Supplemental Materials

Molecular Biology of the Cell

Antoniali et al.

SIRT1 gene expression upon genotoxic damage is regulated by APE1 through nCaREpromoter elements

Giulia Antoniali^a, Lisa Lirussi^a, Chiara D'Ambrosio^b, Fabrizio Dal Piaz^c, CarloVascotto^a, Elena Casarano^a, Daniela Marasco^{d,e}, Andrea Scaloni^b, Federico Fogolari^a and Gianluca Tell^{a,#}

^a Department of Biomedical Sciences and Technologies, University of Udine, 33100 Udine, Italy;

^bProteomics & Mass Spectrometry Laboratory, ISPAAM, National Research Council, 80147 Naples, Italy;

^c Department of Biomedical and Pharmaceutical Sciences, University of Salerno, 84084 Fisciano (Salerno), Italy;

^d Department of Biological Sciences, University of Naples "Federico II", 80134 Naples, Italy;

e Institute of Biostructures and Bioimaging, National Research Council, 80134 Naples, Italy

SUPPLEMENTAL MATERIAL

List of contents:

- Page 2-3, Supplemental Results:
- Page 4-6, Supplemental Figure Legends;
- Page 7, Supplemental Table Legends;
- Page 8-14, Supplemental Figures.

Supplemental Results

Limited proteolysis analyses

A combined limited proteolysis-mass spectrometry analysis was carried out to investigate the APE1 protein region responsible for the nCaRE binding. Experiments were performed on a recombinant APE1 form bearing three additional amino acids at protein N-terminus with respect to the native counterpart. Panels *І* and *ІV* of Figure S5 show the time-course LC-ESI-MS analysis of the endoprotease AspN digestion performed on isolated APE1, as carried out by using an enzyme/substrate of 1:500 w/w. Under these experimental conditions, APE1 remained partly undigested, showing that its native conformation is susceptible to proteolysis at a unique, likely flexible site. After 5 min, only the peptide pair 1-17 and 18-321 was detected (Figure S5, *panel I*), and no further fragments were released after 60 (Figure S5, *panel IV*) and 120 min (data not shown) (Supplemental Table S4). As previously reported (Scaloni *et al.*, 1999), the definition of the primary cleavage sites was inferred by the identification of the complementary peptides released by a single proteolytic event occurring on the intact protein. According to these criteria, non-complexed APE1 was preferentially cleaved at D18 present within the unstructured protein N-terminal domain. When the APE1-SIRT1 nCaRE-B oligonucleotide complex was analyzed under the same conditions, no proteolytic fragments resulting from D18 cleavage were released even after 60 min (Figure S5, *panel II* and *V*) (Supplemental Table S4), demonstrating a tighter conformation of the complex and a DNA-shielding effect for D18. A significantly reduced digestion product at this site was observed only after incubation for 120 min (data not shown). A similar result was obtained also for the APE1-PTH nCaRE-B oligonucleotide complex, which showed absent or negligible proteolysis at D18 after 5 and 60 min, respectively (Figure S5, panel *III* and *VI*). These results clearly demonstrate that upon APE1 interaction with DNA, D18 is no longer accessible to the protease action. Thus, it may be hypothesized that this residue is placed at the protein-nCaRE oligonucleotide complex interface.

The different accessibility of the basic residues in the isolated APE1 protein and in its complex with nCaRE-B oligonucleotides was similarly probed with trypsin, used at 1:5000 w/w enzyme/substrate (Supplemental Table S4). LC-ESI-MS analysis of the proteolytic fragments, released from the isolated protein, led to the identification of the complementary peptide pairs 1-9, 10-321 and 1-10, 11-321 already after 5 min of reaction, which were generated by single hydrolytic events at K9 and K10, respectively. Additional products, resulting from fragment subdigestion reactions, were also observed, but they did not provide information on further accessible amino acids. These results confirmed that APE1 N-terminal portion is very flexible and highly exposed to the proteolytic action. Conversely, appreciable hydrolysis of recombinant APE1 following protein interaction with SIRT1 nCaRE-B or APE1-PTH nCaRE-B oligonucleotides was observed at K9 only after 60 min of reaction (Supplemental Table S4). Due to the nature of the proteolytic products observed, these experiments confirmed the masking effect of both oligonucleotides over the nonstructured protein N-terminal region.

Further experiments were then carried out with broader-specificity proteases, such as chymotrypsin and elastase. In the first case, limited proteolysis of isolated APE1 (enzyme/substrate of 1:1000 w/w) generated only a complementary peptide pair, namely 1-114 and 115-321, which identified L114 as the primary cleavage site (Supplemental Table S4). Additional products resulting from fragment subdigestions were also observed, but they did not provide information on further accessible amino acids. According to X-ray crystallographic data, this residue is exposed on the molecular surface of the globular APE1 domain (Gorman *et al.*, 1997). No significant differences were observed on a time-course basis when peptide maps were characterized for both APE1-nCaRE oligonucleotide complexes (Supplemental Table S4). In fact, both peptides 1-114 and 115-321 were

detected within the APE1-SIRT1 nCaRE-B and APE1-PTH nCaRE-B oligonucleotide products after 5 and 15 min, respectively, similarly to the non-complexed protein sample. These data excluded any involvement of L114 in binding to the nCaRE-B elements and further confirmed the suitability of the strategy used here.

When elastase digestion of isolated APE1 was carried out using an enzyme/substrate of 1:1000 w/w, only two peptide pairs (1-12, 13-321, and 1-14, 15-321) were detected after 5 min of reaction, whereas an additional pair (1-20, 21-321) was observed after 15 min (Supplemental Table S4). These data demonstrated that non-complexed APE1 was preferentially cleaved at A12, A14 and L20. When the recombinant APE1-SIRT1 nCaRE-B oligonucleotide and APE1-PTH nCaRE-B oligonucleotide complexes were analyzed under the same conditions, different proteolytic patterns were obtained, which proved preferential cleavage site at L20, as demonstrated by the identification of the complementary peptides 1-20 and 21-321 after 30 min of reaction (Supplemental Table S4). These results clearly demonstrated that, upon complex formation, A12 and A14 are no longer accessible to elastase, whereas L20 is still partially exposed to the protease action.

Supplemental Figure Legends

Supplemental Figure S1.

Schematic representation of the pipeline steps used for the identification of putative nCaRE-B elements within the human genome.

Supplementary Figure S2.

To test the binding affinity of APE1 for SIRT1 nCaRE sequence, a comparative analysis between the mutated nCaRE sequence and a polyT sequence was performed using SPR. Overlay of sensorgrams relative to the binding of APE1 to immobilized biotinylated ds oligonucleotide corresponding to polyT (A) and nCaRE SIRT1-B (B) and nCaRE SIRT1-B mutated sequence (C). APE1 concentration ranged within 1.0-20 μM for nCaREs and 1.0-10μM for polyT .

Supplemental Figure S3.

To demonstrate that Ku70 is able to stabilize the binding ability of APE1 to SIRT1-B nCaRE sequence we performed EMSA analysis with ds oligonucleotide corresponding to nCaRE SIRT1-B sequence with APE1^{WT} and Ku70 purified recombinant protein. Reactions were performed with 2.5 pmol of nCaRE SIRT1-B oligonucleotide and 10, 20 or 30 pmol of APE1^{WT} (lanes 2, 3, 4), Ku70 (lanes 5, 6, 7) and both APE1^{WT} and Ku70 (lanes 8, 9, 10). Lane 1 represents probe alone; F shows the position of the free oligonucleotide probe. Specific protein/nCaRE interaction is indicated by the arrow. Notably, Ku70 is unable per se to directly bind the SIRT-1 B sequence but significantly enhances APE1 DNA binding activity on this sequence.

Supplemental Figure S4.

To corroborate structural data obtained for SIRT1-B nCaRE-B sequence with T7 endonuclease we performed the same experiment with another enzyme, the mung bean nuclease (Amosava *et al*., 2011), a single-strand specific endonuclease, typically used for the removal of hairpin loops during cDNA synthesis. (A) EMSA analysis with APE1 on SIRT1 nCaRE-B after mung bean endonuclease digestion. 2.5 pmol of ds oligonucleotide corresponding to nCaRE SIRT1-B were digested with mung bean endonuclease at 30°C for the indicated period of time. The reactions then were incubated at 37°C with 10 pmol of recombinant APE1 protein for additional 15 min. Lanes 1 is probe alone; F shows the position of the free oligonucleotide probe. Specific APE1/nCaRE interaction is indicated by arrow. (B) APE1 and mung bean endonuclease compete for the same nCaRE binding site. EMSA analysis of APE1 binding to nCaRE sequence after digestion with mung bean endonuclease or after preincubation with APE1 and subsequent digestion with mung bean. 2.5 pmol of ds oligonucleotide corresponding to nCaRE SIRT1-B were first digested with mung bean endonuclease at 30°C for 4 h and subsequently incubated with increasing amounts of recombinant APE1 (10, 30 pmol) at 37°C for 15 min (lanes 7, 8). Alternatively, 2.5 pmol of SIRT1- B nCaRE probe were first incubated with APE1 at 37°C for 15 min and sequentially digested at 30°C for 4 h with mung bean endonuclease (lanes 9, 10). Lanes 1 is probe alone; $1st$ and $2nd$ indicate if APE1 incubation with the probe was performed temporally before $(1st)$ or after $(2nd)$ mung bean digestion; F shows the position of the free oligonucleotide probe. Specific APE1/nCaRE interaction is indicated by arrow.

Supplemental Figure S5.

Comparative limited proteolysis experiments on isolated or DNA-complexed APE1. Time-course analysis of simultaneous trials performed on non-complexed recombinant APE1 (І, ІV), recombinant APE1-SIRT1 nCaRE-B complex (ІІ, V) or recombinant APE1-PTH nCaRE complex (ІІІ, VІ) are shown. LC-ESI-MS profiles from samples taken at 5 (І-ІІІ) and 60 min (ІV-VІ) are reported. Identified peptides are reported on the corresponding chromatographic peaks; 1-321 denotes the intact protein. See Supplemental Table S4 for detailed information. Experiments were performed on a recombinant APE1 form bearing three additional amino acids at protein N-terminus with respect to the native counterpart.

Supplemental Figure S6.

To determine the optimal time for a detectable SIRT1 mRNA and protein accumulation upon 1 mM $H₂O₂$ treatment, we harvested cells at different time of release as indicated. (A) Data shown in the histogram represent the amount of SIRT1 mRNA normalized to the amount of GAPDH. Statistical tests were estimated by Student's t test. A p-value < 0.001 was considered as statistically significant (*). (B) The induction of SIRT1 protein levels were estimated in HeLa cells treated with 0.1 mM $H₂O₂$ for the indicated times. SIRT1 protein levels were measured on whole cell lysate (WCE) using a specific antibody; tubulin was used as a loading control. Histogram reports the densitometric quantification of Western blotting signals from at least three independent replicates. SIRT1 amounts are expressed as mean \pm SD of the signal, considering the untreated sample as reference. A *p*-value < 0.05 was considered as statistically significant (*). (C) To finally corroborate our model of SIRT1 activation after oxidative treatment, the p53 acetylation status of K382, a known target of SIRT1 deacetylation (Vaziri *et al*., 2001), has been estimated. Western blotting analysis of SIRT1 protein levels and of p53 acetylation in clones expressing $APE1^{WT}$, $APE1$ silenced cells APE1^{CL.3} and control clones APE1^{SCR-1} after 1 mM H_2O_2 treatment, for 1 h. Data shown in the histogram are normalized to the amount of tubulin and represent the densitometric quantification of SIRT1 protein level and of p53 acetylation. Data are expressed as average fold of induction relative to the untreated cells in at least three independent Western blotting analysis.

Supplemental Figure S7.

Recruitment of BER enzymes on hSIRT1 promoter after oxidative condition. HeLa cells treated with 1 mM H_2O_2 for 10, 15, 20, 40, 60 minutes, were analyzed by qChIP using specific antibodies recognizing 8-oxodG, OGG1 and APE1. The histogram represent the amount of hSIRT1 promoter sequence immunoprecipitated. Data were presented as percent of input and normalized to the quantity of DNA immunoprecipitated by α -tubulin (α -tub) and further normalized on the amount of immunoprecipitated protein.

Supplemental Figure S8.

Time-course interaction of RNA Polymerase II and Ku70 with APE1 after H_2O_2 treatment. HeLa cells were transfected with a plasmid expressing APE1^{WT} and challenged with 1 mM H_2O_2 , for different times (as reported). Immunoprecipitated material and whole cell lysate were separated onto SDS-PAGE, immunoblotted and analyzed for their Ku70 (*upper panel*) and RNA Polymerase II (*bottom panel*) content, as associated with APE1 after H_2O_2 stimulus. Ponceau S staining was used for loading control. Normalized co-immunoprecipitated amounts of Ku70 and RNA Polymerase II are indicated under each relative bar. Mean values of two independent experiments are reported for Ku70.

Supplemental Figure S9.

To further support the evidences of SIRT1 mRNA and protein induction upon oxidative stress $(H₂O₂)$, we also challenged the cells with another genotoxic agent, methyl methanesulfonate (MMS). (A) Analysis of SIRT1 mRNA level with Q-PCR in clones expressing $APE1^{WT}$ or APE1 silenced cells $APE1^{CL.3}$ after 0.5 mM MMS treatment for 8 h. Data shown in the histogram are normalized to the amount of GAPDH. The significance of sample average difference observed was estimated with Schaffè test. A p-value < 0.05 was considered as statistically significant (*). Below, Western blotting analysis on protein extract of clones used. Tubulin protein level was used to normalize sample. (B) HeLa cells were treated with 0.5 mM MMS for the indicated times. SIRT1 protein levels were measured on whole cell lysate (WCE) using a specific antibody; tubulin was

used as a loading control. Histogram reports the densitometric quantification of Western blotting signals from at least three independent replicates. SIRT1 amounts are expressed as mean \pm SD of the signal, considering the untreated sample as reference. A p -value < 0.05 was considered as statistically significant (*).

Supplemental Table Legends

Supplemental Table S1.

Significant GO terms associated to genes bearing nCaRE elements and present in expression profile data obtained from experiment in which APE1 was knocked down. Here we are reported only the most significant GO annotation terms obtained when performing the exact Fisher's test. The column titled "n_annotation" correspond to the total number of genes associated to that GO term in Gene Ontology database; instead, column "n_both" represents the amount of genes in our dataset associated to the same GO term. "Expect" column reports for all GO terms the number of genes expected by chance in a set made of the same number of dataset genes, but selected at random from database. The columns titled "C", "F" and "P" correspond to the three branches of the Gene Ontology: cellular component, molecular function and biological process respectively.

Supplemental Table S2.

The final 57 genes extracted from the application of Gene Ontology and phylogentic footprinting analysis that can be considered *bona fide* as bearing the candidate nCaRE sequences within their regulatory elements and potentially regulated by APE1.

Supplemental Table S3.

Complete data reporting all the significant functional enrichments obtained performing GeneCodis analysis. 'GO term ID' and 'Annotations' columns represent the Gene Ontology codes of annotations and the textual description of annotations, respectively. BP refers to 'biological process' category of Gene Ontology annotation. Third and fourth columns represent the number of genes in the input list and the reference list for a given annotation, respectively. P-values calculated using hypergeometric distribution and its correction using the stimulation-based approach are reported. The 'Genes' column identifies the set of genes in the input list showing a given annotation.

Supplemental Table S4.

Comparative limited proteolysis experiments on isolated or DNA-complexed APE1. Experiments were performed on a recombinant APE1 form bearing three additional amino acids at protein Nterminus with respect to the native counterpart. Time-course analysis of simultaneous trials performed on non-complexed recombinant APE1, recombinant APE1-SIRT1 nCaRE-B complex or recombinant APE1-PTH nCaRE complex are shown. Proteolytic enzyme, protein to protease ratio (w/w), sampling time, peptide, theoretical and experimental mass value, and primary cleavage site in the recombinant and the native protein (in parenthesis) are shown.

Flow-chart of the pipeline

Supplemental Figure S2

A B

Supplemental Figure S4

9

C

A

B

