

Supplementary Materials

Tenascin C protects aorta from acute dissection in mice

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Supplementary Figure 1. Effects of Ca+AngII treatment.

(a) Images of EVG staining of the infrarenal aortae of wild type (WT) and TNC-KO (KO) mice with the indicated treatment. (b) Diameters of infrarenal aortae from WT or TNC-KO mice with the indicated treatment. The number of animals are indicated in parentheses. (c) Systolic blood pressure in WT (open circles) and TNC-KO (closed circles) mice before and after the Ca+AngII treatment. Data are shown as mean \pm SD.

Supplementary Figure 2. AAD in TNC-KO mice.

(a) Representative ultrasonographic image of an enlarged aorta in TNC-KO mice. Bars, 0.5 mm. (b) Images of EVG staining of AAD in TNC-KO mice 6 weeks after Ca+AngII treatment. Insets are enlarged images of the rectangles in the upper panels and show disrupted elastic lamellae (arrows). The true lumen (tr) and the false lumen (fl) of the aorta are indicated.

Supplementary Figure 3. Cytokine profile of mice treated with AngII or Ca+AngII.

Quantitative analysis of cytokines (Bio-plex, Bio-Rad) in the sera of mice without (Cont, n = 4) and with AngII (n = 5) or Ca+AngII (n = 8) treatment. Data are shown as mean \pm SD.

Supplementary Figure 4. Physiological measurements in live mice.

(a) Blood pressure measurements by aortic catheterization at various positions in aortae as illustrated at the bottom of the panel. Mice were untreated (open circles) or treated with Ca (closed circles), AngII (open squares), or Ca+AngII (closed squares) for 1 week. Data are the mean \pm SD of eight independent observations. Systolic blood pressures were deliberately adjusted to approximately 100 mmHg for the measurement of dP/dt as shown in Figure 2. (b) The descent rates of the anterior aortic walls as determined by ultrasonography in live mice with no treatment (open columns) and Ca+AngII treatment for 1 week (closed columns). Data (% end diastolic diameter/sec) are the mean \pm SD of six independent observations. *P < 0.05 and ***P < 0.001 as compared to control (Cont).

Supplementary Figure 5. TNC expression in aorta treated with AngII or Ca+AngII.

(a) β -galactosidase staining of aortae from *Tnc* reporter mice 0 (Cont), 4 or 6 weeks after starting the indicated treatment. Note that CaCl₂-treated infrarenal aortae

(brackets) developed fibrosis that interfered with penetration of the chromogenic substrate and thus with color development. **(b)** Cross-sectional images of the aorta 4 weeks after the Ca+AngII treatment. The tissues were stained for β -galactosidase activity (β gal), smooth muscle α -actin (α SMA) and hematoxylin (Nuclei). Bar, 50 μ m. **(c)** Serum levels of TNC are shown. Data are mean \pm SD of five independent observations for each group. *P < 0.05 compared to control (Cont).

Supplementary Figure 6. Transcriptome of the suprarenal aorta before AAD onset. Functional annotation clusters obtained by DAVID are shown as heat maps of the average linkage hierarchical clustering with the corresponding gene symbols of subclusters. Functional annotation clusters are shown in the order of the enrichment scores from panels **(a)** to **(d)**.

Supplementary Figure 7. Gene expression profile of aortae and smooth muscle cells. **(a)** Induction of a panel of proinflammatory genes in the suprarenal aorta was analyzed by quantitative RT-PCR using the RT² Profiler PCR Array System (SA Biosciences, PAMM-077). Total RNA samples were obtained from the suprarenal aortae of WT mice (blue bars) or TNC-KO mice (red bars) after 1 week of Ca+AngII treatment. Data are the mean \pm SD of eight independent observations and are expressed as the log ratio of induction from baseline. Data are plotted in order from greatest to least induction in TNC-KO mice. **(b)** The same set of genes as in **(a)** were analyzed in TNC-KO aortic smooth muscle cells (SMC) cultured in the presence (blue bars) or absence (red bars) of exogenously added TNC (10 μ g/mL). Induction of the genes by 10 ng/mL TNF α are shown as the mean \pm SD of four independent observations and are shown as log ratios. **(c)** Induction of collagen family genes was analyzed using GeneChip (Mouse Genome 430 2.0, Affymetrix) in the samples in **(b)**. Genes with the detection call “absence” in all samples were omitted. Data are plotted in order of greatest to least induction in aortic SMC in the absence of exogenous TNC for **(b)** and **(c)**.

Supplementary Figure 8. Histological analyses of aortae from WT or TNC-KO mice with or without 1 week of Ca+AngII treatment. **(a)** Representative images of picrosirius red staining of aortae are shown. Branches (br) were excluded from the analysis. **(b)** Quantitative analysis of adventitial sirius red staining is shown. Animal numbers are

indicated in parentheses. (c) Representative images of immunohistochemical stainings are shown for type I collagen, lysyl oxidase and fibronectin. Bars, 200 μ m.

Supplementary Figure 9. Imaging cytometric analysis for cell populations. Aortae were obtained from WT or TNC-KO mice with or without 1 week of Ca+AngII treatment. The number of animals are indicated in parentheses. Subpopulation without α SMA staining was analyzed for CD45 expression in the aortic tissue sections.

Supplementary Figure 10. Imaging cytometric analyses for NF κ B, phospho-Stat3 (P-Stat3) and phospho-Smad2 (P-Smad2) are shown for total populations and subpopulations with or without α SMA expression from the same dataset for Figure 5a. Blue and red lines indicate the histograms of WT and TNC-KO, respectively. Gray arrows indicate the changes in cell populations in TNC-KO.

Supplementary Movie 1. OCT imaging of AAD in mice.

A series of optical sections obtained by optical coherence tomography (OCT) of an enlarged aorta from a Ca+AngII-treated TNC-KO mouse. The aorta was scanned from the distal to the proximal portion, and the scans spanned the enlarged region.

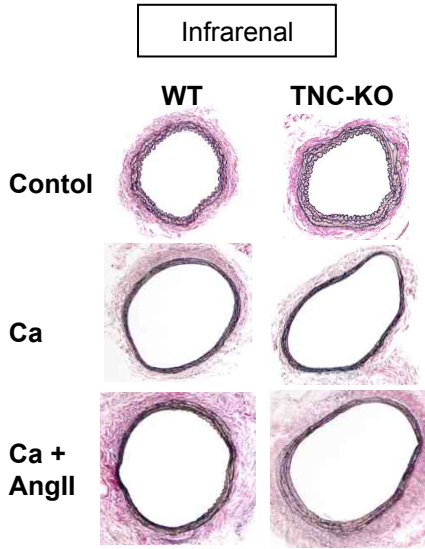
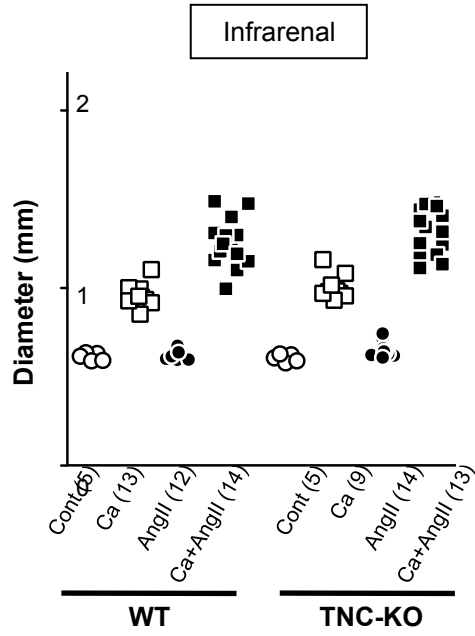
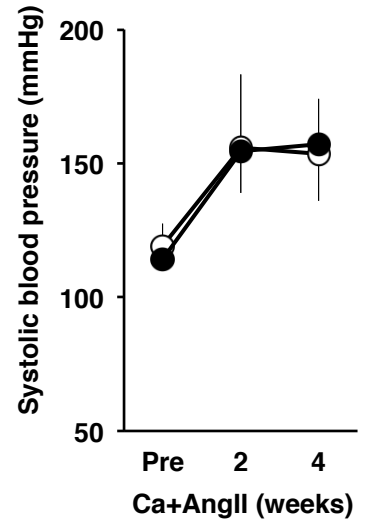
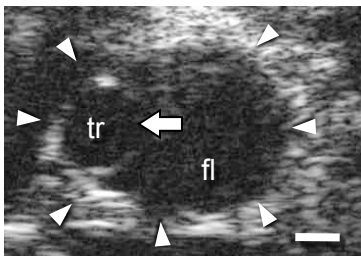
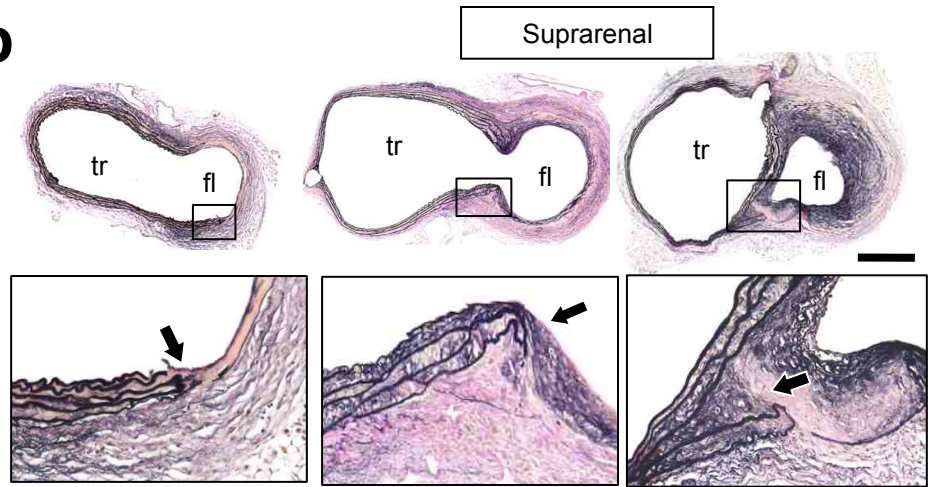
Supplementary Movie 2. Three-dimensional reconstruction of OCT images.

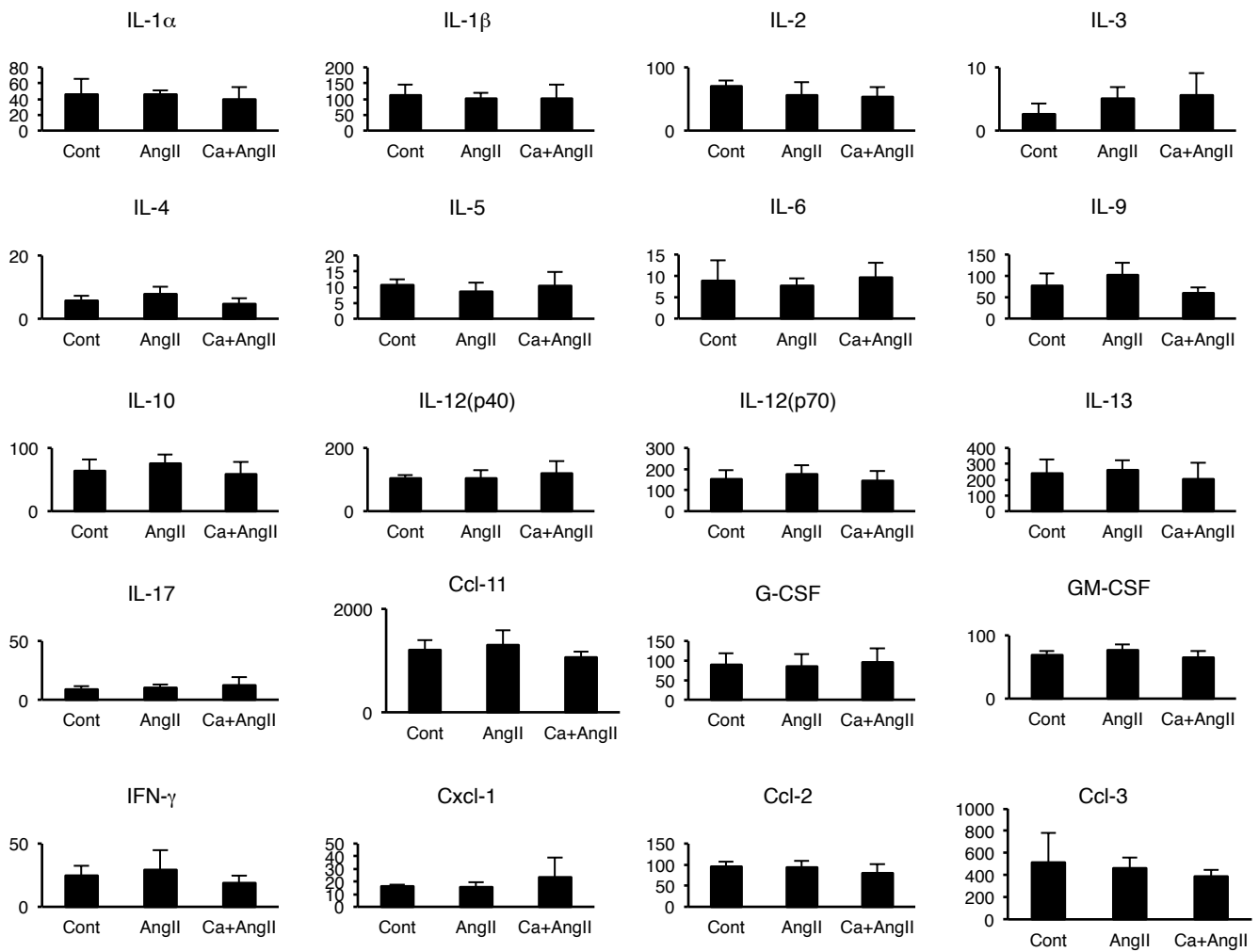
3D reconstruction of a mouse AAD from a series of OCT images using OsiriX Imaging Software (version 5.0).

External Databases 1. The dataset for transcriptome analysis.

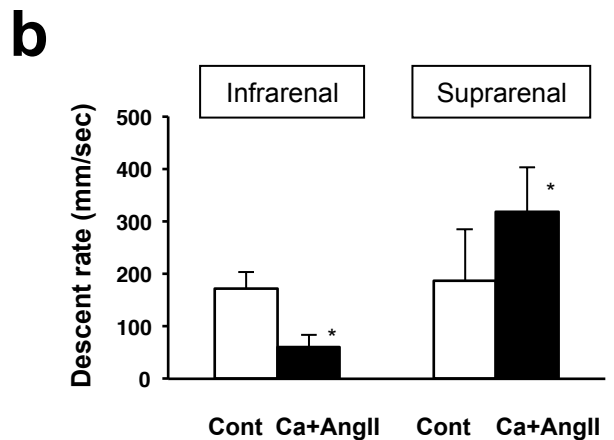
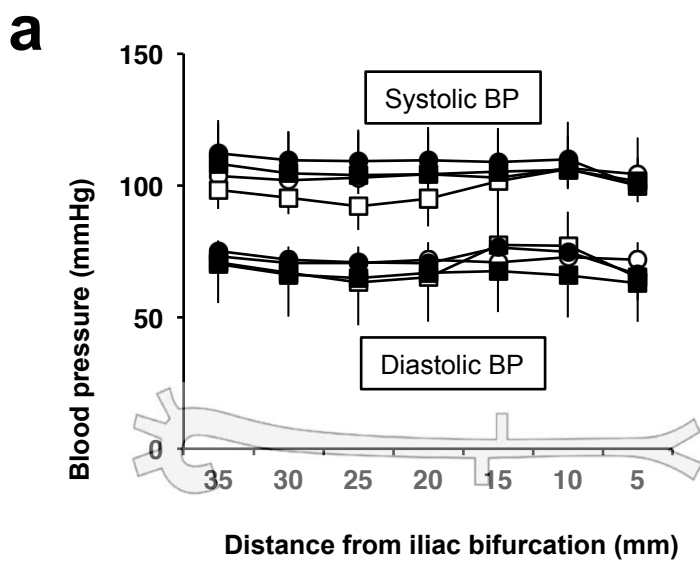
The dataset for the transcriptome analysis can be found in Gene Expression Omnibus at the following link:

<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?token=fxgvrosykcciedq&acc=GSE3689>

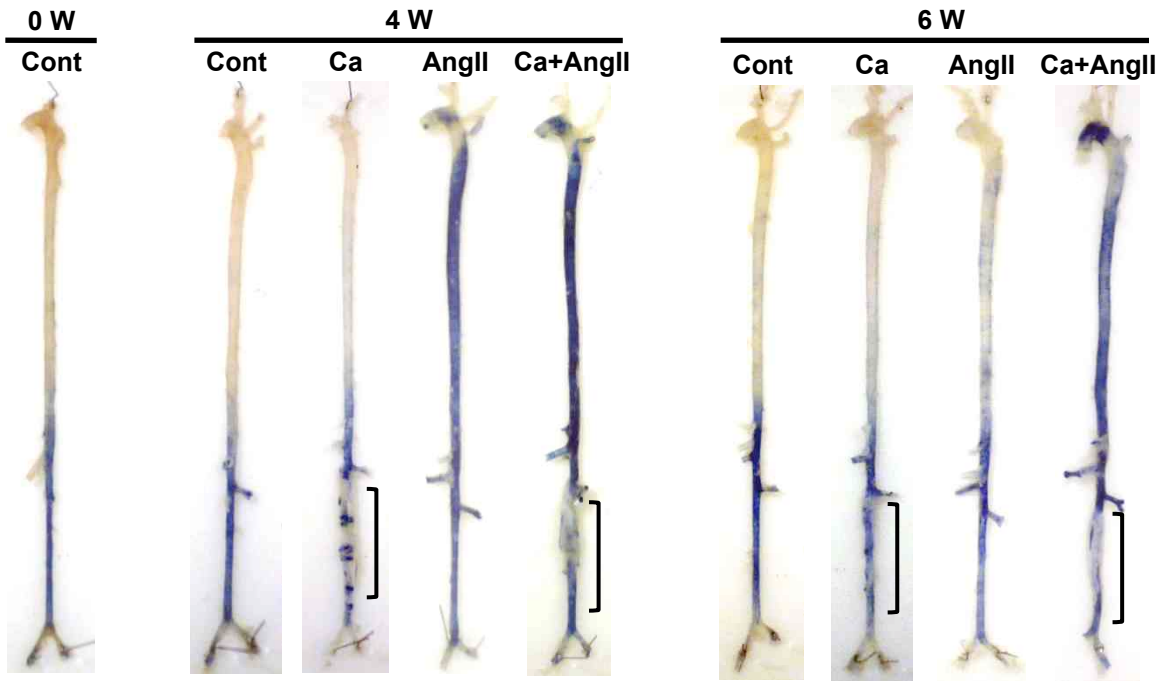
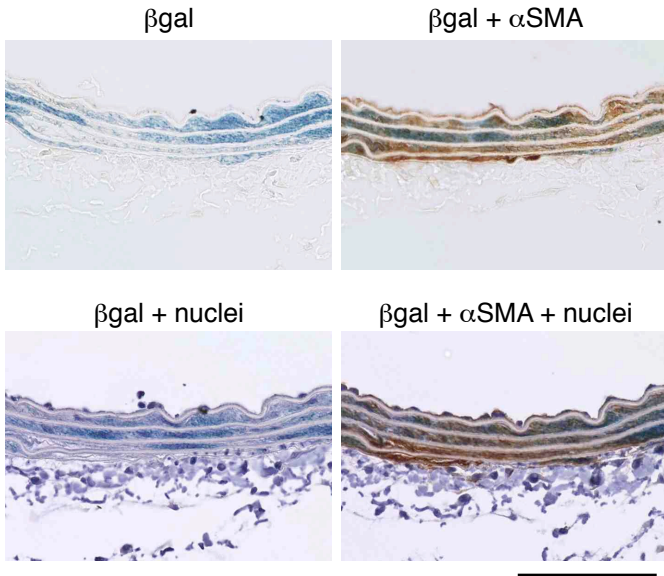
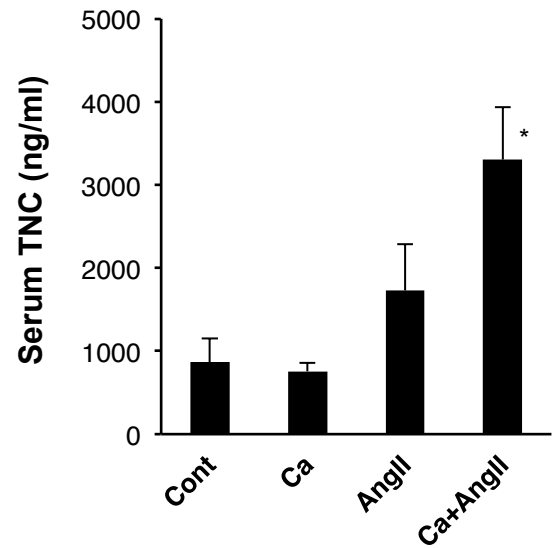
a**b****c****Supplementary Figure 1****a****b****Supplementary Figure 2**

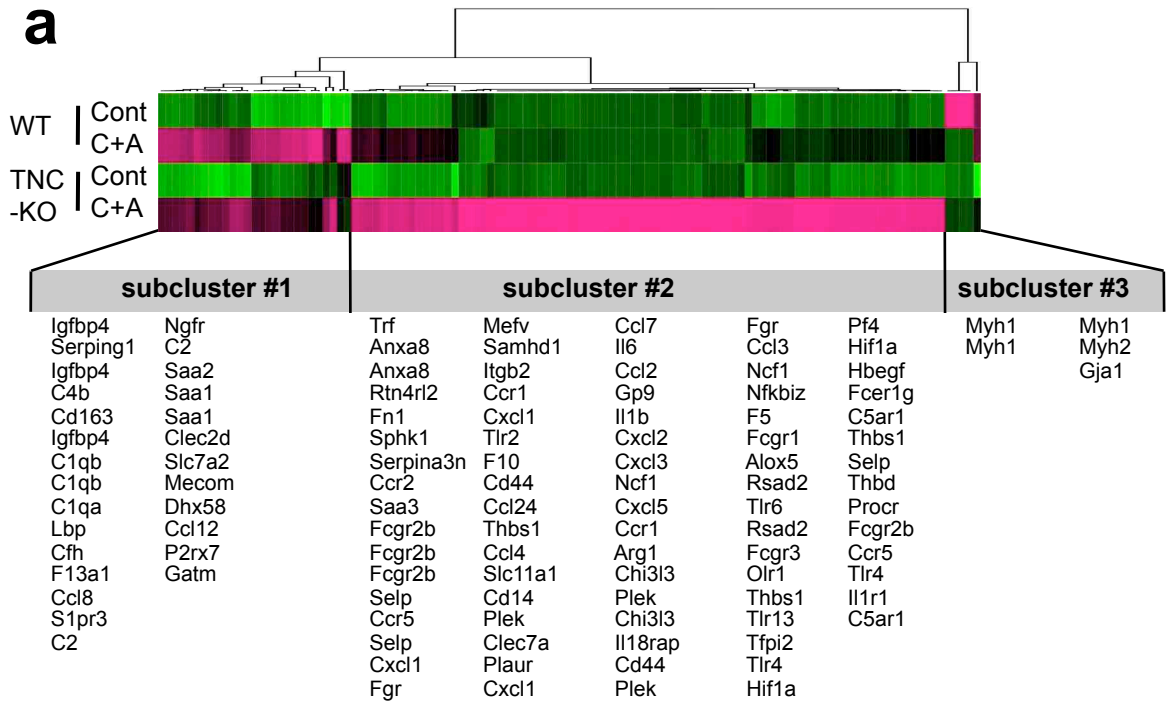


Supplementary Figure 3

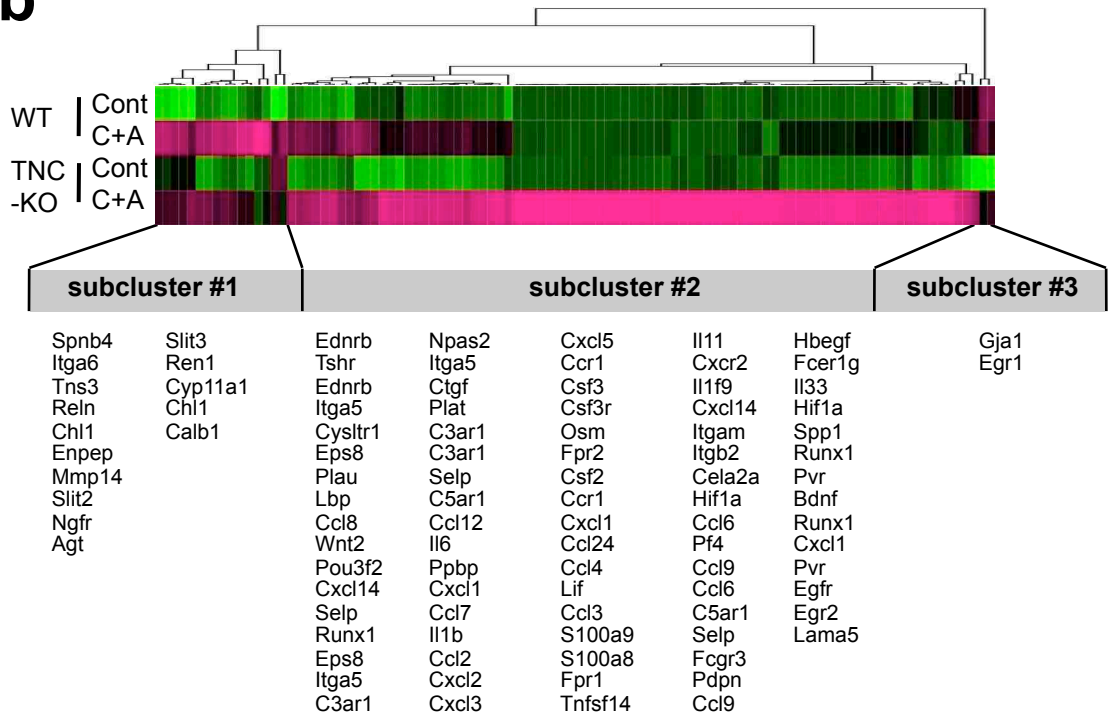


Supplementary Figure 4

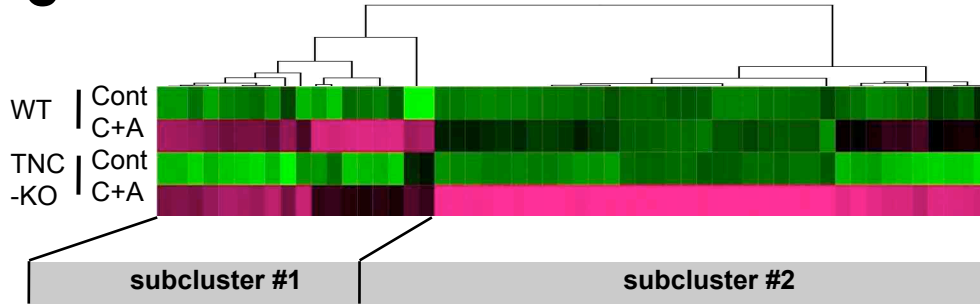
a**b****c****Supplementary Figure 5**

a

Annotation Cluster 2 Enrichment Score: 24.71
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 GOTERM_BP_FAT GO:0006952~defense response

b

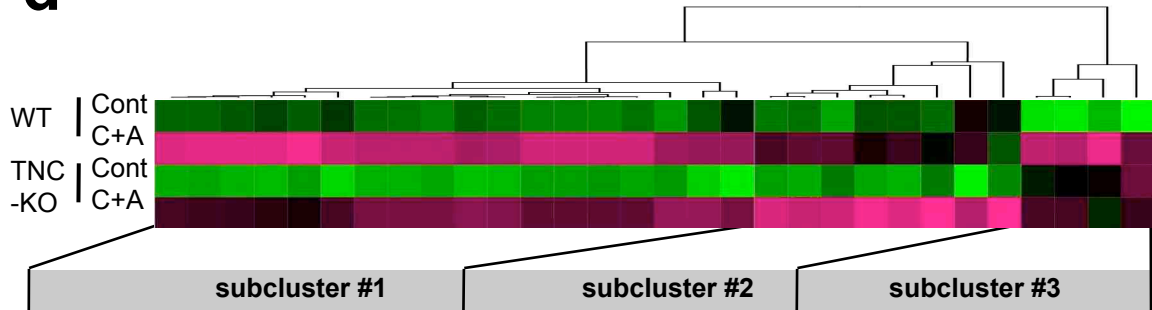
Annotation Cluster 3 Enrichment Score: 20.08
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c

Clec10a	Fbln7	Selp	Olr1	Clec5a	Ctgf
Clec11a	Clec2d	Thbs1	Tnfaip6	Cd44	Itih4
Fbn1	Col5a1	Hbegf	Ccl7	Clec7a	Cd93
Cd209a	Comp	Lgals3	Clec4d	Thbs1	Gfpt2
Cfh	Clec4a2	Klra2	Clec4e	Ptx3	Fgf2
Clec4a3	Col5a1	Clec4e	Clec4n	Clec1b	Selp
Ccl8	Gpnmb	Thbs1	Chi3l3	Fn1	
Mrc1	Col5a3	Itgam	Cd44	Sele	
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Annotation Cluster 4 Enrichment Score: 10.37

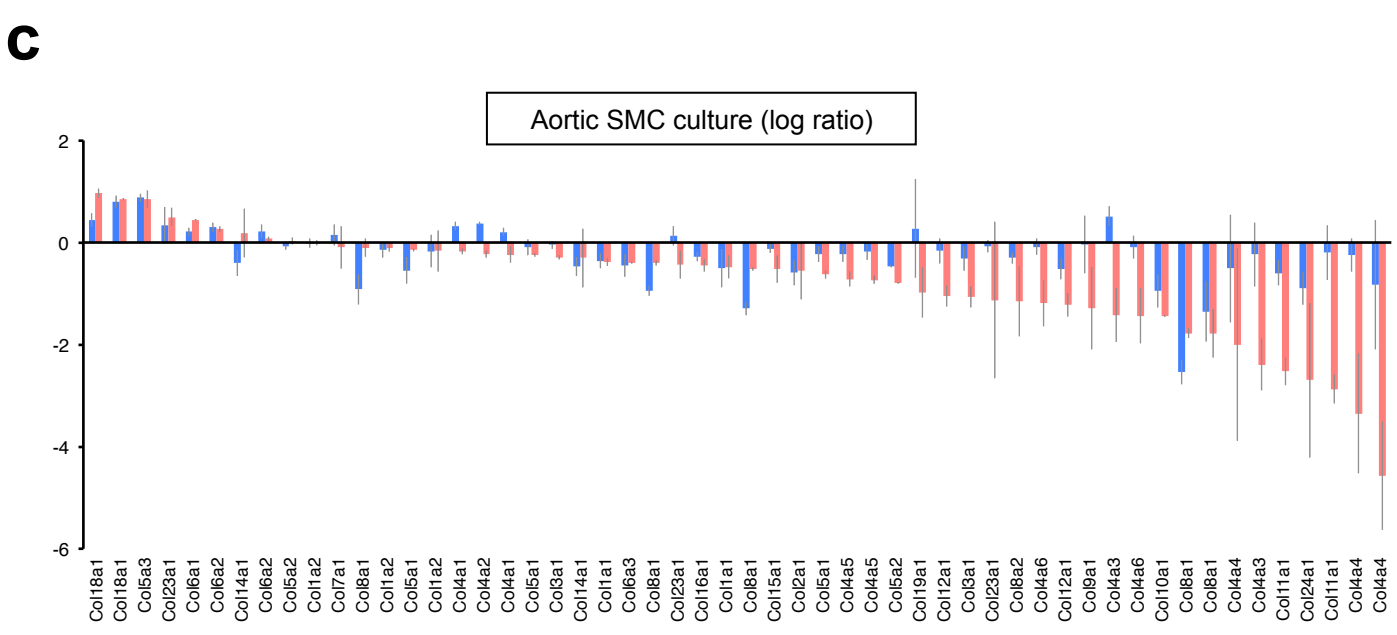
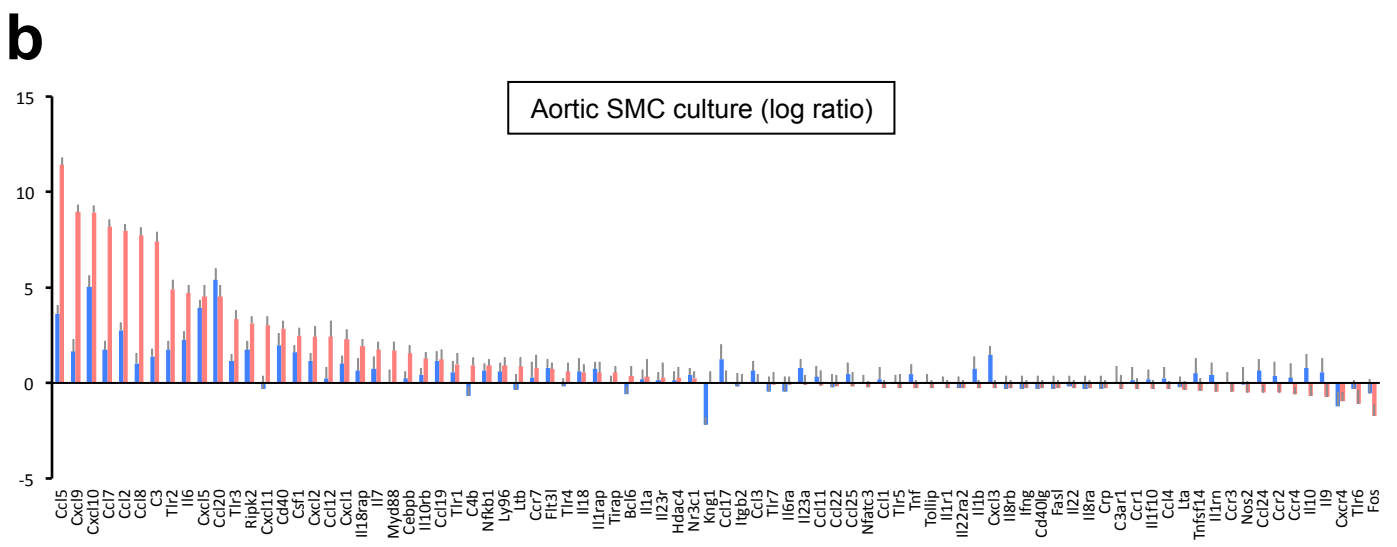
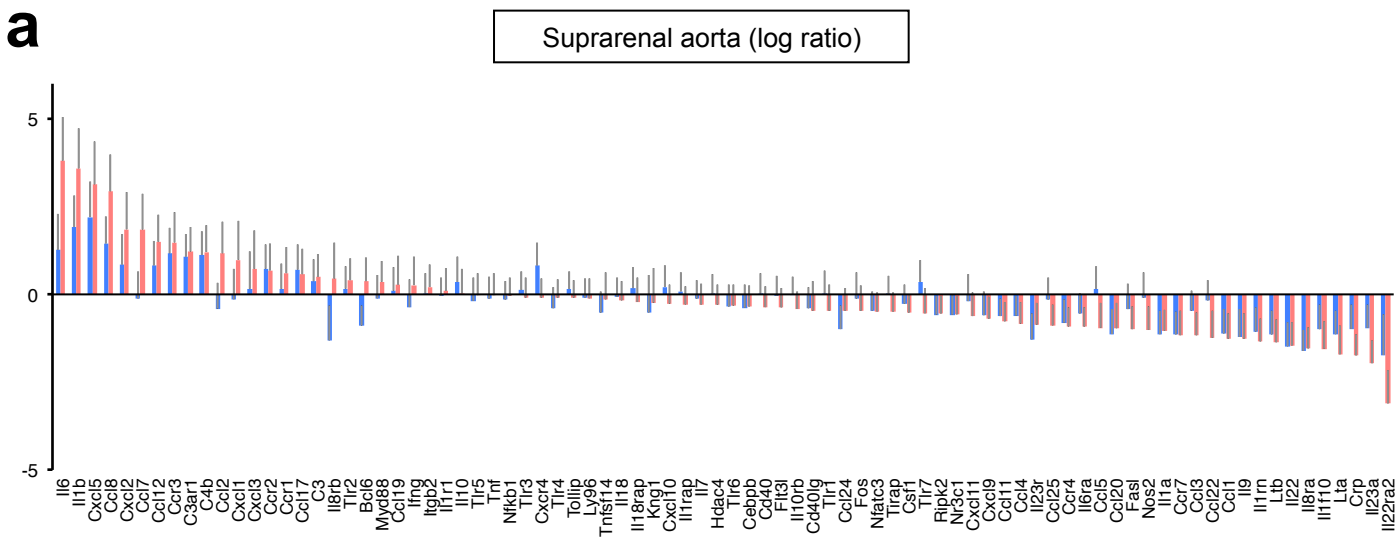
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 GOTERM_BP_FAT GO:0060326~cell chemotaxis
 GOTERM_BP_FAT GO:0030595~leukocyte chemotaxis
 GOTERM_BP_FAT GO:0050900~leukocyte migration
 GOTERM_BP_FAT GO:0007626~locomotory behavior
 GOTERM_MF_FAT GO:0005125~cytokine activity
 GOTERM_BP_FAT GO:0007610~behavior
 GOTERM_BP_FAT GO:0030593~neutrophil chemotaxis
 GOTERM_BP_FAT GO:0016477~cell migration
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 GOTERM_BP_FAT GO:0051674~localization of cell
 GOTERM_BP_FAT GO:0006928~cell motion

d

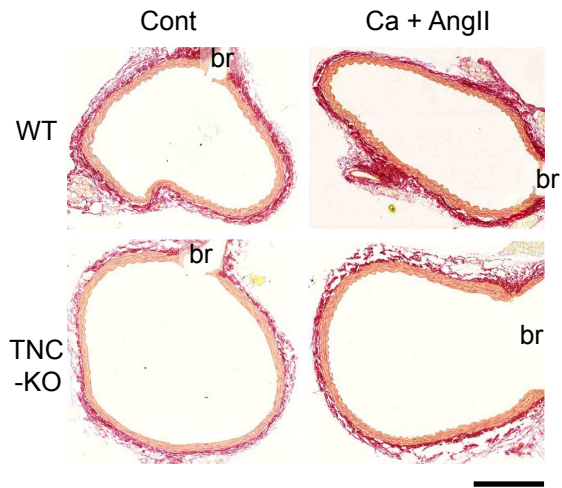
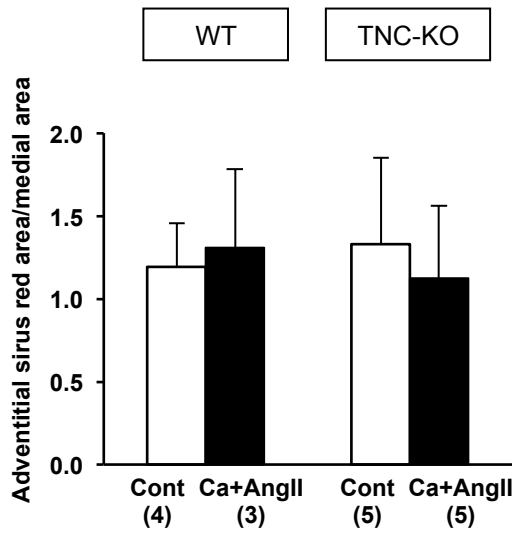
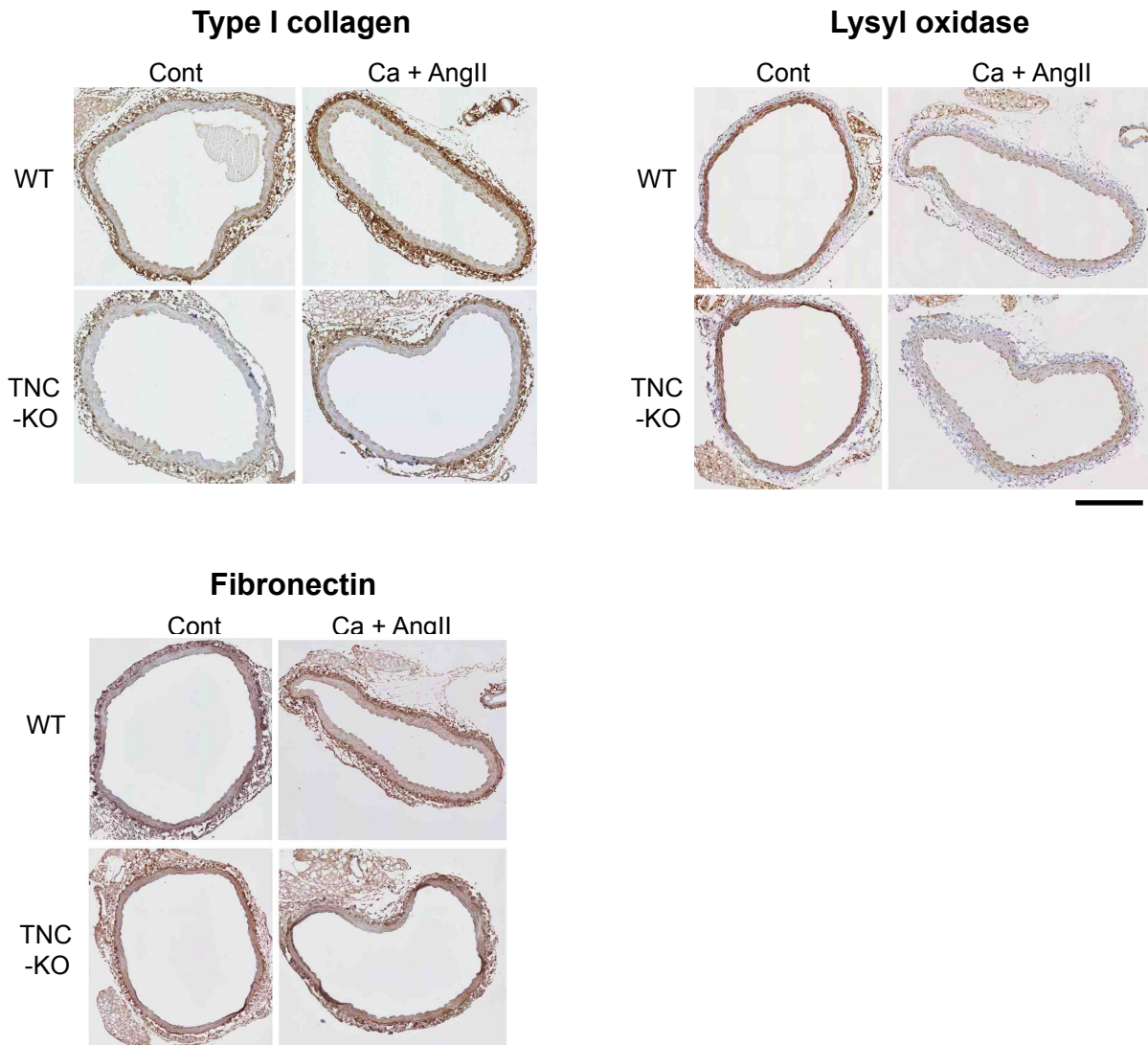
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Col1a1	Col4a2	Fbln1	Lox	Timp1	Col5a3
Col5a1	Col5a2	Col1a2	Smoc2	Lama5	Col11a1
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Col8a1	Col4a1	Eln			

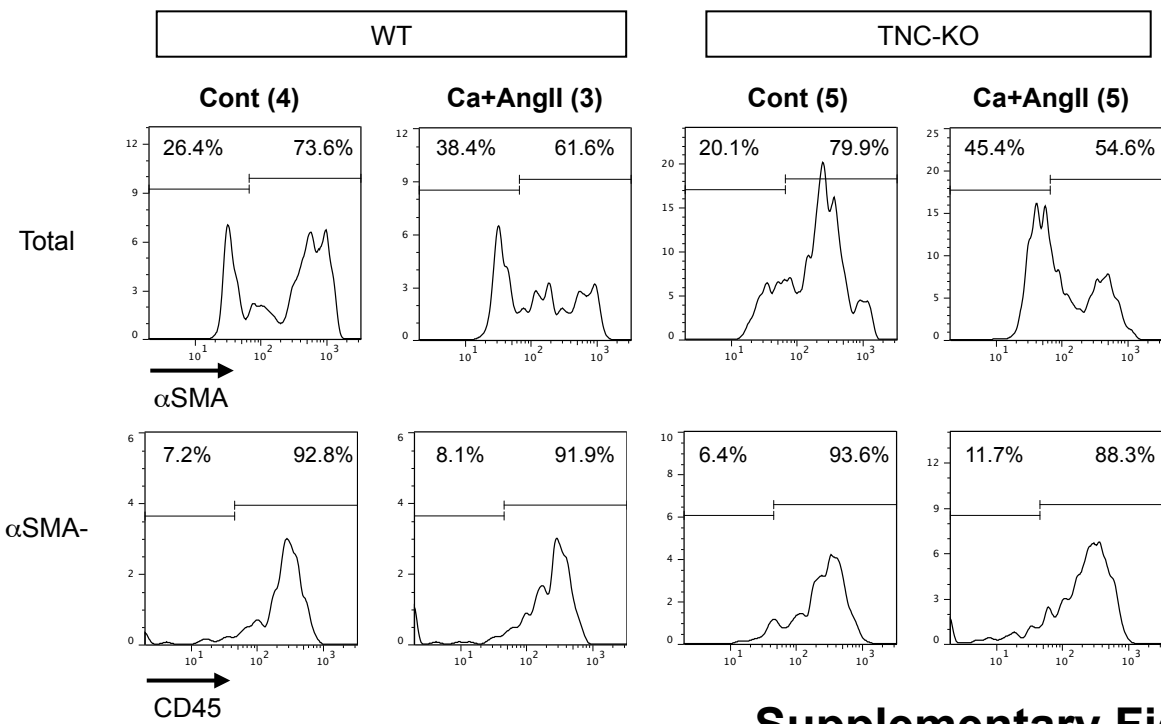
Annotation Cluster 5 Enrichment Score: 7.86

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 GOTERM_MF_FAT GO:0005539~glycosaminoglycan binding
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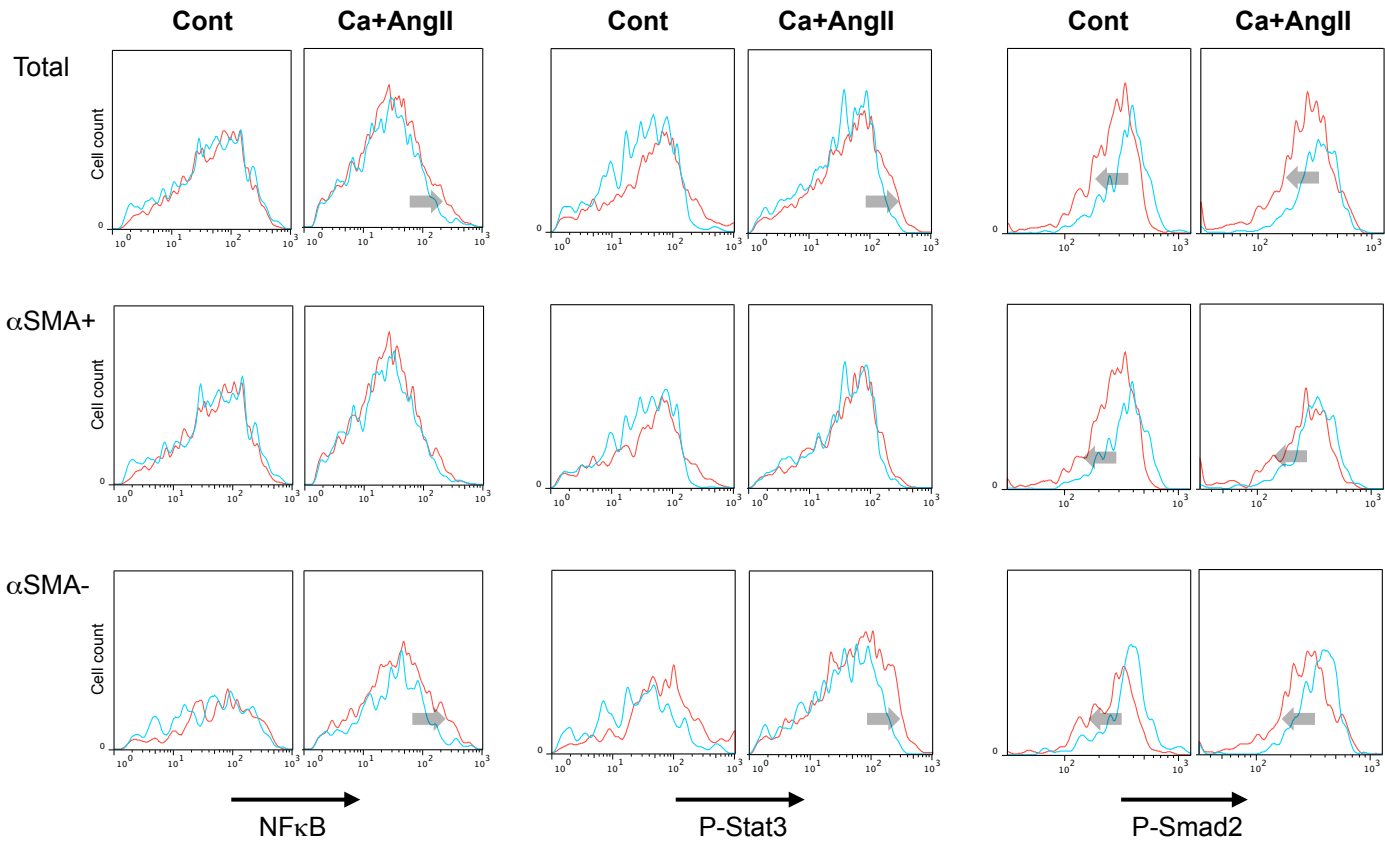


Supplementary Figure 7

a**b****c**



Supplementary Figure 9



Supplementary Figure 10