

Quantum and All-Atom Molecular Dynamics
Simulations of Protonation and Divalent Ion
Binding to Phosphatidylinositol 4,5-
Bisphosphate (PIP₂)

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Supporting Information

Using the Gaussian '09 program, the molecular geometry of PIP₂ was obtained using the Hartree-Fock scheme in conjunction with the 6-31+G(d) basis set. The initial structure contained a proton placed equidistant between oxygen atoms from the 4-phosphate and 5-phosphate groups. The final Z-matrix is as follows:

```
c
c 1 cc2
c 2 cc3          1 ccc3
c 3 cc4          2 ccc4          1 dih4
c 4 cc5          3 ccc5          2 dih5
c 5 cc6          4 ccc6          3 dih6
o 2 oc7          3 occ7          4 dih7
p 7 po8          2 poc8          3 dih8
o 8 op9          7 opo9          2 dih9
c 9 co10         8 cop10         7 dih10
c 10 cc11        9 cco11         8 dih11
c 11 cc12        10 ccc12        9 dih12
o 12 oc13        11 occ13        10 dih13
c 13 co14        12 coc14        11 dih14
o 14 oc15        13 oco15        12 dih15
o 3 oc16         2 occ16         7 dih16
o 4 oc17         3 occ17         2 dih17
o 5 oc18         4 occ18         3 dih18
p 18 po19        5 poc19         4 dih19
o 19 op20        18 opo20        5 dih20
o 6 oc21         5 occ21        18 dih21
p 21 po22        6 poc22         5 dih22
o 22 op23        21 opo23        6 dih23
o 1 oc24         2 occ24         7 dih24
o 8 op25         7 opo25        2 dih25
o 8 op26         7 opo26        2 dih26
o 19 op27        18 opo27        5 dih27
o 19 op28        18 opo28        5 dih28
o 22 op29        21 opo29        6 dih29
o 22 op30        21 opo30        6 dih30
o 11 oc31        10 occ31        9 dih31
c 31 co32        11 coc32        10 dih32
c 32 cc33        31 cco33        11 dih33
o 32 oc34        31 oco34        11 dih34
c 14 cc35        13 cco35        12 dih35
h 16 ho36        3 hoc36         2 dih36
h 17 ho37        4 hoc37         3 dih37
h 24 ho38        1 hoc38         2 dih38
h 2 hc39         7 hco39         8 dih39
h 3 hc40         2 hcc40         7 dih40
h 4 hc41         3 hcc41         2 dih41
h 5 hc42         18 hco42        19 dih42
h 6 hc43         21 hco43        22 dih43
h 1 hc44         2 hcc44         7 dih44
h 33 hc45        32 hcc45        31 dih45
h 35 hc46        14 hcc46        13 dih46
h 29 ho47        22 hop47        21 dih47
h 11 hc48        31 hco48        32 dih48
h 33 hc49        32 hcc49        31 dih49
h 33 hc50        32 hcc50        31 dih50
h 35 hc51        14 hcc51        13 dih51
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h	35	hc52	14	hcc52	13	dih52
h	10	hc53	11	hcc53	31	dih53
h	10	hc54	11	hcc54	31	dih54
h	12	hc55	11	hcc55	31	dih55
h	12	hc56	11	hcc56	31	dih56

cc2	1.526344
cc3	1.525584
ccc3	110.464
cc4	1.534422
ccc4	107.977
dih4	-63.226
cc5	1.536272
ccc5	109.996
dih5	65.784
cc6	1.550422
ccc6	112.130
dih6	-54.297
oc7	1.430472
occ7	114.252
dih7	178.785
po8	1.609872
poc8	129.570
dih8	-76.014
op9	1.654558
opo9	100.649
dih9	89.510
co10	1.390493
cop10	118.339
dih10	56.942
cc11	1.525172
cco11	108.169
dih11	-175.973
cc12	1.516607
ccc12	112.077
dih12	53.050
oc13	1.444620
occ13	109.903
dih13	169.421
co14	1.307801
coc14	121.324
dih14	88.633
oc15	1.194599
oco15	126.101
dih15	1.910
oc16	1.415805
occ16	111.903
dih16	-54.700
oc17	1.396342
occ17	108.994
dih17	-171.724
oc18	1.397510
occ18	116.117
dih18	178.192
po19	1.724291
poc19	127.832
dih19	30.224
op20	1.508970
opo20	104.741
dih20	50.797
oc21	1.424589
occ21	112.141
dih21	-64.455
po22	1.622718
poc22	126.247
dih22	-43.036
op23	1.496990
opo23	109.001
dih23	-30.862
oc24	1.418164
occ24	109.016

dih24	-63.339
op25	1.484818
opo25	106.531
dih25	-159.329
op26	1.474582
opo26	112.354
dih26	-22.804
op27	1.504387
opo27	102.301
dih27	173.149
op28	1.529475
opo28	102.611
dih28	-68.029
op29	1.621747
opo29	100.814
dih29	84.774
op30	1.485743
opo30	108.720
dih30	-162.053
oc31	1.445320
occ31	104.664
dih31	170.387
co32	1.300480
coc32	123.522
dih32	142.520
cc33	1.510033
cco33	110.400
dih33	176.609
oc34	1.200221
oco34	126.449
dih34	-3.727
cc35	1.510545
cco35	110.753
dih35	-177.884
ho36	0.955083
hoc36	111.918
dih36	-68.963
ho37	0.977482
hoc37	105.034
dih37	170.673
ho38	0.948667
hoc38	105.953
dih38	52.346
hc39	1.082596
hco39	108.137
dih39	45.153
hc40	1.082127
hcc40	109.459
dih40	61.497
hc41	1.086439
hcc41	107.608
dih41	-53.191
hc42	1.080006
hco42	104.771
dih42	145.460
hc43	1.080634
hco43	105.331
dih43	-159.036
hc44	1.080878
hcc44	107.794
dih44	54.135
hc45	1.083874
hcc45	109.720
dih45	56.197
hc46	1.083719
hcc46	110.223
dih46	-49.020
ho47	0.961524
hop47	112.864
dih47	-26.260
hc48	1.079439

hco48	108.416
dih48	22.809
hc49	1.084019
hcc49	109.203
dih49	-61.054
hc50	1.082332
hcc50	110.002
dih50	177.802
hc51	1.082120
hcc51	109.751
dih51	-170.839
hc52	1.084700
hcc52	108.552
dih52	69.339
hc53	1.086291
hcc53	109.176
dih53	-68.761
hc54	1.081245
hcc54	108.631
dih54	49.990
hc55	1.075921
hcc55	110.080
dih55	175.558
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hcc56	111.112
dih56	-62.016

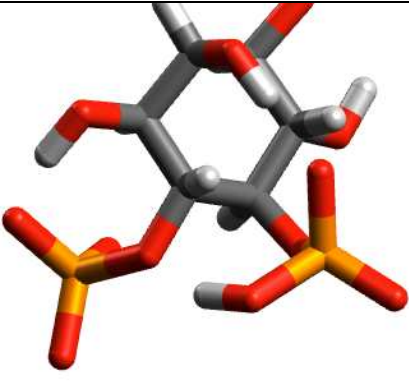
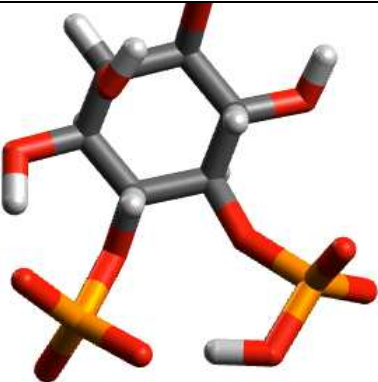
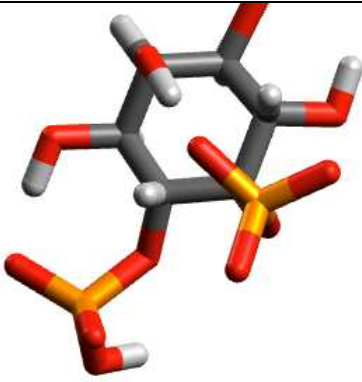
Proton on 5-phosphate	Proton shared between 4 and 5	Proton on 4-phosphate
		
+15.4 kcal/mol	0 kcal/mol	+22.0 kcal/mol

Figure S1. Relative energies of protonation isomers. The relative stabilities of protonated PIP₂ isomers were computed with Gaussian 09 at the HF/3-21G level of theory with implicit water solvent using a Polarizable Continuum Model¹⁻³. Relaxed dihedral potential energy scans were used to identify energetic minima among isomers. In the lowest energy structure (middle panel), the proton is shared between the 4 and 5-phosphates. The distances from the proton to the 5 and 4-phosphate oxygens are 1.11 Å and 1.30 Å, respectively. This shared state is 15.4 kcal/mol and 22.0 kcal/mol more stable than localization to the 5-phosphate (left panel) or the 4-phosphate (right panel), respectively.

Our results are obtained from all-atom molecular dynamics simulations of membrane bilayers of 800 total lipids containing PIP₂ in the presence of phosphatidylcholine (PC), phosphatidylserine (PS), phosphatidylethanolamine (PE), and cholesterol with 150 mM NaCl in a solvent box containing approximately 80,000 waters. The outer leaflet contains 300 POPC [3-palmitoyl-2-oleoyl-D-glycero-1-phosphatidylcholine] molecules and 100 cholesterol molecules. The inner leaflet contains 200 DOPE [2,3-dioleoyl-D-glycero-1-phosphatidylethanolamine] molecules, 100 cholesterol molecules, 60 DOPS [2,3-dioleoyl-D-glycero-1-phosphatidylserine] molecules, 40 SA-PIP₂ [1-stearoyl-2-arachadonyl-*sn*-glycero-1-phosphatidylinositol-4,5-bisphosphate] molecules. The lipids were packed at approximately 65 Å² per lipid and randomly seeded with PIP₂ typically about 10% of the total lipids on one leaflet. The system was equilibrated for 10 ns before the production run, for which the analysis is presented below. We used the CHARMM36 lipid forcefield in the GROMACS simulation package⁴⁻⁷ to prepare membrane bilayers in an NPT ensemble and have simulated for 31 ns so far, using a 2 fs timestep. Pressure is maintained using the Parrinello-Rahman barostat^{8,9}, coupled independently (semi-isotropic) in directions normal and perpendicular to the bilayer with a time constant of 2.0 ps and a pressure of 1 bar. Electrostatics are treated with the particle mesh Ewald (PME)¹⁰ method with cubic periodic boundaries, using a real space cutoff of 1.2 nm and a Fourier grid spacing of 0.16 nm with cubic interpolation. Temperature coupling is accomplished using the thermostat due to Bussi¹¹, in which velocity is rescaled with a stochastic term. These simulations employ the CHARMM-specific TIP3P (i.e., TIPS3P) water model (with van der Waals interactions on the water hydrogen atoms) and van der Waals interactions which are smoothly switched off between 0.8 and 1.2 nm¹².

Histograms were computed (Figure S2) for the QM/MM simulations presented in the main text (a single PIP₂ molecule in a water sphere) and for each lipid every 20 frames of the classical all-atom trajectory. The head-tail angle was measured as described in the main text. Our QM/MM and classical bilayer simulation results quantitatively agree with each other, but disagree if we employ the parameters of Lupyan, et al.

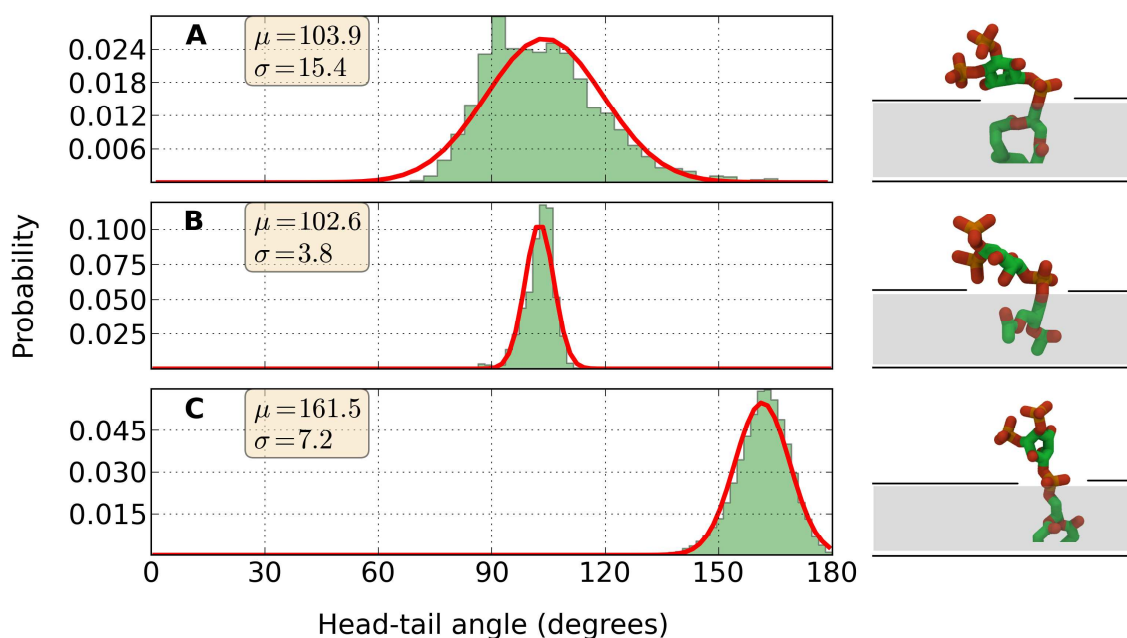


Figure S2. **A.** The probability distribution for 80 PIP₂ lipids in one leaflet of a bilayer also containing phosphatidylserine, phosphatidylethanolamine, and cholesterol. The histogram was averaged over each lipid and every 20 frames of the all-atom simulation for 15 ns using CHARMM36 lipid parameters. The mean head-tail angle is 104 degrees. **B.** The probability distribution of the angle between the PIP₂ headgroup and the PIP₂ acyl chains as measured from an isolated PIP₂ molecule in a water sphere over 10 ps of a QM/MM simulation. Sodium ions were present for charge neutrality but fixed to be ~ 10 Å away from the PIP₂. The mean head-tail angle is 103 degrees. **C.** The probability distribution from a simulation of a bilayer containing pure PIP₂ using the same system parameters as panel A, but with the parameters of Lupyan, et al. used instead of the CHARMM36 lipid parameters¹³. The average head-tail angle is 162 degrees.

We computed the average PIP₂ molecular area in membrane bilayer systems to compare with the QM/MM results presented in the main text. Since there was only a single molecule in the QM/MM simulations, we used the separation of the phosphate groups as a proxy for molecular area. To calculate the molecular area in the bilayer simulations, we projected the center of gravity of each lipid onto a plane and created a Delaunay triangulation. We summed the area created by forming a polygon from the bisector of each line meeting a vertex corresponding to PIP₂. There are subtleties in defining the average molecular area of lipids in a heterogeneous bilayer simulation and these methods are still evolving.

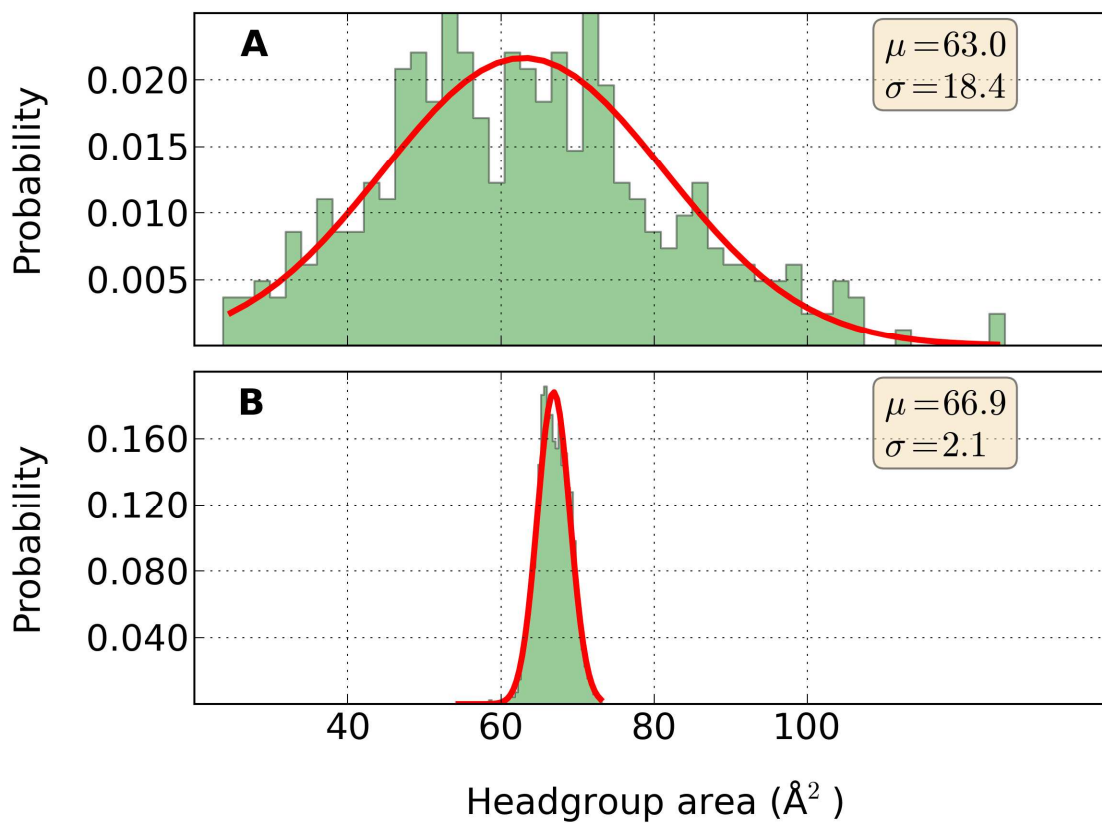


Figure S3. **A.** The probability distribution for 80 PIP₂ lipids in a bilayer also containing phosphatidylcholine, phosphatidylserine, and phosphatidylethanolamine. **B.** The probability distribution of the approximate molecular area of PIP₂ as measured from an isolated PIP₂ molecule in a water sphere over 10 ps of a QM/MM simulation. Sodium were present for charge neutrality but fixed to be ~10 Å away from the PIP₂.

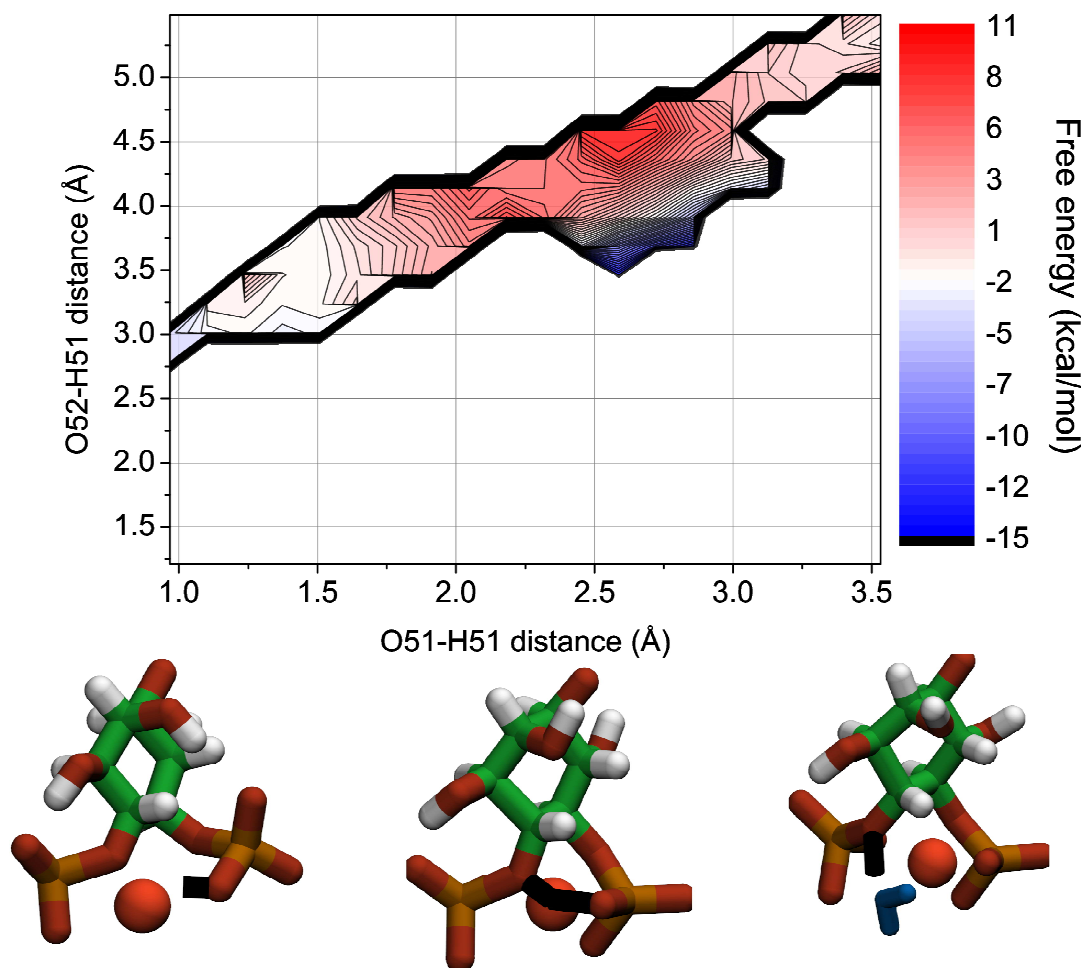


Figure S4. The free energy landscape for dissociating a proton from the 5-phosphate group of PIP₂. Full deprotonation in the presence of magnesium has not been achieved. Insets show magnesium (in red) between the two phosphomonoester groups of PIP₂ and the proton to be dissociated in black. The initial state of the QM/MM 2D umbrella sampling is on the left, with proton H51 bound to oxygen O51. The inset in the middle corresponds to the energy well on the contour plot (blue), where proton H51 is shared between oxygen O51 and oxygen O4P. The inset on the right corresponds to the final window of the trajectory, where proton H51 is shared between oxygen O4P of PIP₂ and a QM water (in blue).

It is possible to calculate pKa values from the free energy plots presented in the main text. To do this, we consider the canonical definition of the association constant, K_a , between a ligand, L, and its binding partner, R, in terms of the ratio of the probabilities of being not bound or bound. In this case, we take the ligand to be a proton and its binding partner to be a PIP₂ molecule. So we write,

$$K_a = \frac{1}{[L]} \times \frac{\mathcal{P}_{\text{bound}}}{\mathcal{P}_{\text{unbound}}}$$

where $\mathcal{P}_{\text{bound}}$ and $\mathcal{P}_{\text{unbound}}$ can be determined by integrating over the degrees of freedom and the potentials of mean force. pKa is related to K using the following relationship:

$$\text{pKa} = \log_{10} \left| K_a \times \frac{1}{1661 \text{ \AA}^3} \right| \text{ using the standard concentration of } 1 \text{ mol/liter} = 1/1661 \text{ \AA}^3.$$

We can perform the pKa calculation in two ways. In the first method, we integrate the bound state until $r = r^*$ which is the point where the proton leaves the domain of influence of the oxygen to which it was originally bound. This is judged by viewing the simulation trajectories and looking at the minima of the potentials of mean force. The unbound state is integrated from $r = r^*$ until the final point in the energy landscape.

In the second method, we integrate the bound state until $r = r^*$ as before. To evaluate the unbound state, we integrate from $r = r^*$ until the next metastable minimum at $r = r^{**}$. In this technique, events that we categorize as dissociation, such as proton transfer to other atoms on PIP₂, may not correspond to dissociation events judged from experiments.

Values for r^* and r^{**} and the results of these calculations are shown in Figure S5. Error

bars in the pKa calculation represent the outcome of changing the value of r^* by 0.25 Å or approximately the width of a well in the potential of mean force.

We stress that the absolute value of these numbers should not be compared to the results of experiments due to the approximations necessary in obtaining absolute free energy curves from the potentials of mean force calculated along a single reaction coordinate. Kooijman et al. calculate the pKa of PIP₂ in lipid vesicles, which is different from our case in several respects (e.g., the effect of a negative surface charge density of the vesicle membrane) so a direct comparison is not strictly valid¹⁴. Moreover, the pKa values reported by Kooijman et al. correspond to PI(3,5)P₂ and PI(3,4,5)P₂ which could have different ionization properties due to the positioning of the phosphate groups. In fact, for PI(4,5)P₂, Kooijman et al. note that a biphasic pH-dependent ionization behavior that cannot be explained by a Henderson-Hasselbach equation was obtained. Our calculations are consistent with this; our free energy landscape does not simply represent two states (bound or unbound) but displays intermediate metastable states. These states correspond to sharing of the proton between the vicinal phosphomonoester groups. Hence, our pKa should be regarded as an apparent pKa for proton transfer from the host oxygen (O51 in the main text) to the next energetically favorable state (rather than a free unbound state).

The subtleties caused by a complex (rugged) free energy landscape and non-independence of proton binding to vicinal sites cause problems in defining an absolute pKa; moreover, the apparent pKa is subject to the definitions of vicinal binding sites as metastable states (or our choice of r^{**}). Another complexity is a computational one. In the QM/MM potentials of mean force, the energy landscape is only accurate for short

excursions from the initial bound state along the reaction coordinate. For longer excursions, the wave functions from the QM region are spread over larger volumes causing an imbalance in basis set superposition. Also for these large distances along the reaction coordinate, close interaction between classical charges and the QM wave function become significant.

In general, we find that the pKa for protonation of PIP₂ in the presence of calcium is the smallest, implying strong dissociation of the proton. The pKa in the presence of magnesium is similar to that of a pure water environment, where we find protonation of PIP₂ on the 5-phosphate group to be the most stable conformation.

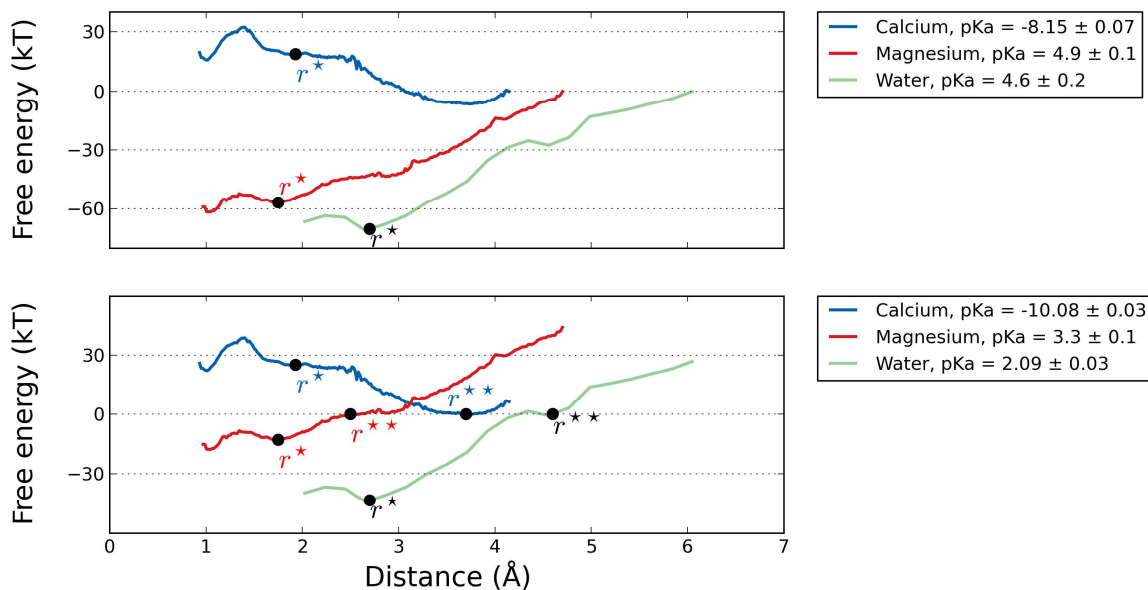


Figure S5. The potentials of mean force for proton dissociation from PIP₂ in a water sphere with either calcium, magnesium, or pure water. In the latter case, sodium ions are present for charge neutrality but confined ~ 10 Å away from the PIP₂.

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