

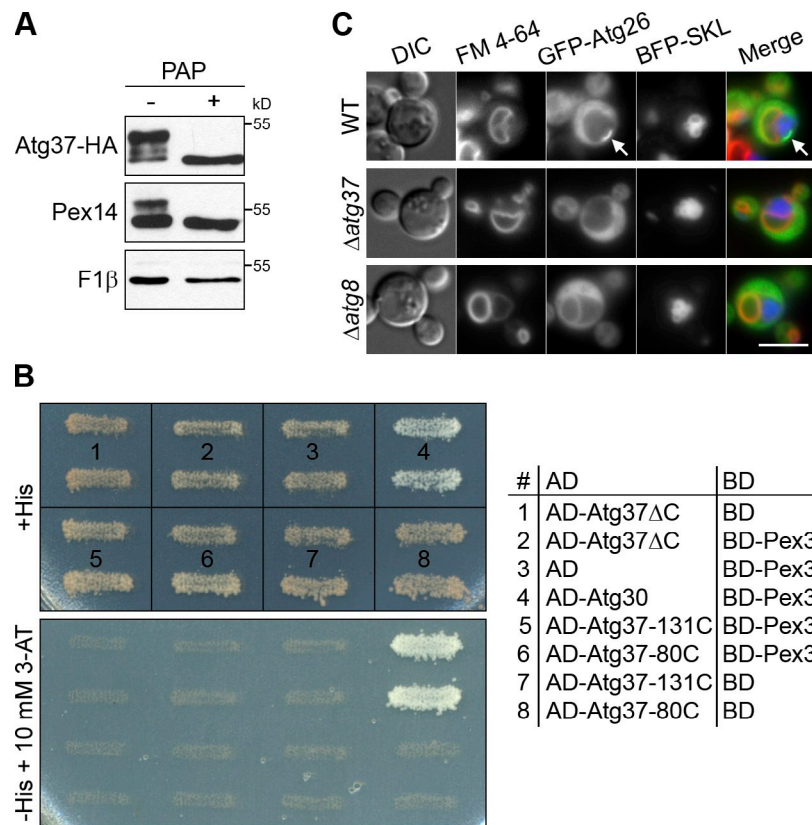
Nazarko et al., <http://www.jcb.org/cgi/content/full/jcb.201307050/DC1>

Figure S1. **Supplemental data on Atg37 modification, interaction, and the role in phagophore formation.** (A) Atg37 is a phosphoprotein. Cell lysates were incubated with (+) or without (-) PAP. (B) Atg37 does not interact with Pex3 in Y2H. Interaction was measured by the growth of cells on -His plates with 10 mM 3-AT. 3-AT, 3-amino-1,2,4-triazole; AD, activation domain; BD, binding domain. (C) Atg37 is essential for MIPA formation. Methanol-grown cells were adapted to glucose medium. Arrows point to the GFP-Atg26 labeled MIPA in the WT cell. DIC, differential interference contrast. Bar, 5  $\mu$ m.

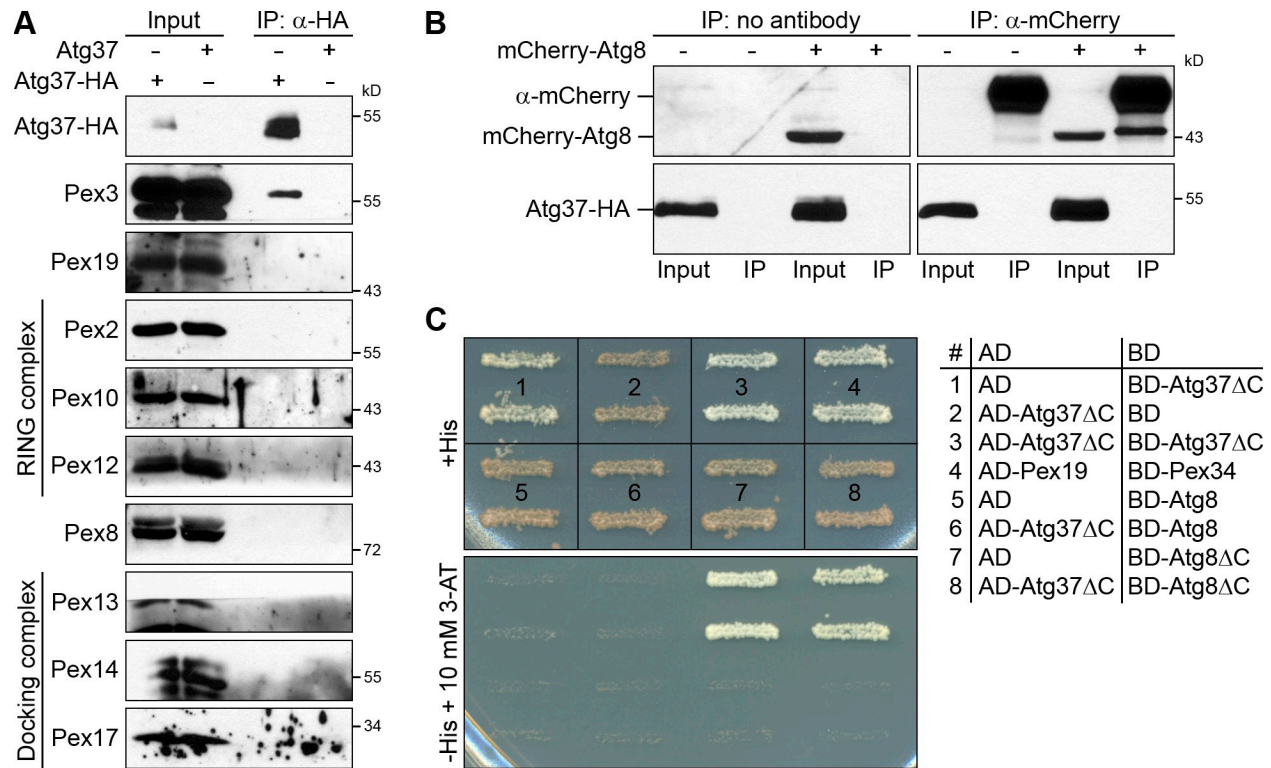


Figure S2. **Atg37 binds Pex3, but not Atg8, under pexophagy conditions.** (A) Pex3 coimmunoprecipitates with Atg37 under pexophagy conditions. Methanol-grown cells were adapted to glucose medium for 0.5 h. (B) Atg37 does not coimmunoprecipitate with Atg8. Methanol-grown cells were adapted to glucose medium for 0.5 h. The  $\alpha$ -mCherry band represents the heavy chain of the antibody. (C) Atg37 does not interact with Atg8 in Y2H. Atg37 $\Delta$ C self-interacted, but did not interact with either Atg8 or Atg8 $\Delta$ C (aa 1–115).

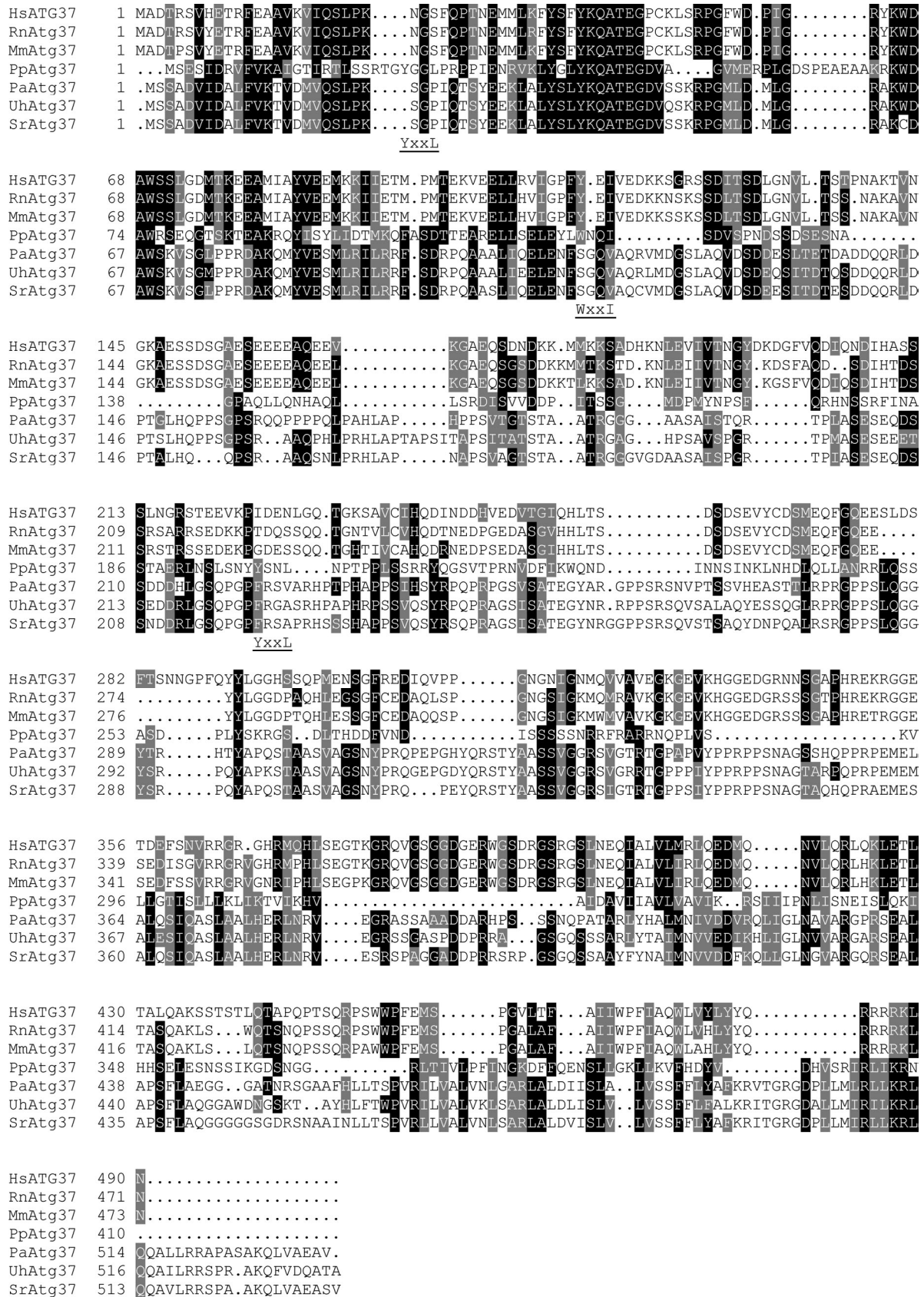


Figure S3. Multiple sequence alignment of Atg37 orthologues. Identical residues are indicated with black boxes and similar residues with gray boxes. HsATG37, *Homo sapiens* ACBD5 isoform 2 (NP\_001035938.1); RnAtg37, *Rattus norvegicus* Acbd5 (NP\_001071103.1); MmAtg37, *Mus musculus* Acbd5 (AAH61484.2); PpAtg37, *Komagataella* (formerly *Pichia*) *pastoris* GS115 Atg37 (CAY71862.1); PaAtg37, *Pseudozyma antarctica* T-34 Atg37 (GAC77071.1); UhAtg37, *Ustilago hordei* Atg37 (CCF51006.1); SrAtg37, *Sporisorium reilianum* SR22 Atg37 (CBQ71469.1). The potential AIMS of PpAtg37 are underlined below the aligned sequences.



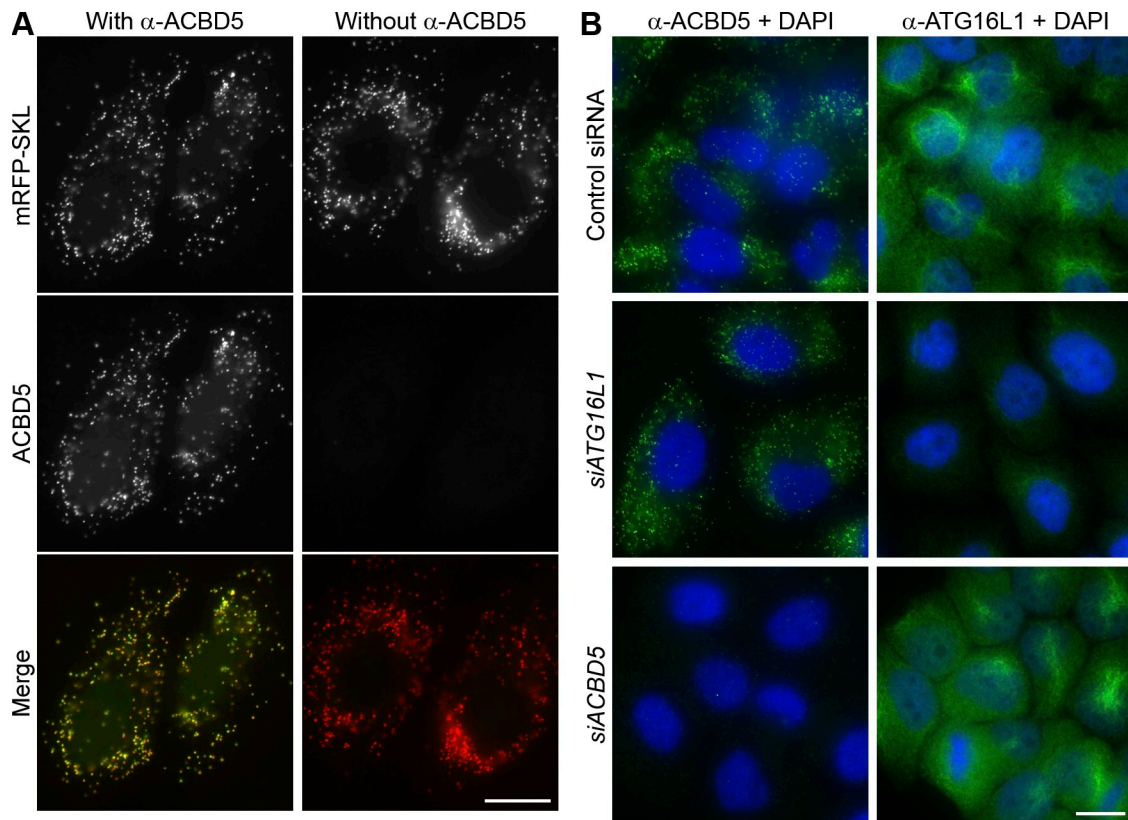


Figure S4. **ATG37/ACBD5 localizes to peroxisomes in human cells.** (A) ACBD5 is localized to peroxisomes in HeLa cells. Cells expressed peroxisomally targeted mRFP-SKL and were immunostained with antibody to ACBD5. Secondary antibody alone did not show any signal in the respective channel (right). Bar, 10  $\mu$ m. (B) siRNAs efficiently down-regulate ATG16L1 and ACBD5 in HeLa cells. Cells were transfected with nontargeting control siRNA or siRNA targeting ACBD5 or ATG16L1 for 3 d and immunostained with antibodies to ACBD5 (left) or ATG16L1 (right). DAPI stained nuclei. Bar, 10  $\mu$ m.

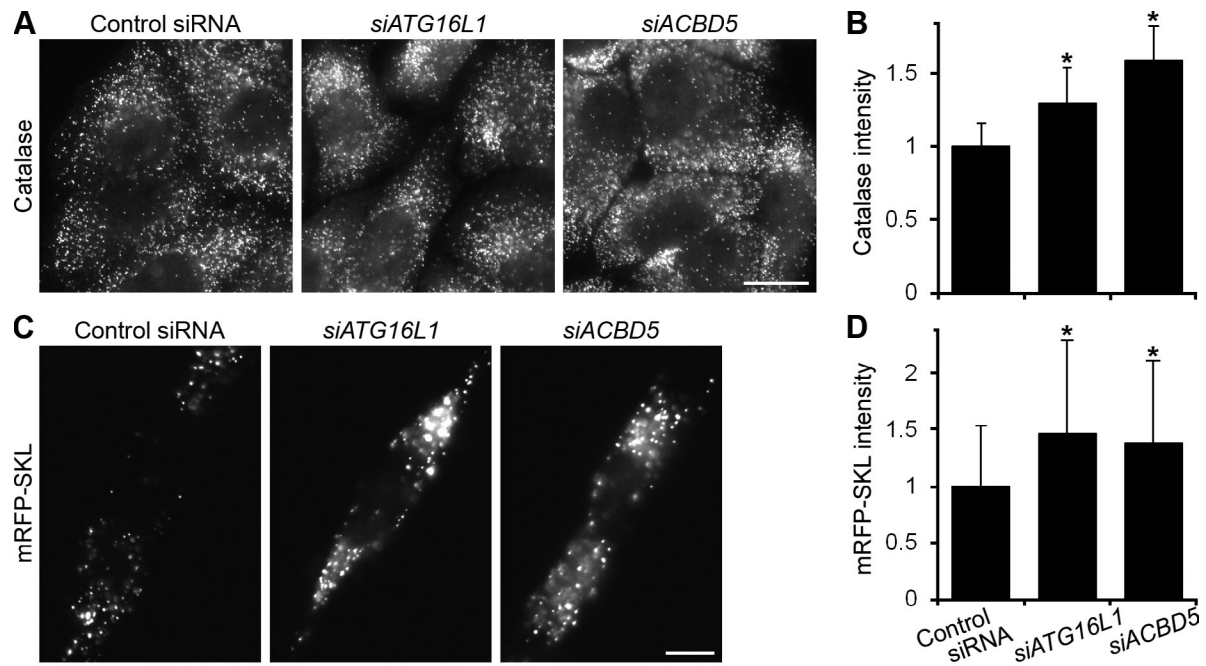


Figure S5. **Role of ATG37/ACBD5 in pexophagy.** (A) Knockdown of ACBD5 increases the peroxisome abundance in HeLa cells. Immunostaining with antibodies to peroxisomal catalase is shown. Bar, 10  $\mu$ m. (B) Quantitation of data in A. Catalase intensity of cells transfected with control siRNA was set as 1. Relative catalase intensity is shown as the mean + SD (error bars;  $n \geq 250$ ; \*,  $P < 0.05$  vs. control siRNA). (C) Knockdown of ACBD5 increases the peroxisome abundance in primary fibroblasts of a patient with a PBD caused by a missense mutation in the *PEX16* gene. Bar, 10  $\mu$ m. (D) Quantitation of data in C. mRFP-SKL intensity of cells transfected with control siRNA was set as 1. Relative mRFP-SKL intensity is shown as the mean + SD (error bars;  $n \geq 20$ ; \*,  $P < 0.05$  vs. control siRNA).

Table S1. *P. pastoris* strains and plasmids used in this study

Name	Description	Genotype and plasmid	Source
PDG2d	<i>Δatg28</i>	GS200 <i>Δatg28::ScARG4</i>	Stasyk et al., 2006
PPY12h	WT	<i>arg4 his4</i>	Gould et al., 1992
PPY12m	WT	<i>arg4 his4</i>	Gould et al., 1992
R8	<i>Δatg11</i>	GS115 <i>atg11-2::Zeocin<sup>R</sup></i>	Kim et al., 2001
SEW1	<i>Δpex3</i>	PPY12h <i>Δpex3::ARG4</i>	Wiemer et al., 1996
SKF13	<i>Δpex19</i>	PPY12h <i>Δpex19::Zeocin<sup>R</sup></i>	Snyder et al., 1999
SJCF51	WT	PPY12m <i>his4::pTW51 (P<sub>AOX1</sub>-GFP-SKL, HIS4)</i>	Nazarko et al., 2009
SJCF320	WT	SJCF247 <i>his4::pJCF208 (P<sub>ATG8</sub>-GFP-ATG8, HIS4)</i>	Farré et al., 2008
SJCF376	<i>Δatg30</i>	SJCF332 <i>his4::pJCF208 (P<sub>ATG8</sub>-GFP-ATG8, HIS4)</i>	Farré et al., 2008
SJCF547	WT	PPY12m <i>his4::pJCF208 (P<sub>ATG8</sub>-GFP-ATG8, HIS4)</i>	Farré et al., 2008
SJCF925	<i>Δatg8</i>	PPY12h <i>Δatg8::Geneticin<sup>R</sup> (pJCF182)</i>	Nazarko et al., 2009
SJCF936	<i>Δatg30</i>	PPY12h <i>Δatg30::Zeocin<sup>R</sup> (pJCF56)</i>	Nazarko et al., 2009
SJCF1821	<i>Δatg30</i>	SJCF959 <i>P<sub>AOX1</sub>::pPICz-BFP-SKL (P<sub>AOX1</sub>-BFP-SKL, Zeocin<sup>R</sup>)</i>	Farré et al., 2013
SMD1163	<i>pep4 prb1 his4</i>	<i>pep4 prb1 his4</i>	Tuttle and Dunn, 1995
SMY154	WT	PPY12h <i>PEX3::pMY126 (P<sub>PEX3</sub>-PEX3-TAP, Zeocin<sup>R</sup>)</i>	This paper
SMY431	<i>Δpmp47</i>	PPY12m <i>Δpmp47::Zeocin<sup>R</sup> (pMY243)</i>	This paper
SMY432	<i>Δatg37</i>	PPY12m <i>Δatg37::Zeocin<sup>R</sup> (pMY92)</i>	This paper
SMY433	WT	SMY432 <i>his4::pMY244 (P<sub>GAPDH</sub>-GFP-ATG37, HIS4)</i>	This paper
SMY437	<i>Δatg37</i>	SMY432 <i>his4::pTW51 (P<sub>AOX1</sub>-GFP-SKL, HIS4)</i>	This paper
SMY448	WT	PPY12m <i>his4::pMY244 (P<sub>GAPDH</sub>-GFP-ATG37, HIS4)</i>	This paper
SRRM197	<i>Δypt7</i>	PPY12h <i>Δypt7::Geneticin<sup>R</sup></i>	Manjithaya et al., 2010
STN29	<i>Δatg1</i>	R12 <i>his4::pTW51 (P<sub>AOX1</sub>-GFP-SKL, HIS4)</i>	Nazarko et al., 2007
STN48	<i>Δatg37</i>	SMY432 <i>P<sub>AOX1</sub>::pRDM054 (P<sub>AOX1</sub>-BFP-SKL, Hygromycin<sup>R</sup>)</i>	This paper
STN50	<i>Δatg37</i>	STN48 <i>his4::pJCF208 (P<sub>ATG8</sub>-GFP-ATG8, HIS4)</i>	This paper
STN52	<i>Δatg37</i>	SMY432 <i>his4::pJCF208 (P<sub>ATG8</sub>-GFP-ATG8, HIS4)</i>	This paper
STN66	<i>Δatg1</i>	R12 <i>his4::pJCF208 (P<sub>ATG8</sub>-GFP-ATG8, HIS4)</i>	Nazarko et al., 2009
STN95	<i>Δatg30 Δatg37</i>	SJCF936 <i>Δatg37::Geneticin<sup>R</sup> (pMY92Kan)</i>	This paper
STN96	<i>Δatg37</i>	PPY12h <i>Δatg37::Geneticin<sup>R</sup> (pMY92Kan)</i>	This paper
STN117	WT	PPY12h <i>his4::pJCF392 (P<sub>GAPDH</sub>-GFP-ATG26, HIS4)</i>	This paper
STN119	<i>Δatg8</i>	SJCF925 <i>his4::pJCF392 (P<sub>GAPDH</sub>-GFP-ATG26, HIS4)</i>	This paper
STN126	WT	SJCF936 <i>his4::pJCF213 (P<sub>ATG30</sub>-ATG30-GFP, HIS4)</i>	This paper
STN128	WT	STN96 <i>his4::pMY323 (P<sub>ATG37</sub>-ATG37-GFP, HIS4)</i>	This paper
STN130	WT	SJCF936 <i>his4::pJCF45 (P<sub>GAPDH</sub>-ATG30-GFP, HIS4)</i>	This paper
STN132	WT	STN96 <i>his4::pTYN4 (P<sub>GAPDH</sub>-ATG37-GFP, HIS4)</i>	This paper
STN139	WT	STN96 <i>his4::pMY244 (P<sub>GAPDH</sub>-GFP-ATG37, HIS4)</i>	This paper
STN148	<i>Δatg30</i>	STN95 <i>his4::pTYN4 (P<sub>GAPDH</sub>-ATG37-GFP, HIS4)</i>	This paper
STN149	<i>Δatg37</i>	STN95 <i>his4::pJCF45 (P<sub>GAPDH</sub>-ATG30-GFP, HIS4)</i>	This paper
STN155	<i>Δatg30</i>	STN95 <i>his4::pMY323 (P<sub>ATG37</sub>-ATG37-GFP, HIS4)</i>	This paper
STN161	WT	STN128 <i>P<sub>AOX1</sub>::pRDM054 (P<sub>AOX1</sub>-BFP-SKL, Hygromycin<sup>R</sup>)</i>	This paper
STN163	<i>Δatg30</i>	STN155 <i>P<sub>AOX1</sub>::pRDM054 (P<sub>AOX1</sub>-BFP-SKL, Hygromycin<sup>R</sup>)</i>	This paper
STN199	WT	STN126 <i>P<sub>AOX1</sub>::pRDM054 (P<sub>AOX1</sub>-BFP-SKL, Hygromycin<sup>R</sup>)</i>	This paper
STN201	<i>Δatg37</i>	STN95 <i>his4::pJCF213 (P<sub>ATG30</sub>-ATG30-GFP, HIS4)</i>	This paper
STN203	<i>Δatg37</i>	STN201 <i>P<sub>AOX1</sub>::pRDM054 (P<sub>AOX1</sub>-BFP-SKL, Hygromycin<sup>R</sup>)</i>	This paper
STN215	WT	STN96 <i>his4::pMY320 (P<sub>ATG37</sub>-ATG37-2xHA, HIS4)</i>	This paper
STN245	WT	PPY12h <i>ATG37::pMY328 (P<sub>ATG37</sub>-ATG37-GFP, ARG4)</i>	This paper
STN249	<i>Δpex19</i>	SKF13 <i>ATG37::pMY328 (P<sub>ATG37</sub>-ATG37-GFP, ARG4)</i>	This paper
STN255	WT	STN245 <i>his4::pKSN183 (P<sub>PEX3</sub>-PEX3-mRFP1, HIS4)</i>	This paper
STN260	<i>Δpex19</i>	STN249 <i>his4::pKSN183 (P<sub>PEX3</sub>-PEX3-mRFP1, HIS4)</i>	This paper
STN286	WT	STN288 <i>P<sub>AOX1</sub>::pRDM054 (P<sub>AOX1</sub>-BFP-SKL, Hygromycin<sup>R</sup>)</i>	This paper
STN288	WT	STN128 <i>arg4::pJCF477 (P<sub>ATG8</sub>-mCHERRY-ATG8, ARG4)</i>	This paper
STN292	<i>acbs</i>	STN96 <i>his4::pTYN21 (P<sub>ATG37</sub>-ATG37<sup>Y40E, K44A</sup>-2xHA, HIS4)</i>	This paper
STN294	<i>Atg37<sup>ΔTMD</sup></i>	STN96 <i>his4::pTYN24 (P<sub>ATG37</sub>-ATG37<sup>Δ290,339</sup>-GFP, HIS4)</i>	This paper
STN300	<i>acbs</i>	STN96 <i>his4::pTYN22 (P<sub>ATG37</sub>-ATG37<sup>Y40E, K44A</sup>-GFP, HIS4)</i>	This paper
STN352	<i>acbs</i>	STN300 <i>P<sub>AOX1</sub>::pRDM054 (P<sub>AOX1</sub>-BFP-SKL, Hygromycin<sup>R</sup>)</i>	This paper
STN356	<i>Atg37<sup>ΔTMD</sup></i>	STN294 <i>P<sub>AOX1</sub>::pRDM054 (P<sub>AOX1</sub>-BFP-SKL, Hygromycin<sup>R</sup>)</i>	This paper
STN363	WT	STN215 <i>arg4::pJCF477 (P<sub>ATG8</sub>-mCHERRY-ATG8, ARG4)</i>	This paper
STN556	WT	STN96 <i>his4::pKO8 (P<sub>ATG37</sub>-GFP-ATG37, HIS4)</i>	This paper
STN563	<i>Δatg30</i>	STN95 <i>his4::pMY320 (P<sub>ATG37</sub>-ATG37-2xHA, HIS4)</i>	This paper

Table S1. *P. pastoris* strains and plasmids used in this study (Continued)

Name	Description	Genotype and plasmid	Source
STN566	$\Delta atg30 \Delta ypt7$	SJCF1736 ATG37::pMY328 (P <sub>ATG37</sub> -ATG37-GFP, ARG4)	This paper
STN567	$\Delta ypt7$	SJCF1809 ATG37::pMY328 (P <sub>ATG37</sub> -ATG37-GFP, ARG4)	This paper
STN605	$\Delta atg37$	STN96 <i>his4</i> ::pJCF392 (P <sub>GAPDH</sub> -GFP-ATG26, HIS4)	This paper
STN612	WT	STN117 P <sub>AOX1</sub> ::pRDM054 (P <sub>AOX1</sub> -BFP-SKL, Hygromycin <sup>R</sup> )	This paper
STN614	$\Delta atg8$	STN119 P <sub>AOX1</sub> ::pRDM054 (P <sub>AOX1</sub> -BFP-SKL, Hygromycin <sup>R</sup> )	This paper
STN616	$\Delta atg37$	STN605 P <sub>AOX1</sub> ::pRDM054 (P <sub>AOX1</sub> -BFP-SKL, Hygromycin <sup>R</sup> )	This paper
STN618	WT	PPY12h P <sub>AOX1</sub> ::pPICz-BFP-SKL (P <sub>AOX1</sub> -BFP-SKL, Zeocin <sup>R</sup> )	This paper
STN620	$\Delta atg37$	STN96 P <sub>AOX1</sub> ::pPICz-BFP-SKL (P <sub>AOX1</sub> -BFP-SKL, Zeocin <sup>R</sup> )	This paper
STN622	$\Delta ypt7$	SRRM197 <i>arg4</i> ::pJCF340 P <sub>ATG11</sub> -2xFLAG-ATG11, ARG4)	This paper
STN630	$\Delta ypt7$	STN622 <i>his4</i> ::pJCF343 (P <sub>GAPDH</sub> -ATG30-TAP, HIS4)	This paper
STN632	$\Delta ypt7$	STN630 <i>his4</i> ::pTYN57 (P <sub>GAPDH</sub> -GFP-ATG17, Hygromycin <sup>R</sup> )	This paper
STN634	$\Delta ypt7$	STN622 <i>his4</i> ::pTYN57 (P <sub>GAPDH</sub> -GFP-ATG17, Hygromycin <sup>R</sup> )	This paper
STN636	WT	STN618 <i>arg4</i> ::pJCF291 P <sub>ATG11</sub> -GFP-ATG11, ARG4)	This paper
STN639	$\Delta atg37$	STN620 <i>arg4</i> ::pJCF291 P <sub>ATG11</sub> -GFP-ATG11, ARG4)	This paper
STN640	$\Delta atg30$	SJCF1821 <i>arg4</i> ::pJCF291 P <sub>ATG11</sub> -GFP-ATG11, ARG4)	This paper
STN641	$\Delta ypt7 \Delta atg37$	STN632 $\Delta atg37$ ::Zeocin <sup>R</sup> (pMY92) colony #2	This paper
STN642	$\Delta ypt7 \Delta atg37$	STN632 $\Delta atg37$ ::Zeocin <sup>R</sup> (pMY92) colony #3	This paper

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