

***Bacillus subtilis* RecA and its accessory factors, RecF, RecO, RecR and RecX, are required for spore resistance to DNA double strand break**

Ignacija Vlašić^{1,2}, Ramona Mertens¹, Elena M. Seco³, Begoña Carrasco³, Silvia Ayora³, Günther Reitz¹, Fabian M. Commichau⁴, Juan C. Alonso^{3,*}, and Ralf Moeller^{1,*}

German Aerospace Center, Institute of Aerospace Medicine, Radiation Biology Department, Cologne (Köln), Germany¹, Ruđer Bošković Institute, Division of Molecular Biology, Laboratory of Evolutionary Genetics, Zagreb, Croatia², Centro Nacional de Biotecnología, CSIC, Department of Microbial Biotechnology, Madrid, Spain³, University of Göttingen, Department of General Microbiology, Göttingen, Germany⁴

Supplementary Table S1 *Bacillus subtilis* strains used in this study

Strain	Relevant genotype	Source or reference ^a
168	<i>trpC2</i>	Laboratory stock
WN463 ^a	+ <i>recA</i>	(1)
LAS600 ^b	Δupp	(2)
LAS523 ^b	+ <i>dinR3</i> [<i>lexA</i> (Ind ⁻)]	(2)
BG214 ^c	<i>trpCE metA5 amyE1 ytsJ1 rsbV37 xre1 xkdA1 att</i> ^{SPB} <i>att</i> ^{ICEBsI}	Laboratory stock
BG809 ^c	+ $\Delta ykoV$	(3)
BG849 ^c	+ $\Delta ykoV \Delta recA$	(3)
BG190 ^c	+ $\Delta recA$	(4)
BG277 ^c	+ $\Delta recN$	(5)
BG125 ^c	+ <i>addA5</i>	(6)
BG189 ^c	+ <i>addA5 addB72</i>	(5)
BG705 ^c	+ $\Delta recQ$	(7)
BG425 ^c	+ $\Delta recS$	(8)
BG713 ^c	+ <i>addA5 \Delta recJ</i>	(7)
BG675 ^c	+ $\Delta recJ$	(7)
BG129 ^c	+ <i>recF15</i>	(6)
BG439 ^c	+ $\Delta recO$	(9)
BG128 ^c	+ $\Delta recR$	(10)
BG1065 ^c	+ $\Delta recX$	(11)
BG1053 ^c	+ $\Delta recX recF15$	(11)

^aStrains isogenic with 168 (DSMZ, DMS402). ^bStrains derived from prototrophic 168 bearing a null *upp* mutation. ^cStrains isogenic with BG214 (a 168 derivative).

References

1. Moeller, R., Stackebrandt, E., Reitz, G., Berger, T., Rettberg, P., Doherty, A.J., Horneck, G. and Nicholson, W.L. (2007) Role of DNA repair by nonhomologous-end joining in *Bacillus subtilis* spore resistance to extreme dryness, mono- and polychromatic UV, and ionizing radiation. *J Bacteriol*, **189**, 3306-3311.
2. Simmons, L.A., Goranov, A.I., Kobayashi, H., Davies, B.W., Yuan, D.S., Grossman, A.D. and Walker, G.C. (2009) Comparison of responses to double-strand breaks between *Escherichia coli* and *Bacillus subtilis* reveals different requirements for SOS induction. *J Bacteriol*, **191**, 1152-1161.

3. Mascarenhas, J., Sanchez, H., Tadesse, S., Kidane, D., Krishnamurthy, M., Alonso, J.C. and Graumann, P.L. (2006) *Bacillus subtilis* SbcC protein plays an important role in DNA inter-strand cross-link repair. *BMC Mol Biol*, **7**, 20.
4. Ceglowski, P., Luder, G. and Alonso, J.C. (1990) Genetic analysis of *recE* activities in *Bacillus subtilis*. *Mol Gen Genet*, **222**, 441-445.
5. Alonso, J.C., Stiege, A.C. and Luder, G. (1993) Genetic recombination in *Bacillus subtilis* 168: effect of *recN*, *recF*, *recH* and *addAB* mutations on DNA repair and recombination. *Mol Gen Genet*, **239**, 129-136.
6. Alonso, J.C., Tailor, R.H. and Luder, G. (1988) Characterization of recombination-deficient mutants of *Bacillus subtilis*. *J Bacteriol*, **170**, 3001-3007.
7. Sanchez, H., Kidane, D., Cozar, M.C., Graumann, P.L. and Alonso, J.C. (2006) Recruitment of *Bacillus subtilis* RecN to DNA double-strand breaks in the absence of DNA end processing. *J Bacteriol*, **188**, 353-360.
8. Fernández, S., Sorokin, A. and Alonso, J.C. (1998) Genetic recombination in *Bacillus subtilis* 168: effects of *recU* and *recS* mutations on DNA repair and homologous recombination. *J Bacteriol*, **180**, 3405-3409.
9. Fernandez, S., Kobayashi, Y., Ogasawara, N. and Alonso, J.C. (1999) Analysis of the *Bacillus subtilis* *recO* gene: RecO forms part of the RecFLOR function. *Mol Gen Genet*, **261**, 567-573.
10. Alonso, J.C., Shirahige, K. and Ogasawara, N. (1990) Molecular cloning, genetic characterization and DNA sequence analysis of the *recM* region of *Bacillus subtilis*. *Nucleic Acids Res*, **18**, 6771-6777.
11. Cardenas, P.P., Carrasco, B., Defeu Soufo, C., Cesar, C.E., Herr, K., Kaufenstein, M., Graumann, P.L. and Alonso, J.C. (2012) RecX facilitates homologous recombination by modulating RecA activities. *PLoS Genet*, **8**, e1003126.

Supplementary Figures

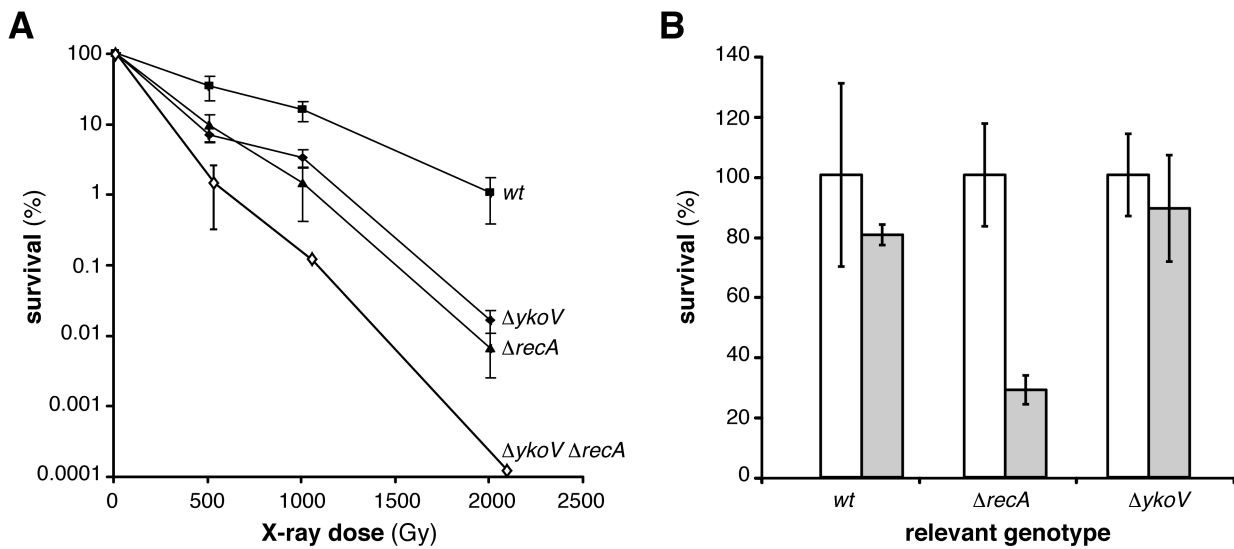


Figure S1. Survival of *B. subtilis* wt and mutant spores deficient in HR or NHEJ after X-ray radiation (A) and UHV (B). (A) X-ray dose dependent survival of wt (■), $\Delta recA$ (▲), $\Delta ykoV$ mutant (◆) or $\Delta ykoV \Delta recA$ (◇) mutant spores. (B) Survival of untreated (white column) and treated spores (gray column) after 7 days UHV exposure.

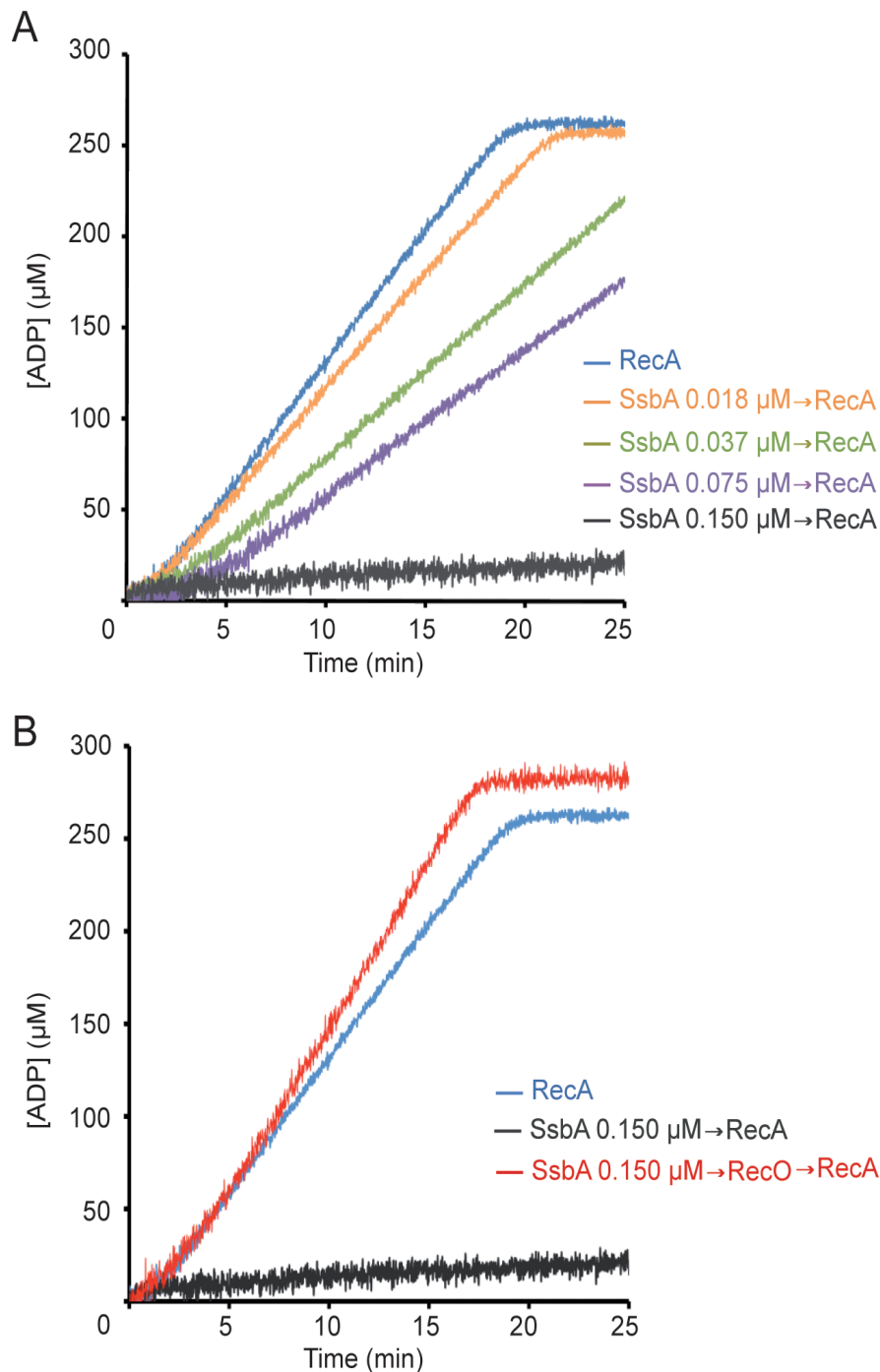
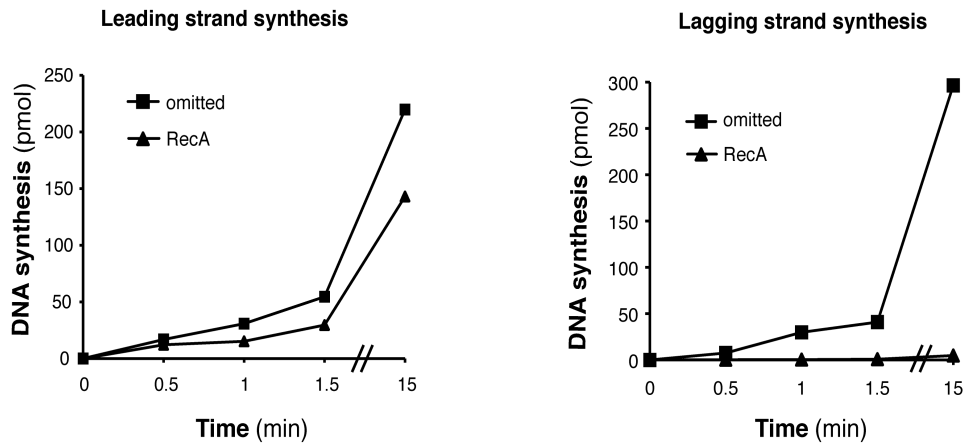


Figure S2. SsbA blocks RecA assembly and RecO overcomes this kinetic barrier. (A) Circular ssDNA (10 μM in nt) was pre-incubated with increasing SsbA concentrations (0, 0.018, 0.037, 0.075 and 0.150 μM) for 5 min at 37 $^{\circ}$ C in buffer A containing 5 mM ATP. Then, RecA (1 μM) was added, and the ATPase activity measured for 25 min. (B), circular ssDNA was pre-incubated with SsbA for 5 min at 37 $^{\circ}$ C, and then RecO (0.2 μM) was added to the preformed SsbA:ssDNA complexes and incubation continued for 5 min at 37 $^{\circ}$ C. Finally RecA (1 μM) was added, and the ATPase activity was measured for 25 min. All reactions were repeated three or more times with similar results. The amount of ATP hydrolysed was calculated as described in Materials and methods.

A. In absence of SsbA



B. In presence of SsbA

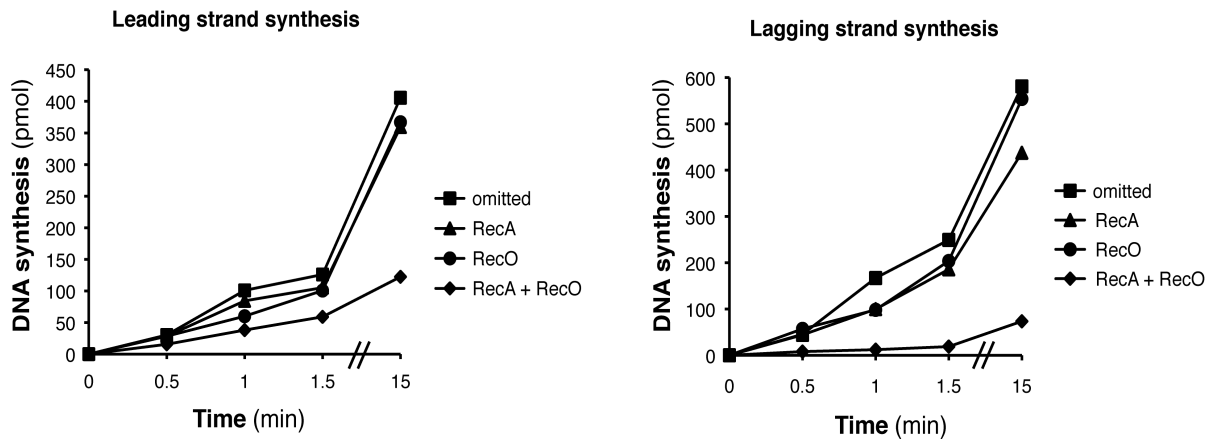


Figure S3. RecA inhibits *in vitro* the *B. subtilis* replisome in concert with RecO. Quantification of leading and lagging strand synthesis obtained measured by radioactivity incorporation as a function of time in the absence of SsbA (A) or in the presence of SsbA (B). The reactions and the measurement of DNA synthesis on both leading and lagging strands were performed as described in Materials and methods.