genitalium* is not routinely tested for in most countries, there is interest in introducing testing and treatment. However, before this is done there is a need to gain a better understanding of the infection to avoid repeating the problems encountered with *C. trachomatis* screening.¹⁵ In the United Kingdom (UK) there are few data on the **prevalence frequency** of *M.genitalium* infection in different population groups of women. Oakeshott *et al.* found that *M.genitalium* prevalence was 3.3% among young women in a community based sample who took part in a *C. trachomatis* screening trial in the UK.¹⁶ Estimates from studies in other countries indicate that **the prevalence levels** of *M. genitalium* **isare** 40% to 60% lower than **the prevalence of** *C. trachomatis*, with little co-infection.^{17;18} The recommended treatment for uncomplicated chlamydia infection is a single dose of azithromycin 1g stat. There is growing evidence of considerably lower *M.genitalium* cure rates with this dose of

1
2
3 azithromycin compared with *C. trachomatis* (79-87% vs. 92-97%, respectively).¹⁹⁻²¹ ~~This may~~
4 ~~be due to genotypic variations in *M.genitalium* resistance to antibiotic treatment and an~~
5 ~~extended course of azithromycin or moxifloxacin has been shown to have superior cure~~
6 ~~rates.^{6,11}—Resistance has been shown to develop following 1g of azithromycin and macrolide~~
7 ~~resistance is endemic in some populations.~~²²⁻²⁴
8
9
10
11
12

13
14
15 We investigated ~~the prevalence of *M.genitalium* infection~~ by real-time PCR and
16 determined its correlates in the largest ~~*M. genitalium* prevalence cross-sectional~~ study of *M.*
17 *genitalium* among women screened for *C. trachomatis* in the National Chlamydia Screening
18 Programme (NCSP) and STI clinics in the United Kingdom (UK).
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

METHODS

Patients and specimens

We used an unlinked anonymised method to test routinely collected and stored cervical swabs, self-taken vaginal swabs and first catch urine samples for *M.genitalium*. The samples were from 2180 women aged 15 to 64 years who had *C. trachomatis* screening when they attended two STI clinics in central and North London and 2455 women aged 15 to 24 years who participated in the NCSP in London in a four month period in 2009. Each clinic offers comprehensive STI screening, treatment and partner notification services to symptomatic and asymptomatic women and men, irrespective of age. Samples from all female clinic attendees were eligible for the study. The NCSP is a national screening programme for chlamydia in the UK among women and men who are under 25 years old in age. The NCSP samples were from a variety of low and high STI risk settings within two London boroughs. In 2009 the majority of participating sites from which the samples were tested were family planning clinics (47%), universities (17%) and general practices (16%). Other testing sites included pharmacies, abortion services, outreach, young persons' services, schools and postal testing (Tina Sharp, NCSP Chlamydia Co-ordinator, personal communication).

The samples were originally collected from the NCSP and clinics and transported to the microbiology laboratory at University College London Hospital in 3 mL (self-taken vaginal and cervical swabs) or 4 mL (urine samples diluted 1:1) of APTIMA transport medium (Gen-Probe Inc., San Diego, USA) for routine *C. trachomatis* testing. After *C. trachomatis* testing the negative samples were stored for 6 weeks at -20°C and positive samples were stored for 3 months at -20°C before they were released for testing as part of this study. Available demographic, sexual behaviour, clinical PID diagnosis and sexually

1
2
3 transmitted infections data were recorded before samples were unlinked from all personal
4
5 identifiers prior to *M.genitalium* testing.
6
7

8 ***M. genitalium* testing**

9
10 Samples were thawed and DNA from 200 µL of the APTIMA transport medium was purified
11
12 by BioRobot 9604 automated workstation using the QIAamp® Virus BioRobot® 9604 Kit
13
14 (QIAGEN, Hilden, Germany). Before freezing and storing the eluate at -20°C it was tested by
15
16 quantitative PCR (qPCR) adapted from a method by Jensen *et al.*^{17;25} The qPCR targeted the
17
18 MgPa adhesion gene (MG191) using MgPa-355FW and MgPa-432R primers and MgPa-380
19
20 MGB probe (primers and probes were provided by Applied Biosystems, Warrington, UK).
21
22 Pilot laboratory work showed no difference in Aptima transport medium and PBS spiked
23
24 with *M.genitalium* DNA in different concentrations.
25
26
27
28

29 We introduced a degenerate oligonucleotide ('wobble') in the forward primer to
30
31 account for a frequent detected base substitution that has previously been shown to be
32
33 successful in another study by Chalker *et al.*²⁶ As an internal control for PCR inhibition we
34
35 used murine CMV (mCMV) and primers mCMVTAQ1 (forward primer) and mCMVTAQ2
36
37 (reverse primer) and mCMVTAQPR probe labelled with JOE (Primers and probe were
38
39 provided by Eurofins MWG Operon) designed by Garson *et al.*²⁷. The qPCR assays were
40
41 performed in 25 µL volumes; comprising 1x EXPRESS qPCR Supermix (Universal,
42
43 Invitrogen™, Life technologies Ltd. Paisley, UK), 0.4 µM forward and reverse primers, 0.2
44
45 µM probes and 7.5 µL of samples, and nuclease-free water (Promega UK Ltd., Southampton,
46
47 UK).
48
49
50
51

52 Thermal cycling was performed on an ABI 7500 Real-time PCR instrument using the
53
54 following conditions: hotstart at 95°C for 2 min and 1 cycle, denaturation at 95°C for 15 sec,
55
56 annealing and extension at 60°C for 1 min and 45 cycles. The data was analysed using
57
58
59
60

1
2
3 Sequence Detection Software (SDS) version 1.4 with manual baseline/threshold settings to
4
5 estimate quantification cycle.
6
7

8 Positive samples were re-extracted and retested by qPCR. If these tested negative the
9
10 samples was re-extracted and tested by qPCR a third time. If negative again the sample was
11
12 considered equivocal and was excluded from the analysis.
13

14 ***M. genitalium* genotyping**

15
16 *M. genitalium* PCR positive samples were sequenced by MgPa1-3 typing assay
17
18 according to Hjort *et al.* 2006.²⁸ The assay was modified with respect to PCR reagents and
19
20 PCR conditions. In a total volume of 50 µL the following were mixed: 25 µL of Taq PCR
21
22 Master Mix kit (QIAGEN, Hilden, Germany), 0.4 µM of mgpa-1 and mgpa-3 primers, 5 µL
23
24 of template, and nuclease-free water. To increase the sensitivity of the assay 10 µL of the
25
26 template was used in cases where the bacterial load was less than 1 genome copy per µL.
27
28
29
30
31

32 The PCR was performed on an ABI9700 instrument and in 3- step cycling conditions:
33
34 denaturation at 94 °C for 1 min, annealing at 60 °C for 1 min and extension at 72 °C for 1 min
35
36 and 50 cycles.
37
38

39 The amplified product were purified manually by QIAquick PCR purification kit (QIAGEN,
40
41 Hilden, Germany) and sent to the UCL sequencing service for sequencing of both the forward
42
43 and reverse strand.
44
45
46
47

48 **Statistical analysis**

49
50 We have only included data from women who are at least 15 years old in the analysis.
51
52 Data were analysed using SPSS® 14.0 for Windows. Paired sample T-test was used to
53
54 compare the difference of mean values. Multiple logistic regression analysis was used to
55
56
57
58
59
60

investigate the relationship between *M. genitalium* or *C. trachomatis* infection and demographic and sexual behaviour characteristics in women attending NCSP or STI clinics.

Categorical variables in the NCSP model included participant age, specimen type, a new sexual partner within three months, more than one partner within 12 months and ethnicity. The categorical variables included in the STI model were participant age, specimen type, current STI infections and ethnicity. ~~Prevalence~~Frequency, odds ratios adjusted for age (aOR) and 95% confidence intervals (CIs) were calculated and values of $p < 0.05$ were considered statistically significant.

Ethics approval

On the advice of the chair of the local ethics committee, ethical approval was not required since the study team received anonymised samples for testing in the study from the laboratory and no other identifiable data were available.

RESULTS

Of 4635 samples, we excluded 21 samples for which the *M. genitalium* test result was equivocal and included 4613 samples in our analysis (figure 1).

NCSP participants were aged 15 to ~~25~~24 years whereas STI clinic attendees were aged 15 to 64 years. Women attending the two clinics had significantly different mean ages (~~26.3~~20.1 years, SD 7.72.5 vs. ~~28.6~~27.8 years, SD 7.64 years, $p < 0.0001$). The highest prevalence of *M. genitalium* and *C. trachomatis* was in age groups 15 to 24 years in NCSP and the STI clinics. As we only had ethnicity data for 39% (851/2172) of the STI clinic attendees, we did not compare ethnicity across the clinics.

***M. genitalium* and *C. trachomatis* prevalence-infection**

As shown in table 1, the overall prevalence-frequency of *M. genitalium* and *C. trachomatis* was 3.0% (138/4613, 95% CI 2.5-3.5%) and 5.4% (249/4613, 95% CI 4.8-6.1%), respectively. The overall co-infection rate was 0.5% (23/4613, 95% CI 0.3-0.7%). Of 249 women with *C. trachomatis*, 23 (9%) women had *M. genitalium* infection.

Among NCSP participants, *M. genitalium* and *C. trachomatis* frequency were 2.3% (57/2441, 95% CI 1.7-2.9%) and 6.8% (166/2441), 95% CI 5.8-7.8%), respectively.

Table 1. *M. genitalium* and *C. trachomatis* prevalence-infection among NCSP and STI clinic attendees

Infection	Clinic 2 N=716 N (% , 95% CI)	Clinic 1 N=1456 N (% , 95% CI)	NCSP N=2441 N (% , 95% CI)	Total N=4613 N (% , 95% CI)
<i>M. genitalium</i> and <i>C. trachomatis</i>	3 (0.4, 0-0.9)	4 (0.3, 0-0.6)	16 (0.7, 0.4-1.0)	23 (0.5, 0.3-0.7)
Total <i>M. genitalium</i>	38 (5.3, 3.7-7.0)	43 (3.0 , 2.0-3.9)	57 (2.3, 1.7-2.9)	138 (3.0, 2.5-3.5)
<i>M. genitalium</i> only	35 (4.9, 3.3-6.5)	41 (2.8, 2.0-3.7)	39 (1.6, 1.1-2.1)	115 (2.5, 2.0-2.9)
Total <i>C. trachomatis</i>	23 (3.2, 1.9-4.5)	60 (4.1, 3.1-5.1)	166 (6.8, 5.8-7.8)	249 (5.4, 4.8-6.1)
<i>C. trachomatis</i> only	20 (2.8, 1.6-4.0)	56 (3.8, 2.9-4.8)	150 (6.1, 5.2-7.1)	226 (4.9, 4.3-5.5)

M. genitalium infection significantly differed between the two clinics (5.3%, 95% CI 3.7-7.0% and 3.0%, 95% CI 2.1-3.8%, $p < 0.01$) but the difference was not significant after adjusting for age ($p = 0.16$). *C. trachomatis* did not differ significantly between the two clinics (3.2%, 95% CI 1.9-4.5% and 4.1%, 95% CI 3.1-5.1%, $p = 0.30$).

1
2
3 **Association of *M. genitalium* and *C. trachomatis* with sexual behaviour and demographic**
4 **characteristics of participants in the National Chlamydia Screening Programme (NCSP)**
5
6
7

8 Table 2 shows the association of *M. genitalium* and *C. trachomatis* with sexual
9 behaviour and demographic characteristics among NCSP participants. *M. genitalium* was less
10 frequently detected than *C. trachomatis* in both age groups (15-19 years old 2.8%, 29/1045 vs.
11 8.3%, 83/1045 and 20-24 years old 2.0%, 28/1396 vs. 5.7%, 79/1396, respectively). When
12 adjusted for age *M. genitalium* was significantly more common in black/black British women
13 compared with white women (aOR 2.3, 95% CI 1.2-4.5, p=0.01). Women who reported
14 multiple sexual partners in the past twelve months were twice as likely to have both
15 *M. genitalium* and *C. trachomatis* infections compared with women who reported only one
16 partner (aOR 2.4, 95% CI 1.3-4.4, p=0.01) and (aOR 2.0, 95% CI 1.4-2.8, p<0.01),
17 respectively. Women who reported new sexual partners in the previous three months were
18 also more likely to have *C. trachomatis* infection (aOR 1.6, 95% CI 1.1-2.3, p=0.01). Those
19 who did not self-identify as white, black/black British, Asian/Asian British or mixed ethnicity
20 were less likely to be infected with *C. trachomatis* compared with white women (aOR 0.6,
21 95% CI 0.4-0.9, p=0.01).
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Table 2. Association of characteristics with *M. genitalium* and *C. trachomatis* in NCSP

attendees

Characteristic	(N=2441) % of women with characteristic	<i>M.genitalium</i> % (proportion of women)	aOR ^a (95% CI)	P-value	<i>C.trachomatis</i> % (proportion of women)	aOR ^a (95% CI)	P-value
Age:							
15-19	41.6	2.8 (29/1045)			8.3 (87/1045)		
20-24	56.5	2.0 (28/1396)			5.7 (79/1396)		
Ethnicity							
White	46.6	2.0 (23/1138)	1		7.4 (84/1138)	1	
Black or Black British	12.8	4.8 (15/314)	2.3 (1.2-4.5)	0.01	8.3 (26/314)	1.1 (0.7-1.7)	0.83
Asian or Asian British	4.4	1.9 (2/108)	0.9 (0.2-4.0)	0.93	6.2 (5/108)	0.6 (0.3-1.6)	0.33
Mixed	7.7	3.7 (7/187)	1.8 (0.8-4.3)	0.18	10.2 (19/187)	1.3 (0.8-2.3)	0.29
Other Ethnic Groups	28.4	1.4 (10/694)	0.7 (0.3-1.5)	0.35	4.6 (32/694)	0.6 (0.4-0.9)	0.01
New sexual partner in previous 3 months							
Yes	31.5	3.2(25/770)	1.5 (0.8-2.6)	0.20	9.2 (71/770)	1.6 (1.1-2.3)	0.01
No	39.3	2.2 (21/959)	1		5.8 (56/959)	1	
Don't want to answer	0.2	0.0 (0/6)	-	-	0.0 (0/6)	-	-
Not filled in	28.9	1.6 (11/706)	0.7 (0.3-1.4)	0.33	5.5 (39/706)	0.9 (0.6-1.4)	0.69
Sex with > 1 partner within 12 months							
Yes	30.8	3.9 (29/751)	2.4 (1.3-4.4)	0.01	10.0 (75/751)	2.0 (1.4-2.8)	<0.01
No	39.5	1.7 (16/963)	1		5.4 (52/963)	1	
Don't want to answer	0.3	0.0 (0/8)	-	-	0.0 (0/8)	-	-
Not filled in	29.5	1.7 (12/719)	1.0 (0.5-2.1)	0.99	5.4(39/719)	1.0 (0.6-1.5)	0.99
Specimen							
Cervical/	1.3	3.2 (1/31)	3.3 (0.4-25.8)	0.26	9.7 (3/31)	2.0 (0.6-7.4)	0.21
Self-taken vaginal	40.4	4.2 (41/986)	4.2 (2.3-7.6)	<0.001	9.3 (92/986)	2.0 (1.5-2.8)	<0.001
First catch urine	58.3	1.0 (15/1424)	1		5.0 (71/1424)	1	

aOR^a odds ratios adjusted for age only

Association of *M. genitalium* and *C. trachomatis* with sexual behaviour and demographic characteristics of STI clinic attendees

Table 3 shows the association of *M. genitalium* and *C. trachomatis* with sexual behaviour and demographic characteristics among STI clinic attendees. The age distribution for both *M. genitalium* and *C. trachomatis* was similar with infections more frequently detected in younger women (15 to 19 years 9.7%, 18/186 vs. 6.4%, 12/186, respectively and 20 to 24 years 6.2%, 41/665 vs. 6.0%, 40/665) than other age groups. *M. genitalium* was more frequently detected in 15 to 19 year old women than *C. trachomatis* although this was not statistically significant ($p=0.28$).

Table 3. Association of characteristics with *M. genitalium* and *C. trachomatis* in women attending two London STI clinics

Characteristic	(N=2172) % of women with characteristic	<i>M. genitalium</i> (%) (proportion of women)	aOR ^a (95% CI)	P-value	<i>C. trachomatis</i> proportion of women)	aOR ^a (95% CI)	P-value
Age:							
15-19	8.6	9.7 (18/186)			6.4 (12/186)		
20-24	30.6	6.2 (41/665)			6.0 (40/665)		
25-29	28.6	1.6 (10/621)			2.9 (18/621)		
30-34	15.6	2.3 (9/339)			3.2 (11/339)		
35- 64	16.6	0.8 (3/361)			0.6 (2/361)		
Ethnicity							
White	23.0	6.0 (30/499)	1		7.0 (35/499)	1	
Black or Black British	6.9	7.4 (11/149)	1.2 (0.6-2.5)	0.60	4.0 (6/149)	0.5 (0.2-1.3)	0.54
Asian or Asian British	1.7	17.6 (6/36)	3.1 (1.2-8.1)	0.19	5.6 (2/36)	0.8 (0.2-3.4)	0.73
Mixed	3.9	4.8 (4/84)	0.7 (0.2-2.1)	0.54	7.1 (6/84)	0.9 (0.4-2.3)	0.91
Other Ethnic groups	3.9	9.5 (8/83)	1.6 (0.7-3.7)	0.24	3.6 (3/83)	0.5 (0.1-1.6)	0.49
Unknown	60.8	1.7 (22/1321)	0.5 (0.2-1.1)	0.09	2.3 (31/1321)	0.7 (0.3-1.4)	0.66
Specimen							
Cervical/	90.3	3.8 (75/1961)	1.4 (0.6-3.2)	0.48	3.4 (38/1130)	0.7 (0.4-1.6)	0.44
Self-taken vaginal					4.3 (36/831)		
First catch urine	9.7	2.8 (6/211)	1		4.3 (9/211)	1	

aOR^a odds ratios adjusted for age only

Specimen type and bacterial load

Overall *M. genitalium* was detected in 3.7% (43/1161), 4.0% (74/1817) and 1.3% (21/1635) of cervical swabs, self-taken vulval swabs and first-void urine samples, respectively. Since *M. genitalium* frequency in cervical and self-taken swabs was similar ($p=0.86$), the results for the two groups of swabs were merged and tested against first-void urine samples in the statistical model. *M. genitalium* was significantly more likely to be detected in swabs compared with urine specimens (3.9% vs. 1.3%, $P<0.001$).

The overall ~~prevalence~~ frequency of *C. trachomatis* in cervical swabs, self-taken vulval swabs and first-void urine samples was 3.5% (41/1161), 7.0% (128/1817) and 4.9% (80/1635), respectively. *C. trachomatis* significantly differed between cervical and self-taken swabs ($p<0.001$) and the two groups were separately tested against the urine samples in the statistical model.

The majority (58%, 1424/2441) of specimens provided by the women in NCSP were urine samples. However swab samples were almost four times more likely to test positive for *M. genitalium* compared with urine samples (aOR 3.6, 95% CI 1.9-6.7, $p<0.001$) and *C. trachomatis* ~~prevalence~~ infection was almost twice as high among swabs compared with urine samples (aOR 1.8, 95% CI 1.2-2.4 $p=0.001$). Conversely the majority (90.3%, 1961/2172) of clinic specimens were swabs. *M. genitalium* and *C. trachomatis* in the clinic swab and urine specimens also differed (*M. genitalium* 3.8%, 75/1961 vs. 2.8%, 6/211 and *C. trachomatis* 3.8%, 74/1961 vs. 4.3%, 9/211, respectively).

In quantitative analysis of *M. genitalium* positive specimens, mean *M. genitalium* bacterial load in swab and urine samples did not significantly differ between the clinics or NCSP. Clinic data were therefore combined for comparison of the mean bacterial load in different specimen types. There was no difference in overall cervical and self-taken vaginal

1
2
3 swab bacterial loads (3.72 (CI 3.39-4.05) vs. 3.91 (CI 3.66-4.17) \log_{10} genome copies/mL,
4
5 equivalent to geometric means of 5,218 (CI 2,438-11,171) and 8,192 (CI 4,575-14,669)
6
7 organisms/mL, respectively) ($p=0.349$). The overall mean bacterial load in swabs 3.84 (CI
8
9 3.52-4.11) equivalent to 6,705 (CI 3506-12,920) organisms/mL was significantly higher than
10
11 in first-void urine samples (3.14 (CI 2.87-3.41) equivalent to 1386 (CI 740-2,597)
12
13 organisms/mL) ($p<0.0001$, equal variances not assumed).
14
15

16 17 **Genetic diversity** 18

19
20 The absence of false positive results was confirmed by the presence of 57 different
21
22 genotypes by sequence analysis of 127 *M. genitalium* positive specimens and 13 sequences
23
24 from previously isolated strains (figure 2). The discriminatory index by Hunter and Gaston *et*
25
26 *al.* 1988²⁹ was calculated to be 0.94 both with and without inclusion of the previously
27
28 isolated strain sequences. None of the sequenced samples were identical with the type strain
29
30 G37 used as a PCR standard control. Genetic diversity data are available in FASTA format
31
32 for download in the supplementary material.
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

DISCUSSION

Overall *M.genitalium* was relatively common ~~at with a prevalence of~~ 3.0% among NCSP participants and STI clinic attendees. *M.genitalium* was more likely to be found in swabs compared with urine samples (3.9% vs. 1.3%, respectively) and the mean bacterial load was also much higher (6,705 (CI 3,506-12,920) organisms/mL vs. 1386 (CI 740-2,597) organisms/mL, respectively).

Only 0.5% of all the women had both *C. trachomatis* and *M.genitalium* infections. Among women who had *C. trachomatis*, 9% were co-infected with *M.genitalium* compared with <5% in population based studies.^{16;18;30;31} Among NCPSP participants the age-adjusted odds of detecting *M.genitalium* were twice as high among women of black/black British ethnicity (aOR 2.3) and those reporting multiple sexual partners in the past year (aOR 2.4) compared with women of white ethnicity or those who reported only one partner, respectively. After adjusting for age, *C. trachomatis* was also significantly more likely to be diagnosed in women with multiple partners (aOR 2.0) and new sexual partners in the previous three months (aOR 1.6) but was less likely to be detected in women who did not give a self-identified ethnic group (aOR 0.6) compared with reporting only one partner, not reporting new partners or being of white ethnicity, respectively. No significant associations were observed for either infection among STI clinic attendees. ~~However among STI clinic attendees *M.genitalium* was as, if not more likely, to be detected as *C. trachomatis* among women aged 15-24 years (15-19 years 9.7% vs. 6.4% and 20-24 years 6.2% vs. 6.0%, respectively). It was also more likely to be detected among STI clinic attendees aged 15-24 years compared with NCSP participants (15-19 years 9.7% vs. 2.8% and 20-24 years 6.2% vs. 2.0%, respectively).~~

1
2
3 This is the largest UK based *M. genitalium* study to date to provide prevalence
4 estimates of infection among for both community and STI clinic based populations. Transport
5 media may affect the sensitivity of DNA based PCR tests. The study samples were originally
6 collected in Aptima medium. We therefore tested Aptima and PBS media with *M. genitalium*
7 DNA and did not find any differences. We confirmed positive *M. genitalium* PCR results by
8 genotype sequencing. Our analysis of *M. genitalium* and *C. trachomatis* correlates is limited
9 by availability of data: only age and ethnicity were available for both clinic and NCSP
10 datasets and ethnicity data was missing for 61% of STI clinic attendees. There is also a
11 possibility that some young women may have had chlamydia tests through both the NCSP
12 and the STI clinics during the sample collection period. It is not possible to quantify this
13 although we speculate that the numbers are likely to be low given the relatively short time
14 frame.
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29

30 Our STI clinic *M. genitalium* prevalence-frequency is similar to that found in several
31 studies of female STI clinic attendees (4.5% to 7%)^{32,33} although other studies have reported a
32 much higher prevalence-frequencies (19.3% to 38.2%).^{34,35} In lower risk non-STI clinic
33 attendees such as college students the prevalence-infection has been shown to range from <1%
34 to 5%.^{5,36} which is in keeping with our estimate in the chlamydia screening population. In one
35 clinic in our study *M. genitalium* prevalence was higher than *C. trachomatis* and the lower *C.*
36 *trachomatis* prevalence may reflect variation during the short study period as may the higher
37 *M. genitalium* prevalence. We speculate that it may also be due to variations in chlamydia
38 screening uptake and therefore *C. trachomatis* prevalence in different parts of London.
39
40
41
42
43
44
45
46
47
48
49
50

51 The higher prevalence-frequency of *M. genitalium* in women attending clinics than the
52 NCSP (3-5.3% vs. 2.3%, respectively) may in part reflect the higher proportion of swabs
53 taken in clinics than in NCSP settings. Urine samples have been shown to be less sensitive for
54 *M. genitalium* diagnosis than swabs (61% to 65% compared with 74% to 91%).^{37,38} It is
55
56
57
58
59
60

1
2
3 therefore likely that our NCSP *M.genitalium* prevalence-frequency is an underestimation.
4
5 Although urine sample sensitivity may be increased by up-concentrating the samples by
6
7 centrifugation this is not a practical step for large scale testing. A higher bacterial load may be
8
9 associated with symptoms as has been shown for men.²⁵ This may also explain the difference
10
11 in prevalence-infection between the two populations since STI clinic attendees are more likely
12
13 to be symptomatic than NCSP participants. The association of *M.genitalium* with multiple
14
15 sexual partners and black ethnicity has been previously observed.^{16,31} Additional risk factors
16
17 include younger age as observed in our STI clinic attendees, bacterial vaginosis, being
18
19 symptomatic, cervicitis, douching, smoking, prior miscarriage, menstrual cycle, social class
20
21 and marital status.^{12;16;31;35;39-41}

22
23
24
25
26 *M.genitalium* appears to be a relatively common infection among women in London.
27
28 The low level of *M.genitalium* and *C. trachomatis* co-infection (0.5%) suggests that
29
30 diagnosing and treating chlamydia will have little impact on *M.genitalium*. However
31
32 Azithromycin 1g used to treat uncomplicated *C. trachomatis* infection appears to be sub-
33
34 optimal for *M.genitalium* treatment²⁴ This treatment dose has also been associated with the
35
36 development of *M.genitalium* macrolide resistance in some studies of predominantly
37
38 symptomatic men.^{22 24} The risk of inadvertent *M.genitalium* antibiotic resistance in co-
39
40 infected women who are treated for chlamydia with 1g of azithromycin is therefore
41
42 potentially a cause for concern although further research is required to confirm this.
43
44
45

46
47 To avoid the problems encountered with *C. trachomatis* screening and *M.genitalium*
48
49 antimicrobial resistance, prior to introducing routine testing for *M.genitalium*, further research
50
51 is needed to better understand its natural history, the role of asymptomatic and symptomatic
52
53 *M.genitalium* in PID and determine optimum treatment guidelines.
54
55
56
57
58
59
60

Acknowledgements

The authors would like to thank Menelaos Pavlou (Centre for Sexual Health and HIV Research, Mortimer Market Centre, London, UK) and Tina Sharp (Mortimer Market Centre, London, UK, previously NCSP Chlamydia Co-ordinator, Camden Primary Care Trust) for extracting clinical data from the Sexually Transmitted Disease Clinics and the National Chlamydia Screening Programme, respectively. We thank Dr. Stephane Hue, Center for Virology, UCL for aligning the sequenced fragments and create the resulting phylogenetic tree.

Authors Contribution

All authors contributed to conception and design of the study and / or to acquisition of data. HS performed the experiments. HS SSD and JS drafted the paper and all authors contributed to critical revision of the paper.

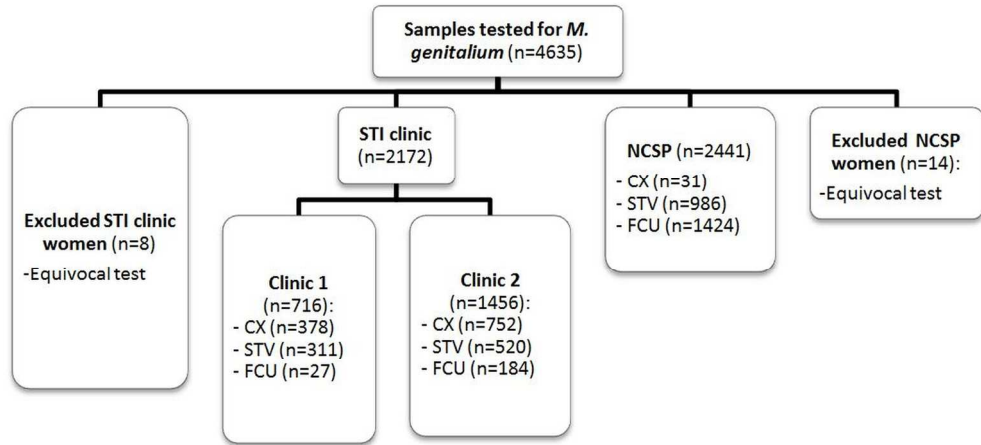
Reference List

- 1
2
3
4
5
6
7 (1) Barrett S, Taylor C. A review on pelvic inflammatory disease. *Int J STD AIDS* 2005; 16(11):715-
8 720.
- 9
10 (2) Simms I, Stephenson JM. Pelvic inflammatory disease epidemiology: what do we know and
11 what do we need to know? *Sex Transm Infect* 2000; 76(2):80-87.
- 12
13 (3) Haggerty CL, Ness RB. Diagnosis and treatment of pelvic inflammatory disease. *Womens*
14 *Health (Lond Engl)* 2008; 4(4):383-397.
- 15
16 (4) Haggerty CL, Taylor BD. Mycoplasma genitalium: an emerging cause of pelvic inflammatory
17 disease. *Infect Dis Obstet Gynecol* 2011; 2011:959816.
- 18
19 (5) McGowin CL, Anderson-Smits C. Mycoplasma genitalium: an emerging cause of sexually
20 transmitted disease in women. *PLoS Pathog* 2011; 7(5):e1001324.
- 21
22 (6) Taylor-Robinson D, Jensen JS. Mycoplasma genitalium: from Chrysalis to multicolored
23 butterfly. *Clin Microbiol Rev* 2011; 24(3):498-514.
- 24
25 (7) Bjartling C. The association between Mycoplasma genitalium and pelvic inflammatory
26 disease after termination of pregnancy. 2010.
- 27
28 (8) Bjartling C. Mycoplasma genitalium in cervicitis and pelvic inflammatory disease among
29 women at a gynecologic outpatient service. 2012.
- 30
31 (9) Cohen CR, Manhart LE, Bukusi EA, Astete S, Brunham RC, Holmes KK et al. Association
32 between Mycoplasma genitalium and acute endometritis. *Lancet* 2002; 359(9308):765-766.
- 33
34 (10) Svenstrup HF, Fedder J, Kristoffersen SE, Trolle B, Birkelund S, Christiansen G. Mycoplasma
35 genitalium, Chlamydia trachomatis, and tubal factor infertility - a prospective study. *Fertility*
36 *and Sterility* 2008; 90(3):513-520.
- 37
38 (11) Manhart LE, Broad JM, Golden MR. Mycoplasma genitalium: should we treat and how? *Clin*
39 *Infect Dis* 2011; 53 (s3): s129-42.
- 40
41 (12) Mobley VL, Hobbs MM, Lau K, Weinbaum BS, Getman DK, Sena AC. Mycoplasma genitalium
42 infection in women attending a sexually transmitted infection clinic: diagnostic specimen
43 type, coinfections, and predictors. *Sex Transm Dis* 2012; 39(9):706-709.
- 44
45 (13) Jensen JS. *Mycoplasma genitalium* - the etiological agent of urethritis and other Sexually
46 Transmitted Diseases. *Sex Transm Dis* 2003.
- 47
48 (14) Mavedzenge SN, Van Der PB, Weiss HA, Kwok C, Mambo F, Chipato T et al. The association
49 between Mycoplasma genitalium and HIV-1 acquisition in African women. *AIDS* 2012;
50 26(5):617-624.
- 51
52 (15) Low N, Bender N, Nartey L, Shang A, Stephenson JM. Effectiveness of chlamydia screening:
53 systematic review. *International Journal of Epidemiology* 2009; 38(2):435-448.
- 54
55
56
57
58
59
60

- 1
2
3 (16) Oakeshott P, Aghaizu A, Hay P, Reid F, Kerry S, Atherton H et al. Is *Mycoplasma genitalium* in
4 women the "New Chlamydia?" A community-based prospective cohort study. *Clin Infect Dis*
5 2010; 51(10):1160-1166.
6
7 (17) Anagrus C, Lore B, Jensen JS. *Mycoplasma genitalium*: prevalence, clinical significance, and
8 transmission. *Sex Transm Infect* 2005; 81(6):458-462.
9
10 (18) Andersen B, Sokolowski I, Ostergaard L, Moller JK, Olesen F, Jensen JS. *Mycoplasma*
11 *genitalium*: prevalence and behavioural risk factors in the general population. *Sexually*
12 *Transmitted Infections* 2007; 83(3):237-241.
13
14 (19) Jensen JS, Bradshaw CS, Tabrizi SN, Fairley CK, Hamasuna R. Azithromycin Treatment Failure
15 in *Mycoplasma genitalium* Positive Patients with Nongonococcal Urethritis Is Associated
16 with Induced Macrolide Resistance. *Clinical Infectious Diseases* 2008; 47(12):1546-1553.
17
18 (20) Lau CY, Qureshi AK. Azithromycin Versus Doxycycline for Genital Chlamydial Infections: A
19 Meta-Analysis of Randomized Clinical Trials. *Sexually Transmitted Diseases* 2002; 29(9).
20
21 (21) Horner PJ. Azithromycin antimicrobial resistance and genital Chlamydia trachomatis
22 infection: duration of therapy may be the key to improving efficacy. 2012.
23
24 (22) Anagrus C, Lore B, Jensen JS. Treatment of *Mycoplasma genitalium*. Observations from a
25 Swedish STD clinic. *PLoS One* 2013; 8(4):e61481.
26
27 (23) Tagg KA, Jeoffreys NJ, Couldwell DL, Donald JA, Gilbert GL. Fluoroquinolone and macrolide
28 resistance-associated mutations in *Mycoplasma genitalium*. *J Clin Microbiol* 2013;
29 51(7):2245-2249.
30
31 (24) Twin J. Transmission and selection of macrolide resistant *Mycoplasma genitalium* infections
32 detected by rapid high resolution melt analysis. 2012.
33
34 (25) Jensen JS, Bjornelius E, Dohn B, Lidbrink P. Use of TaqMan 5' nuclease real-time PCR for
35 quantitative detection of *Mycoplasma genitalium* DNA in males with and without urethritis
36 who were attendees at a sexually transmitted disease clinic. *J Clin Microbiol* 2004; 42(2):683-
37 692.
38
39 (26) Chalker VJ, Jordan K, Ali T, Ison C. Real-time PCR detection of the mg219 gene of unknown
40 function of *Mycoplasma genitalium* in men with and without non-gonococcal urethritis and
41 their female partners in England. *J Med Microbiol* 2009; 58(Pt 7):895-899.
42
43 (27) Garson JA, Grant PR, Ayliffe U, Ferns RB, Tedder RS. Real-time PCR quantitation of hepatitis B
44 virus DNA using automated sample preparation and murine cytomegalovirus internal
45 control. *J Virol Methods* 2005; 126(1-2):207-213.
46
47 (28) Hjorth SV, Bjornelius E, Lidbrink P, Falk L, Dohn B, Berthelsen L et al. Sequence-based typing
48 of *Mycoplasma genitalium* reveals sexual transmission. *J Clin Microbiol* 2006; 44(6):2078-
49 2083.
50
51 (29) Hunter PR, Gaston MA. Numerical index of the discriminatory ability of typing systems: an
52 application of Simpson's index of diversity. *J Clin Microbiol* 1988; 26(11):2465-2466.
53
54
55
56
57
58
59
60

- 1
2
3 (30) Manhart LE, Holmes KK, Hughes JP, Houston LS, Totten PA. Mycoplasma genitalium among
4 young adults in the United States: an emerging sexually transmitted infection. *Am J Public*
5 *Health* 2007; 97(6): 1118-1125.
6
7 (31) Walker J, Fairley CK, Bradshaw CS, Tabrizi SN, Chen MY, Twin J et al. 'The difference in
8 determinants of Chlamydia trachomatis and Mycoplasma genitalium in a sample of young
9 Australian women'. *BMC Infect Dis* 2011; 11:35.
10
11 (32) Manhart LE, Critchlow CW, Holmes KK, Dutro SM, Eschenbach DA, Stevens CE et al.
12 Mucopurulent cervicitis and Mycoplasma genitalium. *J Infect Dis* 2003; 187(4):650-657.
13
14 (33) Moi H, Reinton N, Moghaddam A. Mycoplasma genitalium in women with lower genital tract
15 inflammation. *Sex Transm Infect* 2009; 85(1):10-14.
16
17 (34) Casin I, Vexiau-Robert D, De La SP, Eche A, Grandry B, Janier M. High prevalence of
18 Mycoplasma genitalium in the lower genitourinary tract of women attending a sexually
19 transmitted disease clinic in Paris, France. *Sex Transm Dis* 2002; 29(6):353-359.
20
21 (35) Mobley VL, Hobbs MM, Lau K, Weinbaum BS, Getman DK, Sena AC. Mycoplasma genitalium
22 infection in women attending a sexually transmitted infection clinic: diagnostic specimen
23 type, coinfections, and predictors. *Sex Transm Dis* 2012; 39(9):706-709.
24
25 (36) Jensen AJ, Kleveland CR, Moghaddam A, Haaheim H, Hjelmevoll SO, Skogen V. Chlamydia
26 trachomatis, Mycoplasma genitalium and Ureaplasma urealyticum among students in
27 northern Norway. *J Eur Acad Dermatol Venereol* 2013; 27(1):e91-e96.
28
29 (37) Lillis RA, Nsuami MJ, Myers L, Martin DH. Utility of urine, vaginal, cervical, and rectal
30 specimens for detection of Mycoplasma genitalium in women. *J Clin Microbiol* 2011;
31 49(5):1990-1992.
32
33 (38) Wroblewski JK, Manhart LE, Dickey KA, Hudspeth MK, Totten PA. Comparison of
34 transcription-mediated amplification and PCR assay results for various genital specimen
35 types for detection of Mycoplasma genitalium. *J Clin Microbiol* 2006; 44(9):3306-3312.
36
37 (39) Short VL, Totten PA, Ness RB, Astete SG, Kelsey SF, Murray P et al. The demographic, sexual
38 health and behavioural correlates of Mycoplasma genitalium infection among women with
39 clinically suspected pelvic inflammatory disease. *Sex Transm Infect* 2010; 86(1):29-31.
40
41 (40) Oakeshott P, Hay P, Taylor-Robinson D, Hay S, Dohn B, Kerry S et al. Prevalence of
42 Mycoplasma genitalium in early pregnancy and relationship between its presence and
43 pregnancy outcome. *BJOG* 2004; 111(12):1464-1467.
44
45 (41) Vandepitte J, Muller E, Bukonya J, Nakubulwa S, Kyakuwa N, Buve A et al. Prevalence and
46 correlates of Mycoplasma genitalium infection among female sex workers in Kampala,
47 Uganda. *J Infect Dis* 2012; 205(2):289-296.
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60



198x90mm (300 x 300 DPI)

For peer review only



Figure 2: Phylogenetic tree showing clustering of 127 DNA sequences from the *M. genitalium* positive specimens of the study (marked with grey font) and 13 DNA sequences from *M. genitalium* strain from patients with no known sexual relationship (marked with black font) 139x198mm (300 x 300 DPI)