

SUPPLEMENTAL DATA

Table S1. Baseline clinical characteristics of all treated patients.

	Nonresponders		Responders		Total		<i>P</i>
	N	%	N	%	N	%	
Total	7	100	22	100	29	100	
Age median (range)	48 (30 - 52)		48 (27 - 62)		48 (27 - 62)		0.81
Gender (male)	3	43	10	45	13	45	1
Sex mismatch (male patient, female donor)	2	29	4	18	6	21	0.61
Prior Transplant	2	29	1	5	3	10	0.14
Transplant Type							0.24
Myeloablative	6	86	22	100	28	97	
Non-myeloablative	1	14	-	-	1	3	
Cell Source							0.24
Bone Marrow	6	86	22	100	28	97	
Peripheral Blood Stem Cells	1	14	-	-	1	3	
Donor Type							1
Matched Related	7	100	21	95	28	97	
Matched Unrelated	-	-	1	5	1	3	

Table S2. Baseline clinical characteristics of microarray samples

Patient	Age at DLI	Gender	Transplant Type	Time from Transplant to Relapse (mo)	Time from Transplant to DLI (mo)	Timepoints after DLI (mo)
R1	59	F	Myeloablative	17	18	0 and 3
R2	63	F	Myeloablative	37	38	0 and 6
R3	40	F	Myeloablative	24	27	0 and 3
R4	40	F	Myeloablative	22	24	0 and 12
NR1	48	M	Non-myeloablative	13	14	0 and 6
NR2	50	M	Myeloablative	48	49	0 and 3

Table S3. Gene probes differentially regulated between responders and nonresponders before and after DLI therapy.

Pretreatment		Response to treatment	
<i>Gene Probe</i>	<i>Fold Change</i>	<i>Gene Probe</i>	<i>Fold Change</i>
CD247	5.4	SNHG3	3.3
TNFAIP3	5.2	TSEN2	3.2
LAIR2	3.8	ZBED2	3
PSMB9	2.9	GPM6B	2.7
NCALD	2.6	SACS	2.6
VAMP5	2.6	CBLB	2.3
PPDPF	2.5	PDSS1	2.2
PRKX	2.5	PTTG2	2.2
LITAF	2.4	ZNF451	2
AMZ2	2.4	TMEM177	2
LY6E	2.4	KIAA0317	1.9
EIF5A2	2.4	CAD	1.9
NUAK1	2.4	HIFX	-2.3
ANKRA2	2.3	TPPP3	-2.4
RABAC1	2.3	AP1S2	-2.5
PPP1R2	2.2	DUSP3	-2.6
WWP1	2.1	HLA-DQB1	-2.6
CD81	2	PTAFR	-2.6
UBL3	2	EMR2	-2.6
IFITM2	1.9	LST1	-2.7
HLA-F	1.9	LGALS1	-2.7
AKIRIN1	1.9	HCK	-2.8
TBCB	1.8	TSPO	-2.8
TAP2	1.8	FCGR2A	-2.8
TNIP2	1.7	LILRB4	-2.9
PRLR	-1.5	LY86	-3
KRT13	-1.7	TGFBI	-3.1
MPL	-1.7	LILRA1	-3.1
LMNB1	-1.8	FGL2	-3.1
KIF2C	-1.8	TNFAIP2	-3.1
ENTPD7	-1.9	TMEM176A	-3.2
LTA4H	-2	AIF1	-3.4
TMEM177	-2	BST2	-3.4
RAB3D	-2.1	CYBB	-3.4
TKT	-2.1	SLC15A3	-3.5
SMAD1	-2.2	GPX1	-3.6
ASGR2	-2.4	TLR8	-4
FPR2	-2.4	TYROBP	-4.4
ACPP	-2.4	CSF1R	-4.6
KIF14	-2.4	FCER1G	-4.6
POLQ	-2.5	CST3	-5.4
ZNF193	-2.7	CD9	-6
TROAP	-2.8		
PADI2	-3.7		
LOC100272216	-3.8		
HP	-4		
PADI4	-4.1		
ASPM	-4.2		
P2RY2	-4.2		
MOSC1	-5.3		
CHI3L1	-5.5		

Table S4. Description of curated T cell exhaustion gene sets.

Gene Set	Description	References	Genes
Exhaustion	<i>Exh vs rest in LCMV</i>	26	Genes up in LCMV-specific CD8+ T cells in chronic infection (exhausted) vs acute (effector, memory)
			<i>Exh vs mem in LCMV</i>
	<i>PD-1 ligation</i>	27	
			<i>PD-1 NN</i>
	<i>BATF NN</i>	27	
			<i>CTLA4</i>
<i>HIV Prog</i>	Up in HIV progressors down in HIV elite controllers	27	RASA1 SPOCK2 RASA1 CDC14A TNFRSF25 PTGER4 PCDC4 TNFAIP3 GADD45A RGS1 BIRC3 SLC7A5 PDE4B NELL2 GNAO1 TNFAIP3 HSPH1 LOC728198 TAF9B RGS1 SESN1 CNH1 IFRD1 MED14 MRPS22 M2C2R ACVR1B PDE4B SYNJ2 WRN ZNF238 SNX4 CDC14A RGDPA RGDPS RGDPE RGDPH PIK3IP1 CD28 JMD6 KIAA0999 TACCI SYPL1 C9orf97 PABPC3 ABCG1 TAPT1 YPEL5 FLJ32679 GOLGA8 GOLGGA8 RBP1 ZNF136 PDE4B ARHGAP5 ZRF1 EIF1 LONRF3 AREG LOC727738 CTSO MASP2 TMEM184B DUSP4 SLC39A8 PLXDC1 BCAS2 MAP4K3 MED4 RIF1 PLXDC1 ZBTB10 LOC442215 SPATA2 CDK5R1 LRIG1 FAM102A RAB6P1 DNAJB12 EIF4I CD44 OSBP ID1 LOC146053 LOC391688 LOC391706 LOC3400652 LOC349922 LOC644932 LOC644972 LOC646216 LOC646527 RPS3A TCAL4 ZNF395 KLHL11 CBLB IL7R YIPF5 PRDX1 GULP1 CCERNA1 APOM4P2A WDR48 RMDN5A ETV3 ZMYM2 LOC151162 FLJ32679 GOLGA8 GOLG88 PCDC4 THOC1 ARNTL1 C12orf41 TNF TMEM30B MBOAT5 SLC6A13 KIR2DS3 KIAA0831 C10BP EIF4G1 MSH6 RIG1 N4BP3 FLIPL1 NRA42 IDH1 KLF9 C6orf64 PELO PFN2 TGIF1 RABGGTB CAND2 LIMK2 RWD2A DKK3 IFN25 P12 CDCA9 PRKCI AT2P21 SFRS3 LEPROTL1 PPP1R2 SYPL1 FAIM CD44 LOC653256 RABL3 LRRC40 CXCR3 NRA42 SATB1 YTHDC2 CCT6B CKAP2 PABPC1 ADAM9 KLF7 RPAIN SNRPA1 WDR48 DKB29 SSX2IP C9orf82 CFHR2 GNAL3 TSM FBXL11 PPP1CB NVL EIF4G2 INPP5A MRP9 TMS6F1 LOC92482 ISCA1 SFRS18 RANBP10 ALG5 FBKBP1 TSN POLR2J SCML2 GOLGA8A SF3B1 SLC7A6 EML4 ITSN2 HGSNAT MOAP1 CXCR4 MED14 LSR KTEL1 TBC1D2B ELL2 ISG20L1 SLC25A13

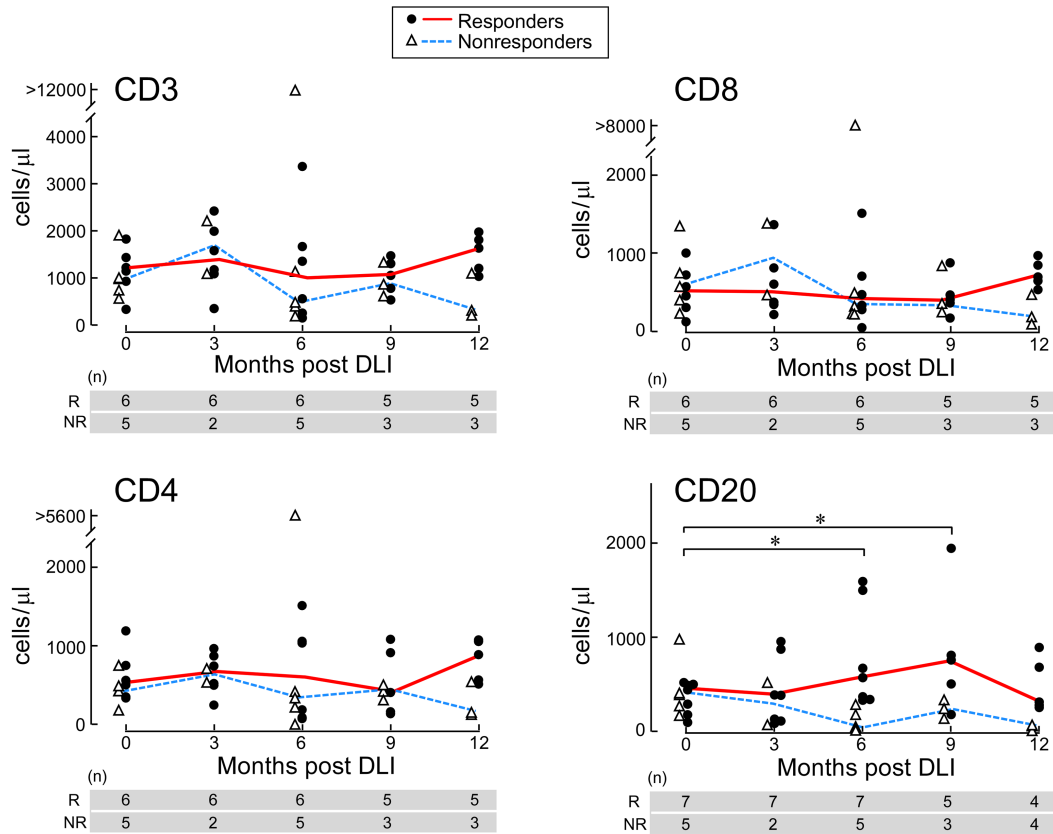


Figure S1. Characterization of immune cell subsets in peripheral blood over time following DLI. Flow cytometric analysis of immune populations in the peripheral blood from responders and nonresponders before and 3, 6, 9, and 12 months after DLI. For each immune cell subset, significant differences between 0 and 3, 6, 9, or 12 month timepoints are noted. * $P < 0.05$; Wilcoxon signed-rank test.

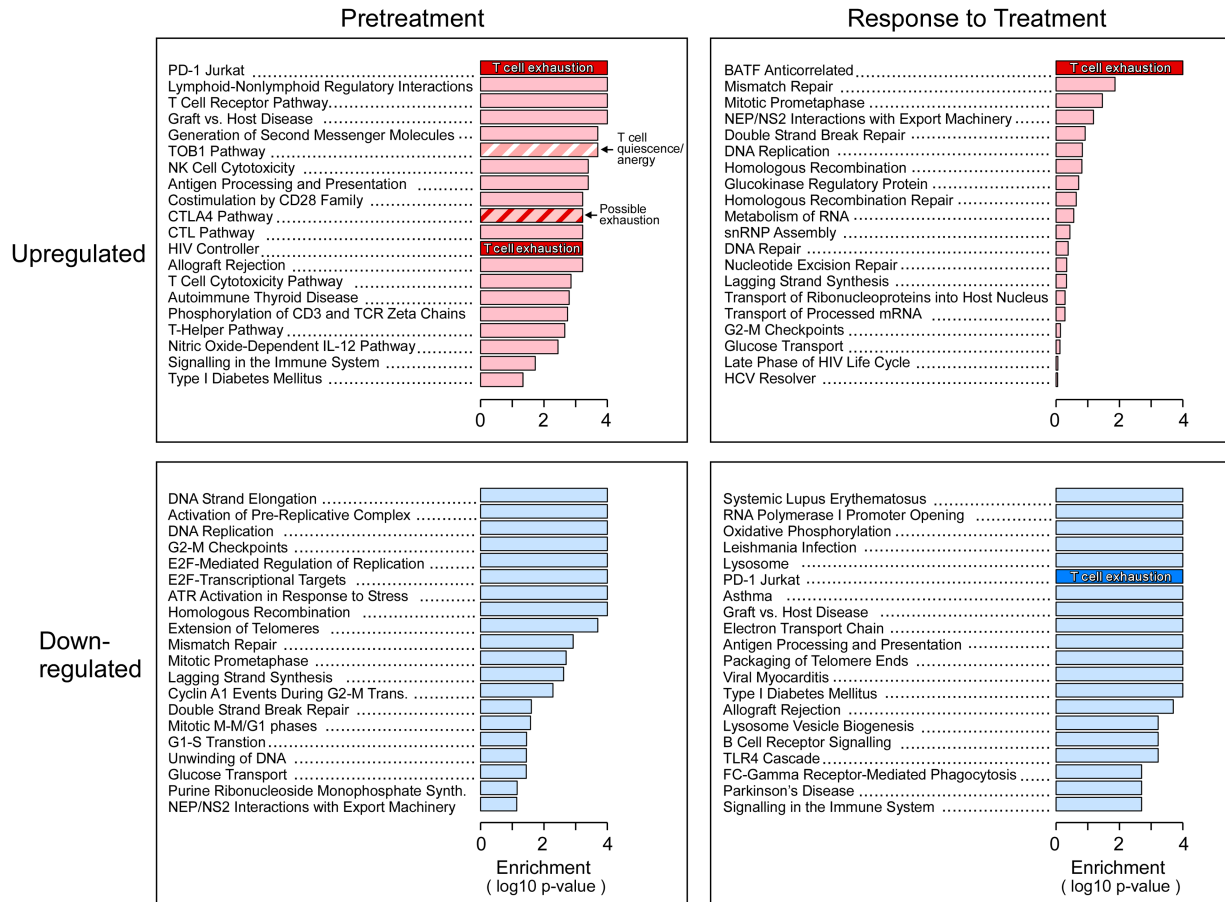
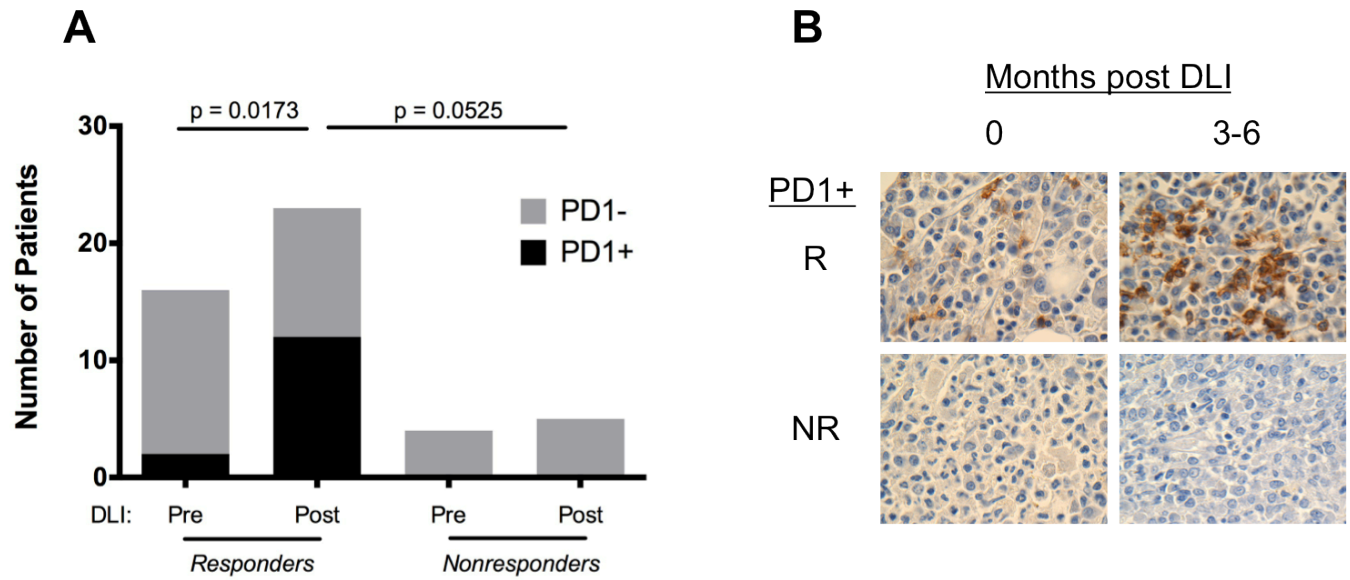


Figure S2. Gene set enrichment analysis (GSEA) of top 15 enriched gene sets in both ‘pretreatment’ and ‘response to treatment’ analyses. Gene sets are derived from ~880 curated gene sets from the CP (canonical pathways) collection of the Molecular Signatures Database, spiked with 11 specifically curated gene sets for T cell exhaustion, detailed in Table S4. Bar graphs show *P*-value of enrichment (signed according to enrichment score) for distinct gene sets. Dark red/blue bars indicate exhaustion gene sets; hatched bars indicate possible associate with T cell exhaustion.



Clinical Cohorts

Figure S3. Association of PD1 expression and clinical response. (A) IHC staining analysis of PD1⁺ cells in the marrow from responders and nonresponders before and after DLI. (B) Representative IHC of marrow samples from one responder and one nonresponder demonstrates robust marrow infiltration by PD1⁺ cells in the responder before and subsequent increase after DLI. Fisher's exact test used for p-value calculation. The images were captured from an Olympus QColor 5 camera on an Olympus BX51 microscope with a 100X/1.30 objective at RT using acquisition software QCapture Pro.