

Table S2. **List of strains used in this study.**

strain	genotype	reference
BR-F	MATa/MAT α	(Kuthan et al., 2003)
BR-S	MATa/MAT α	(Kuthan et al., 2003)
BR-F-RF	MATa/MAT α	this study
BR-F-Flo11p-GFP	MATa/MAT α <i>FLO11/FLO11-EGFP</i>	this study
BR-F-RF-Flo11p-GFP	MATa/MAT α <i>FLO11/FLO11-EGFP</i>	this study
BR-F-RF-p _{GAL1} -GFP	MATa/MAT α <i>HIS3/his3Δ::nat1-p_{GAL1}-GFP</i>	this study
BR-F- <i>sir2</i>	MATa/MAT α <i>sir2Δ::kanMX/sir2Δ::nat1</i>	this study
BR-S- <i>hda1</i>	MATa/MAT α <i>hda1Δ::kanMX/hda1Δ::nat1</i>	this study

Table S3. List of primers and plasmids used in the strain construction.

primer	sequence	purpose
SIR2-del-for	AGACACATTCAAACCATTTTTCCCTCATCGGCACATT AAAGCTGGCAGCTGAAGCTTCGTACGC	deletion of <i>SIR2</i> gene
SIR2-del-rev	ATTGATATTAATTTGGCACTTTTAAATTATTAATTGC CTTCTACGCATAGGCCACTAGTGGATCTG	deletion of <i>SIR2</i> gene
HDA1-del-for	GAGAAAGGGAAAGTTGAGCACTGTAATACGCCGAAC AGATTAAGCCAGCTGAAGCTTCGTACGC	deletion of <i>HDA1</i> gene
HDA1-del-rev	TTCTTCATCACTCCATTCTTCAAACGAATCCAGTATA AAGTCTGTGCATAGGCCACTAGTGGATCTG	deletion of <i>HDA1</i> gene
FLO11-eGFP-for	GCTTATTTGGTCCTTTTCGCTTCTATTTAACTCGGCTT TGGGTTTTTGCAGGTCGACAACCCTTAAT	internal fusion of GFP with <i>FLO11</i> gene
FLO11-eGFP-rev	ACAGCTAGTTCCTTCGGAGGATCCTCTTGGAAGTAG TGCAGTTGGGCGGCCGCATAGGCCACT	internal fusion of GFP with <i>FLO11</i> gene
P_{GAL1}-eGFP-for	CTTCGAAGAATATACTAAAAAATGAGCAGGCAAGATA AACGAAGGCAAAGATGCGTACGCTGCAGGTCGAC	P _{GAL1} -GFP cassette for genomic integration
P_{GAL1}-eGFP-rev	TATACACATGTATATATATCGTATGCTGCAGCTTTAA ATAATCGGTGTCACACTACGATGAATTCTCTGTCTG	P _{GAL1} -GFP cassette for genomic integration

plasmid	application	reference
pAG25	deletion cassette (nat marker)	(Goldstein and McCusker, 1999)
pUG6	deletion cassette (kanMX marker)	(Guedener et al., 2002)
pYM-N25	P _{GAL1} -GFP cassette	(Janke et al., 2004)
pOM40	internal GFP fusion	(Gauss et al., 2005)
pSH65	marker excision	(Gauss et al., 2005)

Table S4. List of primers used in qPCR.

Primer	ORF	Gene	Sequence	Tm	% GC	nt
MAM3Fd	YOL060C	<i>MAM3</i>	tctagggatggtgccgattc	64.3	50	20
MAM3Rv	YOL060C	<i>MAM3</i>	tacaccaccagcagcacta	64.8	55	20
SHR5Fd	YOL110W	<i>SHR5</i>	cgtctcgctctgtcagaat	65	55	20
SHR5Rv	YOL110W	<i>SHR5</i>	ccgaacacctcaccttcttc	64.1	55	20
TOA1Fd	YOR194C	<i>TOA1</i>	acaaaggcgagatggaaatg	64.3	45	20
TOA1Rv	YOR194C	<i>TOA1</i>	tatacccactccgcttcac	64.2	55	20
CLP1Fd	YOR250C	<i>CLP1</i>	gtcaaggtaaactcgggcat	63.2	50	20
CLP1Rv	YOR250C	<i>CLP1</i>	tcgacagcgtatatgggaaa	63.4	45	20
MPD1Fd	YOR288C	<i>MPD1</i>	agcttcgataaagcgatcca	64	45	20
MPD1Rv	YOR288C	<i>MPD1</i>	gccttgcggaacgtactaga	64.5	55	20

Fd: forward primer, Rv: reverse primer, Tm: melting temperature, nt: nucleotides

Supplemental references

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Janke, C., Magiera, M.M., Rathfelder, N., Taxis, C., Reber, S., Maekawa, H. et al. (2004) A versatile toolbox for PCR-based tagging of yeast genes: new fluorescent proteins, more markers and promoter substitution cassettes. *Yeast* **21**: 947-962.

Kuthan, M., Devaux, F., Janderová, B., Slaninová, I., Jacq, C., and Palková, Z. (2003) Domestication of wild *Saccharomyces cerevisiae* is accompanied by changes in gene expression and colony morphology. *Mol Microbiol* **47**: 745-754.