

1 SUPPLEMENTARY MATERIAL

2 Material and Methods

3 Analysis of the amount of HA incorporated into virions.

4 Wildtype and glycosylation mutant viruses in the background of strain A/Netherlands/602/2009
5 were grown in MDCK cells in the presence of TPCK-trypsin. The supernatants were harvested,
6 cleared by low-speed centrifugation and loaded onto 20% sucrose cushions for ultracentrifugation.
7 The resulting virus pellets were resuspended in PBS and subjected to PNGaseF (New England
8 Biolabs) treatment according to the manufacturer's protocol. The deglycosylated virus preparations
9 were analysed by SDS-Page and Coomassie staining of the gel. The intensities of the HA1 and NP
10 bands were quantified by Multigauge software and plotted as ratio of HA/NP.

11

12 Oseltamivir resistance tests.

13 *a. NA star assay.* Viruses were grown in MDCK cells, supernatants were harvested and cleared by
14 low-speed centrifugation. The NA star assay (Invitrogen) was performed using 1:10 dilutions of the
15 virus-containing supernatants according to the manufacturer's instructions.

16 *b. Inhibition of virus replication in tissue culture.* MDCK were infected with the different viruses
17 at an MOI of 0.1 in the presence of different concentrations of Oseltamivir (0/10/100 nM).

18 Supernatants containing virus were harvested at 24h post infection and virus titers were determined
19 on MDCK cells.

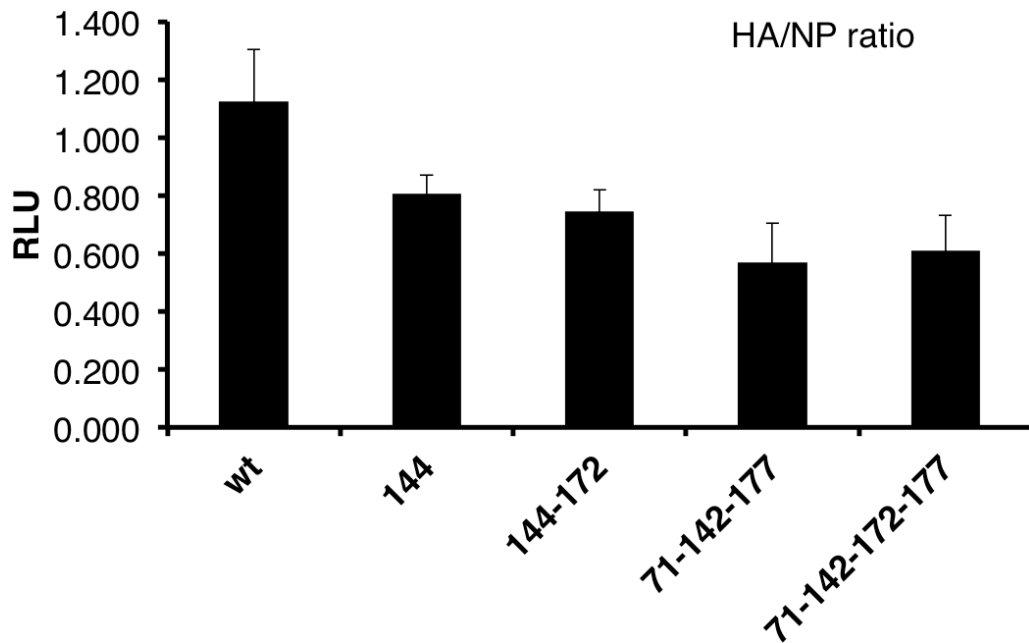
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21 Neutralization Assay.

22 Two-fold serial dilutions of heat (56°C) inactivated mouse sera were mixed and pre-incubated with
23 100 times the 50% tissue culture infectious dose (TCID₅₀) of each indicated virus per well of a 96-
24 well plate in a final volume of 100 µl. The plates were then incubated for 1hr at 37°C. The serum-

25 virus mixture was then added onto monolayers of MDCK cells that had been seeded at a density of
26 2×10^4 cells/well/100 μ l 18hrs earlier. Infection was allowed to proceed for 18-20hrs. The media was
27 removed and the cells were washed with PBS and then fixed with 80% ice-cold acetone/PBS.
28 Infections were detected by incubation with a rabbit polyclonal anti-NP antibody. Positive wells
29 were visualized by utilizing a secondary antibody labeled with peroxidase and the plates read in a
30 plate reader. Neutralizing titers were determined as the highest dilution that displayed neutralization
31 activity.
32

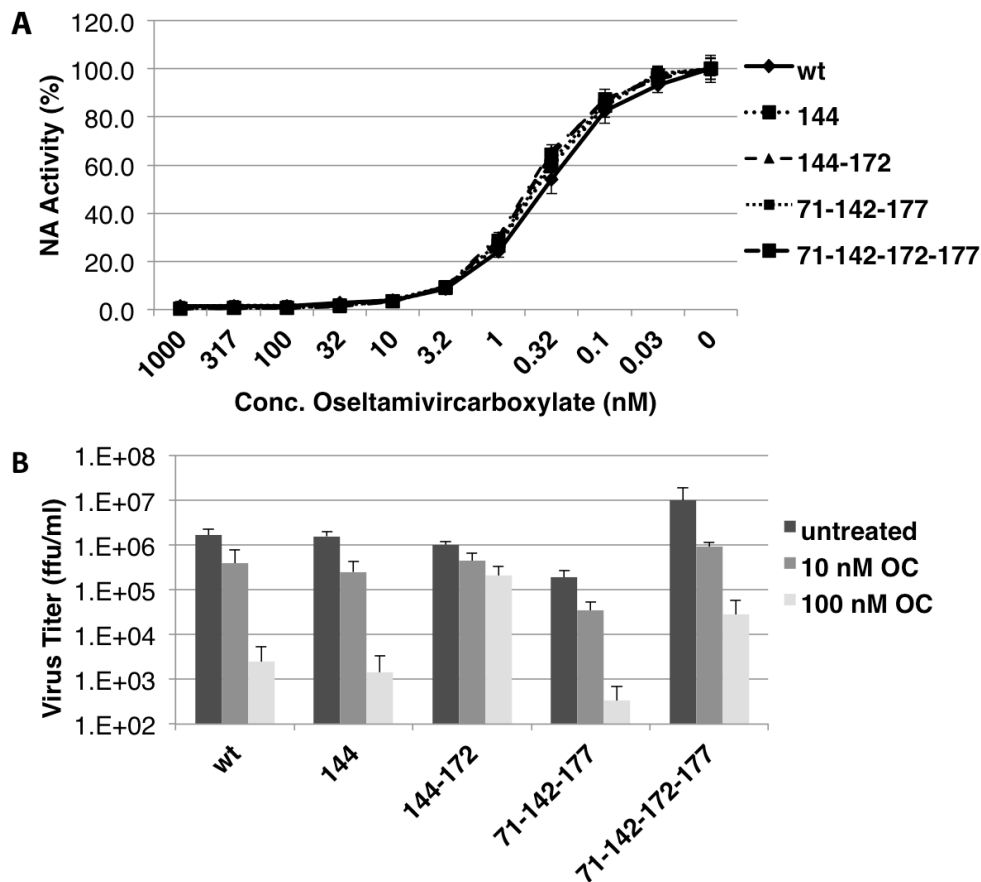
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35 **Figure S1. HA/NP ratio in virions of glycosylation mutant viruses.** Viruses (WT, 144, 144-172,
36 71-142-177, 71-142-172-177) were grown in MDCK cells and purified by ultracentrifugation
37 through a 20% sucrose cushion. The viruses were deglycosylated by PNGaseF treatment and
38 analysed by SDS-Page. Coomassie stained gels were used to quantify the HA1 band in comparison
39 to the NP band for each virus. The average ratio from three independent gels is graphed.

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41

42 **Figure S2. Sensitivity of the glycosylation mutant viruses to the Oseltamivir antiviral. A.**

43 Neth/09 glycosylation mutant viruses do not differ from WT in their sensitivity to Oseltamivir in an

44 *in vitro* assay. Log dose inhibition curves for the WT Neth/09 and the 144, 144-172, 71-142-177

45 and 71-142-172-177 viruses were determined in an NA Star chemiluminescent substrate cleavage

46 assay. Data points represent the average number of relative fluorescent units (RLU) at each drug

47 concentration (0-1000nM in 3-fold dilutions), expressed as a percentage of the RLUs obtained in

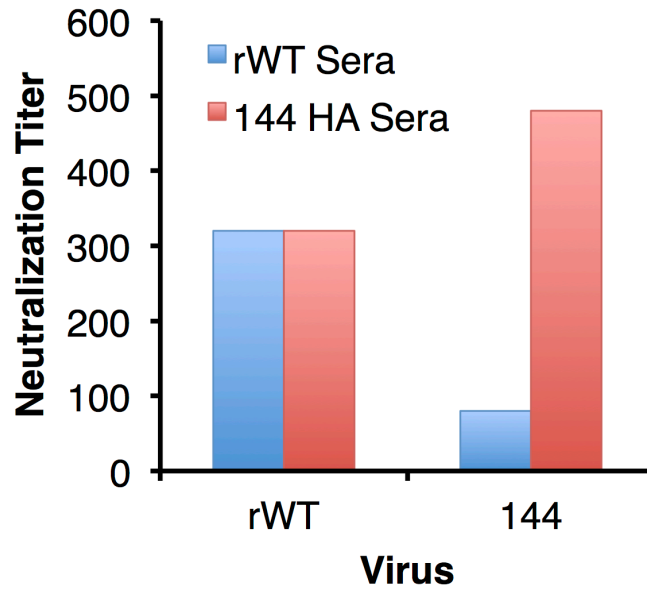
48 the absence of drug; error bars indicate standard deviations. 2B. Glycosylation mutant viruses show

49 minor differences in their sensitivity to Oseltamivir in a tissue culture replication assay. MDCK

50 were infected with the different viruses at an MOI of 0.1 in the presence of different concentrations

51 of Oseltamivir (0, 10 and 100 nM). Supernatants containing virus were harvested at 24h post

52 infection and virus titers were determined on MDCK cells by standard plaque assay.



Virus	Sera	
	rWT	144
rWT	320	320
144	80	480

53

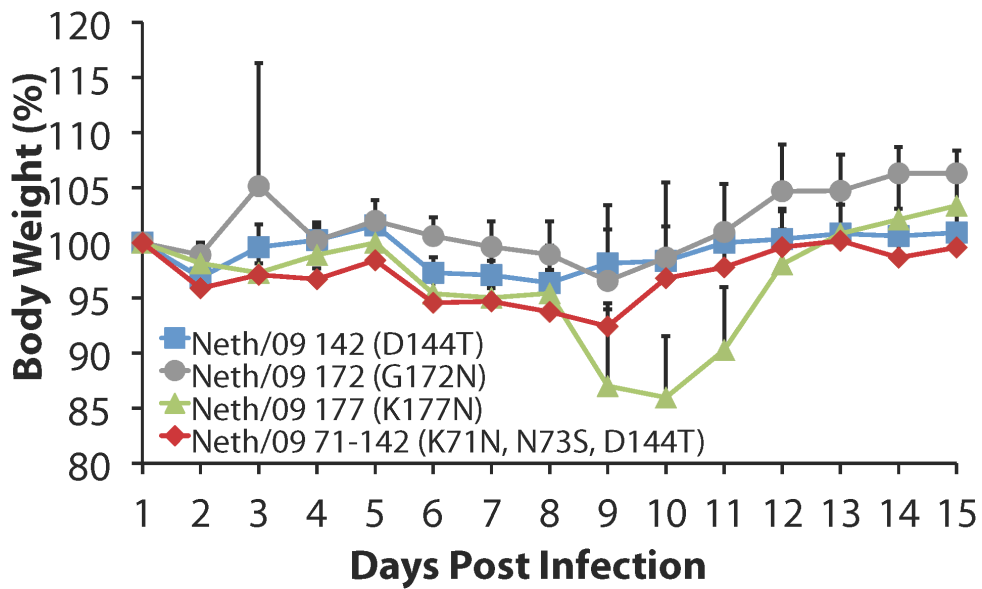
54 **Figure S3: Neutralization activity of sera from mice infected with rWT and 144 glycosylation**

55 **mutant viruses.** Sera from mice infected with either rWT or a virus containing glycosylation 144,

56 obtained at day 28 p.i. were analyzed by micro-neutralization assays. Titers represent the

57 geometrical mean of the highest dilution that displayed neutralization activity.

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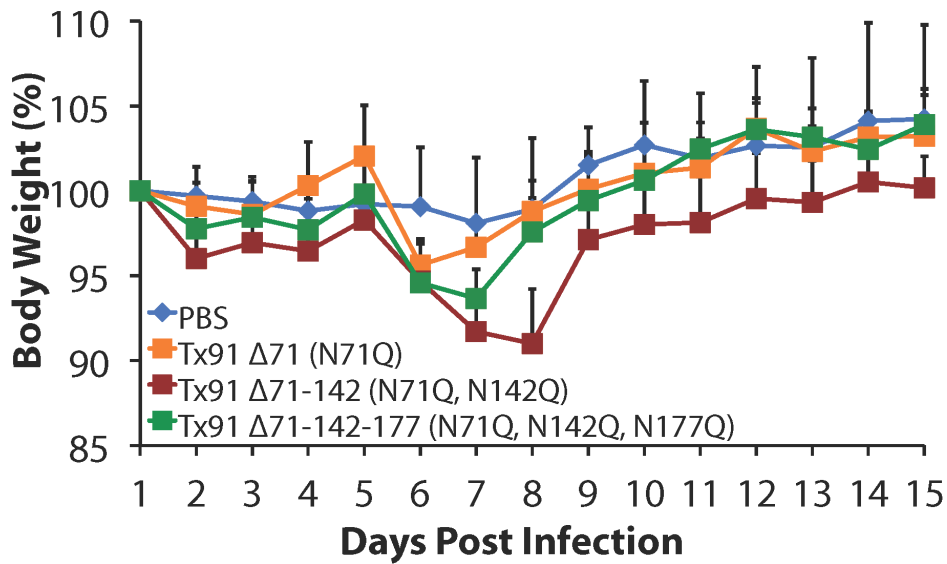


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60 **Figure S4: Pathogenesis of Neth/09 viruses containing single and double glycosylations.**

61 Additional viruses containing single or double glycosylations were generated to evaluate their
 62 individual contribution to virulence and cross-reactivity (or lack of) seen with the viruses containing
 63 the multiple glycosylations. 8-week-old C57B/6 female mice were infected with 1×10^4 pfu of each
 64 of the indicated viruses and the average body weight of mice $n=5$ per group was determined for 15
 65 days. Error bars denote the s.d. for each time point. Mice were allowed to seroconvert for 29 days,
 66 and convalescent sera were used to test their HI activity against the virus containing the naturally
 67 occurring glycosylations. Results are shown in Table 2.

68



69

70 **Figure S5: Pathogenesis of Tx91 viruses containing glycosylation deletions by alternative**

71 **mutations.** To fully evaluate the contribution of glycosylations in cross reactivity, we generated

72 Tx91 glycosylation mutants that removed the glycosylation motif and introduced a non-relevant

73 amino acid within or close to the antigenic site (amino acid changes are shown). 8-week-old

74 C57B/6 female mice were infected with 1×10^4 pfu of each of the indicated viruses and average

75 body weights of mice $n=5$ per group were obtained for 15 days. Error bars denote the s.d. for each

76 time point. Mice were allowed to seroconvert and were then challenged at 29 days p.i with

77 100LD50 of WT Neth/09. Results are shown in Fig 6.

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79 **Table S1. HAI titers of adult and pediatric human sera after vaccination with 2009 pH1N1**
 80 **inactivated vaccine.**

Sera	Age (Years)	Virus														Vaccine	
		rWT		144		144-172		71-142-177		71-142-172-177		Bris/59		Bris/10		2009 H1N1	2009 TIV
		Pre	Post	Pre	Post	Pre	Post	Pre	Post	Pre	Post	Pre	Post	Pre	Post		
1 ^a	1	<10	20	<10	10	<10	10	<10	20	<10	40	20	20	20	20	Yes	No
2 ^b	5	<10	20	<10	10	<10	<10	<10	10	<10	20	40	40	<10	<10	Yes	No
3 ^a	8	<10	<10	<10	<10	<10	<10	<10	10	<10	10	80	80	<10	<10	Yes	No
4 ^b	9	<10	20	<10	10	<10	<10	<10	10	<10	20	10	10	40	160	Yes	No
5 ^b	10	<10	80	<10	20	<10	40	<10	80	<10	320	80	1280	40	1280	Yes	No
6 ^a	12	10	80	10	80	10	40	10	160	10	160	10	10	<10	10	Yes	No
7 ^a	13	<10	80	<10	40	<10	40	<10	160	<10	320	<10	10	10	10	Yes	No
8 ^a	14	<10	10	<10	<10	<10	<10	<10	10	<10	20	80	80	10	10	Yes	No
9 ^c	14	<10	160	<10	160	<10	80	10	640	20	640	<10	40	10	40	Yes	Yes
10 ^a	16	<10	10	<10	<10	<10	<10	<10	20	<10	10	160	160	20	20	Yes	No
11 ^c	16	<10	20	<10	10	<10	<10	<10	20	<10	40	10	80	10	80	Yes	Yes
12 ^a	17	<10	40	<10	10	<10	10	20	160	20	160	10	20	<10	<10	Yes	No
13 ^c	17	<10	160	<10	40	<10	40	<10	640	<10	640	20	20	<10	<10	Yes	Yes
14 ^b	17	<10	80	<10	20	<10	20	10	160	40	320	<10	<10	10	320	Yes	No
15 ^a	18	<10	80	<10	<10	<10	10	<10	80	10	320	80	80	10	10	Yes	No
16 ^b	19	20	160	<10	40	<10	20	20	160	20	160	<10	80	<10	320	Yes	Yes
17 ^a	20	<10	1280	<10	320	<10	160	10	1280	20	2560	40	80	160	160	Yes	No
18 ^b	21	10	640	10	320	<10	160	20	1280	20	1280	80	80	80	160	Yes	Yes
19 ^a	31	<10	40	<10	20	<10	40	<10	40	10	160	<10	<10	<10	<10	Yes	No
20 ^b	33	<10	80	<10	20	<10	40	<10	20	<10	80	20	20	10	20	Yes	No
21 ^b	34	<10	2560	<10	1280	<10	1280	<10	80	<10	640	<10	20	<10	10	Yes	Yes
22 ^b	34	10	160	10	<10	10	<10	20	320	10	640	<10	160	40	1280	Yes	Yes
23 ^a	36	<10	160	<10	80	<10	80	<10	160	<10	320	<10	<10	<10	<10	Yes	No
24 ^a	40	<10	160	<10	80	<10	80	<10	160	10	320	<10	10	<10	<10	Yes	No
25 ^a	42	<10	40	<10	20	<10	20	<10	40	<10	160	20	20	<10	<10	Yes	No
26 ^a	42	<10	640	<10	640	10	640	<10	80	80	640	10	20	10	10	Yes	No
27 ^b	42	20	640	10	320	10	160	20	320	20	640	10	20	20	40	Yes	Yes
28 ^b	43	10	640	10	320	10	160	<10	640	10	640	10	20	80	40	Yes	Yes
29 ^a	44	<10	40	<10	20	<10	40	<10	10	<10	80	<10	10	160	160	Yes	No
30 ^a	46	<10	1280	10	1280	<10	1280	<10	160	40	160	<10	<10	20	20	Yes	No
31 ^a	46	<10	20	<10	<10	<10	<10	<10	20	<10	80	80	80	10	10	Yes	No
32 ^a	47	<10	160	<10	160	<10	160	<10	80	<10	80	<10	<10	<10	<10	Yes	No
33 ^a	48	<10	40	<10	10	<10	40	<10	40	<10	160	<10	10	<10	<10	Yes	No
34 ^b	49	<10	40	<10	20	<10	20	<10	40	<10	160	<10	10	<10	20	Yes	Yes
35 ^a	52	<10	640	<10	320	<10	320	<10	160	<10	320	80	80	<10	<10	Yes	No
36 ^a	54	<10	20	<10	<10	<10	<10	<10	40	<10	80	10	10	20	20	Yes	No
37 ^b	55	<10	640	<10	320	<10	640	<10	320	<10	640	<10	80	<10	<10	Yes	Yes
38 ^a	57	<10	20	<10	10	<10	<10	<10	20	<10	40	<10	<10	<10	<10	Yes	No
39 ^a	59	40	160	40	80	40	80	40	80	40	160	10	10	20	20	Yes	No
40 ^d	60	20	1280	10	320	20	320	20	320	20	160	<10	<10	<10	10	Yes	Yes
41 ^a	63	10	40	<10	20	<10	20	<10	40	10	40	<10	<10	<10	<10	Yes	No
42 ^a	65	<10	20	<10	<10	<10	<10	<10	10	<10	40	<10	20	<10	80	Yes	No
43 ^a	66	20	160	10	160	10	80	20	80	40	160	40	40	80	80	Yes	No
44 ^a	67	<10	40	<10	10	<10	<10	<10	20	<10	20	<10	<10	10	10	Yes	No
45 ^d	67	10	1280	<10	640	<10	320	20	640	10	320	20	40	40	80	Yes	Yes
46 ^d	67	20	640	<10	40	20	320	20	640	20	2560	10	10	40	320	Yes	Yes
47 ^a	68	10	160	10	40	10	40	10	160	40	320	40	40	20	20	Yes	No
48 ^a	73	<10	80	<10	20	<10	40	10	80	20	320	<10	<10	<10	<10	Yes	No
49 ^a	75	10	320	10	320	<10	160	10	160	10	160	10	20	10	10	Yes	No
50 ^d	82	40	2560	40	2560	40	160	40	1280	40	160	10	20	40	160	Yes	Yes
51 ^d	51	<10	<10	<10	<10	<10	<10	<10	<10	<10	<10	40	20	<10	40	No	Yes
52 ^d	74	<10	10	<10	<10	<10	10	<10	<10	<10	10	<10	320	20	80	No	Yes

82 ^a Human serum samples obtained on days 0 (Pre) and 42 (Post) after vaccination, respectively.

83 Subjects received the 2009 H1N1 inactivated vaccine on days 0 and 21.

84 ^b Human serum samples obtained on days 0 (Pre) and 21 (Post) after vaccination, respectively.

85 Subjects received the 2009 H1N1 inactivated vaccine on day 0 only.

86 ^c Human serum samples obtained on days 0 (Pre) and 42 (Post) after vaccination, respectively. The

87 2009 TIV: trivalent inactivated influenza virus vaccine which included strains A/Brisbane/57/2007

88 (H1N1), A/Brisbane/10/2007 (H3N2) and B/Brisbane/60/2008, was given on day 0 and the 2009

89 H1N1 vaccine was given on day 21.

90 ^d Human serum samples obtained on days 0 (Pre) and 63 (Post) after vaccination, respectively. 2009

91 H1N1 inactivated vaccine was given on days 0 and 21 and the 2009 TIV: trivalent inactivated

92 influenza virus vaccine, which included strains A/Brisbane/57/2007 (H1N1), A/Brisbane/10/2007

93 (H3N2) and B/Brisbane/60/2008, was given on day 42.