Genome-wide Nucleosome Map and Cytosine Methylation Levels of an Ancient Human Genome

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Section SI1. Sequence datasets

Most of the analyses were performed on previously released sequence data sets, consisting of: 1) the first complete ancient human genome [Rasmussen et al. 2010] (section SI1.1); 2) a modern control data set based on a panel of modern samples [Green et al. 2010, Reich et al. 2010] (section SI1.2). 3) the first ancient Aborigine genome [Rasmussen et al. 2011] (section SI1.3); and 4) genomic reads of a 110,000-130,000 year-old polar bear [Miller et al. 2012] (section SI1.5). In addition, we generated genomic reads from an ancient horse dating back to ca. 4.5 ka (section SI1.4) and from modern human hairs (section SI1.6). Those were analyzed to check for the specificity of the patterns observed in hairs and their possible extension to other types of ancient samples, such as calcified bones that represent the vast majority of fossil remains excavated.

1.1. Saqqaq Palaeo-eskimo sequence dataset

The sequence data have been originally generated as part of our characterization of the Saggag Palaeo-Eskimo genome [Rasmussen et al. 2010]. Overall, ancient DNA extracts from a 4.000-year-old permafrost-preserved hair were built into Illumina libraries and deep-sequenced at about a 20X-coverage on a GAIIx sequencing platform and Single End sequencing. The vast majority of the sequencing reads (242 lanes out of a total of 245, corresponding to ca. 3.5 billions of reads) were recovered following library amplification with Phusion DNA polymerase (Finnzymes, hereafter referred to as Phusion). A total of 3 GAIIx lanes, however, were generated following library amplification with a Tag platinum high-fidelity DNA polymerase (Life technologies, hereafter referred to as Hifi) and resulted in a total number of 50.1 millions of reads [Ginolhac et al. 2011]. Contrarily to Hifi, Phusion has been demonstrated to show poor activity at uracil residues (Figure S3.1) [Fogg et al. 2002]. The latter are generated following post-mortem deamination of cytosine residues, mainly at single-stranded overhanging ends. As chemical analogs of thymines. Uracils yield to the mis-incorporation of AT base pairs (instead of GC) during library preparation and amplification, resulting in an excess of $GC \rightarrow AT$ mismatches when ancient genomes are compared to modern reference genomes [Stiller et al. 2006; Brotherton et al. 2007; Gilbert et al. 2007; Briggs et al. 2007]. Thus, that Phusion, rather than Hifi, has been used for generating the first complete sequence of an ancient genome greatly contributed to the overall quality of the genome by limiting the impact of damage-driven misincorporations in downstream analyses [Rasmussen et al. 2010].

For the nucleosome analysis the original set of 999.3 millions of reads was used, as defined in [Rasmussen et al. 2010]. For the methylation analysis, a mapping procedure allowing for indels was followed as described below (this was performed because the alignments available from the original study were performed with SESAM and were indel-free; [Rasmussen et al. 2010]). Read ends were trimmed for potentially residual adapter sequences, and after the first base showing the poorest quality score (C) using the software AdapterRemoval available for download at http://code.google.com/p/adapterremoval/ [Lindgren 2012]. Trimmed reads were subsequently mapped against nuclear human genome reference (hg18) using bwa 0.5.9 [Li and Durbin 2010] and default values for optional parameters. Unmapped sequences, together with the high-quality reads previously identified, were remapped against the available mitochondrial genome sequence of this individual [Gilbert et al. 2008] using similar parameters; in addition, sequencing reads with a minimal length of 25 nucleotides, a minimal mapping quality of 30, and showing no alternative hits

were further selected using a combination of the samtools suite and awk command lines. Finally, sequence duplicates were collapsed using the sort and rmdup commands available in SAMtools [Li and Durbin 2010], resulting in a final BAM file of unique mitochondrial reads that was used for some of the analyses presented below aiming at detecting cytosine methylation-driven nucleotide mis-incorporation patterns (section SI3). The final dataset consisted of a final number of 696.0 millions and 26.3 millions of non-clonal unique sequences originating from Phusion and *Taq* Platinum high fidelity (Hifi) library amplification, respectively.

1.2. Control genome dataset

We constructed a data set with the same properties as the Saqqaq data set in terms of number of reads and read length distribution, except reads were randomly selected (and truncated to match the Saqqaq size distribution) from a panel of sequencing runs from modern genomes. This data set allowed us to (1) evaluate if the Saqqaq data set observations could be ascribed to mapping biases and also to (2) more generally represent a null set to evaluate the significance of the statistical measurements made during the analysis.

To construct this set, we used the combined set of modern genomes sequenced at low read depth as part of the Neanderthal and Desinova projects, which were downloaded from UCSC [Green et al. 2010, Reich et al. 2010]. All samples originate from lymphoblastoid cell lines of the Human Genome Diversity Project (HGDP). A main criterion for the data sets used for the Control was that they were made using the same technology, but based on modern samples that were not degraded or fragmented. All reads, including unmapped, were extracted from the downloaded bam files and converted to fastq format. These were then shuffled to avoid any ordering relative to genomic coordinates. We then constructed a data set with the same length distribution as the Saggag, by truncating individual modern reads to the appropriate size, though we initially included about 1.2 times more reads than present in the Saggag data set. We then mapped each fragment size separately to hg18 using BWA [Li et al. 2010]. For each fragment-size set of unsorted mapped reads, we then extracted the exact same number of reads of the given size as present in the Saggag data set. Finally, all the reads were combined to make our final Control data set.

1.3. Ancient Aborigine sequence dataset

A total number of 311.0 millions of Illumina reads recovered from a *ca*. 100-years old hair sample originating an ancient Aborigine individual [Rasmussen et al. 2011]. These reads were generated following a procedure similar to the one used for characterizing the complete genome of the *Saqqaq* individual, except that DNA libraries were amplified using *Taq* Gold DNA polymerase. The initial unprocessed data set consisted of 2.1 billions of reads (100 nucleotides long), which was reduced by 85.2% following index filtering, adaptor trimming, and selection of uniquely mapping and non-clonal reads. These span 60% of the human genome with an average read depth of 11x (6.4x overall). The average length of endogenous trimmed and unique reads consisted of 64 nucleotides and mapping was done using BWA and standard parameters.

1.4. Ancient horse sequence dataset

The ancient horse sample was provided by one of the co-authors (Dr. Alexei Tikhonov). It has been found in Yakutia at Batagai (Republic of Sakkha) and showed an uncalibrated radiocarbon date of 4,450±35 BP (Gr 50842). A total of 0.225 grams of ulna bone was drilled to powder at low speed in ancient DNA lab facilities available at the Centre for GeoGenetics. DNA was extracted following the silica-based procedure described in [Rohland and Hofreiter 2007], with slight modifications following [Orlando et al. 2009]. DNA was released from silica pellets using a final elution volume of 205ul of EB (QIAgen) and 15 minutes of incubation at 37°C. We constructed one Illumina DNA library following the standard truSeg procedure, except that 16.5ul of DNA extract were used as input and that the End Repair reaction was performed using End-It End Repair kit reagents from Epicentre (catalog reference ER0720). A first incubation at 4°C for 2 minutes was followed by a second incubation at 37°C for 45 minutes. During those incubation steps, 5'-overhangs were filled-in by T4 DNA polymerase and 5'-ends are phosphorylated by PNK. Endrepaired DNA templates were purified using QIAgen minelute purification kit and 10 volumes of PN buffer instead of 5 volumes of PB buffer. The whole elution volume was used for klenow exo- polyA-tailing (37°C for 30 min) and adapter ligation using standard indexed truSeq adapter. The DNA library was further purified using Ampure XP beads with 1.8:1 as a volume ratio between beads and DNA, and eluted into 20ul of EB solution. We amplified 10ul of DNA library using 300nM Illumina PCR primers (PCR primer 1.0: 5'-AAT GAT ACG GCG ACC ACC GAG ATC TAC ACT CTT TCC CTA CAC GA ; PCR primer 2.0: 5'-CAA GCA GAA GAC GGC ATA CGA GAT), 15 cycles of PCR amplification, a concentration of 1mg/ml BSA, 1µM dXTP (Invitrogen) and 5U of Tag Gold (invitrogen). Final PCR volume was 25µl and cycling conditions consisted of a first DNA denaturation at 95°C for 10 minutes, followed by 15 cycles of denaturation (95°C, 30 sec), annealing (60°C, 30 sec) and elongation (72°C, 60 sec). A final elongation was performed at 72°C for 7 minutes before amplified DNA was purified into 25ul EB using QIAgen minelute purification kit. The DNA library was sequenced on a hiseg2000 platform available at the Danish National DNA Sequencing Centre using 100 cycles Paired-End reactions.

Illumina Paired-end reads were trimmed for adapter sequences at their 3'-end following a pair-wise alignment procedure and using the software AdapterRemoval [Lindgreen 2012]. A minimal sequence identity of 90% over 5-10 nucleotides was required for adapter alignment (this threshold was lowered to 67% for longer alignments). In addition, reads showing stretches of bases with low quality scores and/or Ns at sequence ends were trimmed from the first nucleotide position showing the low quality score. Reads starting with stretches of at least one undetermined base were trimmed, resulting in a final sequence starting at the first position showing a determined base. Following adapter removal, read pairs were aligned and collapsed as long as showing a maximum of one mismatch over a sequence of 11 nucleotides at read ends. A larger fraction of mismatches (33%) were tolerated for longer alignments (>11 nucleotides). Sequence collapse was performed by calling a consensus based on the nucleotide showing the highest quality score; when the two reads showed identical nucleotides, a quality score was re-assigned based on the product of the two previous quality scores but with a maximal limit set-up to a Phred score 41 (e.g. when the two nucleotides where sequenced with a 10% chance of error, the base quality score of that position in the consensus was re-assigned to 1%). Read-pairs showing no overlap were discarded. Collapsed-pairs were further

mapped as single-end reads against the horse reference genome EquCab2.0 available for download at UCSC

(<u>http://hgdownload.cse.ucsc.edu/downloads.html#horse</u>) using BWA and standard parameters, except that the seed was disabled following the recommendations of [Schubert et al. 2012].

1.5. Ancient polar bear sequence dataset

We followed the full procedure described in section SI1.4 to characterize the size distribution of endogenous DNA inserts using 812,006,446 Illumina Paired-end reads (SRS347019) generated from the DNA extract of a 110,000-130,000 years old polar bear from Svalbard (Norway) [Miller et al. 2012]. The *de novo* assembled scaffolds of the polar bear genome (available for download at

ftp://climb.genomics.cn/pub/10.5524/100001_101000/100008/Ursus_maritimus.scaf.f a.gz) were used as a reference for read mapping.

1.6. Modern human hair sequence dataset

We followed the same experimental procedure as for characterizing the Saqqaq genome. We generated a total number of 10.0 millions correctly indexed, adaptor trimmed, uniquely mapping and non-clonal Illumina reads from a single DNA library prepared on modern hairs from a British born Caucasian living in Denmark at present. Average read length was of 61 nucleotides. These sequences span 15.7% of the human genome with an average read depth of 1.4X (0.22X overall). This dataset was generated in order to evaluate if the nucleosome-driven signal detected on previous datasets was specific to ancient DNA. Single-End sequencing was performed to a lower read depth, sufficient to statistically confirm that the same signal is present. DNA extraction and downstream bioinformatics analysis followed the same protocol as for the ancient Saqqaq Palaeo-Eskimo genome [Rasmussen et al. 2010].

1.7. Size distributions

A subset of 116.1 millions of Paired-End reads generated from 4 Saggag: indexed libraries was considered for analyzing the size distribution of ancient DNA inserts. Pairs of reads showing a minimum overlap of 11 bases (with 100% identity) were collapsed into single reads using AdapterRemoval [Lindgren 2012] following the procedure described in section SI1.4. This resulted in a total number of 83.6 millions of collapsed reads that were mapped against the human reference genome hg18 using bwa, following the same parameters as described in section SI1.4. We then randomly selected 1.5 million of unique hits per library (for a total of 6.0 millions of reads) and further filtered for collapsed reads 1) ending with base quality scores of 40 and 41 when shorter than the size of sequencing reads (75 nucleotides) and for longer sizes, 2) those that end with base quality scores of 33-40. We found a total number of 4.8 millions selected collapsed reads, corresponding to endogenous library inserts sequenced over their full length, showing unique high-quality alignments against polar bear scaffolds. The resulting size distribution is shown as Figure 2.d, bottom.

Ancient Horse: We focused on collapsed reads showing a sequence length longer than 25 nucleotides and a minimal mapping quality (MQ) of 25. We filtered PCR duplicates using 5'- and 3'-coordinates and keeping the read showing the best

average base quality. This procedure was implemented using a python script kindly provided by Dr. Martin Kircher (*FilterUniqueBAM.py*; for a SAM compatible version, see <u>https://bioinf.eva.mpg.de/fastqProcessing/</u>). The size distribution of those collapsed reads mapping uniquely the horse reference genome was plotted using R [R Development Core Team 2012] and a subset of 100,000 reads randomly selected. Corresponding reads are available for download at SRA (SRA105533). Horse inserts showed a remarkable 10-bp size-periodicity spanning 38bp to 142bp (Figure 2.d, top). This periodicity corresponds to the average size of a DNA helix turn. No 10-bp periodicity could be observed for inserts larger than the size of nucleosome (146bp). This pattern suggests that DNA fragmentation in spacer regions occurs randomly in contrast to nucleosome cores where DNA strands facing away from nucleosome protection are exposed to hydrolytic reactions and fragmentation.

We noticed that within pairs, the quality of the second read was Polar bear: generally of a lower quality than the first read, with a significant fraction showing lowest quality scores over the full read length. This could result in artifact while collapsing reads, with increased false positive and false negative identification of overlapping read ends. In order to avoid such bias and select collapsed reads with starting and ending coordinates reflecting the full length of DNA library inserts, we further selected from the population of collapsed reads 1) those that end with base quality scores of 40 and 41 when shorter than the size of sequencing reads (101 nucleotides) and for longer sizes, 2) those that end with base quality scores of 38-40. We found a total number of 3.8 millions of selected collapsed reads showing unique high-quality alignments against polar bear scaffolds. Those were considered for plotting insert size distribution (Figure 2.d, middle). A periodicity, similar to the one observed using the ancient horse sequence dataset, although admittedly of a lower magnitude, was observed on 110-130 ka polar bear DNA inserts (38bp-153bp), suggesting long term preservation of nucleosomal positioning signal in ancient DNA extracts. We found the size class of 101 nucleotides slightly over-represented. This likely reflects a sequencing or data-handling artifact, as this class of length corresponds to the number of sequencing cycles performed on the hiseg2000 platform.

Of note, size distributions showing similar ca. 10-bp periodicity were observed for a series of ancient DNA extracts from Holocene and Pleistocene bone material preserved in the permafrost (data not shown).

We constructed similar size distributions for the modern hair data generated in this study as well as for a range of other modern genomes generated from fresh blood using a similar methodology (from different horse breeds; Orlando et al. 2013). Those size distributions are presented for 30-60bp in Figure S1.1 in order to facilitate comparison across samples. The top graphs provide size distributions for the Saggag, the ancient polar bear, the ancient horse and the modern hair. The bottom graphs provide size distributions for modern horses. Frequencies have been normalized to peak at 1 for the size showing maximum frequency. The ancient horse show a short-range periodicity at 38, 49 and 59 bp whereas the ancient polar bear show a short-range periodicity at 39, 50 and 60 bp. Similarly, the modern hair also shows a short-range periodicity with peaks at 49 and 60 bp (no reads shorter than 39bp and longer than 60bp were experimentally generated, precluding direct observation of a larger size range). As the horse and polar bear datasets have been generated in absence of size-selection, this clearly rules out size selection as a driver for the short-range periodicity observed in fragment size distributions. Given that no short range periodicity is observed in sequence datasets generated from fresh blood, we conclude that the short range periodicity observed in the modern hair sample originates from active degradation of hair DNA following apoptotic processes

occurring during hair differentiation (Botchkavera et al. 2006). The periodicity observed in Saqqaq, the horse and the polar bear, exactly matches the average turn of a B-DNA helix, with nucleotides facing away from nucleosomes becoming more susceptible to *post-mortem* degradation than nucleotides in contact with nucleosomes. In a similar way, apoptotic enzymes are well known to cause DNA laddering by cleaving chromatin DNA into inter-nucleosomal fragments (Nagata et al. 1998). During this process, nucleosome protection also creates ~ 10-bp periodicity patterns (Aruscavage et al. 2010). This is responsible for the short-range periodicity observed in the modern hair data.

That 1-bp offsets are sometimes observed between the Saqqaq, the ancient horse, the ancient polar bear and the modern hair (with peaks either at 38bp or 39bp, 48-50bp), most likely comes from two causes. First, one helix turn is around 10.4 bases, which is expected to create a mixture of both 10-bp and 11-bp periodicities. Second, our trimming procedure removes any stretch of Ns at read starts and depending on the dataset considered, we observed variable fractions of sequencing starting with one N. Read trimming results in the shortening of the sequence by one nucleotide, creating a proportion of fragments offset by 1 nucleotide in their size distribution.



Figure S1.1. Size distribution of ancient and modern sequence datasets. Top: Ancient and modern hair sequence datasets. Bottom: Modern blood sequence datasets.

Size, bp

Section SI2. Nucleosomes

2.1. Read depth

We evaluated the average read depth in different genomic regions where read depth is defined as the number of mapped reads covering that region normalized by the region size. We first evaluated read depth at different genic regions, by segmenting the entire genome according to the protein-coding genes of the UCSC Gene set [Fujita et al. 2011]. When multiple genes or isoforms overlapped a region, annotation priority was given as: protein-coding exons (CDS) > 5'UTR > 3'UTR > introns > intergenic regions. In addition, read depth was also evaluated at CpG islands (CGI) as defined by the UCSC Genome Browser.

Read depth was found to vary dramatically between genomic regions for the Saqqaq genome, with the highest read depth observed for CGI. In contrast the Control set displayed a nearly constant read depth across genomic regions, with the lowest read depth observed at CGI. We note that for a wider variety of other genomic datasets, CGI also showed lower mapping rates than the other genomic regions investigated (data not shown). Both the Aboriginal sample and the Modern hair sample largely follow the same read depth pattern as Saqqaq. Finally, a modern human MNase-based nucleosome occupancy data set generally also follows the same pattern, though with less variation between regions.

Region	genome	coding	intergenic	intron	3'UTR	5'UTR	CpG island
Saqqaq	19.4(1.0)	53.8(2.8)	17.2(0.9)	21.1(1.1)	26.9(1.4)	76.8(4.0)	126.1(6.5)
Control	19.4(1.0)	20.7(1.1)	18.7(1.0)	20.3(1.0)	20.8(1.1)	18.1(0.9)	11.5(0.6)
Aboriginal	6.9 (1.0)	17.9(2.6)	6.1 (0.9)	7.6 (1.1)	9.9 (1.4)	22.0(3.2)	32.7(4.8)
Modern hair	0.2 (1.0)	0.5 (2.1)	0.2 (0.9)	0.2 (1.0)	0.3 (1.3)	0.7 (3.2)	1.0 (4.5)
Schones	2.7 (1.0)	3.9 (1.5)	2.6 (1.0)	2.7 (1.0)	3.1 (1.2)	3.9 (1.5)	3.8 (1.4)

Table S2.1.

Regional read depth for main data sets. The average read depth across genomic regions is given for each data set. Read depth normalized by genomic average (enrichment) is given in parenthesis. Schones refers to a mapping of human nucleosomes using MNase [Schones et al. 2008].

Region	genome	coding	intergenic	intron	3'UTR	5'UTR	CpG island
Saqqaq	0.2	5.9	0.0	0.2	-0.2	10.1	22.8
Control	0.6	1.0	0.5	0.8	1.3	0.4	-0.6

Table S2.2.

GC-normalized read depth for Saqqaq and Control.

2.2. Control of mapping and sequencing biases

To eliminate the effect of any potential mapping biases due to the repetitive structure of the human genome, we defined a set of genomically unique regions (Unique). For all mapping procedures used, we retained only uniquely mapping reads. Using ExactRepeats, a mapping program based on suffix arrays (developped by one of us: Anders Krogh), we defined all regions that are uniquely mappable by reads of length 26 or greater. These regions were further trimmed by 100 bases at either end, ensuring that the observed read depth within these is unaffected by mapping issues. The resulting set spans 51.3% of the genome.

The PCR amplification step of NGS library generation is known to introduce biases in the frequency of occurrence of different fragments dependent on GC-content and length [Benjamini and Speed 2012]. To rule out that such biases could create or contribute to the observed variation and to improve the specificity of the nucleosome signal in the downstream analysis, we evaluated their strength at each position along the genome and corrected the read depth signal accordingly. We achieved this by estimating what the read depth would be at every genomic position given the read length distribution; the location of uniquely mappable regions for every read length; and the GC-content for all possible fragments overlapping a given position. The estimated read depth composed of contributions from all available read lengths were then subtracted from the observed read depth. The resulting read depth was thus positive when more read depth is observed than expected from GC content and negative when less read depth was observed than expected from GC content.

More specifically, we based this evaluation on the GC correct package [Benjamini and Speed 2012] implemented in R [R Development Core Team 2012]. Briefly, GCcorrect learns the correlation between GC-content and read depth across the genome and uses it to predict the expected number of reads covering every position of the genome. It disregards non-unique regions based on a mappability file. Since mappability is dramatically different depending on the read length (in Saqqaq ranging from length 20 to 76 bp), we supplied a separate mappability file for each length. ExactRepeats (see above) was used to create these. Reads shorter than 26 bp were discarded from the analysis as there were too few reads for GCcorrect to learn the correlation between read depth and GC content. For each read length, we supplied GC correct with the corresponding mappability file and the overall number of observed reads. GCcorrect then returned the expected number of reads starting at any given position for the given read length. The resulting expected read depth summed across all read lengths was calculated using an extension to the UCSC Genome Browser command line tool bedItemOverlapCount, which allowed to sum up genome-wide real-valued data across multiple data sets. This overall expected read depth was subtracted from the observed read depth (disregarding reads shorter than 26), to produce the final GC-corrected read depth. The same procedure was applied to the Control data set, while taking the original fragment sizes in the panel of underlying data sets into account.

2.3. Correlation with other nucleosome data and predictions

We evaluated the correlation between different data sets across various genomic regions. Briefly, the following data sets were included in the analysis, (1) The original uncorrected Saqqaq read depth; (2) GC-corrected Saqqaq read depth; (3) The original uncorrected Control read depth; (4) The GC-corrected Control read depth; (4) The original Aboriginal read depth; (5) The read depth of the modern hair sample; (6) GC content in 5 bp windows as available from the UCSC Genome Browser [Meyer et al. 2012]; (7) Computational predictions of nucleosome occupancy as based on SVM models trained on MNase digestion data from A375 (A375 set) and MDA-kb2 (Dennis set; [Dennis et al. 2007) cell lines; 8) Experimental genome-wide MNasebased occupancy data sets obtained from CD4+ cells [Schones et al. 2008] and lympoblastoid cell lines [Gaffney et al. 2012]. The Schones data is based on read depth of 25 bp long singleend reads, while the Gaffney data is based on midpoints of paired-end reads (126bp < fragment length < 184bp) smoothed by 30bp, following the authors instructions.

Both Pearson's and Spearman's (data not shown) correlation coefficients were evaluated across large subsets of the genome. As the results were highly similar, we only report Pearson's correlation coefficients (PCC).

We first focused on the conservatively defined Unique regions (see Section SI2.2), which span 51% of the genome. These were represented by a random sample of 10,000 regions, in total spanning 7.7 Mb. The results are reported in the main article text.

We next evaluated the correlation across a 20 Kb subsection of a known nucleosome array region (chr12:34,331,000-34,451,000; hg18), where nucleosomes are thought to be consistently and specifically positioned independently of tissue [Gaffney et al. 2012]. To rule out the influence of common mapping biases, we included only the subset of Unique regions (~13 Kb). The resulting correlation matrix (Figure S2.1 a) allows several observations:

- 1. For both Saqqaq and Control the GC corrected and original read depths are highly correlated.
- 2. The Saqqaq and Aboriginal data sets are also highly correlated (0.74).
- 3. Similar levels of correlation are seen for Saqqaq versus the computational nucleosome occupancy predictions (0.77 and 0.74 for uncorrected read depth).
- 4. The experimental MNase data sets, which are based on related cell types (CD4+ and lymphoblastoid), show a correlation of 0.38 between them and a lower correlation with the computational predictions (0.16-0.28) than Saqqaq. This could suggest some level of noise in state-of-the art MNase-based occupancy maps, potentially from the cutting biases of the MNase enzyme [Gaffney et al. 2012].
- 5. The correlation of the Saqqaq and Aboriginal data sets against the experimental data sets (pcc of 0.23 to 0.43) is of the same magnitude as the correlation between the two experimental data sets (pcc=0.38).
- 6. Generally the original, uncorrected Saqqaq data set shows higher levels of correlation with the computational and experimental occupancy maps than the GC corrected Saqqaq data set. This may be attributed to "over-correction" at inherently GC rich nucleosome sites, as GC content is known to be part of the nucleosome-positioning signal [Collings et al. 2010; Valouev et al. 2011]. However, it is also likely that the MNase based data sets, which also underlie the computational predictions, are subject to some level of GC-dependent PCR bias, which is not corrected for.

Finally, we evaluated the correlation in the +/-1Kbp regions around the TSSs of all UCSC transcripts. To illustrate the variance in the correlation coefficients between TSS regions, we plotted the full distribution of PCCs for these. Separate plots were made for the correlation of select data sets against the GC-corrected Saqqaq read depth (Figure 1.f); the original Saqqaq read depth (Figure S2.1.b); and the Schones data set (Figure S2.1.c). Again, the computational predictions stand out with consistently strong positive correlation against the uncorrected

Saqqaq data set across the TSS regions (Figure S2.1.b). The Schones data set shows stronger correlation with the Gaffney data set than with either Saqqaq or the computational predictions (Figure S2.1.c). The nucleosome positioning patterns in TSS regions have been shown to correlate with gene expression levels and hence be cell type specific [Valouev et al. 2011]. The relatively high correlation of Schones and Gaffney in TSS regions may thus be explained by the underlying cell types being related.



Correlation matrix for nucleosome array

Figure S2.1

Correlation between data sets.

a, Correlation matrix for known nucleosome array region (20 Kb). Pearson's correlation coefficients were calculated for each pairwise combination of data sets and color-coded according to magnitude. All correlations coefficients are significantly different from zero (all p-values < 1.2e-15). **b**, Distributions of Pearson correlation coefficients across TSS regions for original, uncorrected Saqqaq read depth *versus* uncorrected Control set, GC content (GC), two computational occupancy maps (Dennis and A375), and an experimental occupancy map (Schones). **c**, Distribution of Pearson correlation coefficients across TSS regions for Schones versus a subset of data sets from (b) and another experimental occupancy map (Gaffney).

2.4. Nucleosome occupancy at anchor sites

Nucleosome occupancy, represented by the read depth, was plotted around multiple anchor points in the genome: TSSs, CTCF sites and splice sites. All instances of each group of anchor points were aligned and a meta plot of the accumulated read depth in the surrounding regions divided by the number of anchors was plotted. For TSSs and splice sites we used the UCSC genes set.



Figure S2.2

Nucleotide and methylation around donor and acceptor splice sites. Methylation signal is described in sections SI3.2 and SI3.3.

2.5. Fourier Transform

We predicted that nucleosome protection would cause a periodicity in read depth variation corresponding to the size of the nucleosome and the spacer region (~200 bp). To unbiasedly evaluate if such periodicity could be observed, we performed a Fourier transform (FT) analysis using Welch's FT method (a windowed Fourier transform). More specifically, we performed a spectral density plot (periodogram) genome-wide across CpG islands, TSS±1000 bp, gene bodies, and CTCF sites ±1000 bp. The raw spectral density was corrected for the background frequency distribution, to remove low frequency variations and constant offsets, by subtracting a background estimated by exponential curve modeling. To visualize the spatial distribution of frequency information, short time Fourier transforms (spectrograms) over known anchor sites, such as Transcription Start Sites (TSS) (source: UCSC genes) and CTCF binding sites (source: [Fu et al. 2008], see section SI3.7), were generated. For Saqqaq and Control the GC-corrected read depth was used.



Fourier Transform periodicity analyses of transcription start site regions.

a, Read depth periodogram for Saqqaq and Control. Saqqaq peak periodicity is 193 bp. **b**, Spectrogram for GC-corrected Saqqaq read depth. The strongest periodicity signal exists downstream of the TSS. **c**, Spectrogram for Control. Note that some periodicity is found around 200bp in the region, though much weaker than for Saqqaq. **d**, GC content does show any datable periodicity in the range.



Fourier Transform periodicity analyses of CpG islands.

a, Read depth periodogram for Saqqaq and Control. Saqqaq peak periodicity is 189 bp. **b**, Spectrogram for GC-corrected Saqqaq read depth. **c**, Spectrogram for Control. Note that some periodicity is found around 200bp in the region, though much weaker than for Saqqaq. **d**, GC content does show any datable periodicity in the range.



Fourier Transform periodicity analyses of CTCF regions.

a, Read depth periodogram for Saqqaq and Control. Saqqaq peak periodicity is 182 bp. **b**, Spectrogram for GC-corrected Saqqaq read depth. **c**, Spectrogram for Control. Note that some periodicity is found around 200bp in the region, though much weaker than for Saqqaq. **d**, GC content does show any datable periodicity in the range.



Periodogram of gene bodies.

Read depth periodogram for Saqqaq and Control. Saqqaq peak periodicity is 192 bp. Note that Control that exhibits some signal, albeit much weaker than Saqqaq.



Figure S2.7

Periodograms of CpG islands for Modern hair and Aboriginal data sets.

a, Periodogram for Modern hair data set. Peak periodicity is at 186 bp. Note that modern hair was low-coverage and hence less signal was detected. **b**, Periodogram for Aboriginal data set. Peak periodicity is at 182 bp.

2.6. Phasograms

We predicted that in regions where nucleosomes are consistently positioned, the distance between two successive nucleosomes should equal the size of the nucleosome plus the size of the spacer region (~200 bp). Phasograms were produced in a similar manner as described in [Valouev et al. 2011] in order to test for the presence of such signal.

Using the raw uncorrected data, we counted the distance of pairs of 5' ends on the same strand including only positions with at least at least 5 reads. For both the short and long range plots we used gene bodies (+1Kbp to the termination site) of UCSC protein coding genes

The signal overall follows an exponential distribution due to the local variation in read depth. To estimate the period, we therefore fit and exponential background distribution (Figure S2.8a), which we subtract from the signal to better reveal the periodicity (Figure S2.8b). Finally we analyzed the autocorrelation of these regions identifying the dominant short range phasing to be 10 bp and the long range phasing to be 185 bp (Figure S2.8c).



Figure S2.8

Phasogram of Gene bodies.

a, Raw signal of Saqqaq and Control. **b**, Background subtracted. **c**, Autocorrelation plot of phasogram signal with background removed. Peak periodicity is 10 bp for short range and 185 bp for long range (gray lines and blue numbers). Note that the autocorrelation for Control is based on a very weak signal.

2.7. Genome-wide map of nucleosomes

To generate a genome-wide map of nucleosome positions we applied a sliding window to the GC-corrected read depth and called the center position if it has the maximal read depth in that window (Figures 3.a, S2.9). We refer to these as peaks. The window size was chosen to be 147-bp based on the size of a single nucleosome. If the peak score is negative, which is possible when the expected read depth is greater than the observed read depth, the peak is discarded and no call is made. Though some (overcorrected) true calls are likely missed based on this, it avoids calls based simply on variation in the expected read depth.

Each call was assigned a score, defined as the read depth of the peak (p) minus the average read depth of the left flanking region (lf) and right flanking region (rf): score= p - (lf + rf / 2) (Figures 3.a, S2.9). The score is constructed to capture aspects of both occupancy (peak height) and positioning (linker region depletion).

Importantly the expected read depth from our GC-correct procedure takes the place of a control experiment in the peak calling procedure. It is designed to capture and reflect any bias resulting from size and/or GC-content. Furthermore, the mappability of each read length is carefully taken into account and thus modeled in the final expected read depth (Section S2.2). The subtraction of the expected read depth thus corrects for peaks that would be wrongly called due to these effects, similar to the role of a control experiment in say ChIP-seq analysis. To further eliminate any risk mapping biases, the downstream analysis of the nucleosome calls are all restricted to the Unique regions (Section S2.2).

Nucleosome calls based on the original, uncorrected read depth were made using the same approach. However, instead of the difference score used for the GC corrected read depth, we used a log-odds score, which better captures the positioning signal of linker region depletion: score = $\log(p / ((If + rf) / 2 + eps))$. Eps was chosen to be 1/25. The analyses of these calls gave similar results, as exemplified by their nucleotide and di-nulceotide distrbutions (Figure S2.13).



Figure S2.9

Nucleosome calls and scores.

(Similar to Figure 3.a and included for convenience of reference.) Nucleosome center positions (dyads) are called as read depth peaks if maximal at center of running window of nucleosome length (147 bp). Calls are scored by the difference in read depth between the peak and the average read depth of the left and right flanking regions.

2.8. Estimating false positives

The Control data set has the same overall genomic read depth and read length distribution as the Saqqaq by construction. However, it shows only a weak nucleosome signal in the periodicity analysis (section SI 2.5). We therefore made nucleosome predictions (section SI 2.7) based on the Control data set, in the assumption they would all effectively be false positives (FP). We subsequently used those to define the expected number of false positive predictions at a given quality score cutoff.

For a given score cutoff, we thus estimate the expected number of true positive calls (TP) as the difference between the number of Saqqaq calls (S) and the number of Control calls (B): TP = S - FP = S - B (Figure 3.b, Figure S2.10).

The false discovery rate (FDR) for a given score cutoff was estimated using a similar procedure as the ratio between the number of Control calls and the number of Saqqaq calls: FDR = B / S.

To investigate if some extreme peaks in the Control could point to problematic regions and lead to spurious calls in the Saqqaq, we evaluated the correlation in ranks between the top-scoring Control peaks and the Saqqaq peaks (figure S2.10c). As can be seen, there is no tendency for the top-ranked Control peaks to be correlated with top-ranked Saqqaq peaks in the Unique regions used for all analysis.



Call score distributions and summary statistics. a, Analyses of all calls. Top-left, difference between Saqqaq and Control calls is taken as an estimate of number true positive calls at given cutoff. Top-right, Saqqaq call distribution, false discovery rate (FDR), and fraction of true positive calls (TP) of the total set are given as a function of cutoff. Bottom, summary statistics for different top-scoring subsets (fractiles). **b**, Analyses of calls in Unique regions as in (a). **c**, The rank of the top 0.1% (left) and top 1% (right) peak calls in the null set plotted against their corresponding rank in the Saqqaq set. A smoothed line has been added for visualization of the general trend (stat_smooth from ggplot2 in R). There is no bias for highly ranked null peaks to receive a high rank in the Saqqaq set.

2.9. Nucleotide and di-nucleotide distributions across nucleosomes

Based on the nucleosome calls (section SI 2.8), we evaluate the nucleotide and di-nucleotide distributions across the nucleosomes. For this, we ranked the calls based on the positioning scores and explored the sequence preference at different thresholds by summing the number of mono-nucleotides or di-nucleotides and dividing by the number of nucleosomes. This gave us an average profile for a given threshold (Figures 3.c, S2.10, S2.11).

The resulting profiles exhibit distinct nucleotide level shifts in nucleotide and di-nucleotide frequencies which indicates that the nucleosome sequences are accurately aligned and hence a large fraction of the nucleosome calls are at nucleotide precision. Furthermore, we note distinct nucleotide preferences at positions close to the center (dyad) and the presence of strand-specific patterns nucleotide (Figure S2.11) and di-nucleotide patterns (Figures 3.c). We hypothesize that these profiles reflect sequence-level positioning determinants and note that the strand-specific patterns that shifts across the dyad suggest a method for precise positioning despite overall weak sequence signal.

For comparison, we also include the nucleotide and pyrimidine/purine di-nucleotide distributions around nucleosome calls based on the uncorrected, original Saqqaq read depth data (Figure S2.13). These mirror the characteristics of the corresponding plots for the GC-corrected read depth (Figure S2.11 and Figure 3.c), showing that the nucleotide distribution results are robust. As expected, the top-scoring uncorrected calls show a stronger degree of GC enrichment among top-scoring calls (Figure S2.13), as these will be penalized by the GC correction scheme.



Mono-nucleotide distributions across sets of top-scoring nucleosome calls for Saqqaq and Control. The average usage of a mono-nucleotide for positions relative to the nucleosome peak call (dyad) for different top-scoring subsets of nucleosome calls from Unique regions (top: top-1%;middle: top-10%; bottom: top-25%).



Strong / weak and CpG / GpC di-nucleotide distributions across sets of top-25% nucleosome calls for Saqqaq and Cotrol. Left, strong (GG, CG, GC, CC) / weak (AA, AT, TA, TT) di-nucleotide profiles. Boundary of peak is denoted with vertical lines (at -30 and +30). Right, CpG and GpC di-nucleotide profiles. Boundary of peak denoted with vertical lines (at -30 and +30).



b Dinucleotide distribution for top-25% of calls on uncorrected Saqqaq and Control read depth



Mono-nucleotide and pyrimidine/purine di-nucleotide distributions across top-25% scoring nucleosome calls for the original, uncorrected Saqqaq and original, uncorrected Control.

a, The average usage of mono-nucleotides at positions relative to the nucleosome peak call (dyad) for top-25% scoring subset of nucleosome calls from Unique regions. **b**,The distribution of purine/pyrimidine di-nucleotide usage across top 25% called nucleosomes from Unique regions.

Section SI3. Methylation footprint of the Saqqaq genome

3.1. Background

While *post-mortem* deamination of cytosine residues results in the formation of uracils, the deamination of methylated cytosines (5mC) results in thymine residues that are adequate templates for both Phusion and *Taq* platinum high-fidelity (Hifi) DNA polymerases (Figure S3.1). Consequently, DNA strands with methylated CpG dinucleotides will partake to the pool of library molecules that will generate sequence data regardless of which of the two DNA polymerases was used, should cytosine residues be deaminated or not (Figure S3.1). Conversely, DNA strands with unmethylated but deaminated CpG dinucleotides (ie. UpG) will provide sequence data following amplification with Hifi but not Phusion (Figure S3.1). In the next sections, we exploited this versatile behavior of 5mCpG and CpG dinucleotides as a direct experimental signature of ancient DNA methylation tracts.



Figure S3.1.

Unmethylated and methylated CpG dinucleotides show different sequencing features following *post-mortem* deamination. A, Biochemical structure of native cytosine (left) and 5mC (right) residues. B, Biochemical structure of deaminated cytosine (ie. uracil, left) and deaminated 5mC (thymine, right) residues. The ability of Phusion and Hifi DNA polymerases to by-pass the different types of nucleotides during PCR amplification is indicated with plus/minus signs.

3.2. Nucleotide mis-incorporation patterns

Nucleotide mis-incorporation patterns were recovered from the mapDamage package [Ginolhac et al. 2011]. This package reports the frequencies of all mismatches observed between sequence reads and the genome used as a reference (here, hg18) for all nucleotidic positions along sequencing reads. Even though Phusion cannot by-pass deaminated cytosine residues, nucleotide mis-incorporation patterns clearly mimicked cytosine deamination, with $C \rightarrow T$ mis-incorporation rates increasing towards the 5'-end of the sequencing reads (Figure 4.a, bottom left), in agreement with previous reports [Briggs et al. 2007; Krause et al. 2010; Ginolhac et al. 2011; Orlando et al. 2011] and the presence of 5'-overhangs that favor cytosine deamination after death. We hypothesized that the signal originated from 5mC residues that were deaminated after death and were replicated

as thymine analogs, hence generating $C \rightarrow T$ mismatches post-sequencing when compared to the hg18 reference.

Encouragingly, this signal was absent from the promoter of the constitutivelyexpressed house-keeping gene GAPDH (Glyceraldehyde 3-phosphate dehydrogenase; uc001qor.1, chr12:6,513,918-6,517,797), in agreement with the documented absence of cytosine methylation on that region [Noer et al. 2006]. Conversely, the signal was present on Phusion reads respectively mapping the genomic regions corresponding to TSH2A (chr6:25,834,270-25,834,769; for a total of 484 reads) and TSH2B (chr6:25,835,116-25,835,552; for a total of 244 reads) (data not shown), two testis-specific genes that are known to be silent in somatic tissues and hypermethylated [Zalensky et al. 2002; Weber et al. 2007].

We further recorded mis-incorporation rates in different genomic regions (mitochondrial DNA; nuclear DNA; exons located outside CGIs; introns located outside CGIs; CGIs). These genomic regions were identified using the IRanges R library [Pages et al. 2010] and UCSC Genome Browser tracks. Since $C \rightarrow T$ mismatch rates decrease from 5' ends of ancient DNA templates due to the presence of 5'overhangs [Briggs et al. 2007], CpN dinucleotides located at sequence starts are expected to show maximal level of cytosine deamination. Following our hypothesis, none of such Phusion reads would show $C \rightarrow T$ misincorporations at the first position unless it was methylated (UpG are not by-passed in contrast to TpG, the postmortem by-product of 5mCpG deamination; Figure S3.1). Therefore at read starts, $CpG \rightarrow TpG$ conversion rates are expected to be significantly increased at CpG due to the methylation background, but not at CpA, CpT and CpC (70-80% of cytosine methylation occurs at CpG sites in the human genome; [Lister et al. 2009]). We tested this prediction by binning sequence reads generated with Hifi and Phusion according to which of those four types of dinucleotides was present in the reference genome at sequencing starts (Figures 4.a and 4.b).

For genomic regions starting with CpG dinucleotides, a higher fraction of Phusion reads were found to start with TpG dinucleotides, especially in exonic and intronic regions located outside CGI (Figure 4.b). Conversely, CGI exhibit reduced proportions of Phusion reads starting with TpG dinucleotides, in agreement with the fact that most, but not all, CGI represent hypomethylated genomic regions [Deaton and Bird 2011]. From all genomic regions considered, the mitochondrial genome appeared as an outlier showing the lowest levels in agreement with the global unmethylated level of this genome [Rebello et al. 2009]. Interestingly, the same trend in CpG \rightarrow TpG conversion rates was observed but with reduced frequencies when considering sequencing reads that covered genomic CpG dinucleotides at positions 20 and 21 (Figure S3.2). Compared to sequencing starts, these positions are affected by lower cytosine deamination rates [Briggs et al. 2007], which results in lower post-mortem 5mCpG to TpG conversion rates, and consequently, lower $CpG \rightarrow TpG$ conversion rates. Importantly, CpA, CpC and CpT dinucleotides exhibit similar levels of TpA, TpC, and TpT conversions, respectively, regardless of the genomic region considered, suggesting that the observed patterns were indeed CpGspecific and therefore driven by the combination of methylation and deamination of cvtosines.

All in all, we found that $C \rightarrow T$ mis-incorporations were preferentially located in genomic regions known to undergo methylation (CpG, resulting when methylated in tractable $5mCpG \rightarrow TpG$ Phusion conversions); as expected (Figure S3.1), the strength of the signal detected appeared directly dependent of *post-mortem* deamination levels (see conversion rates at positions 1-2 and 20-21), suggesting that methylation could only be detected following *post-mortem* deamination of 5mC



residues. This confirmed our hypothesis and suggested that Phusion reads could provide a genuine a signature of methylation patterns in ancient genomes.

Figure S3.2.

Substitution patterns at CpN dinucleotides. We calculated the proportion of alignments showing a CpN dinucleotide in the reference genome and TpN at positions 20-21 within sequencing reads. Such CpG→TpG conversions at positions 20-21 are reported in red and have been calculated for different genomic regions, including: mitochondrial DNA (Mitochondrial); the whole nuclear genome (Nuclear); exons outside of CGI (Exon); introns outside of CGI (Intron); and CGI. Additionally, the rates of other types of dinucleotide conversions were investigated (Other: CpA→TpA, CpT→TpT, CpC→TpC) and are reported in grey. Top: Phusion reads. Bottom: Hifi reads. Figure 4.b provides similar calculations at read starts (ie. positions 1-2).

Overall, relatively high CpG \rightarrow TpG conversion rates (over ca. 3%) were observed with Hifi in all genomic regions, including the mitochondrial genome (Figure 4.b). This appears in striking contrast with what observed on Phusion reads and confirms the ability of Hifi polymerase to by-pass uracil residues, regardless of their methylation status (Figure S3.1). Of note, that higher CpG \rightarrow TpG conversion rates were observed with Hifi in Introns and Exons than in CGIs and in the mitochondrial genome suggests preferential elongation for that enzyme at deaminated 5mC (ie. analog of native thymines) than at deaminated cytosines (ie. uracils).

In this study, we identified a simple means of recovering information about the

methylation status of CpG dinucleotides in ancient genomes by looking at CpG \rightarrow TpG conversions events at starting positions of Phusion reads. At these positions, *post-mortem* cytosine deamination rates are maximal, and methylation preserves our ability to PCR amplify and sequence damaged endogenous ancient templates. Note that the same holds true at sequence ends where complementary G \rightarrow A misincorporations will be observed [Briggs et al. 2007], resulting in CpG \rightarrow CpA conversion events. However, as the Saqqaq sequence data mostly corresponded to Single End reads, the large fraction of DNA inserts were not sequenced over their full length. In addition, a significant drop in base qualities was observed towards read ends. We therefore decided to disregard CpG \rightarrow CpA conversion events at read ends (those result from the deamination of 5mCpG on the complementary strand), despite the information those carry about methylation. We anticipate that this information could be fully used in case where extensive Paired-End sequence data is available.

3.3. Quantifying ancient regional methylation levels

We used CpG \rightarrow TpG at read starts in order to define M_s, a proxy for regional methylation levels (Figure S3.3). Specifically, M_s is defined as the total counts of CpG→TpG conversions at read starts normalized by the number of CpG dinucleotides present within a given genomic region of the reference human genome hg18. We disregarded conversions that possibly colocated with positions described as polymorphic in dbSNP130 in order to filter for SNP hotspots, which provide a possible source of methylation-unrelated CpG→TpG conversions. Of note, even if $CpG \rightarrow TpG$ conversions represent a footprint of CpG methylation at a given location of the human genome, such conversions could only be detected in situations where the 5mC residue is affected by post-mortem deamination (see sections SI3.1 and SI3.2). For the Saqqaq data, this occurs only in ca. 4% of the cases (Figure 4.a), representing on average one chance event every 25 observations. This means that in 96% of the cases a given 5mCpG will not be deaminated and will therefore be read as a regular CpG, leaving no CpG \rightarrow TpG conversion mark. Therefore, the absence of $CpG \rightarrow TpG$ conversions at a given CpG cannot be used to demonstrate the absence of methylation, unless this position shows an extremely high depth-ofcoverage. Our methodology will consequently be most generally innapropriate for reconstructing genome-wide methylation maps at single nucleotide levels. However, by summing over particular genomic regions instead of unique CpGs (eg. promoters, gene bodies, exons and introns; see below), we could increase the chances to observe $CpG \rightarrow TpG$ conversions and predict regional methylation levels. Since CpGmethylation levels have been shown to be significantly correlated within 1,000-2,000 bp [Eckart et al. 2006; Down et al. 2008], our method provides a means to scan genomes for differences in regional methylation levels, despite its relative lack of sensitivity at the single nucleotide level.

We first validated M_s as a methylation proxy by looking at known methylation patterns in specific genomic regions: gene promoters and exon-intron boundaries. As promoter methylation levels have been reported to be negatively correlated with their CpG density, we first binned promoters into three groups according to CpG density following [Ball et al. 2009]. A promoter region (PM) was defined following [Ball et al. 2009] as spanning 500 nucleotides upstream and 2,000 nucleotides downstream of the Transcription Start Site (TSS). High CpG promoters were defined to contain a 500-bp interval with a GC content of at least 0.55 and a CpG observed/expected ratio of at least 0.75. Low CpG promoters were defined as containing no 500-bp interval with a CpG observed/expected ratio of at least 0.48; all remaining promoters were defined as intermediate CpG promoters. Of the 51,939 genes analyzed, 16,186 (31.2%) were found to have high CpG promoters, 11,830 (22.7%) had low CpG density promoters, and the remaining 23,923 (46.1%) had intermediate CpG promoters (Figure 4.c). For each category we calculated the distribution of M_s values. Given the relatively low *post-mortem* deamination rates observed in the Saqqaq genome (leading to ~4% C \rightarrow T mismatch rates at sequencing starts; Figure 4.a), we required each promoter to have a minimum of 50 CpG dinucleotides in the reference in order to increase the likelihood of detecting deamination-driven 5mCpG \rightarrow TpG conversions. In addition, we required a minimal number of 2 C \rightarrow T mismatches per promoter in order to limit the impact of sequencing error in our estimates. In total, we considered a final number of 2,444, 5,973 and 10,026 promoters in the three classes of promoters (low, intermediate and high CpG, respectively).

We found that on average, genes having promoters with high CpG densites exhibited lower M_s values than genes having promoters with intermediate CpG densities (median R_s values for the global gene dataset: 1.35% *versus* 2.06%; Figure 4.c). Similarly, genes having promoters with intermediate CpG densities showed lower M_s values than promoters from the low CpG class (median M_s values of 2.06% *versus* 5.46%). Overall, this supported the expected inverse relationship between CpG density and promoter methylation levels as reported in Ball et al. [2009] and therefore confirmed M_s as a good proxy for gene methylation at the regional level.



Figure S3.3

Defining M_s, a proxy for methylation levels of a given genomic region. As cytosines at sequencing starts show maximal but still moderate deamination levels in the Saggag sequence dataset (Figure 4.a, bottom left), we decided to focus only on the first position of sequencing reads, outlined in grey, and ignore possible CpG→TpG conversions located further down in the sequence. The direction of sequencing reads is indicated with arrows. For Phusion reads mapping the human reference hg18 on the (+) strand and at a location starting with a CpG dinucleotide, we first counted N1 and N2 as the number of reads starting with CpG and TpG. respectively (panel A). The latter results from *post-mortem* deamination of 5mC. We next focused on reads mapping the (-) strand and counted the number of Phusion reads starting with CpG and TpG. These were flagged as ending with CpG and CpA when reverse completed and aligned to the (+) strand (panel B). The M_s statistics of a given region consists of the ratio of methylation-derived mis-incorporation signatures by the total number of reads starting/ending at a CpG location in hg18. Note that this measure could be extended to other sequence positions in cases ancient DNA templates show higher deamination rates than what observed on the Saggag individual (and to sequencing ends if full length sequence information is available).

We next performed similar analyses at splice sites boundaries, in an attempt to recapitulate the known enrichment in methylated Cytosines at the splice site boundary and the overall decrease in methylation rates from exons to introns [Laurent et al. 2010]. Exon coordinates of all genes from the hg18 reference genome were retrieved from UCSC Genome browser database [Meyer et al 2012] resulting in 95,746 splice sites. We calculated M_s for each position within 100 bp of each splice site. Final M_s profiles were plotted as a function of their position relative to the distance to the splice site boundary using a smoothing procedure based on 10bp long sliding windows (Figure S2.2). M_s successfully recovered the expected decline from relatively higher methylation levels in exons *versus* introns, as well as a sharp local increase in methylation at the splice site. This further confirmed M_s as a good proxy for monitoring methylation levels.

We further focused on CGIs, which function as promoters for about 60% of all human genes. Recently, Ginno et al. [2012] reported that CGI promoters showing significant strand asymmetry in the distribution of guanines and cytosines (GC skew) immediately downstream of their transcription start (TSS) are methylation-resistant. This feature capacitates the formation and the thermodynamic stability of R loops during transcription (with G-rich RNA strands reannealing to template C-rich DNA strands, forcing G-rich DNA strands into a single strand conformation) and is highly predictive of the unmethylated state of CGIs. Consequently, at CGI promoters showing high GC skew, methylation is expected to show minimal levels at TSS and to gradually increase upstream and downstream the TSS before reaching average levels ca. 1,000-2,000 bp away [Ginno et al. 2012]. This pattern is virtually absent at CGI promoters showing no strand asymmetry in the distribution of guanines and cytosines immediately downstream from their TSS. We therefore retrieved all 7,820 genomic coordinates covered by GC skew CGI promoters as defined from Ginno et al. [2012] and binned each position relative to the distance to the TSS. We then calculated Ms levels at each of those positions summing across all GC skew promoters. Final M_s profiles were plotted using a smoothing procedure based on 10bp long sliding windows (Figure S3.4). GC skew CGI promoters were found to show the expected methylation profile, with minimal methylation at TSS followed by a gradual increase upstream and downstream the TSS, suggesting that M_s provides a good measure of ancient cytosine methylation levels.



Figure S3.4

$\rm M_{s}$ methylation profiles at GC skew CGI promoters.

GC skew CGI promoters show significant strand asymmetry in the distribution of guanines and cytosines immediately downstream from their TSS [Ginno et al. 2012]. We calculated M_s values across GC skew CGIs at each position located within 2,000 bp upstream and downstream of the TSS. The final plot was generated following a smoothing procedure. A characterization of the promoters showing a pattern similar to GC skew CGI promoters is provided in Supplemental Section SI3.5 and Table S3.5.

We next followed the procedure described in Straussman et al. [2009] to predict CGIs that are protected from *de novo* methylation and that thus remain constitutively under-methylated throughout development. At such regions, M_s is expected to show lower values that at methylation-rich CGIs. The latter were defined using the data presented in Straussman et al. [2009] as CGIs showing methylation in every cell type tested. Although no specific data were available for hair shafts and hair follicles in this study, the methylation levels were monitored across a wide range of tissues as diverse as brain, liver, bone, colon, sperm and blood and CGIs showing methylation in every cell type tested were recorded. Similar levels of methylation at those CGIs are likely to be found in hairs. Predicted constitutively under-methylated CGIs represented a total number of 13,372 genomic regions while methylation-rich CGIs represented 2,583. As expected, we found a ca. 7.4-fold reduction in methylation levels for predicted under-methylated CGIs as compared to ubiquitously methylated CGIs (Figure S3.5). This again validated M_s as a good proxy for regional methylation levels.



Figure S3.5

M_s methylation levels at predicted under-methylated and ubiquitously methylated CGIs. GCIs were binned in two main classes following Straussman et al. [2009]. The first corresponds to a class of CGIs that remain significantly under-methylated during development while the second corresponds to CGIs showing significant methylation levels across a full range of somatic tissues.

Finally, we focused on two specific genes for which methylation levels have been quantified in a range of hairs sampled from human individuals at different stages of their life (from birth to >80 years). The first gene, SOX10, has been shown to reach high methylation levels within its promoter region (chr22:36,710,485-36,711,485) following two years of age [Kim et al. 2006]. Similarly, the second gene, CSX, follows similar and synchroneous age-dependent methylation changes, however reaching lower methylation levels than SOX10 at its 3'-untranslated region [Kim et al. 2006]. We therefore used M_s to calculate methylation levels within both regions. For CSX, coverage was limited, precluding restricting M_s calculation to the sole 3'-UTR. Ms was therefore calculated for a larger region (chr5:172,591,951-172,592,951) where we found evidence for methylation in both genes. We estimated ca. 18-fold greater methylation levels at SOX10 than CSX. This suggests that besides large-scale trends at different classes of genomic regions, M_s can also capture hair-specific methylation information at specific genes.

3.4. Methylation-based unsupervised hierarchical clustering

Convinced that M_s could capture genuine information regarding ancient regional methylation levels of the human genome, we next compared the methylation levels as estimated from the Saggag data to known methylation levels of different human tissues, including hairs, blood, buccal, fat, liver, muscle, omentum, pancreas and saliva [Slieker et al. 2013]. For hair, blood, buccal and saliva, those methylation levels were surveyed in five human donors (PT1, PT2, PT3, PT4 and PT5) ranging in age from 22 to 40 years. For blood, fat, liver, muscle, omentum and pancreas, we restricted our comparisons to the only two individuals from the original study surveyed for all tissues (IT13 and IT15 of age 64 and 65, respectively). This comparative dataset corresponded to normalized methylation levels at 450k CpG sites as available from the Illumina 450k array [Dedeurwaerder et al. 2011; Roessler et al. 2012]. Taking advantage of the known correlation between CpG sites located within 1,000bp-2,000 bp [Eckart et al. 2006; Down et al. 2008], we first calculated M_s for 2,000 bp-wide regions centered at each CpG. Of note, at least 50% (95%) of such regions include 124 (24) CpGs a large proportion of CpGs and are therefore suitable for M_s calculations.

Hereafter (Supplemental Sections SI3.4-SI3.7, SI4), we will refer to coverage as the total number of counts for reads starting at a CpG location within a given genomic region (N1+N2+M1+M2: see Figure S3.3). This represents the total number of reads observed within a given region that enabled us to calculate a regional M_s value for each CpG represented on the Illumina 450k array. We excluded CpG sites located on sexual chromosomes in order to avoid possible bias related to chromosomal X inactivation in females (individuals PT1, PT2 and PT3) and/or coverage differences (the Saggag individual was a male). We then filtered CpG regions for a range of minimal coverage (every 50, from 100 to 500) and tested for a possible correlation between M_s and methylation levels observed in each tissue and each individual described above. Linear models were built using the Im() function in R [R Development Core Team 2012] and adjusted-R squared values were calculated for three different models. In the first model, M_s and methylation levels known for a given individual and a given tissue were the two measurements considered ($y \sim x$). In the second model, we also introduced the coverage observed as an extra measurement, as the genomic regions considered show different ranges of CpG densities; this measurement was considered as independent from M_s (y~x+z) Finally, in the third model, this measurement was allowed to covary with M_s (y~x*z). Tables S3.1 and S3.2 report the models that received higher support in each of the tests performed together with respective adjusted-R squared values. Of note, we found high adjusted R-squared values across all tissues and individuals investigated (0.440-0.785). supporting significant correlation levels between known biological methylation data and the Saqqaq M_s levels. Importantly, the Saqqaq data showed a higher fit with models built with the methylation data originating from hair tissues (adjusted-R squared = 0.618-0.785 versus 0.440-0.778 for non-hair tissues; Tables S3.1-S3.2), in agreement with the nature of the sample used for reconstructing the Saggag genome [Rasmussen et al. 2010].

Individual	Coverage	Model	Blood	Model	Buccal	Model	Saliva	Model	Hair	Number
PT5	500	v~x*z	0.707	v~x*z	0.770	v~x*z	0.738	v~x*z	0.778	937
	450	v∼x+z	0.656	v~x+z	0.730	v~x*z	0.706	v~x*z	0.743	2139
	400	y~x+z	0.623	y∼x+z	0.694	y∼x+z	0.664	v∼x+z	0.707	4827
	350	v~x+z	0.603	v~x+z	0.666	v~x+z	0.637	v~x+z	0.675	10421
	300	y∼x*z	0.577	y∼x*z	0.636	y∼x*z	0.606	y∼x*z	0.659	20830
	250	v∼x*z	0.555	v∼x*z	0.618	v~x*z	0.588	v~x*z	0.644	390007
	200	y∼x*z	0.558	y∼x*z	0.613	y∼x*z	0.588	y∼x*z	0.625	68873
	150	y∼x*z	0.577	y∼x*z	0.623	y∼x*z	0.601	y∼x*z	0.635	114561
	100	y∼x*z	0.573	y∼x*z	0.613	y∼x*z	0.593	y∼x*z	0.621	182064
PT4	500	y∼x*z	0.717	y∼x*z	0.768	y∼x*z	0.734	y∼x*z	0.775	937
	450	y∼x+z	0.677	y∼x+z	0.715	y∼x+z	0.696	y∼x*z	0.750	2139
	400	y∼x+z	0.640	y∼x+z	0.681	y∼x*z	0.656	y∼x+z	0.710	4827
	350	y∼x+z	0.621	y∼x+z	0.658	y∼x*z	0.628	y∼x+z	0.694	10421
	300	y∼x*z	0.592	y∼x*z	0.634	y∼x*z	0.601	y∼x*z	0.672	20830
	250	y∼x*z	0.577	y∼x*z	0.621	y∼x*z	0.581	y∼x*z	0.654	390007
	200	y∼x+z	0.573	y∼x*z	0.613	y∼x+z	0.584	y∼x*z	0.633	68873
	150	y∼x*z	0.590	y∼x*z	0.622	y∼x*z	0.596	y∼x*z	0.638	114561
	100	y∼x*z	0.582	y∼x*z	0.611	y∼x*z	0.591	y∼x*z	0.627	182064
PT3	500	y∼x*z	0.698	y∼x*z	0.762	y∼x*z	0.739	y∼x*z	0.785	937
	450	y∼x+z	0.648	y∼x+z	0.715	y∼x+z	0.702	y∼x*z	0.752	2139
	400	y∼x*z	0.613	y∼x+z	0.681	y∼x+z	0.662	y∼x+z	0.709	4827
	350	y∼x+z	0.594	y∼x+z	0.658	y∼x+z	0.638	y∼x+z	0.692	10421
	300	y∼x*z	0.569	y∼x*z	0.630	y∼x*z	0.612	y∼x*z	0.669	20830
	250	y∼x*z	0.555	y∼x*z	0.615	y∼x*z	0.580	y∼x*z	0.644	390007
	200	y∼x+z	0.558	y∼x+z	0.612	y∼x+z	0.587	y∼x*z	0.635	68873
	150	y∼x*z	0.575	y∼x*z	0.619	y∼x*z	0.606	y∼x*z	0.641	114561
	100	y∼x*z	0.570	y∼x*z	0.610	y∼x*z	0.598	y∼x*z	0.624	182064
PT2	500	y∼x*z	0.691	y∼x*z	0.778	y∼x*z	0.755	y∼x*z	0.776	937
	450	y∼x+z	0.639	y∼x+z	0.720	y∼x+z	0.715	y∼x*z	0.747	2139
	400	y∼x+z	0.607	y∼x+z	0.683	y∼x+z	0.675	y∼x+z	0.706	4827
	350	y∼x+z	0.591	y∼x*z	0.635	y∼x*z	0.618	y∼x+z	0.688	10421
	300	y∼x*z	0.569	y∼x*z	0.625	y∼x*z	0.606	y∼x*z	0.665	20830
	250	y∼x+z	0.555	y∼x*z	0.606	y∼x*z	0.601	y∼x*z	0.643	390007
	200	y∼x+z	0.554	y∼x*z	0.609	y∼x*z	0.601	y∼x*z	0.633	68873
	150	y∼x*z	0.573	y∼x*z	0.624	y∼x*z	0.613	y∼x*z	0.638	114561
	100	y∼x*z	0.569	y∼x*z	0.613	y∼x*z	0.603	y∼x*z	0.622	182064
PT1	500	y∼x*z	0.705	y∼x*z	0.769	y∼x*z	0.718	y∼x*z	0.782	937
	450	y∼x+z	0.650	y∼x+z	0.703	y∼x+z	0.687	y∼x+z	0.741	2139
	400	y∼x+z	0.617	y∼x+z	0.670	y∼x+z	0.655	y∼x+z	0.703	4827
	350	y∼x+z	0.600	y∼x*z	0.646	y~x	0.542	y∼x+z	0.686	10421
	300	y∼x*z	0.575	y∼x*z	0.626	y∼x*z	0.568	y∼x*z	0.661	20830
	250	y∼x*z	0.562	y∼x*z	0.612	y∼x*z	0.537	y∼x*z	0.632	390007
	200	y∼x+z	0.560	y∼x*z	0.607	y∼x+z	0.560	y∼x*z	0.618	68873
	150	y∼x*z	0.577	y∼x*z	0.620	y∼x*z	0.553	y∼x*z	0.632	114561
	100	v~x*z	0.572	v~x*z	0.607	v~x*z	0.548	v~x*z	0.622	182064

Table S3.1

Saqqaq M_s methylation levels show stronger correlation with modern hair methylation levels than blood, buccal and saliva tissues.

We considered the normalized methylation data available from [Slieker et al. 2013] for individuals PT1, PT2, PT3, PT4 and PT5. Parameters x, y, and z are described as follows: y = observed methylation levels in a given tissue for a given modern human individual; x = Saqqaq M_s values calculated for a 2,000 bp-wide genomic region centered on a CpG site; z = coverage. The model showing the best fit was selected amongst three different models (y~x; y~x+z, and; y~x*z). Corresponding adjusted-R squared values are provided as well as the total number of CpG loci considered in each analysis.

Individual	Coverage	Model	Blood	Model	Liver	Model	Muscle	Model	Omentum	Model	Pancreas	Model	Fat	Number
IT15	500	y∼x*z	0.680	y∼x*z	0.661	y∼x*z	0.727	y∼x*z	0.714	y∼x*z	0.685	y∼x*z	0.722	937
	450	y∼x+z	0.615	y∼x+z	0.612	y∼x+z	0.686	y∼x+z	0.659	y∼x+z	0.639	y∼x+z	0.697	2139
	400	y∼x+z	0.588	y∼x+z	0.465	y∼x+z	0.626	y∼x+z	0.525	y∼x*z	0.620	y∼x+z	0.470	4827
	350	y∼x+z	0.539	y~x	0.455	y~x	0.629	y∼x+z	0.534	y∼x+z	0.601	y∼x+z	0.542	10421
	300	y∼x+z	0.482	y∼x+z	0.485	y∼x*z	0.582	y∼x*z	0.548	y∼x*z	0.482	y∼x*z	0.562	20830
	250	y∼x+z	0.498	y∼x+z	0.440	y∼x*z	0.547	y∼x*z	0.491	y∼x*z	0.486	y∼x*z	0.543	390007
	200	y∼x+z	0.496	y∼x+z	0.458	y∼x*z	0.521	y∼x*z	0.519	y∼x*z	0.521	y∼x+z	0.528	68873
	150	y∼x*z	0.506	y∼x*z	0.487	y∼x*z	0.545	y∼x*z	0.542	y∼x*z	0.543	y∼x*z	0.525	114561
	100	y∼x*z	0.499	y∼x*z	0.485	y∼x*z	0.530	y∼x*z	0.534	y∼x*z	0.541	y∼x*z	0.535	182064
IT13	500	y∼x*z	0.683	y∼x*z	0.663	y∼x*z	0.723	y∼x*z	0.703	y∼x*z	0.695	y∼x*z	0.722	937
	450	y∼x+z	0.629	y∼x+z	0.628	y∼x+z	0.682	y∼x*z	0.643	y∼x+z	0.652	y∼x+z	0.696	2139
	400	y∼x+z	0.574	y∼x*z	0.609	y∼x+z	0.623	y∼x*z	0.625	y∼x*z	0.632	y∼x+z	0.673	4827
	350	y∼x*z	0.581	y∼x*z	0.589	y~x	0.572	y∼x*z	0.593	y~x	0.616	y∼x+z	0.538	10421
	300	y∼x+z	0.518	y∼x+z	0.576	y∼x*z	0.560	y∼x+z	0.554	y∼x*z	0.584	y∼x+z	0.540	20830
	250	y∼x+z	0.481	y∼x+z	0.550	y∼x*z	0.546	y∼x*z	0.546	y∼x*z	0.571	y∼x*z	0.524	390007
	200	y∼x*z	0.503	y∼x*z	0.556	y∼x*z	0.509	y∼x+z	0.542	y∼x*z	0.568	y∼x+z	0.515	68873
	150	y∼x+z	0.515	y∼x*z	0.548	y∼x*z	0.537	y∼x*z	0.532	y∼x*z	0.580	y∼x*z	0.549	114561
	100	y∼x*z	0.515	y∼x*z	0.538	y∼x*z	0.521	y∼x*z	0.525	y∼x*z	0.573	y∼x*z	0.534	182064

Table S3.2

Levels of Saqqaq M_s methylation with the methylation levels measured in a variety of somatic tissues from modern human individuals.

We considered the normalized methylation data available from [Slieker et al. 2013] for individuals IT13 and IT15. See legends of Table S3.1 for further details.

We next performed unsupervised hierarchical clustering analyses in order to evaluate whether methylation levels as measured in the Saggag sequence data would support clustering with modern hair tissues. Analyses were first restricted to individuals PT1, PT2, PT3, PT4 and PT5 where methylation levels were surveyed across four different tissues, including hairs (this information was not available for any other individual from the original study, including IT13 and IT15) [Slieker et al. 2013]. We selected the most supported linear model described above in order to predict Saqqaq methylation values from M_s. We then selected from the Illumina 450k array data all CpG showing at least a 2-fold average difference in methylation levels between pairs of tissues (note that we also found similar clustering results while investigating a more conservative selection scheme where CpGs showing at least a 2-fold average differences between one tissue and all the other tissues altogether; Figure S3.10 and S3.11). Interestingly, CpG showing at least 2-fold higher methylation levels in modern hairs than other tissues ('High') showed greater Ms values for the Saggag than CpG showing at least 2-fold higher methylation levels in other tissues than in modern hairs ('Low'; Figures S3.6-S3-7), regardless of the threshold considered as minimal coverage (even though Figures S3.6-S3-7 refer 100 or 500 coverage thresholds, the full range of possible values from 100 to 500, iterating every 50, was investigated and resulted in similar findings; data not shown). This suggests that for those CpG, the Saggag hair M_s values better fit an expected hair methylation profile than any other tissue profile investigated. Finally, we used the subset of CpG showing at least a 2-fold average difference in methylation levels between pairs of tissues to perform unsupervised clustering using the heatmap() function from R [R Development Core Team 2012]. Predicted Saggag methylation values were ignored whenever the absolute values of residuals were greater than 0.1 and forced to zero when negative and to one when superior to one (note that similar clustering was also found in absence of filtering on residuals; data not shown). The analyses were repeated over a full range of minimal coverage (every 50, from 100 to 500), and resulted in similar outcomes. Therefore, the analyses performed with the lower coverage investigated are shown as Figure 5 (information about the specific of CpG sites considered is available upon request). For all coverage threshold

investigated, the Saggag specimen was found to cluster together with hair tissues from 5 modern human individuals, in agreement with the nature of the biological material used for reconstructing the Saggag genome [Rasmussen et al. 2010]. GREAT version 2.0.2 analysis [McLean et al. 2010], where the top-10% CpGs showing a maximal absolute methylation difference between Saggag hairs and the closest of the five modern individuals was challenged against the background of remaining CpGs, revealed no significant annotation categories related to hair biology (hypergeometric test; data not shown). Interestingly, the top-10% array CpGs that showed divergent methylation profiles apparently seemed more often located 50-500 bp away from the TSS than other array CpGs passing our filtering criteria (29.6% vs 27.5%; Figure S3.8). However, this difference was not found significant (chi-square p-value = 0.396). This suggests that the top-10% most divergent CpG sites represent a random sample of the other array CpG sites. That lower coverage did not disrupt this clustering, despite fewer chance events of observing deaminated 5mC, indicates that the *post-mortem* levels of cytosine deamination observed here are compatible with the recovery of a tissue-specific methylation proxy as long as coverage values are superior or equal to 100. Lower minimal coverage requirements were not investigated but could likely be achieved in situations where post-mortem deamination levels are higher than those observed in the Saggag individual.

Performing the full set of analyses described above but using 1,500 bp-wide regions centered at each CpG (instead of 2,000 bp), we found similar support for stronger correlations and clustering between the Saqqaq and hair methylation levels (minimal coverage investigated ranged from 100 to 400, iterating every 50; Figure S3.9). Using the same approach, we also found that the clustering of the Saggag together with modern hair tissues was maintained when considering all CpGs. regardless of possible methylation differences across tissues (one example corresponding to regions with coverage greater or equal to 100 is provided in Figure S3.12). This suggests that CpG sites showing no strong tissue specificity in their methylation patterns do not preclude genuine clustering. Finally, since M_s provides regional methylation levels as opposed to the single-base methylation values gathered from the Illumina 450K array, we calculated regional methylation values from the Illumina 450K array data. We achieved this by parsing the genomic coordinates of all CpGs from the array and collapsing those that are distant by at best 1,000bp (i.e. the physical distance used on each side of the CpG when calculating M_s values). We then calculated a regional methylation value for those CpGs by averaging normalized methylation levels. Applying a minimal coverage threshold of 100 for M_s calculations, such regions exhibited 4.76 CpGs on average (median = 4, min = 2, max = 28). Only regions showing a minimum number of 4 CpGs were considered in order to insure that the average values could represent regional methylation, thereby limiting the impact of potentially aberrant single CpGs (eg. a hyper-methylated CpG located on the shore of an otherwise unmethylated island). We then used M_s values for Saggag at the first array CpG present in a group of collapsed CpGs and performed unsupervised clustering following the same procedure as above. The results are shown in Figure S3.13. This analysis supports the clustering of the Saggag data together with modern hairs despite the presence of a fraction of loci where the Saggag exhibits estimated methylation values apparently outside the range of the variation observed amongst hair tissues of the five modern individuals analyzed here (Figure S3.13). Such loci were identified as the top-10% CpGs showing a maximal absolute methylation difference for hairs between the Saggag and the closest of the five modern individuals. GREAT analysis was performed against the background of CpGs used for clustering in order to test for possible enrichment in specific GO molecular functions (hypergeometric test) [McLean et al. 2010]. No significant GO category was identified following Bonferroni correction for multiple testing (data not shown). All in all, those analyses support the validity of our methylation estimates and demonstrate that genuine methylation
information can be recovered from past individuals using next-generation sequencing information in absence of bisulfite treatment.







Figure S3.6

Comparing methylation levels in the Saqqaq hairs at CpG showing 2-fold difference methylation in modern tissues.

We identified CpG from the Illumina 450k array showing at least 2-fold higher methylation in modern hairs than other tissues (blood, buccal or saliva; 'High') and CpG from the Illumina 450k array showing at least 2-fold lower methylation in modern hairs than other tissues ('Low'). M_s values were greater for the 'High' CpG set and lower for the 'Low' CpG set, in agreement with known methylation levels in modern hairs. We considered a minimal coverage threshold of 500. The total number of CpG regions from the array is reporting above the boxplots. Median M_s values are also indicated.

Figure S3.7

Comparing methylation levels in the Saqqaq hairs at CpG showing 2-fold difference methylation in modern tissues.

We identified CpG from the Illumina 450k array showing at least 2-fold higher methylation in modern hairs than other tissues (blood, buccal or saliva; 'High') and CpG from the Illumina 450k array showing at least 2-fold lower methylation in modern hairs than other tissues ('Low'). M_s values were greater for the 'High' CpG set and lower for the 'Low' CpG set, in agreement with known methylation levels in modern hairs. We considered a minimal coverage threshold of 100. The total number of CpG regions from the array is reporting above the boxplots. Median M_s values are also indicated.

Figure S3.8

Distribution of the absolute distance of array CpGs from TSS.

Red: top-10% CpG sites with a methylation profile divergent to modern hair methylation profiles. Black: other CpG sites. CpG sites correspond to the clustering analysis presented in Figure 5.



Unsupervised hierarchical clustering of the Saqqaq and modern human tissues based on methylation levels.

See Figure S3.10 for details, except that the regions considered were 1,500 bp-wide. A minimal coverage of 100 was required. A total number of 4,939 CpG showing at least a 2-fold difference in methylation levels between pairs of tissues for modern humans PT1, PT2, PT3, PT4 and PT5 were considered for unsupervised hierarchical clustering. The list of CpGs considered is available upon request.



Unsupervised hierarchical clustering of the Saqqaq and modern human tissues based on methylation levels.

Same as Figure 5, except that a more conservative selection scheme was considered where CpGs showing at least a 2-fold average differences between one tissue and all the other tissues. This represented a total number of 2,607 CpG sites.



Unsupervised hierarchical clustering of the Saqqaq and modern human tissues based on methylation levels.

Same as Figure S3.9, except that a more conservative selection scheme was considered where CpGs showing at least a 2-fold average differences between one tissue and all the other tissues. This represented a total number of 1,722 CpG sites.



Unsupervised hierarchical clustering of the Saqqaq and modern human tissues based on methylation levels.

Same as Figure S3.10, except that all CpGs passing coverage threshold were considered, regardless of their respective level of methylation difference across tissues. A minimal coverage threshold of 100 was applied and a total number of 45,016 CpG was considered.



Unsupervised hierarchical clustering of the Saqqaq and modern human tissues based on methylation levels.

Same as Figure S3.10, except that array CpGs located within 1,000 bp were collapsed in order to estimate regional methylation levels for modern individuals. A minimal coverage threshold of 100 was applied and a total number of 685 collapsed regions containing 4,102 CpGs was considered.

3.5. Promoter and first exon methylation levels

We downloaded from the UCSC genome browser all gene annotations available for the human reference genome hg18 and calculated the distribution of M_s values for each gene within their promoter region. Promoters were defined as the 1,000 bp located upstream of the transcription starting site (TSS). The top-1% promoters showing highest coverage but M_s values equal to zero are provided in Table S3.3. Those represent a total number of 266 UCSC candidate promoters (of which 168 are not redundant) virtually devoid of cytosine methylation in the Saggag genome (this list could be extended to 1,311 genes if the top-5% was considered instead of the top-1%). The lack of detectable methylation levels despite extensive coverage excludes promoter cytosine methylation as one important mechanism controlling the expression of those genes. Likewise, we identified the top-1% promoters showing highest methylation levels (Table S3.4). In order to correct for possible coverage issues, we first selected all genes showing a minimum coverage of 75 over their full gene body. We also required a minimum coverage of 25 for the promoter region (given the observed deamination levels, ca. 4% at read starts, this gives on average one chance to observe one CpG \rightarrow TpG conversion). We then selected genes where coverage at promoters was at least equivalent to that observed within gene bodies (correcting for length differences between both regions). Our list of 300 UCSC candidates for highest methylation levels is provided in Table S3.4 (214 are not redundant). This list likely includes a substantial fraction of loci (but not uniquely) whose expression was silenced in Saggag hairs.

Additionally, and encouraged by the overall methylation profile observed at GC skew CGI promoters (Supplementary Section S3.4; Figure S3.4), we also attempted to identify which promoters were closely matching such a pattern. M_s values were calculated within 4 successive 1kb windows located within 2kb upstream and downstream of TSS. We then identified which promoters showed minimal M_s values in the second region (i.e. the first kb upstream of TSS) and were associated with (i) at least doubled M_s values in the preceding kb, (ii) a 50% M_s increase in the third region (the first kb downstream of TSS) and (iv) with at least a further 2-fold increase in the following region. This pattern was in agreement with average methylation differences observed across those regions at GC skew CGI promoters (Figure S3.4). Requiring a minimum coverage of 25 for each of the four regions, we identified a total number of 402 promoters matching the expected profile, which represented ca. 5.6% (402/7,214) of the promoters considered (Table S3.5).

Finally, we calculated the distribution of M_s values for each gene within their first exonic region, following criteria similar to those used for identifying promoters with highest methylation levels. Interestingly, M_s values at the first exon were found to be significantly correlated with promoter M_s values (Spearman correlation coefficient = 0.318, p-value < 2.2 10⁻¹⁶), in agreement with recent reports [Brenet et al. 2011]. Given the tight association between transcriptional silencing and methylation levels at the first exon [Brenet et al. 2011], the 214 genes showing highest (top-1%) methylation in this region (159 of which are not redundant; Table S3.6) could potentially provide insights about downregulated cellular functions in the Saqqaq hairs. Potential functional insights inferred from methylation levels and nucleosomal positionning are further discussed in an independent section (SI4).

Name	UCSC	Coordinates	Cov(PM)
ZNF362	uc001bxc.1	chr1:33493761-33494761	254
AK3L1		chr1:65385474-65386474	261
SYDE2	uc009wcm.1,uc001dku.2	chr1:85439316-85440316	289
VVINT3A		CNF1:226260375-226261375	234
		chr1:229539305-229540305	242
		chr1:245161346-245162346	220
GEN1		chr2.17797895-17798895	232
SOS1		chr2:30201108_30202108	230
PRKCE		chr2:45731547-45732547	275
RTN4	uc002rvd 1	chr2:55129830-55130830	231
BCL2L11	uc002tat.1.uc002tau.1.uc002tay.1	chr2:111593962-111594962	232
DLX1	uc010faj.1.uc002uhl.1.uc002uhm.1	chr2:172657454-172658454	241
OSBPL6	uc002ulw.2,uc002ulx.1,uc002uly.1	chr2:178766620-178767620	246
GULP1	uc002ugg.1	chr2:188864804-188865804	227
SATB2	uc002uuy.1	chr2:200031064-200032064	253
FAM119A	uc002vce.2,uc010fuk.1	chr2:208198218-208199218	243
DNPEP	uc002vlf.1	chr2:219960777-219961777	235
GBX2		chr2:236741391-236742391	298
CRELD'I	UCUU3DUT. 1, UCUU3DUQ. 1, UCUU3DUN. 1, UCUU3DUI. 1	CNF3:9949506-9950506	234
SIACZ-C		CIII 3: 3982 3982 - 3982 0982	208
		chr3:50696676 50697676	250
THOC7		chr3:63824637-63825637	270
SCA7		chr3:63824273-63825273	332
ATXN7	uc003dly 2 uc010hny 1 uc003dly 2	chr3:63824273-63825273	332
CDV3	uc003epr 1	chr3 ¹ 34774991-134775991	326
TMEM175	uc010ibl.1.uc003aba.1.uc003abr.1.uc003abs.1.uc003abt.1.uc003abu.1.uc003abv.1	chr4:915262-916262	260
DKFZp547K246	uc003gbp.1	chr4:915262-916262	260
NOP14	uc003ggj.1,uc003ggk.2,uc003ggl.2,uc010icq.1	chr4:2934916-2935916	228
RGS12	uc003ggu.2,uc010ics.1	chr4:3263553-3264553	246
KLF3	uc003gtg.2,uc003gth.2	chr4:38341185-38342185	317
FAM47D	uc003hjv.1	chr4:77353217-77354217	248
PGRMC2	uc003igg.1	chr4:129428398-129429398	232
PDGFC	uc003iph.1,uc003ipi.1	chr4:158111996-158112996	251
ANKRU37		CNF4:186553834-186554834	231
PLNZ SEDS12		CIII5.5/791070-57792070	200
		chr5:122452016 122452016	229
VDAC1		chr5.133368332_133369332	263
TCF7		chr5.133478499-133479499	260
FIF4F1B		chr5 175989289-175990289	253
BMP6	uc003mxu.2	chr6:7671010-7672010	238
HS3ST5	uc003pwh.2	chr6:114770233-114771233	282
TCBA1	uc010kep.1,uc003pzp.1,uc010keq.1	chr6:124165768-124166768	323
NKAIN2	uc003pzn.1,uc003pzo.1	chr6:124165768-124166768	323
MAP3K7IP2	uc003qmj.1	chr6:149679756-149680756	228
FGFR10P	uc003qvj.1,uc003qvk.1	chr6:167331806-167332806	227
CCR6	uc003qvI.1	chr6:167331806-167332806	227
C7orf23		chr7:86686967-86687967	255
RELN		CNF7:103417198-103418198	263
KPRA1		chr7:149042035-149043035	270
WHSC1L1		chr8:38358947-38359947	260
XKR4		chr8:56176571-56177571	236
PLAG1	uc003xsr.2.uc010lvi.1	chr8:57286413-57287413	271
MAFA	uc003yyc.1	chr8:144583719-144584719	343
EEF1D	uc003yyr.1,uc003yys.1,uc003yyt.1,uc003yyu.1,uc003yyv.1	chr8:144750726-144751726	329
TIGD5	uc003yyx.1	chr8:144750364-144751364	328
SCRIB	uc003yzo.1,uc003yzp.1	chr8:144969537-144970537	355
PUF60	uc003yzq.1,uc003yzr.1	chr8:144983197-144984197	302
pp9320	uc010mtm.1	chr8:144994389-144995389	263
		cillo: 140001040-140002046	235
	ατουσεσμ.2,ατο τοπιμε. τ,ατουσετί.2 μοθοβοστί 1	chr9:36300206_36301206	204
PIP5K1R	uc004221.1,0000221.1	chr9:70509436-70510436	269
C9orf85	uc004ain.1.uc010mot.1.uc010mou.1.uc010mov.1	chr9:73715243-73716243	268
PTCH1	uc010mrg.1.uc004avl.2	chr9:97310143-97311143	285
PPP2R4	uc004bxm.1.uc004bxn.1.uc004bxo.1	chr9:130912434-130913434	240
DKFZp781M17165	uc010myr.1	chr9:130912434-130913434	240
CAMSAP1	uc004cgr.2	chr9:137938826-137939826	290
SEC16A	uc004chx.1,uc010nbo.1	chr9:138497328-138498328	357
PFKP	uc001igq.1	chr10:3099819-3100819	264
ARHGAP21	uc009xkl.1	chr10:25052175-25053175	300
		chr10:33200020-3320/028	209
FAM25A		chr10.88843033_99944033	200
		chr10.000+0303-000+4333	200
GALNAC4S-6ST		chr10.125841898-125842898	254
STK32C	uc001lld.1	chr10:133970706-133971706	368
PDDC1	uc001lrc.1,uc001lrd.2,uc001lrf.1,uc009vcg.1	chr11:767487-768487	227
PHLDA2	uc009yds.1,uc001lxa.1	chr11:2907170-2908170	263
TP53I11	uc001myi.1	chr11:44928287-44929287	325
SYT7	uc001nrv.1,uc009ynr.1	chr11:61104874-61105874	238
GPR137	uc001nzg.1	chr11:63808907-63809907	242
LRFN4	uc001ois.2,uc001oir.1	chr11:66380572-66381572	287
PITENM1		cnr11:67028416-67029416	245
		cnF12:12/09130-12//0130	229
		ohi 12.12/09130-12//0130	229
		chr12.13403/02-13404/02	230
BICD1	uc001rku,1 uc001rkv 1	chr12:32150452-32151452	263

SCN8A	uc001ryw.1	chr12:50270287-50271287	277
DKFZp761C241	uc001tdw.1	chr12:94134972-94135972	238
APAF1	uc001tfy 1 uc001tfz 1 uc001tga 1 uc001tgb 1 uc001tgc 1	chr12.97562209-97563209	238
TMEM116	uc001ttc 1 uc001ttd 1 uc001tte 1 uc001ttf 1 uc001ttb 1 uc001tti 1	chr12:110935318-110936318	240
PAR35		chr12:110038082-110030082	270
		chr12:123023116 123024116	240
		ohr12.123023110-123024110	217
		CIII 12. 123022733-123023733	317
PAMPZ		CHI12.131773205-131774205	201
TSC22D1	uc001uzo.1	chr13:44048392-44049392	288
PCDH9	uc001vik.1,uc001vil.1,uc001vin.2	chr13:66702464-66703464	237
KLF12	uc010aeq.1	chr13:73606395-73607395	245
ARHGAP5	uc001wrm.1,uc001wrn.1,uc001wro.1,uc001wrp.1	chr14:31615246-31616246	232
CCNK	uc001yqi.2,uc001yqq.2	chr14:99016492-99017492	324
EVL	uc001vat.1	chr14:99506904-99507904	263
CDC42BPB	uc001vmi 1	chr14 102593495-102594495	289
EIE5		chr14:102869469-102870469	242
CKB		chr14:102003403-102070403	38/
TMEM121		chr14:105065217-105066217	265
		obr15:24560244 24570244	200
GADRDS		clii 15.24509544-24570544	209
PARO	uco012/0.2	CIII 15:38331071-38332071	242
LOC196993	ucu02ata.2	chr15:69194893-69195893	250
PKM2	uc010bit.1	chr15:70309117-70310117	234
KIAA1024	uc002bew.1	chr15:77510913-77511913	295
AXIN1	uc002cgp.1,uc002cgg.1	chr16:342465-343465	322
SOX8	uc002ckn.1	chr16:970809-971809	314
CACNA1H	uc002cks.1.uc002ckt.1	chr16:1142242-1143242	298
ADCY9	uc002cvx 1	chr16:4106187-4107187	300
DKF7n564M1672		chr16:4792164-4793164	235
LIBN1		chr16:4836913-4837913	396
MAP1LC3B		chr16:85082320_85083320	250
SPC7		obr16:99101306 99103306	230
3567		CIII 10.00101300-00102300	2/1
ZNF276	UC002f0s.2	CNF16:88314453-88315453	322
ZMYND15	uc002tyt.1	chr17:4589086-4590086	270
KCTD11	uc002gge.2	chr17:7194932-7195932	232
MYH10	uc002gll.1,uc002glm.1,uc010cnx.1	chr17:8474761-8475761	239
ZSWIM7	uc002gpe.1,uc002gpf.1	chr17:15843731-15844731	263
CDK5R1	uc002hhn.1	chr17:27837218-27838218	392
SMARCD2	uc010dea.1	chr17:59273558-59274558	249
MESD11	uc002itd 2 uc010dbb 1 uc002ite 1	chr17.72244723-72245723	243
USP36	uc002iwa 1	chr17.74348204-74349204	266
CBX8		chr17:75385485-75386485	250
RAC3		chr17:77581821_77582821	334
DENC		obr17:77602020 77602020	224
REING		chi 17.77002939-77003939	234
ASALS		CIII 18:29411539-29412539	247
SMADZ		CIII 18.437 10924-437 11924	300
DOK6		CNF18:65218271-65219271	247
NET01	uc002lkw.2,uc002lky.1	chr18:68685790-68686790	290
I XNL4A	uc010drg.1	chr18:75894903-75895903	309
ADAMTSL5	uc002ltd.1	chr19:1464019-1465019	247
PLK5	uc002ltg.1	chr19:1474958-1475958	332
SF3A2	uc002lvg.1	chr19:2186816-2187816	247
SAFB2	uc002mcd.1	chr19:5573938-5574938	289
NFIX	uc002mwd.1	chr19:12966584-12967584	230
CCNF1	uc002nsn 1	chr19:34993741-34994741	265
TSHZ3		chr19:36532030-36533030	274
CERPA		chr10:38/85160_38/86160	360
		chr10:46461826-46462826	230
		chr10:60461950 60462950	233
5AF31		chi 19.00401000-00402000	233
LUC284296	uccouzqkz.z	CIII 19:00030112-00037112	234
SIRPA	ucuuzwits.1,ucuuzwitt.1	CNF2U:1822826-1823826	230
ADRA1D	uc002wkr.2	chr20:4177659-4178659	273
FOXA2	uc002wsm.1	chr20:22513101-22514101	238
C20orf24	uc002xfo.1	chr20:34635370-34636370	242
DBNDD2	uc002xoe.1	chr20:43467932-43468932	246
CTSA	uc002xqk.2	chr20:43952607-43953607	242
PARD6B	uc002xvo.1	chr20:48780488-48781488	266
LSM14B	uc010aiz.1	chr20:60130588-60131588	342
SLC2A4RG	uc002var 1	chr20:61841103-61842103	383
TIAM1	uc002vow 1	chr21:31853161-31854161	233
SERS15		chr21.32026133_32027133	249
KIAA1172	u = 0.02 ypc. 1, $u = 0.02$ ypc. 1, $u = 0.02$ ypc. 1, $u = 0.02$ ypc. 1	chr21.32020133-32027133	240
		GIIZ 1.32020133-3202/133	249
AGPAIS		CITE 1:44 108544-44 109544	332
		CNF22:17/99036-17800036	310
ZNRF3	ucuu3aeg.2	cnr22:27608890-27609890	266
CARD10	ucuu3asx.1,ucuu3asy.1	cnr22:36245170-36246170	285

Top-1% mostly covered Saqqaq promoters virtually devoid of cytosine methylation.

Name	UCSC	Coordinates	Cov(PM)	Ms
TTLL10	uc001acv.1	chr1:1098146-1099146	76	0.105
		chr1:12004477-12005477	53	0.113
		chr1:17166285-17167285	30	0.111
		chr1:21749394-21750394	38	0.122
MAP3K6	uc001bnz.1	chr1:27560945-27561945	38	0.105
UBE2U	uc001dbn.1	chr1:64441078-64442078	58	0.138
GPR61	uc001dxy.2	chr1:109883017-109884017	27	0.148
HORMAD1	uc001evk.1,uc001evl.1,uc001evm.1	chr1:148959976-148960976	52	0.115
MUC1	uc001fhz.1	chr1:153428022-153429022	29	0.103
CACNA1E	uc001gox.1,uc009wxt.1	chr1:179967143-179968143	36	0.111
		CNF1:203293356-203294356	44	0.114
OBSCN		chr1:226461484-226462484	27 48	0.105
COLEC11	$\mu c 002 q x z = 1 \ \mu c 002 q x a = 1 \ \mu c 002 q x b = 1 \ \mu c 002 q x c = 1 \ \mu c 010 e w c = 1$	chr2:3619512-3620512	47	0.104
ALLC	uc010ewt.1	chr2:3682661-3683661	47	0.128
ADCY3	uc002rfr.2	chr2:24918040-24919040	55	0.109
ABCG5	uc002rtn.1,uc002rto.1,uc002rtp.1	chr2:43919462-43920462	29	0.207
ACTG2	uc010fex.1,uc002sjw.1,uc010fey.1	chr2:73972601-73973601	27	0.111
CYP27C1	uc002tod.2	chr2:127679813-127680813	58	0.103
RAB3GAP1	ucou2tuj.1,uco10fnt.1	chr2:135525323-135526323	36	0.111
DRFZP434A012 DCKD		chr2.233001626-233002626	26	0.111
KI HI 30		chr2:238713156-238714156	58	0.113
FLJ00133	uc002wak.2	chr2:241652388-241653388	60	0.117
C2orf85	uc010fzu.1	chr2:242459559-242460559	67	0.104
C3orf32	uc003bqz.1	chr3:8761726-8762726	34	0.118
VGLL4	uc010hdv.1	chr3:11585398-11586398	36	0.139
CAND2	uc003bxk.1	chr3:12831873-12832873	34	0.118
FGD5		CNF3:14834810-14835810	28	0.107
JESSD2		chr3:46850580-46851580	20 38	0.179
KIAA0540		chr3.47020679-47021679	45	0.103
RNF123	uc010hkv.1	chr3:49710905-49711905	25	0.120
NT5DC2	uc003dem.1	chr3:52537822-52538822	52	0.154
MORC1	uc003dxl.1	chr3:110319658-110320658	76	0.132
IGSF11	uc003eby.1,uc003ebz.1,uc010hqs.1	chr3:120347588-120348588	57	0.123
SLC41A3	uc003eii.1	chr3:127258308-127259308	39	0.154
		chr3:128393664-128394664	/6	0.118
		chr3:105553350 105554350	47	0.100
TNK2		chr3:197097073-197098073	27	0.125
KIAA1530	uc010iby.1	chr4:1348865-1349865	32	0.156
TACC3	uc010ica.1	chr4:1701697-1702697	28	0.143
ZFYVE28	uc003gew.1	chr4:2312606-2313606	78	0.154
RGS12	uc003ghb.2	chr4:3383754-3384754	45	0.133
TBC1D14	uc003gju.2	chr4:7052696-7053696	41	0.122
ACOX3	ucuu3gle.1	CNr4:8468626-8469626	48	0.125
C4orf32		chr4:113285002-113286002	20 38	0.107
SI C6A18		chr5·1277470-1278470	61	0.105
CLPTM1L	uc003ica.1	chr5:1394975-1395975	51	0.137
ANKH	uc003jfl.2	chr5:14799111-14800111	29	0.172
LOC153328	uc003laz.1,uc003lba.2	chr5:135197264-135198264	28	0.107
CAMK2A	uc003lrt.1,uc003lru.1,uc010jhe.1	chr5:149649529-149650529	29	0.103
	uc010jhl.1,uc010jh0.1	chr5:150423501-150424501	39	0.128
1 NIP 1 AK 127224	uconogmil 1	chr5:177479705-177480705	39	0.128
KIAA0676		chr5:179230101_179231101	38	0.100
IGnT2	uc010iol.1	chr6:10599442-10600442	68	0.103
AX747210	uc003nak.1	chr6:13403797-13404797	28	0.107
GRM4	uc003oio.1,uc010jvj.1	chr6:34138994-34139994	26	0.115
TCP11	uc003okd.2,uc003oka.2,uc003okb.2,uc003okc.2	chr6:35217165-35218165	118	0.127
IFEB	uc003oqu.1,uc003oqy.1	chr6:41811975-41812975	29	0.138
		CIII / 3054083-855683	31	0.129
C7orf27	ucoosik. i uc003smh 2	chr7:2548679-2549679	40 94	0.150
WIPI2	uc003snz.2.uc003soa.1	chr7:5219426-5220426	37	0.120
PON1	uc003uns.1	chr7:94791780-94792780	34	0.118
AZGP1	uc003ush.1	chr7:99411623-99412623	31	0.129
FLJ23834	uc003vdl.2,uc003vdm.2	chr7:105389921-105390921	27	0.148
ASZ1	uc003vjb.2	chr7:116854813-116855813	67	0.104
		CNF7:142203493-142204493	30	0.139
		chr8:27343310-27344310	93	0.100
CLU		chr8:27525085-27526085	35	0.113
PTP4A3	uc003ywg.1,uc003ywh.1	chr8:142500189-142501189	85	0.106
hPRL-3	uc010met.1	chr8:142500189-142501189	85	0.106
SPATC1	uc003zaq.1	chr8:145157595-145158595	26	0.115
WNK2	uc004atl.1	chr9:95089891-95090891	26	0.115
CORO2A	uc004ayl.1	cnr9:99974995-99975995	45	0.111
ANKSO	UCUU4ayV.1	CDF9:1005/955/-100580557	29	0.241
	ucoo4big. i ucoo4bra 1	chr9.120206207_120207207	20 20	0.115
SH2D3C	uc004brz 2	chr9:129564518-129565518	37	0.216
ODF2	uc004bvf.1	chr9:130261650-130262650	26	0.115
KIAA0515	uc004cao.2	chr9:133334996-133335996	41	0.122
DBH	uc004cel.1	chr9:135490306-135491306	38	0.105
FLJ00280	uc004cfd.1	chr9:136447825-136448825	44	0.114
MAN1B1		cnr9:139116621-139117621	41	0.122
NFLF	ucoo4cmj. 1,ucoo4cmk. 1,ucoo4cmi. 1,uco 10nca. 1 uc010nci 1	chr9.139204001-139200051 chr9.139470759-139471759	38	0.108
	400 10101.1	0110.1004/0100-1004/11/08	50	0.100

ERCC6	uc009xod.1	chr10:50354362-50355362	28	0.107
LDB3	uc001kdr.1,uc001kds.1,uc009xsy.1,uc001kdu.1,uc001kdv.1	chr10:88417301-88418301	31	0.161
KIAA0613	uc009xsz.1	chr10:88417301-88418301	31	0.161
KCNK18	uc001ldc.1	chr10:118945990-118946990	25	0.120
C10orf141	uc001lju.1,uc009yap.1	chr10:128864690-128865690	30	0.167
INPP5A	uc001llg.1	chr10:134270409-134271409	121	0.116
FLJ00228	uc001lov.2	chr11:282823-283823	48	0.104
EFCAB4A	uc001lrw.2	chr11:818297-819297	60	0.117
KIAA0899	uc009vca.1	chr11:981000-982000	50	0.140
HCCA2	uc001lto 1 uc001ltp 1	chr11.1458691-1459691	65	0 108
TNNT3	uc001lun 2 uc001luu 2 uc001lun 2 uc001luw 2 uc001luo 2 uc001lug 2	chr11.1896375-1897375	53	0 113
C11orf21		chr11:2279866-2280866	53	0 113
APBR1	μ_{c}	chr11:6382668-6383668	49	0 122
NR1H3		chr11:47226043-47227043	46	0.122
TNKS1RD1		chr11:56827520_56828520	35	0.100
AR420224		chr11:64417407 64419407	50	0.114
EL 100043		chr11:65104605 65105605	34	0.119
CARD2		chr11:67047475 67049475	56	0.110
		chr11:60005572 60006572	24	0.143
BUIZ/ 192		CIII 1 1.09995572-09990572	34	0.118
		CHITT.73033330-73030330	37	0.100
CAPN5		CNF11:76507101-76508101	45	0.156
FATUZ		CITITITITZ00009-117201009	50	0.120
IFFU1	ucourdoz.1,ucourdpa.1	CNF12:0528535-0529535	50	0.120
HOM-TES-103	ucu01qpg.2	chr12:6528535-6529535	50	0.120
LAG3	uc001qqs.2,uc001qqt.2,uc001qqu.2	chr12:6750931-6751931	30	0.133
C1RL	uc001qsn.1,uc009zft.1,uc001qso.1	chr12:7153069-7154069	25	0.120
STYK1	uc001qys.1	chr12:10717906-10718906	64	0.125
COL2A1	uc009zkw.1	chr12:46668521-46669521	36	0.111
POU6F1	uc001rxy.1,uc001rxz.1	chr12:49877085-49878085	64	0.141
FIGNL2	uc001rzc.1	chr12:50502475-50503475	25	0.120
NR4A1	uc001rzr.2,uc009zmc.1	chr12:50722784-50723784	32	0.125
GEFT	uc009zpy.1,uc001soz.1	chr12:56289230-56290230	55	0.109
BC061638	uc009zrx.1	chr12:72972678-72973678	70	0.114
NOS1	uc001twm.1	chr12:116283965-116284965	26	0.192
MLXIP	uc001ubt.1	chr12:121182783-121183783	26	0.192
limkain	uc001ufy 1	chr12.122996061-122997061	35	0 1 1 4
KIAA1517		chr12.124003069-124004069	27	0 222
PGAM5		chr12:131800517-131801517	27	0 111
DNA IC15		chr13:42494362-42495362	26	0.192
		chr13:113155840-113156840	47	0.102
		chr13:113360502-113361502	107	0.100
C14orf104		obr14:40171609 40172609	26	0.140
		obr14:70706170 70707170	20	0.104
		chi 14.72790170-72797170	30	0.105
KIAA 1509		chi 14:908 19720-90820720	20	0.120
SLC24A4		CNF14:91974432-91975432	31	0.129
SLC25A29	uculoavv.1	CNF14:99831824-99832824	26	0.115
PPP2R5C	uc001ykg.1	chr14:10141/251-101418251	35	0.114
AHNAK2	ucu01ypx.2	chr14:104507865-104508865	36	0.111
FAM82A2	uc001zmq.1	chr15:38835341-38836341	29	0.103
MAPKBP1	uc010bcl.1	chr15:39891006-39892006	28	0.107
CAPN3	uc001zpq.1	chr15:40480880-40481880	38	0.105
BCL2L10	uc002abq.1	chr15:50192264-50193264	38	0.105
TIPIN	uc002apr.2	chr15:64436108-64437108	57	0.105
NR2E3	uc002ath.1,uc002ati.1	chr15:69888948-69889948	30	0.133
SEMA4B	uc010bnv.1	chr15:88564850-88565850	34	0.118
CLCN7	uc002clu.2	chr16:1439799-1440799	140	0.107
IFT140	uc002clz.1	chr16:1548136-1549136	43	0.116
DNASE1	uc002cvr.1	chr16:3641941-3642941	34	0.118
JMJD5	uc010bxw.1	chr16:27127915-27128915	28	0.107
MYST1	uc002eba 2	chr16:31045011-31046011	33	0 121
ABCC12	uc002efe 1	chr16:46747430-46748430	54	0 130
SNX20	uc002eqi 2 uc002eqk 1	chr16:49272667-49273667	28	0 143
EDC4		chr16:66467893-66468893	38	0 105
BCAR1		chr16:73827902-73828902	97	0.165
FAM38A	ucol2ftr 2	chr16:87330160-87331160	110	0.136
Mib		chr16:97330160 97331160	110	0.100
AK005795		chr16:99406950 99407950	37	0.100
		chr16:00526021 00527021	47	0.100
	100021pt. 1	chr16:88556740 99557740	41 50	0.149
		chil 10.00000749-00007749	55	0.132
		chi 17.3007023-3000023	21	0.111
ALUXIS		CIII 17.4491709-4492709	34	0.110
DHX33		CNF17:5305407-5306407	25	0.120
CACNB1	ucuu2hri.1	chr17:34603662-34604662	34	0.118
CALCOCO2	uc002iof.1	chr17:44262371-44263371	66	0.106
CACNA1G	uc002irx.1,uc002iry.1,uc002irz.1,uc002isa.1,uc002isb.1,uc002isc.1,uc002isd.1	chr17:46000230-46001230	32	0.188
	uc002ise.1,uc002isf.1,uc002isa.1.uc002ish.1.uc002isi.1			
6DK2		obr17:69045502 60046502	22	0 404
SUKZ		CHI 17:00940003-00940003	33	0.121
		chi 17.7 1907597-7 1908597	25	0.120
AN 120072		CHI 17.7 3044514-73045514	50	0.120
RINE213		CHF17:75927321-75928321	59	0.136
AK098403	ucu10dhy.1	cnr17:76701355-76702355	67	0.104
SLC38A10	uc002jzy.1	cnr17:76874109-76875109	65	0.108
C17orf70	uc002kao.1	chr17:77125977-77126977	44	0.114
FN3K	uc002kfw.1	chr17:78287866-78288866	55	0.127
LOXHD1	uc002lcf.2	chr18:42393250-42394250	48	0.104
MOBKL2A	uc002luu.1	chr19:2029718-2030718	90	0.133
TMPRSS9	uc002lvv.1,uc002lvw.1	chr19:2339784-2340784	29	0.138
TBXA2R	uc002lye.1	chr19:3551665-3552665	27	0.111
FSD1	uc002maa.1	chr19:4261593-4262593	43	0.116
ALKBH7	uc002meo.1	chr19:6322444-6323444	29	0.172
KLF1	uc002mvo.1	chr19:12859017-12860017	28	0.143
CC2D1A	uc002mxa.1	chr19:13889317-13890317	41	0.146
INSL 3	uc002nhm.1.uc010ebf 1 uc010ebg 1	chr19:17793320-17794320	31	0 129
KIAA0892	uc010ece 1	chr19:19318459-19319459	33	0 152
GRAMD14		chr19:40176528_40177528	35	0.102
SI AND IA		01110.70170020-40177020	55	0.114

	up002pmf 2	abr10-54258026 54250026	21	0 161
	ucoo2pmt 1	chr10:54236930-34239930	20	0.101
	ucoo2print. I	CIII 19.0400070-040070	29	0.103
ASPDH	ucouzpsi.z	CIII 19:557/09759-557 10759	12	0.222
KLK11		CNF19:56223102-56224102	27	0.148
SIGLEC12	UCUU2pwx.1	CNF19:56696855-56697855	35	0.171
UNQ9215	ucu10eoy.1	chr19:56696855-56697855	35	0.171
LILRA2	uc002qqq.2,uc010ero.1	chr19:59776110-59777110	26	0.154
KIAA1398	uc010gcm.1	chr20:17565374-17566374	42	0.119
SGK2	uc002xkv.1	chr20:41627151-41628151	33	0.121
TNFRSF6B	uc002yfy.1	chr20:61795538-61796538	45	0.178
AIRE	uc002zej.1,uc010gpr.1	chr21:44533576-44534576	62	0.113
APECED	uc010gpq.1	chr21:44533576-44534576	62	0.113
C21orf2	uc010gps.1	chr21:44578835-44579835	46	0.109
COL18A1	uc002zhk.1	chr21:45750842-45751842	45	0.111
ARVCF	uc002zra.1	chr22:18358335-18359335	31	0.129
KIAA0330	uc010aul.1	chr22:22896681-22897681	27	0.111
CRYBB1	uc003acv.1	chr22:25343991-25344991	33	0.121
CYB5R3	uc003bcw.2	chr22:41362840-41363840	27	0.111
PARVB	uc010gzn.1	chr22:42752517-42753517	37	0.108
TBC1D22A	uc003bif.1	chr22:45547488-45548488	26	0.115
C22orf34	uc003bit 2	chr22:48437194-48438194	45	0 111
TRABD	uc003biy 1 uc003biw 1	chr22:48972570-48973570	72	0 111
d.I402G11 5	uc010han 1 uc003hiv 1	chr22:48985873-48986873	77	0 117
MAPK12	uc003hkn 1	chr22:49036709-49037709	41	0 146
FINA		chrX:153235605-153236605	45	0 111
	0000+INI.Z	01173-10020000-100200000	75	V.111

Top-1% methylated Saqqaq promoters.

Ms(-2000;-1000)	Cov.	Ms(-1000;TSS)	Cov.	Ms(TSS;1000)	Cov.	Ms(1000;2000)	Cov.	TSS
0.1200	25	0.0000	25	0.0256	31	0.0909	36	chr16:66075217
0.0250	40	0.0000	26	0.0227	39	0.0800	54	chr11:60450096
0.0250	40	0.0000	26	0.0227	39	0.0800	54	chr11:60450096
0.0682	44	0.0000	27	0.0328	88	0.1111	34	chr12:107510288
0.0294	34	0.0000	29	0.0078	81	0.0588	29	chr11:62363707
0.0750	40	0.0000	29	0.0204	46	0.0784	44	chr14:23/23809
0.0280	35 27	0.0000	29	0.0526	35	0.1111	33 37	chr2:96980965
0.0638	47	0.0000	30	0.0693	77	0.1429	33	chr1:202493620
0.0638	47	0.0000	30	0.0693	77	0.1429	33	chr1:202493620
0.0638	47	0.0000	30	0.0693	77	0.1429	33	chr1:202493620
0.0638	47	0.0000	30	0.0693	77	0.1429	33	chr1:202493620
0.0638	47	0.0000	30	0.0693	//	0.1429	33 22	chr1:202493620
0.0638	47	0.0000	30	0.0693	77	0.1429	33	chr1:202493620
0.0286	35	0.0000	31	0.0194	64	0.0758	85	chr19:4509501
0.0968	31	0.0000	31	0.0114	103	0.0513	27	chr19:53663423
0.1200	50	0.0000	31	0.0221	162	0.0579	76	chr7:719269
0.0571	35	0.0000	32	0.0208	45	0.0500	6Z 49	CNF21:44012431
0.0270	55	0.0000	33	0.0213	53	0.0521	80	chr19:5885473
0.0492	61	0.0000	33	0.0323	40	0.0714	59	chr21:42578058
0.0714	42	0.0000	34	0.0222	43	0.1136	50	chr14:72797810
0.0667	45	0.0000	35	0.0120	56	0.0300	63	chr17:78634237
0.0294	34 34	0.0000	37	0.0417	34 34	0.1304	33	CDF19:38052523
0.0294	34	0.0000	37	0.0417	34	0.1304	33	chr19:38052523
0.0200	50	0.0000	37	0.0146	185	0.0400	57	chr7:733813
0.0571	35	0.0000	38	0.0081	79	0.0588	27	chr10:103805922
0.0571	35	0.0000	38	0.0081	79	0.0588	27	chr10:103805922
0.0323	31	0.0000	38	0.0128	141	0.0400	31 21	Chr15:8152/110
0.0323	25	0.0000	38	0.0128	88	0.0400	49	chr19:1388362
0.0714	28	0.0303	38	0.0645	38	0.1321	50	chr8:21957128
0.0220	91	0.0000	40	0.0189	51	0.0758	65	chr16:66255916
0.0364	55	0.0000	41	0.0375	67	0.0851	80	chr11:1811609
0.0339	59	0.0000	41	0.0467	/0	0.10/1	59	chr9:129205140
0.0520	57	0.0000	42	0.1369	52	0.3333	20	chr19:56583009
0.0185	54	0.0000	44	0.0286	49	0.1000	39	chr12:50872051
0.0185	54	0.0000	44	0.0286	49	0.1000	39	chr12:50872051
0.0339	59	0.0000	44	0.0147	72	0.0625	45	chr12:54902002
0.0339	59	0.0000	44	0.0290	74	0.0845	67	chr19:5773817
0.0189	106	0.0000	44	0.0278	59 42	0.0909	44	CNr1:55236204
0.0323	31	0.0000	44	0.0148	110	0.0536	74	chr2:45021540
0.0211	95	0.0000	45	0.0800	49	0.1818	47	chr11:125645049
0.0577	52	0.0000	45	0.0044	146	0.0449	80	chr19:19600739
0.0303	33	0.0000	46	0.0098	66	0.0714	27	chr11:62363552
0.0606	22	0.0192	40	0.0606	35	0.1538	25 71	CDF3:12/358216 cbr2:128118155
0.0667	45	0.0000	47	0.0106	129	0.0959	69	chr4:969784
0.0667	45	0.0000	47	0.0106	129	0.0959	69	chr4:969784
0.0588	51	0.0000	48	0.0625	62	0.1553	81	chr16:73830059
0.1154	78	0.0000	48	0.0149	66	0.0645	48	chr16:88604030
0.0444	45	0.0000	40	0.0129	98 62	0.0380	04 30	chr7:150283848
0.0847	59	0.0204	48	0.0390	62	0.0833	30	chr7:150283848
0.0645	62	0.0000	49	0.0421	82	0.0860	82	chr22:16654433
0.0238	84	0.0000	50	0.0139	69	0.1000	49	chr12:120740759
0.0385	26	0.0000	50	0.0071	143	0.0588	34	chr16:630849
0.0814	80	0.0000	50	0.0077	106	0.0748	52	chr17:59361606
0.0625	32	0.0000	50	0.0035	156	0.0741	26	chr5:175724107
0.0625	32	0.0000	50	0.0035	156	0.0741	26	chr5:175724107
0.0345	58	0.0000	51	0.0175	63	0.0556	46	chr12:54901971
0.0345	58	0.0000	51	0.0175	63	0.0556	46	chr12:54901971
0.0345	58	0.0000	51	0.0175	63	0.0556	40	chr12:54901971 chr12:54901971
0.0789	38	0.0000	51	0.0056	115	0.1053	29	chr17:7078587
0.0789	38	0.0000	51	0.0056	115	0.1053	29	chr17:7078587
0.0233	86	0.0000	51	0.0278	57	0.0641	82	chr3:130760230
0.0385	26	0.0000	53	0.0066	169	0.0292	81	chr3:185579554
0.0385	26	0.0000	53	0.0066	169	0.0292	81 91	CDF3:185579554
0.0385	26	0.0000	53	0.0066	169	0.0292	81	chr3:185579554
0.0800	25	0.0000	54	0.0155	118	0.0604	81	chr9:20612514
0.0800	25	0.0000	54	0.0155	118	0.0604	81	chr9:20612514
0.1304	69	0.0000	55	0.0128	75	0.1176	45	chr15:38848469
0.0976	41	0.0000	55	0.0252	106	0.0833	40 34	chr4:6061071
0.0233	86	0.0000	57	0.0079	175	0.0435	79	chr11:65413450
0.0625	32	0.0000	57	0.0056	108	0.0588	64	chr16:2194450
0.0625	32	0.0000	57	0.0056	108	0.0588	64	chr16:2194450
0.0625	32	0.0000	57	0.0056	108	0.0588	64	chr16:2194450
0.0625	32 32	0.0000	57 57	0.0056	108	0.0588	64 67	chr16:2194450 chr16:2104450
0.0625	32	0.0000	57	0,0056	108	0.0588	64	chr16:2194450
0.0759	79	0.0000	57	0.0244	67	0.0741	57	chr21:44596911
0.0759	79	0.0000	57	0.0244	67	0.0741	57	chr21:44596911
0.0625	64	0.0000	57	0.0069	124	0.4286	48	chr4:4279522
0.0270	57	0.0000	20	0.0070	00	1000/	20	01119:40/91510

0.0893 56 0.0000 60 0.0156 56 0.1053 34 chr19:45 0.0893 56 0.0000 60 0.0156 56 0.1053 34 chr19:45 0.0103 97 0.0000 62 0.0118 95 0.0714 62 chr16:19	773629 773629 440951
0.0103 97 0.0000 62 0.0118 95 0.0714 62 chr16.19	440951
	4400E1
0.0105 37 0.0000 62 0.0118 35 0.0114 62 0.0119	966172
0.0465 43 0.0114 62 0.0294 40 0.0714 30 chr1:151 0.0465 43 0.0114 62 0.0294 40 0.0714 30 chr1:151	966190 966190
0.0465 43 0.0114 62 0.0294 40 0.0714 30 chr1:151	966190
0.0388 34 0.0000 63 0.0000 100 0.0383 65 cm10.2 0.0189 53 0.0000 63 0.0068 94 0.0909 33 chr19:48	791330
0.0400 25 0.0000 63 0.0118 94 0.0357 26 chr2:11 0.0250 40 0.0000 64 0.0171 100 0.0500 33 chr19:51	211990 941560
0.0250 40 0.0000 64 0.0171 100 0.0500 33 chr19:51	941560
0.0323 31 0.0000 64 0.0189 47 0.0556 33 chr19:55	062325
0.0093 107 0.0000 65 0.0244 88 0.1200 73 $chr11110.0429$ 70 0.0135 67 0.0400 46 0.1111 39 $chr19:45$	774302
0.0517 58 0.0000 67 0.0227 56 0.0714 45 chr19:54 0.0517 58 0.0000 67 0.0227 56 0.0714 45 chr19:54	094380 094380
0.0128 78 0.0000 69 0.0073 184 0.0159 116 chr19:55 0.0256 78 0.0000 69 0.0151 164 0.0444 56 chr21:27	524446 260703
0.0313 160 0.0000 69 0.0291 153 0.1047 113 chr7:27 0.0208 48 0.0000 70 0.0079 83 0.0313 62 chr2:202	152306
0.0208 48 0.0000 70 0.0079 83 0.0313 62 chr2:202	948294
0.0208 48 0.0000 70 0.0079 85 0.0515 62 cli2.202 0.0789 38 0.0000 71 0.0036 149 0.0909 30 chr17:14	412860
0.0789 38 0.0000 71 0.0036 149 0.0909 30 chr17:10 0.0789 38 0.0000 71 0.0036 149 0.0909 30 chr17:10	412860 412860
0.0698 43 0.0116 71 0.0263 56 0.0588 35 chr19:55 0.0698 43 0.0116 71 0.0263 56 0.0588 35 chr19:55	012357 012357
0.0213 47 0.0000 72 0.0050 126 0.0930 51 chr17:71	648240
0.0342 117 0.0000 73 0.0053 153 0.0327 131 chr19:13	221259
0.0230 87 0.0000 74 0.0123 124 0.1071 58 chr17:18 0.0230 87 0.0000 74 0.0123 124 0.1071 58 chr17:18	102780
0.0323 31 0.0080 74 0.0179 65 0.0627 157 chr19:43 0.0323 31 0.0080 74 0.0179 65 0.0627 157 chr19:43	570508 570508
0.0286 35 0.0000 74 0.0213 40 0.0714 28 chr19:50 0.0667 45 0.0000 74 0.0034 164 0.0566 51 chr1:24	517974 51544
0.0645 31 0.0000 75 0.0083 70 0.0608 142 chr19:7 0.0238 42 0.0000 75 0.0125 155 0.0807 54 chr3:108	550706
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0.0667 45 0.0082 76 0.0263 60 0.0741 37 chr22:22 0.0806 62 0.0000 77 0.0050 228 0.0116 163 chr20:62	423279
0.0217 92 0.0000 78 0.0073 177 0.0152 68 chr11:32 0.0217 92 0.0000 78 0.0073 177 0.0152 68 chr11:32	413663 413663
0.0217 92 0.0000 78 0.0073 177 0.0152 68 chr11:32 0.0217 92 0.0000 78 0.0073 177 0.0152 68 chr11:32	413663
0.0217 92 0.0000 78 0.0073 177 0.0152 68 chr11:32 0.0600 58 0.0213 78 0.0073 177 0.0152 68 chr11:32	413663
0.0625 80 0.0156 79 0.0253 92 0.0870 66 chr1:35	96095
0.0625 80 0.0156 79 0.0253 92 0.0870 66 chr1:35 0.0625 80 0.0156 79 0.0253 92 0.0870 66 chr1:35	96095
0.0091 110 0.0000 79 0.0143 78 0.0370 56 chr1:153 0.0091 110 0.0000 79 0.0143 78 0.0370 56 chr1:153	412693 412693
0.0091 110 0.0000 79 0.0143 78 0.0370 56 chr1:153	412693
0.0606 33 0.0000 82 0.0091 61 0.0608 142 chr19:7	50760
0.0400 25 0.0000 82 0.0091 61 0.0008 142 ch19.7 0.0400 25 0.0000 83 0.0028 176 0.0129 107 chr16.88	313893
0.0400 25 0.0000 83 0.0028 176 0.0129 107 chr16:88 0.0400 25 0.0000 83 0.0028 176 0.0129 107 chr16:88	313893
0.0400 25 0.0000 83 0.0028 176 0.0129 107 chr16:88 0.0308 65 0.0000 83 0.0056 102 0.0588 34 chr1:37	313893 63657
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0.0370 27 0.0080 84 0.0128 134 0.0690 30 chr19:50	571463
0.0071 141 0.0000 85 0.0227 89 0.0667 87 chr15:43	192815
0.0071 141 0.0000 85 0.0227 89 0.0667 87 chr15:43 0.0233 86 0.0000 87 0.0090 139 0.0182 69 chr11:12	3068198
0.0233 86 0.0000 87 0.0090 139 0.0182 69 chr11:12 0.0215 93 0.0000 87 0.0056 144 0.0182 73 chr12:10	3068198 1874581
0.0286 70 0.0115 87 0.0244 59 0.0667 105 chr16:6 0.0328 61 0.0064 87 0.0112 125 0.0909 30 chr16:2	20012
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0.0140 143 0.0000 89 0.0441 117 0.1250 86 chr5:141 0.0130 154 0.0000 89 0.0400 72 0.1395 84 chr5:176	040593 875428
0.0189 53 0.0000 90 0.0220 71 0.0882 47 chr17:18 0.0566 53 0.0068 90 0.0117 134 0.0417 40 chr17:18	102595
0.0566 53 0.0068 90 0.0117 134 0.0417 40 chr18:55	091605
0.1200 25 0.0000 91 0.00/0 14/ 0.041/ 32 chr5:179 0.0357 28 0.0000 92 0.0092 71 0.0244 33 chr16:4	792455
0.0444 45 0.0000 93 0.0150 76 0.0370 25 chr16:20 0.0290 69 0.0000 94 0.0045 126 0.0300 74 chr11:5	872318 49957
0.0087 115 0.0000 94 0.0769 64 0.1667 56 chr15:41 0.0087 115 0.0000 94 0.0769 64 0.1667 56 chr15:41	449550 449550
0.0087 115 0.0000 94 0.0769 64 0.1667 56 chr15:41	449550
0.0909 44 0.0070 94 0.0212 145 0.0465 34 chr9:139	152429

0 000	11	0.0070	94	0.0212	145	0.0465	34	chr0:130152/20
0.0909	44	0.0070	94	0.0212	145	0.0465	34	chr9:139152429
0.0909	44 44	0.0070	94 94	0.0212	145	0.0465	34 34	chr9:139152429 chr9:139152429
0.0909	44	0.0070	94	0.0212	145	0.0465	34	chr9:139152429
0.0652	46 46	0.0000	95 95	0.0127	56	0.1000	28 28	chr12:8075625
0.0270	37	0.0068	95	0.0192	97	0.0517	41	chr16:2194742
0.0270	37 48	0.0068	95 95	0.0192	97 110	0.0817	41 48	chr16:2194742 chr17:4656377
0.0417	48	0.0064	95	0.0139	110	0.0800	48	chr17:4656377
0.0110	124	0.0000	95 95	0.0058	125	0.0270	87	chr6:10527783
0.0345	29	0.0121	96	0.0211	68	0.0423	53	chr10:134606054
0.0345	75	0.0121	96	0.0476	73	0.1429	50	chr2:27158968
0.0656	61	0.0000	96	0.0143	199	0.1053	45	chr2:233926891
0.0714	28	0.0000	98	0.0189	58	0.0787	64	chr17:7405042
0.0244	82 176	0.0000	99 99	0.0074	175 208	0.0212	139 155	chr20:22978301 chr22:49049005
0.0056	179	0.0000	99	0.0163	154	0.0330	137	chr4:5762176
0.0093	140	0.0000	100	0.0432	137	0.1270	99 85	chr11:75595222
0.0625	32 30	0.0000	100	0.0103	175	0.0217	33	chr18:490685
0.0333	30	0.0054	100	0.0114	139	0.0247	59	chr20:60228707
0.0333	30 30	$0.0054 \\ 0.0054$	$100 \\ 100$	0.0114 0.0114	139 139	0.0247 0.0247	59 59	chr20:60228707 chr20:60228707
0.0625	48	0.0000	101	0.0109	61	0.1081	33	chr11:8058484
0.0769 0.0250	26 40	0.0062	101 101	0.0205 0.0107	89 200	0.0469 0.0370	41 29	chr11:66914998 chr19:4861019
0.1143	35	0.0000	101	0.0063	147	0.0222	33	chr20:60743241
0.1023	88 88	0.0000	101	0.0095	163	0.1000	79 79	chr7:1510544
0.0392	51	0.0123	102	0.0248	136	0.0968	39	chr9:139068326
0.0244	41	0.0093	102	0.0166	267	0.0390	59	chr11:1366704
0.0244	41 41	0.0093	103 103	$0.0166 \\ 0.0166$	267 267	0.0390	59 59	chr11:1366704 chr11:1366704
0.0244	41	0.0093	103	0.0166	267	0.0390	59	chr11:1366704
0.0244 0.0227	41 44	0.0093	103 103	$0.0166 \\ 0.0080$	267 167	0.0390 0.0301	59 105	chr11:1366/04 chr2:10359490
0.0227	44	0.0000	103	0.0080	167	0.0301	105	chr2:10359490
0.0185	32	0.0000	105	0.0247	49	0.0610	50	chr19:60845797
0.0303	33 47	0.0000	106 107	0.0385	33 82	0.1077	50 33	chr9:129536664 chr16:88250779
0.0233	86	0.0000	109	0.0086	159	0.0571	70	chr19:50373394
0.0103 0.0476	97 42	$0.0000 \\ 0.0192$	$\begin{array}{c} 109 \\ 110 \end{array}$	$0.0179 \\ 0.0309$	70 106	0.0769 0.0811	66 60	chr19:60588529 chr7:718659
0.0560	125	0.0084	110	0.0149	85	0.0400	66	chr9:135283855
0.0455	44 27	0.0000	111	0.0091	59 158	0.1429	50 62	chr8:145714008
0.0123	81	0.0000	113	0.0143	223	0.1250	43	chr2:100802044
0.0588	34	0.0049	115	0.0154	100	0.0488	37	chr16:87450875
0.0588	34 34	0.0049	115 115	$0.0154 \\ 0.0154$	100 100	0.0488	37 37	chr16:87450875 chr16:87450875
0.0588	34	0.0049	115	0.0154	100	0.0488	37	chr16:87450875
0.0769	79 78	0.0000	116	0.0053	79 120	0.0323	34 50	chr13:26231964 chr16:87405843
0.0769	78	0.0000	116	0.0053	120	0.0323	50	chr16:87405843
0.0152	66	0.0068	117	0.0333	78	0.0769	40	chr19:40740589
0.0208	48 34	0.0060 0.0099	118 121	0.0122 0.0488	98 42	0.0357	41 25	chr19:1441164 chr19:63589447
0.0882	34	0.0099	121	0.0476	42	0.2222	25	chr19:63589467
0.0238	42	0.0052	121	0.0075	189 147	0.0255	42	chr5:577447
0.0169	59 50	0.0061	123 124	0.0168	132 147	0.1875	53 39	chr9:139565129
0.0244	82	0.0000	125	0.0048	152	0.1111	67	chr11:47164534
0.0244 0.1200	82 25	0.0000 0.0119	125 125	0.0048	152 99	$0.1111 \\ 0.1304$	67 28	chr11:47164534 chr9:135140451
0.1200	25	0.0119	125	0.0223	99	0.1304	28	chr9:135140451
0.1200	126	0.0000	125	0.0225	168	0.1304	113	chr1:50661729
0.0244	82 82	0.0000	127	0.0181	154	0.0513	85 85	chr16:638363
0.0244	82	0.0000	127	0.0181	154	0.0513	85	chr16:638363
0.0408	98 71	0.0000	129 129	0.0088	165 60	0.0526	65 51	chr11:65442303 chr14:74815098
0.0192	52	0.0000	130	0.0104	179	0.0541	67	chr11:72696316
0.0364 0.0465	55 86	0.0059	130	0.0062	135	0.0488	45 82	chr22:48980534 chr11:617173
0.0667	30	0.0094	131	0.0181	135	0.0465	36	chr8:144887902
0.0612	49	0.0050	132	0.0274	61	0.1000	38	chr16:30680130
0.0612	49 49	0.0050	132 132	0.0274	61 61	0.1000	38 39	chr16:30680130 chr16:30680143
0.0102	197	0.0000	134	0.0040	240	0.0283	162	chr2:14690554
0.0200 0.0200	50 50	0.0051 0.0051	135 135	0.0207 0.0207	131 131	0.0526 0.0526	41 41	chr8:143482444 chr8:143482444
0.0196	51	0.0042	139	0.0180	128	0.0385	35	chr17:2186555
0.0196	51	0.0042	139	0.0180	128	0.2000	35 35	chr7:19123469

0.0148	135	0.0068	142	0.0420	136	0.0943	99	chr11:64878855
0.0342	146	0.0104	142	0.0556	85	0.1333	85	chr17:58111550
0.0210	143	0.0054	143	0.0244	165	0.0625	147	chr22:49332327
0.0349	86	0.0041	144	0.0385	77	0.1143	61	chr6:36915198
0.0568	88 106	0.0000	145	0.0051	134	0.1099	93 76	chr19:41296727
0.0690	29	0.0080	147	0.0168	74	0.0513	28	chr17:959074
0.0055	183	0.0000	148	0.0154	123	0.1429	105	chr19:47079285 chr19:47079285
0.0278	72	0.0047	149	0.0112	140	0.0278	53	chr16:626348
0.0278	72	0.0047	149	0.0112	140	0.0278	53	chr16:626348
0.0278	72 72	0.0047	149	0.0112	140	0.0278	53	chr16:626348
0.0296	135	0.00047	150	0.0441	99	0.1176	72	chr19:18196303
0.0392	102 201	$0.0106 \\ 0.0000$	150 152	$0.0161 \\ 0.0066$	154 177	0.0357	55 121	chr4:7095165 chr14:74814283
0.0149	67	0.0000	154	0.0069	110	0.0526	72	chr17:7194931
0.0357	56 95	0.0000	155	0.0162	142 98	0.0625	49 84	chr11:2907226 chr19:2046867
0.0632	95 41	0.0093	155	0.0198	98	0.1098	84 34	chr19:2046867
0.0732	41	0.0000	155	0.0065	99	0.0303	34	chr1:229539304
0.0732	41 64	0.0000	155 156	0.0065	99 124	0.0303 0.0135	34 64	chr1:229539304 chr1:53565659
0.0263	38	0.0000	157	0.0385	27	0.0833	30	chr17:4589085
0.0167	299 98	0.0000	157	0.0278	159	0.0714	55	chr8:145661545
0.0164	61	0.0081	158	0.0179	66 116	0.0476	43	chr11:65781593
0.1023	88	0.0045	159	0.0157	150	0.0333	87	chr16:549422
0.0194	103 103	0.0050	159 159	0.0212	272 272	0.0522	109 109	chr16:88168676 chr16:88168676
0.0185	54	0.0000	159	0.0041	146	0.0465	63	chr4:3263552
0.0185 0.1000	54 100	0.0000	159 160	0.0041 0.0159	146 92	0.0465 0.0417	63 72	chr4:3263552 chr14:104522660
0.0611	131	0.0057	160	0.0155	172	0.1020	87	chr8:10625432
0.0076	131	0.00047	162	0.0070	136	0.0769	78	chr5:172687330
0.0233	43 100	0.0079	162 164	0.0326	75 163	0.2143	44 107	chr9:139565354
0.0600	100	0.0082	164	0.0137	163	0.0357	107	chr16:778384
0.0600 0.0288	100 104	0.0082 0.0041	164 167	0.0137 0.0089	163 121	0.0357 0.0779	107 97	chr16://8384 chr11:550779
0.0173	173	0.0063	167	0.0110	174	0.0256	103	chr7:126679664
0.0273	273	0.0000	172	0.0435	314	0.0252	180	chr18:75255759
0.0110 0.0110	273 273	0.0000	172 172	0.0056	314 314	0.0252	180 180	chr18:75255759 chr18:75255759
0.0571	35	0.0034	172	0.0131	128	0.0303	91	chr1:37272431
0.0571 0.0250	35 80	0.0034 0.0039	172 176	0.0131 0.0087	128 101	0.0303 0.0882	91 91	chr1:3/2/2431 chr17:76623096
0.0328	61	0.0000	177	0.0191	130	0.0612	54	chr11:2907170
0.0286	105	0.0108	177	0.0216	175	0.1000	48 74	chr16:579107
0.0313	64 288	0.0122	178 179	0.0184	107 323	0.0448	73 186	chr4:7095629 chr19:59666706
0.0556	72	0.0033	184	0.0069	264	0.0245	119	chr4:1232908
0.0556	216	0.0033	184 184	0.0069	264 201	0.0245 0.0187	119	chr7:100331477
0.0093	216	0.0000	184	0.0046	201	0.0187	158	chr7:100331477
0.0093	216	0.0000	184	0.0046	201	0.0187	158	chr7:100331477
0.0565	124 124	0.0042	185 185	0.0179 0.0179	159 159	0.0400 0.0400	96 96	chr16:777622 chr16:777622
0.0565	124	0.0042	185	0.0179	159	0.0400	96	chr16:777622
0.0293	205	0.0042	185	0.0789	125	0.3333	109	chr18:3001945
0.0167	60 29	0.0031	188 188	0.0455	55 194	0.3333	26 70	chr11:63815674 chr16:4106187
0.0042	236	0.0000	188	0.0063	186	0.0164	169	chr3:129689454
0.0385	26 253	0.0067	189	0.0221	130	0.0588 0.0571	68 142	chr11:74818881 chr19:50373335
0.0029	339	0.0000	190	0.0723	201	0.2000	155	chr15:71830869
0.0087	230	0.0000	192	0.0408	164	0.1250	118	chr11:65442261
0.0101 0.0980	99 51	0.0000	193 200	0.0068	194 104	0.0316 0.1212	99 36	chr2:236741391 chr12:122201238
0.0032	308	0.0000	200	0.0435	198	0.0909	159	chr6:109882346
0.0299	321	0.0028	202	0.0417	84 180	0.1429	48 169	chr11:62251527
0.0260	77 87		206	0.0526	63 118	0.1538	49 81	chr2:10505357
0.0194	206	0.0000	209	0.0145	143	0.0333	128	chr11:62251159
0.0274 0.0274	73 73	0.0000 0.0000	210 210	0.0112 0.0112	80 80	0.0294 0.0294	70 70	chr8:144969537 chr8:144969537
0.0076	131	0.0035	211	0.0147	129	0.0690	97	chr14:105027748
0.0076	131	0.0035	211 211	0.0147	129	0.0690	97 97	chr14:105027748 chr14:105027748
0.0076	131	0.0035	211	0.0147	129 211	0.0690	97 142	chr14:105027748
0.0194	103	0.0000	213	0.0087	179	0.0526	75	chr9:137938826
0.0769 0.0182	26 165	0.0078 0.0040	216 216	$0.0149 \\ 0.0115$	44 130	$0.1000 \\ 0.0455$	26 91	chr13:113286713 chr9:137531582
0.0182	165	0.0040	216	0.0115	130	0.0455	91	chr9:137531582
0.0426	235	0.0000	218	0.0040	250	0.0278	140	CULTO:5628345

0 0426	235	0 0000	218	0 0040	250	በ በ278	140	chr16·2958342
0.0442	113	0.0031	223	0.0079	226	0.0263	55	chr10:725606
0.0314	159	0.0077	225	0.0286	101	0.1515	105	chr16:30577743
0.0071	280	0.0000	229	0.0108	265	0.0244	179	chr6:108986718
0.0071	280	0.0000	229	0.0108	265	0.0244	179	chr6:108986718
0.0500	80	0.0089	234	0.0182	75	0.0671	275	chr14:102045325
0.0161	62	0.0026	238	0.0076	109	0.0204	58	chr16:57054705
0.0198	101	0.0047	240	0.0286	103	0.1250	37	chr16:1816225
0.0192	104	0.0047	243	0.0286	106	0.1250	40	chr16:1816230
0.0192	104	0.0047	243	0.0286	106	0.1250	40	chr16:1816230
0.0071	141	0.0031	244	0.0088	205	0.0769	80	chr8:140784481
0.0071	141	0.0031	244	0.0088	205	0.0769	80	chr8:140784481
0.0198	202	0.0000	250	0.0060	270	0.0313	160	chr16:970808
0.0160	125	0.0069	301	0.0179	196	0.0700	113	chr17:77649395
0.0132	228	0.0000	307	0.0122	152	0.0625	128	chr10:133970706

List of promoters presenting a methylation profile matching that of GC skew promoters.

Cov. = Coverage = N1+N2+M1+M2 (see Figure S3.3). M_s (-2000;-1000): 1kb windows starting 2kb upstream of TSS. M_s (-1000;TSS): 1kb windows ending at TSS. M_s (TSS;1000): 1kb windows starting at TSS. M_s (1000;2000): 1kb windows starting 1kb downstream of TSS.

Gene	Entry	Coordinates	Ms	Cov(1 st Exon)
MORN3	uc001uax.1	chr12:120591628-120591943	0.240	25
HNRNPK	uc004ank.2	chr9:85784854-85784917	0.240	25
PINK1	uc001bdn.1	chr1:20844588-20844803	0.231	26
THSD4	uc002atg.1	chr15:69837226-69837468	0.200	35
KRT13	uc002hwu.1	chr17:36914834-36915391	0.200	25
KRT13	uc002hwv.1	chr17:36914834-36915391	0.200	25
KRT13	uc010cxo.1	chr17:36914834-36915391	0.200	25
HIP1R	uc001udk.1	chr12:121906426-121906663	0.194	31
CST7	uc002wtx.1	chr20:24877866-24878211	0.194	31
FLJ00251	uc001mdw.2	chr11:6524472-6525472	0.186	43
ATP8B3	uc002lty.1	chr19:1756887-1757159	0.185	27
9-Sep	uc002jtw.1	chr17:72883760-72884075	0.182	33
TP53	uc002gii.1	chr17:7519096-7519536	0.179	28
TP53	uc010cnf.1	chr17:7519096-7519536	0.179	28
TP53	uc010cng.1	chr17:7519096-7519536	0.179	28
UBXN6	uc010dty.1	chr19:4404927-4405090	0.179	28
C10orf108	uc001ifr.2	chr10:686017-686382	0.175	40
CDC42BPB	uc001ymj.1	chr14:102500592-102500740	0.172	29
PTPRS	uc002mbz.1	chr19:5237061-5237399	0.167	42
BAGE2	uc002yit.1	chr21:10120574-10120796	0.167	36
CR617046	uc001myp.2	chr11:45187118-45187422	0.162	74
KIAA1335	uc002xjz.1	chr20:39475378-39477119	0.161	31
CPNE4	uc003eom.1	chr3:133486774-133486944	0.161	31
MYO1H	uc009zvh.1	chr12:108310871-108311032	0.161	31
DNAJC14	uc001shu.1	chr12:54507303-54508765	0.160	50
KIAA0676	uc003mlk.1	chr5:179229861-179230101	0.160	25
BAGE5	uc002yiu.1	chr21:10120594-10120808	0.159	44
BAGE	uc002yiv.1	chr21:10120594-10120808	0.159	44
GRM2	uc003dbp.1	chr3:51717904-51718489	0.158	57
SH2D4B	uc001kck.1	chr10:82287638-82288251	0.157	70
MICAL2PV1	uc001mkb.2	chr11:12140202-12140542	0.156	32
MICAL2PV2	uc001mkc.2	chr11:12140202-12140542	0.156	32
AKT2	uc002one.1	chr19:45440281-45441049	0.156	32
KIAA0676	uc003mlf.1	chr5:179225444-179226478	0.156	32
MED15	uc002zst.1	chr22:19257965-19259519	0.155	71
NOC2L	uc001aby.2	chr1:882342-883781	0.154	26
KANK4	uc001daf.2	chr1:62501352-62501684	0.154	26
DPH1	uc002ftv.1	chr17:1889009-1889867	0.154	26
LZTR1	uc002ztp.1	chr22:19680252-19680401	0.154	26
MOV10L1	uc003bjj.1	chr22:48870621-48870741	0.154	26
MOV10L1	uc003bjk.2	chr22:48870621-48870741	0.154	26
CNTN2	uc009xbi.1	chr1:203293932-203294414	0.154	26
LZIS1	uc003wzr.1	chr8:20156628-20157083	0.151	53
LZIS1	uc010ltg.1	chr8:20156628-20157083	0.151	53
APBB1	uc001mda.2	chr11:6383193-6383465	0.150	40
APBB1	ucuu1mde.2	chr11:6383193-6383465	0.150	40
ACACB	ucuultoc.1	chr12:108061585-108062246	0.148	27
GRK6	ucou3mgs.1	chr5:176791318-176791714	0.148	27
MID1	uc004ctm.1	chrX:10494928-10495643	0.148	27
MID1	uc004ctn.1	ChrX:10494928-10495643	0.148	27
MID1	ucuu4ctt.2	CNFX:10494928-10495643	0.148	27
MID1	uc004ctu.2	ChrX:10494928-10495643	0.148	27
MID1		CNFX:10494928-10495643	0.148	27
MID1	uculundy.1	CNFX:10494928-10495643	0.148	27
I GMZ	ucuu2xnq.1	CNF2U:36194156-36195576	0.147	68
ATP13A1		CNF19:19626928-19627260	0.146	41
KIAA1640	ucuuzjxn.z	CNF17:75669837-75670452	0.146	48
SBSIN		chi 19:40709380-40711052	0.140	48
pp9904		CIII 1.000/949-0009434	0.143	49
SLC24A4	ucooryan. I	chr14:91988803-91990100	0.143	28
TEKE		obr10:54059147 54059227	0.143	20
7034120	uc002ppm.1	chr6:140927069 140927442	0.143	20
203112D PC075707		chr7:12562712 12562891	0.143	29
CDIV4		CIII7.120026188.120026210	0.143	20
	uc0092ax.1	chr17:62202910 62204109	0.143	20
	uco10bim 1	chr2:47011526 47012127	0.143	20
	uc010hjf1	chr3:42800966-42802304	0.143	50
GPRC5D		chr12:12003601-12004585	0.140	36
CNTN2	uc001bbs 1	chr1:203204356-203205044	0.139	20
MYST1	UC002eaz 1	chr16:31044456-31045910	0.138	29
HDGF2	uc002mag 1	chr19:4450488-4450716	0.138	29
NFAM1	uc003bcn 2	chr22.41158187-41158345	0.138	29
C.7orf46		chr7:23686274_23686427	0.138	20
C.7orf46		chr7:23686274-23686427	0.138	20
C7orf46	uc003swr 2	chr7:23686274-23686427	0.138	29
HDGF2	uc010dua 1	chr19:4450488-4450716	0.138	29
TG	uc010mdw 1	chr8:133979076-133979213	0 138	29
ESPNI	uc002vxg 2	chr2:238673690-238674093	0 137	102
FTV6	uc001raa 1	chr12:11913625-11914170	0.136	44
HNRNPK	uc004anf.2	chr9:85784888-85785004	0.136	44
HNRNPK	uc004ang.2	chr9:85784888-85785004	0.136	44
HNRNPK	uc004anh.2	chr9:85784888-85785004	0.136	44

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chr3:128227018-128220610	
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chr2:98502841-98503049	
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chr6:43373976-43374467	
chr17:4823703-4824640	
chr6:43373976-43374467	
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chr2:2/1653388-2/165368/	
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chr1:201286934-201287628	
chr8:11455251-11456370	
chr7:151142435-151142890	
chr9:139409576-139409677	
chr10:99685917-99686113	
chr12:122993847-122996061	
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chr16:87570566-87570902	
chr6:2904401-2906560	
chr7:150462420-150462588	
chr11:47226803-47226939	
chr11:47226803-47226939	
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chr20:62309391-62310284	
chr11:76568429-76568647	
chr19:1573053-1573222	
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HNRNPK uc004ani.2 HNRNPK uc004anj.2 MST066 uc002dmw.1 ARHGAP17 uc002dmx.1 MST066 uc002dne.1 TACC1 uc003xmh.2 TACC1 uc010lwq.1 DKFZp761P19121 uc003ejh.1 BCR-ABL uc010gtx.1 C13orf35 uc001vsh.1 INPP4A uc002syy.1 SLC22A7 uc003ous.2 uc003out.1 SLC22A7 CAMTA2 uc010ckv.1 SLC22A7 SLC22A7 uc010jyl.1 uc010jym.1 ARHGEF10L uc001baq.1 FLJ00133 uc002wak.2 hCG_31249 GPSM1 uc004bzl.2 uc004che.1 AMPD2 uc001dyd.1 PPFIA4 uc001gyz.1 uc003wua.1 uc003wkj.1 BLK PRKAG2 uc010ncg.1 HBE269 CRTAC1 uc001kot.1 limkain uc001ufv.1 uc001fgh.1 FLAD1 uc001nei.1 ACP2 MYO7A uc001oyd.1 BEAN CBFA2T3 uc002eoq.1 uc002fmm.1 CBFA2T3 uc002fmn.1 SERPINB6 uc003muk.1 AGAP3 uc003wjj.1 uc009ylj.1 ACP2 ACP2 uc009ylk.1 dJ402G11.5 uc010hap.1 MYT1 uc002yij.1 MYO7A uc009vut.1 TCF3 uc002lto.1 MEGF5 uc003mac.1 **ZNF687** uc009wmp.1 SMARCA4 uc010dxt.1 PAX8 uc002tjo.1 TESSP2 uc003cqj.1 FGF17 KIAA1542 uc003xai.1 uc009ybz.1 KRT74 uc001sap.1 CATSPER2 uc001zsh.1 uc001zsi.1 uc001zsk.1 CATSPER2 CATSPER2 SMARCA4 uc002mqg.1 SMARCA4 uc002mqj.2 MED15 uc002zss.1 I YPD2 uc003vwz.1 SMARCA4 uc010dxq.1 SMARCA4 uc010dxr.1 SMARCA4 uc010dxs.1 pp9320 uc003yzv.2 CTBP1 uc003gct.1 SLC6A19 uc003jbw.2 RALGDS uc004ccn.1 TMEM132A uc001nqm.1 FBLIM1 uc001axi.1 DISC1 uc001hvc.2 MCF2L uc001vst.1 CLK3 uc002ayl.2 SPATA22 uc002fvo.1 ZNF335 uc002xqv.1 TRIM7 uc003mmv.1 IRF4 uc003mtc.1 ANKRD6 uc003pnh.2 GLIS3 uc003zic.1 HPCAL1 uc010exf.1 NM 001127438 uc010nmy.1 uc003qtc.1 SLC22A1 SLC22A1 uc003qtd.1 EP400NL uc001ujv.2 UNQ830 uc002vtr 1 KIF1A uc002vzx.1 TBC1D24 uc002cqm.1 PIWIL4 uc001pfa.1 AOP2 uc001rvn.1 MBD6 uc001sok.1 uc001tzb.2 ACADS

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ARHGAP17	uc002dna.1	chr16:24873425-24876358	0.111	54
FFCBP2	uc002fhe 1	chr16:82563017-82563281	0 111	27
RHBDF2	uc002irs.1	chr17:71989052-71989222	0.111	27
MAP4K1		chr19:43800277-43800483	0 111	27
MAP4K1	uc002oiv 1	chr19:43800277-43800483	0 111	27
SGK2	uc002xkg 1	chr20:41602205-41602362	0 111	27
CARMA3	uc003asy 1	chr22:36220039-36220529	0 111	27
RHO	uc003emt 1	chr3:130730172-130730627	0 111	27
AMOTI 2	uc003eqf 1	chr3:135572232-135573026	0 111	72
SI C26A1	uc003abx 1	chr4.977089-977224	0 111	27
SI C26A1	uc003ach 1	chr4:977089-977224	0 111	27
SI C26A1		chr4:977089-977224	0 111	27
KIAA0294		chr8:1795470-1795757	0.111	36
ΔTP2R3	uc004fbs 1	chrX:152454774-152455107	0.111	27
ΔTP2B3	uc004 ft 1	chrX:152454774-152455107	0.111	27
KIAA1033	uc010cfd 1	chr16:66043484_66043813	0.111	27
		chr10:55709941 55709054	0.111	26
		chr8:22180025-22180487	0.111	63
FIVVILZ		chr12:121262142 121262405	0.100	55
	ucoolarb 1	chr15:66294637 66295502	0.109	46
		obr15:00204037-00203302	0.109	40
CALINE4		chi 15.00204037-00203302	0.109	40
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	ucoo lugn.2	CIII 12: 123545040-123545007	0.108	37
MAP3KIU		CITE 19:40390122-40390302	0.108	37
CICFL	ucouzxym.z	CITIZU.55533490-55533589	0.108	37
CICFL	ucuilugiw.1	CNF2U:55533490-55533589	0.108	37
CICFL	uc010giy.1	chr20:55533490-55533589	0.108	37
CICFL	ucu10giz.1	chr20:55533490-55533589	0.108	37
CICFL	uc010gjf.1	chr20:55533490-55533589	0.108	37
CICFL	uc010gjg.1	chr20:55533490-55533589	0.108	37
CICFL	uc010gji.1	chr20:55533490-55533589	0.108	37
INKS1BP1	uc001njp.1	chr11:56826509-56827529	0.108	65
INKS1BP1	uc001njq.1	chr11:56826509-56827529	0.108	65
SDK1	uc003smy.1	chr7:4135843-4136255	0.108	65
PMPCA	uc004chm.1	chr9:138430340-138431487	0.108	65
C1orf167	uc001ata.2	chr1:11762446-11762654	0.107	28
C18orf22	uc002Int.1	chr18:75902290-75903207	0.107	28
MCM5	uc003anw.1	chr22:34136402-34136903	0.107	28
GOLGA2	uc004buh.1	chr9:130062565-130062902	0.107	28
NLRC3	uc010btn.1	chr16:3567151-3567393	0.107	28
IGSF11	uc003eby.1	chr3:120347350-120347588	0.106	47
IGSF11	uc003ebz.1	chr3:120347350-120347588	0.106	47
UBAP2	uc003ztn.1	chr9:33917795-33918833	0.106	47
IGSF11	uc010hqs.1	chr3:120347350-120347588	0.106	47

Top-1% Saqqaq genes showing highest methylation levels in the first exon.

3.6. Predicting the Saqqaq age at death using age-dependent CpG signatures

DNA methylation levels have been shown to change with age in human tissues [Alish et al. 2012; Day et al. 2013; Johansson et al. 2013]. In this section, we explored whether ancient methylation levels as measured through M_s in the Saggag sequence data could be used to propose an age at death for the Saggag individual. Recently, Koch and Wagner [2011] proposed to use methylation levels at four CpG sites associated with the genes TRIM58, KCNQ1DN, NPTX2 and GRIA2 (cg07533148, cg01530101, cg1279989, cg25148589, respectively) and at one additional hypomethylated CpG site (BIRC4BP, cg23571857) to predict the age of modern human individuals. This framework enabled relatively precise predictions, with average differences between predicted and real age of about 11 years and could potentially be used to propose a range for the age at death of the Saggag individual. As cq1279989 showed no overlap with the Illumina 450k array data available from [Siekler et al. 2013], further analyses were restricted to the other four loci. As a positive control, we first used the linear regressions provided in [Koch and Wagner 2011] in order to predict the age of five donor individuals (PT1, PT2, PT3, PT4 and PT5) from their methylation levels at each four loci. Such linear models have been built using methylation levels from two types of tissues (dermis and epidermis) and three cell lines (cervical smear cells, monocytes and T cells) and are likely robust across a large range of somatic tissues. We therefore hypothesized that the linear model was correct for hairs. Predicted ages are plotted in Figure S3.14 together with the known age for each donor individual. Overall, predictions based on observed methylation levels at cg07533148 and cg01530101 were consistent with each other, providing age estimates within 1.7-12.4 years (standard deviation = 5.7 years) around the real age of donor individuals. This is in agreement with the accuracy levels reported in Koch and Wagner [2011]. Inconsistent values showing large differences between predicted and real age values (16.1-39.4 years, standard deviation = 9.4 and 12.4 years, respectively) were found for the other two loci. Consequently, we decided to base our predictions for the age at death of the Saggag individual on the first two CpGs only (cg07533148 and cg0153010).



Figure S3.14

Predictions for the age of five modern human individuals based on methylation levels at four CpG sites. Four CpG sites showing agedependent methylation levels have been considered (cg0733148, red; cg01530101, orange; cg25148589, green, and cg23571857, blue; [Koch and Wagner 2011]). We predicted (coloured crosses) the age of five modern human individuals (PT1, PT2, PT3, PT4 and PT5) for which methylation information was available and compared our predictions to the known age of each individual (black).

We then used the procedure described in Supplemental Section SI3.4 in order to calculate absolute methylation levels for the Saggag at these two loci. Briefly, we relied on a linear model relating M_s values (as calculated within a 2,000 bp-wide region centered on each CpG site) and hair methylation levels measured in any of the five modern human individuals. In this first linear model, different CpG subsets were considered depending on their coverage (a full range of possible coverage was considered, spanning every 50 counts from 100 to 500). Methylation levels were predicted for the Saggag individual using the predict() function in R [R Development Core Team 2012] and a confidence level of 0.999. Saggag methylation values as predicted from the first linear model were further converted into absolute age using a second linear model, namely the linear regression provided in Koch and Wagner [2011]. Predictions are provided in Figure S3.15 for the full range of coverage investigated. The age at death is predicted at around 44.1-69.4 and 52.1-64.2 years for both CpG considered. Given error margins estimated with modern human donors (shifted by up to 12.4 years), these values should not be taken as precise point estimates but rather as an indication that the Saggag individual was most likely old when he died. We should also insist that our estimate depend on a number of strong assumptions that remain to be investigated with further studies: 1) the agedependency of methylation levels at cg07533148 and cg0153010 has remained constant in recent modern human evolution; 2) the age-dependency of methylation levels at cg07533148 and cg0153010 as described in Koch and Wagner [2011] is robust across a range of human populations and environments.



Figure S3.15

Predictions for the age at death of the Saggag individual based on inferred methylation levels at two CpG sites. We predicted the age at death of the Saqqaq individual using on one hand the methylation levels estimated at two CpG sites (cg07533148 and cg0153010) and on the other hand the linear relationship between methylation levels and age as reported in [Koch and Wagner 2011]. Different subsets of CpGs showing a range of minimal coverage were considered to estimate the Saggag methylation levels based on M_s values.

3.7. Methylation profiles across nucleosomes

CTCF (CCCTC-binding Factor) sites play a critical role in shaping the 3D architecture of our genome [Bell and Felsenfeld 2000]. An overall number of 15,000-40,000 of CTCF sites showing quite permissive consensus sequences of 20 nucleotides are interspersed in the human genome [Kim et al. 2007]. Recruitment of the CTCF protein at CTCF sites induces strong chromatin remodelling, positionning arrays of 10 nucleosomes over the ~2kb flanking genomic regions. Within those regions, nucleosomes are strongly phased, showing footprints of ca. 110-170 nucleotides (depending on the nature of histone modifications; 118 bp for mononucleosome data) and are separated by ca. 0-60 bp long spacers (33 bp for mononucleosome data) [Fu et al. 2008].

We retrieved hg18 genomic coordinates of 12,864 CTCF 20-mer binding sites from the Supplemental Table 1 provided in Fu and colleagues [Fu et. al 2008]. We next calculated average M_s values for each 25bp window (1bp increment) within 1kb upstream and downstream of the CTCF binding site and compared these to nucleosome positioning represented by GC-corrected read depth (Figure 4.d). Overall, peaks in methylation levels coincided with sites of low nucleosome occupancy in agreement with independent empirical evidence in modern human cells [Kelly et al. 2012]. We also recovered a 63bp large footprint for the CTCF protein using the central region with lowest methylation levels (<0.024), as methylation is known to suppress CTCF DNA-binding [Bell et al. 2001].

We investigated whether the greater power to detect methylation in positions with greater read depth could be responsible for the pattern observed at CTCF sites. We therefore performed the same analysis by down-sampling sites across the full region. Down-sampling was performed for each position located 1,000 nucleotides upstream and downstream of CTCF binding sites in order to represent each position with similar depth-of-coverage (from 20 to 100, iterating every 10). We generated 100 down-sampled datasets (per depth-of-coverage investigated), calculated M_s at each position then tested for correlation with nucleosome positioning. The results are provided in Table S3.7. While stronger correlation were found for datasets down-sampled at higher depth, we found a significantly negative correlation between M_s and nucleosome occupancy for all dataset investigated (-0.0896 ≤ Pearson correlation coefficient \leq -0.254 ; p-value \leq 6.05x10⁻⁵). This clearly demonstrates that the greater power to detect methylation in positions with greater read depth does not preclude our ability to detect correct methylation and nucleosome positioning signal at CTCF sites.

We then profiled methylation levels at non-CTCF nucleosomes (Figure 4.e). We defined CTCF-nucleosomes as those nucleosomes located within a 1kb of a CTCF binding sites. Conversely, non-CTCF nucleosomes consist of any nucleosome call not overlapping these regions.

We then defined any position within 73bp of a nucleosome center as inside (labeled "In"), while sites located within 85-110bp from the nucleosome centers were considered flanking regions (labeled "Out"; Figure 3.a). We required two consecutive nucleosomes centers to be separated by at least 183bp in order to avoid overlap between In and Out. We calculated M_s for the In and Out group and for each position relative to the nucleosome center (-200;+200) following the procedure described in Figure S3.4. Finally, we plotted raw M_s (without smoothing) as a function of the distance to nucleosome centers.

We found that on average non-CTCF nucleosomes showed similar methylation as that of nucleosomal DNA ("In") and spacer DNA ("Out"; data not shown). We also identified that the 20 nucleotides flanking nucleosome centers were enriched in strong dinucleotides and were strongly depleted of methylation, suggesting a possible functional role for this region in relation with correct nucleosome positioning (Figure 4.e, Figure S2.12).

Depth	20	30	40	50	60	70	80	90	100
Mean cor.	-0.11831	-0.13805	-0.1576	-0.1714	-0.1871	-0.1986	-0.2094	-0.2191	-0.2306
Max cor.	-0.1656	-0.18501	-0.215	-0.22	-0.2203	-0.2381	-0.2401	-0.2605	-0.254
Min cor.	-0.08958	-0.08654	-0.1213	-0.1216	-0.1468	-0.158	-0.1772	-0.1883	-0.1911
Mean p-value	3.01E-06	1.85E-06	1.89E-09	5.74E-10	5.15E-13	1.53E-14	3.06E-17	3.45E-19	6.97E-20
Max p-value	6.05E-05	1.07E-04	5.29E-08	5.06E-08	4.32E-11	1.25E-12	1.51E-15	2.18E-17	6.92E-18
%Data lost	0.1	0.2	0.2	0.25	0.3	0.4	1.9	6.8	18.94

Table S3.7

Nucleosome and methylation patterns at CTCF sites are robust to down-sampling.

Each position located within 1,000 upstream and downstream of CTCF sites was down-sampled to a given depth-of-coverage. We considered a range of possible coverage from 20 to 100, where coverage = N1+N2+M1+M2 (Figure S3.3). Mean (Min) cor. = average (minimum) Pearson correlation coefficient across 100 correlation tests performed between M_s and nucleosome positioning, where M_s has been calculated following down-sampling. Mean (Max) p-value = average (maximum) p-value of Pearson correlation analyses performed across 100 correlation tests performed between M_s and nucleosome positioning, where M_s has been calculated following down-sampling. Mean (Max) p-value = average (maximum) p-value of Pearson correlation analyses performed across 100 correlation tests performed between M_s and nucleosome positioning, where M_s has been calculated following down-sampling. %Data lost indicates the fraction of the positions showing depth-of-coverage inferior to the threshold considered.

Section SI4. Expression analysis

We retrieved Saggag methylation levels at promoters and gene body regions from 280 genes whose proteins have been identified following proteomic analysis of human hair shaft (an extra number of 65 other proteins described in the original publication could not be identified without ambiguity and were therefore disregarded) [Lee et al. 2006]. Those represented a total number of 567 possible transcripts. We then calculated the level of gene body to promoter methylation as a proxy for gene expression, following the procedure described by Ball et al. [2009]. Promoters (PM) were defined as spanning -500 to +2000 around the TSS and gene bodies (GB) as the region from +2000 to the termination site. We then calculated the M_s statistics for PM and GB as indicated in Figure S3.3. We finally calculated the ratio of methylation $M_s(GB)$ over $M_s(PM)$ that we called called R_s . At promoters showing null M_s values, we arbitrarily set R_s to an infinite number, suggesting high expression levels [Ball et al. 2009]. Following a procedure identical to that described in Supplemental Section SI3.4, we first selected all genes showing a minimum coverage of 25 and 75 over their promoter and gene body, respectively. We then selected genes where coverage at promoters was at least equivalent to that observed within gene bodies, correcting for length differences between PM and RE regions. A total number of 140 genes (271 transcripts) from the original list fulfilled those criteria (Table S4.1). Unfortunately, genes for Keratin Associated Proteins, which represent together with keratins major structural components of the hair shaft [Gong et al. 2012], showed not sufficient coverage and could not be investigated for expression patterns.

We compared the distribution of R_s for that list of 140 genes known to be expressed at the protein level in hair shaft and for the full list of genes annotated in the human reference genome (hg18; using similar filtering criteria, this represented a total number of 29,070 genes). We found the former to be significantly greater than the latter (Kolmogorov-Smirnov test, p-value = 0.00152), in agreement with the fact that it consists of known expressed genes in contrast to the whole gene dataset that is a mixture of silenced and expressed genes. This suggests that R_s captures genuine expression information. Of note, we identified amongst the genes showing high to extremely high R_s values a range of transcripts for keratin proteins such as keratins 71 and 85 [Moll et al. 2008] (Table S4.1). Those proteins were likely expressed in the Saggag hairs, in agreement with functional studies of modern hairs. Keratin 85 has been detected at high levels in the hair shaft cortex and medulla [Langbein et al. 2010]. Keratin 71 is also a hair-specific keratin expressed in the inner root sheath [Langbein et al. 2010]. This contrasts with keratin 82, one member of group C type II hair keratins, that is absent from the medulla and restricted to a thin external cuticle layer [Langbein et al. 2010] and for which we also found relatively low R_s value (Table S4.1). Similarly, we found a low R_s value for keratin 79, suggesting low expression levels, if any, in agreement with its classification amongst non hairspecific epithelial keratins [Moll et al. 2008]. In addition to hair-specific keratins, we confirmed the presence of a number of proteins involved in cellular adhesion and cytoskeleton organization. In particular, we predicted the presence of Plakophilin 1 and 3 and, to a lesser extent, Plectin and Desmoplakin (Table S4.1), two proteins associated with cell-cell adhesion via desmosomes, a feature known to be key for the structural organization of hairs [Bazzi et al. 2009]. We also predicted high levels of trichohyalin (TCHH), a protein known to confer mechanical strength to the hair follicle inner root sheath. Altogether, this suggests that the level of gene body over promoter methylation, R_s, could be used as a genuine proxy for predicting gene expression. We therefore stratified genes according to R_s quartiles and we used the first and the top quartiles as provisional lists of genes (7,263-7,264 UCSC entries per quartile considered, representing 4,174 and 3,700 non-redundant entries respectively). In

Supplemental Section SI3.5, we identified a subset of 214 genes showing highest (top-1%) methylation levels in their first exon (159 are not redundant; Table S3.5). This represents a list of candidates for low (null) expression levels given the documented tight association between methylation at first exon and silencing [Brenet et al. 2011]. Therefore, this subset of genes is expected to vastly coincide with genes present in the first R_s quartile (ie. with minimal R_s values), as the latter defines a provisional list of genes whose expression was down-regulated in the Saggag hairs. Interestingly, 100.0% of the genes identified as showing highest methylation levels in their first exon belong to the first R_s quartile. No genes identified as showing highest methylation levels in their first exon belong to other quartiles, which demonstrates the strong consistency in our expression predictions. Extending the list of genes showing highest methylation levels in their first exon to the top-5% (instead of the top-1%; for a total number of 1,089 genes) does not affect those conclusions, as 88.3% and 9.0% intersect the first and second R_s guartiles, respectively, leaving only a number of 20 (2.35%) and 3 (0.35%) genes in the third and fourth quartiles. In tables S4.2 and S4.3, we provide the bottom-1,000 and top-1,000 genes ranked by R_s values, together with their respective coverage in PM and RE regions. Those lists provide candidate genes whose expression was respectively down-regulated and upregulated in the Saggag hairs. We performed functional enrichment analyses of those candidates in DAVID [Huang et al. 2009], considering only those categories with enrichment scores (ES) superior or equal to 1.2 and within each category, the term showing lowest Benjamini-Hochberg p-values (terms with p-values superior to 0.05 were disregarded). Down-regulated candidates were enriched in the following functional categories: Signal (ES = 3.95; Counts = 203; BH p-value = 7.77x10⁻⁶), Cell adhesion (ES = 3.87; Counts = 37; BH p-value = 0.0052) Peptidase S1/S6 -Chymotrypsin/Hap (ES = 3.66; Counts = 20; BH p-value = 6.78x10⁻⁴), Plasma membrane part (ES = 3.10; Counts = 134; BH p-value = 0.0433), Ionic channel (ES = 2.98; Counts = 35; BH p-value = 1.60x10⁻⁴), Homeobox (ES = 2.51; Counts = 23; BH p-value = 0.0432), Glycoprotein (ES = 2.08; Counts = 239; BH p-value = 0.0016), and Muscle protein (ES = 1.27; Counts = 14; BH p-value = 1.34x10⁻⁴). Up-regulated candidates were enriched in the following functional categories: Ubiquitin ligase complex (ES = 3.97; Counts = 19; BH p-value = 6.24×10^{-5}), Phosphorus metabolic process (ES = 2.36; Counts = 78; BH p-value = 0.0231), Ligase (ES = 2.16; Counts = 30; BH p-value = 0.0079), Metal-binding (ES = 2.11; Counts = 173; BH p-value = 0.0110) and Inorganic anion transport (ES = 1.72; Counts = 15; BH p-value = 0.0383).

In order to further assess the validity of our expression predictions, we contrasted microarray expression data from modern hair and R_s values. Mean expression of 10 hair microarray samples from [Kim et al. 2006] (normalized expression data downloaded from GEO (id:GSE3058) was used to partition the represented genes into 10, 20 and 50 quantiles by expression. In addition to R_s (Expression Proxy 1) two other proxies for gene expression were defined based on the literature:

Expression Proxy 2: The read depth over the +1 nucleosome. Calculated as the mean GC-corrected read depth in the region 0 to 200 downstream of the TSS, following [Valouev et al. 2011].

Expression Proxy 3:The strength of the phasing. Signal strength of regularly spaced nucleosomes downstream of the TSS. The mean spectral density was computed over the TSS \pm 1000 for the genes of each quantile by the Welch method and the signal strength at the maximum frequency estimated for each quantile, following [Schones et al. 2008].

The correlation for the proxy and the mean expression of these groups was then calculated using Spearman correlation and significance was determined based on Spearman's rho statistic. Importantly, we found significant correlation between mean expression levels and all our three proxys. This supports the validity of our expression predictions and consequently the quality of both our nucleosome and methylation maps. Spearman's rank correlation coefficient for each of the measures was for 10 groups: **1**) 0.99 (p<2.2e-16), **2**) 0.85 (p=0.00171), **3**) 0.79 (p=0.009844); for 20 groups **1**) 0.95 (p=6.217e-06) **2**) 0.75 (p=0.0002104) **3**) 0.71 (p=0.0006549) and for 50 groups: **1**) 0.87 (p<2.2e-16) **2**) 0.62 (p=3.242e-06) **3**) 0.53 (p=8.968e-05).

Gene	UCSC	Rs	Cov(GB)	Cov(PM)	M _s (GB)	M _s (PM)
GARS	uc003tbm.1	NaN	106	146	0.000	0.000
RBM14	uc001oiz.1	Inf	80	64	0.088	0.000
RBM14	uc001oiy.1	Inf	81	80	0.086	0.000
RBM14	uc001oiw.1	Inf	81	84	0.086	0.000
RBM14		Int	81	84	0.086	0.000
RDIVI 14 RBM14		IIII	00 81	80	0.000	0.000
RBM14	uc0010iy.1	Inf	81	84	0.086	0.000
RBM14	uc001oix.1	Inf	81	84	0.086	0.000
UNC84B	uc010gxr.1	Inf	2013	99	0.039	0.000
RTN4	uc002ryf.1	Inf	117	246	0.017	0.000
RTN4	uc002ryg.1	Inf	117	246	0.017	0.000
RTN4	uc002ryd.1	Inf	114	66	0.018	0.000
RIN4	uc002rye.1	Inf	117	246	0.017	0.000
2-Sep 2-Sep	uc002wbr.1	IIII	260	233	0.035	0.000
2-Sep 2-Sen	uc002wbg.1	Inf	260	233	0.035	0.000
SERPINB5	uc002liz.2	Inf	141	30	0.014	0.000
TSPAN7	uc004deg.2	Inf	154	96	0.039	0.000
ACOT7	uc001ams.1	Inf	2372	34	0.055	0.000
PADI3	uc001bai.1	Inf	371	27	0.065	0.000
AIM1	uc003prh.1	Inf	260	317	0.054	0.000
SELENBP1	ucuulexx.1	Inf	135	32	0.022	0.000
	ucoo rezp.z	In	303	35	0.006	0.000
I MNA	uc001fni 1	Inf	292	38	0.051	0.000
PKP1	uc001awe.1	Inf	536	250	0.076	0.000
PKP1	uc001gwd.1	Inf	536	250	0.076	0.000
VCP	uc003zvy.2	Inf	108	206	0.028	0.000
SND1	uc003vmi.1	Inf	2821	178	0.045	0.000
SEC24C	uc001jux.1	Inf	193	59	0.052	0.000
SEC24C	uc001juw.1	Inf	193	59	0.052	0.000
	ucoo3qrt.2	Int	607	137	0.026	0.000
NME2		Inf	440	127	0.016	0.000
NME2	uc002itj.1	Inf	87	127	0.046	0.000
NME2	uc002ith.1	Inf	87	127	0.046	0.000
NME2	uc002itk.1	Inf	448	127	0.016	0.000
NME2	uc002itj.1	Inf	448	127	0.016	0.000
NME2	uc002iti.1	Inf	87	127	0.046	0.000
NME2	ucuu2ith.1	Inf	87	127	0.046	0.000
PKP3 D/HB	ucoo npc. i	In	106	120	0.048	0.000
FFF1G	uc002kbn.1	Inf	115	85	0.050	0.000
CRAT	uc004bxq.1	Inf	399	49	0.025	0.000
MDH2	uc003ueo.1	Inf	420	125	0.048	0.000
RPS9P4	uc002qed.1	Inf	117	119	0.026	0.000
RPS9P4	uc002qea.1	Inf	117	119	0.026	0.000
RPS9P4	uc002qed.1	Inf	117	119	0.026	0.000
RP59P4 ATP6\/1B2	ucuu2qea.1	Inf	117	119	0.026	0.000
ATP6V1B2	uc003eao 1	Inf	331	51	0.003	0.000
PGD	uc001arc.1	Inf	157	180	0.038	0.000
NPC1	uc002kum.2	Inf	488	182	0.055	0.000
CLTC	uc002ixq.1	Inf	250	152	0.056	0.000
CLTC	uc002ixr.1	Inf	250	152	0.056	0.000
CLTC	uc002ixp.2	Inf	211	152	0.052	0.000
		In	108	85 46	0.019	0.000
TARS	uc003iby 1	Inf	84	103	0.002	0.000
YWHAE	uc002fsj.1	Inf	527	187	0.040	0.000
NOP58	uc002uzb.1	Inf	277	74	0.029	0.000
BLMH	uc002hez.1	Inf	134	201	0.045	0.000
LPCAT3	uc001qsi.1	Inf	201	155	0.050	0.000
KRT85	uc001sag.1	Inf	85	34	0.012	0.000
SARS	uc001dwv.1	Int	146	93	0.048	0.000
USP5	uc001ari 2	Inf	232	118	0.048	0.000
USP5	uc001arh.2	Inf	232	118	0.047	0.000
EIF3E	uc003ymu.1	Inf	125	59	0.056	0.000
OTUB2	uc001yci.1	Inf	417	189	0.041	0.000
PGLS	uc002ngw.1	18.016	189	227	0.079	0.004
HIP1R	uc001udj.1	17.893	1304	307	0.058	0.003
		10.748 15 979	040 1500	303 281	0.047	0.003
EFF2	uc002lze 1	14 509	611	197	0.074	0.005
CSNK1E	uc003avj.1	13.821	776	275	0.050	0.004
MAP7	uc003qgz.1	13.778	1019	312	0.044	0.003
YWHAG	uc003uez.1	13.264	318	222	0.060	0.005
CSNK1E	uc003avk.1	13.230	793	269	0.049	0.004
YWHAZ	uc003yjv.1	13.071	127	332	0.039	0.003
∠-∂ep 2-Sen	uc002wbc.1	12.007 12.607	202 262	367	0.034	0.003
- 004		12.007			0.004	0.000

CSNK1E	uc003avp.1	12.343	557	275	0.045	0.004
CSNK1E	uc003avq.1	12.343	557	275	0.045	0.004
VDAC2	uc001jwz.1	12.333	90	185	0.067	0.005
VDAC2	uc001jxa.1	12.333	90	185	0.067	0.005
PEBP1	uc001twu.1	12.194	93	189	0.065	0.005
PLD3	uc002011J.2	11.667	621	207	0.050	0.005
PLD3	uc002onl.2	11.667	621	207	0.056	0.005
GDI2	uc001iim.2	10.600	225	265	0.040	0.004
GDI2	uc009xid.1	10.600	225	265	0.040	0.004
GDI2	uc001iil.2	10.600	225	265	0.040	0.004
UNC84B	uc003awh.1	10.286	741	206	0.050	0.005
ACTN1	UC001xkm.1	9.721	1581	327	0.059	0.006
CDH1		9.721	797	220	0.039	0.000
CDH1	uc010cfg.1	9.385	797	220	0.043	0.005
EZR	uc003qru.2	9.344	613	358	0.026	0.003
DYNC1H1	uc001yks.1	8.824	1771	329	0.054	0.006
TXNL1	uc002lgg.1	8.777	130	163	0.054	0.006
YWHAB	uc002xmt.1	8.718	78	170	0.103	0.012
	ucoo2xmu. I	8.718	78	170	0.103	0.012
CALM3	uc002eei.2	8 681	235	255	0.079	0.009
CTNNA1	uc003ldh.1	8.679	949	289	0.060	0.007
HNRNPD	uc003hmn.1	8.625	80	230	0.038	0.004
HNRNPD	uc003hmp.1	8.625	80	230	0.038	0.004
HNRNPD	uc003hmo.1	8.625	80	230	0.038	0.004
HNRNPD	uc003hmm.1	8.625	80	230	0.038	0.004
	uco rogxą. r	8.195	745	330	0.050	0.006
ATP50	uc002vtl 1	8 155	103	120	0.050	0.000
YWHAZ	uc003viw.1	8.051	127	409	0.039	0.005
ACOX1	uc002jqe.1	7.924	301	159	0.050	0.006
ACOX1	uc002jqf.1	7.924	301	159	0.050	0.006
ALDH2	uc001tst.1	7.652	615	181	0.042	0.006
YWHAZ	uc010mbr.1	7.469	192	478	0.031	0.004
		7.295	1376	253	0.049	0.007
CTSD	uc001luc.1	7.111	486	216	0.066	0.000
LMNA	uc001fni.2	7.000	441	147	0.048	0.007
LMNA	uc001fnh.2	7.000	441	147	0.048	0.007
LMNA	uc009wro.1	7.000	441	147	0.048	0.007
LMNA	uc001fng.2	6.853	429	147	0.047	0.007
HEXB	uc003kdf.2	6.722	133	149	0.045	0.007
SEC23B	uc00100.1	6.583	319	142	0.047	0.007
SEC23B	uc002wra.1	6.583	319	105	0.063	0.010
CNDP2	uc002llm.1	6.319	771	252	0.075	0.012
ACOT7	uc001amt.1	6.151	2454	227	0.054	0.009
LMNB1	uc003kud.1	6.112	349	474	0.026	0.004
PDIA6	uc002rau.1	5.840	162	172	0.068	0.012
	uc0032ac.1	5 752	3979 1413	301	0.059	0.010
UBE2L3	uc002zva.1	5.649	539	203	0.056	0.007
HADHA	uc002rgy.1	5.494	172	105	0.052	0.010
CANX	uc003mkk.1	5.484	366	223	0.025	0.004
CANX	uc003mkl.1	5.484	366	223	0.025	0.004
GPNMB	uc003swb.1	5.412	153	138	0.039	0.007
		5.412	281	304	0.039	0.007
HSD17B12	uc001mxa.2	5.385	390	105	0.051	0.010
DSP	uc003mxp.1	5.291	326	345	0.015	0.003
CPT1A	uc001oof.2	5.278	1766	329	0.048	0.009
CPT1A	uc00100g.2	5.278	1766	329	0.048	0.009
CRAI	uc004bxh.1	5.034	445	224	0.022	0.004
PCBP2	ucoo1sab.2	4.972	181	150	0.033	0.007
PCBP2	uc001sdc 2	4 972	181	150	0.033	0.007
LAP3	uc003gph.1	4.935	207	227	0.043	0.009
CANX	uc010jlb.1	4.678	286	223	0.021	0.004
PLEC1	uc003zab.1	4.496	3749	217	0.062	0.014
HSP90AA2	uc001ykv.2	4.471	1054	152	0.029	0.007
HSP90AA2	uc001ykv.2	4.471	1054	152	0.029	0.007
	ucoo mili. I	4.470	404 566	193	0.030	0.007
CTNNBIP1	uc001ack.1	4.138	622	198	0.042	0.010
CTNNBIP1	uc001aql.1	4.138	622	198	0.042	0.010
ECHS1	uc001lmu.1	4.016	310	249	0.048	0.012
YWHAZ	uc003yjx.1	3.997	289	385	0.021	0.005
PPIB ENDDD4	uc002and.1	3.900	100	156	0.050	0.013
SINKPU1 PLEC1	ucuu2ktj.1	3.892 3.837	4044	129	0.060	0.016
RPS9P4	uc002adz 1	3.737	118	147	0.025	0.007
RPS9P4	uc002qdy.1	3.737	118	147	0.025	0.007
RPS9P4	uc002qdx.1	3.737	118	147	0.025	0.007
RPS9P4	uc002qdz.1	3.737	118	147	0.025	0.007

RPS9P4	uc002ady 1	3 737	118	147	0.025	0.007
RPS9P4	uc002adx 1	3 737	118	147	0.025	0.007
PDIA3	uc001zsu 1	3 684	152	112	0.033	0.009
PLEC1	uc003zai.1	3.633	5847	292	0.050	0.014
DYNLL1	uc001tvi.1	3.604	323	291	0.037	0.010
FAM83H	uc003vzk.1	3.505	1200	437	0.064	0.018
LMNB1	uc003kuc.1	3.410	278	474	0.014	0.004
RBM14	uc009vrk.1	3.311	474	327	0.051	0.015
RBM14	uc009vrk.1	3.311	474	327	0.051	0.015
PLEC1	uc003zaf.1	3.310	4532	240	0.055	0.017
STX12	uc001bou.2	3.245	208	75	0.043	0.013
ALDOA	uc002dwc.1	3.224	85	274	0.035	0.011
CKAP4	uc001tlk.1	3.202	162	389	0.049	0.015
RAB1B	uc001ohf.1	3,167	90	171	0.056	0.018
FBP1	uc004auw.2	3,109	499	179	0.052	0.017
METAP2	uc001tec.1	2.927	193	113	0.026	0.009
METAP2	uc001tef.1	2.927	193	113	0.026	0.009
PPP1CB	uc002rma.1	2.902	164	272	0.043	0.015
PPP1CB	uc002rmh.1	2.902	164	272	0.043	0.015
DSC3	uc002kwi.2	2.869	107	307	0.009	0.003
DSC3	uc002kwj.2	2.869	107	307	0.009	0.003
PTBP1	uc002lpp.1	2.829	1069	432	0.039	0.014
PTBP1	uc002lps.1	2.829	1069	432	0.039	0.014
PTBP1	uc002lpr.1	2.829	1069	432	0.039	0.014
RTN3	uc001nxm.1	2.812	448	189	0.045	0.016
RTN3	uc001nxq.1	2.812	448	189	0.045	0.016
RTN3	uc001nxp.1	2.812	448	189	0.045	0.016
RTN3	uc001nxo.1	2.812	448	189	0.045	0.016
RTN3	uc001nxn.1	2.812	448	189	0.045	0.016
PLEC1	uc003zah.1	2.784	5688	331	0.050	0.018
PKM2	uc002atw.1	2.707	238	451	0.042	0.016
PKM2	uc002atx.1	2.707	238	451	0.042	0.016
PKM2	uc002aty.1	2.707	238	451	0.042	0.016
PPA1	uc001jqv.1	2.675	106	189	0.028	0.011
SEC23B	uc002wrc.1	2.671	313	88	0.061	0.023
FASN	uc002kdu.1	2.577	2175	568	0.068	0.026
CAPN12	uc002ojd.1	2.476	458	54	0.046	0.019
TOLLIP	uc001lte.1	2.476	1703	278	0.053	0.022
TOLLIP	uc009ycu.1	2.476	1703	278	0.053	0.022
PHGDH	uc001ehz.1	2.438	273	121	0.040	0.017
G6PD	uc004fly.1	2.409	274	165	0.044	0.018
C3	uc002mfm.1	2.404	892	64	0.075	0.031
ANXA2	uc002agn.1	2.373	236	140	0.017	0.007
ANXA2	uc002agl.1	2.373	236	140	0.017	0.007
ANXA2	uc002agm.1	2.373	236	140	0.017	0.007
ANXA2	ucuu2agn.1	2.373	236	140	0.017	0.007
ANXAZ		2.373	236	140	0.017	0.007
	ucoo2agm. I	2.373	230	140	0.017	0.007
		2.300	900	154	0.035	0.015
GOFD KDT22	uc004llx.1	2.200	200	150	0.045	0.020
	ucoo2fiwi.i	2.247	09	40	0.050	0.025
		2.234	500	120	0.041	0.018
	uc0020VX.1	2.204	240	235	0.018	0.008
YWHAO		2.200	153	200	0.000	0.017
RBM14		1 972	100	327	0.000	0.015
RBM14		1 972	199	327	0.030	0.015
RBM14	uc009vrh 1	1 972	100	327	0.030	0.015
RBM14		1 972	199	327	0.030	0.015
RBM14		1 972	199	327	0.030	0.015
RBM14	uc009vrh 1	1 972	199	327	0.030	0.015
KRT80	uc001rzw.1	1.972	289	57	0.035	0.018
IQGAP1	uc002bpl.1	1.936	550	213	0.036	0.019
VCL	uc001iwe.1	1.934	547	230	0.042	0.022
VCL	uc001iwd.1	1.934	547	230	0.042	0.022
PLEC1	uc003zag.1	1.868	4793	282	0.053	0.028
CSNK1E	uc003avm.1	1.827	1943	150	0.037	0.020
ACAA1	uc003chu.1	1.748	90	118	0.044	0.025
ACAA1	uc003cht.1	1.748	90	118	0.044	0.025
ENO1	uc001apj.1	1.722	151	208	0.033	0.019
YWHAQ	uc002qzw.1	1.710	153	157	0.033	0.019
ME1	uc003pjy.1	1.691	510	138	0.049	0.029
ARF4	uc003dix.2	1.640	150	123	0.013	0.008
EFHD1	uc010fyf.1	1.631	1156	138	0.035	0.022
ATP5A1	uc002lbr.1	1.558	107	100	0.047	0.030
RAB1A	uc002sdn.1	1.553	229	97	0.048	0.031
RAB1A	uc002sdm.1	1.553	229	97	0.048	0.031
RAB1A	uc002sdo.1	1.553	229	97	0.048	0.031
EFHD1	uc002vtd.1	1.497	191	26	0.058	0.038
PLEC1	uc003zae.1	1.477	4153	233	0.057	0.039
CD9	uc001qnq.1	1.337	449	191	0.049	0.037
CD9	uc001qnp.1	1.265	450	207	0.049	0.039
TAGLN2	uc001fun.1	1.232	79	146	0.025	0.021
PDIA6	uc002rav.1	1.223	485	113	0.043	0.035
C1orf204	ucuu1tuh.1	1.198	248	33	0.036	0.030
C 10IT204	ucountuh.1	1.198	248	33	0.036	0.030

LRRC15	uc003ftu.1	1.179	311	55	0.064	0.055
ACOT7	uc001amr.1	1.090	2144	38	0.057	0.053
PHGDH	uc001eib.1	1.078	232	25	0.043	0.040
ZWILCH	uc002aqb.1	0.798	188	50	0.032	0.040
KRT80	uc001rzx.1	0.794	423	48	0.033	0.042
KRT80	uc001rzy.1	0.794	423	48	0.033	0.042
ATP5A1	uc002lbt.1	0.766	248	95	0.032	0.042
ACOT7	uc001amq.1	0.719	2115	38	0.057	0.079
TGM3	uc002wfx.2	0.680	441	25	0.054	0.080
EIF2S3	uc004dbc.1	0.660	97	32	0.021	0.031
TGM1	uc001wod.1	0.592	224	106	0.045	0.075
KRT82	uc001sai.1	0.553	152	42	0.079	0.143
PLD3	uc002onn.1	0.476	455	50	0.057	0.120
ATG9B	uc010lpv.1	0.297	768	48	0.025	0.083
PDIA6	uc002raw.1	0.277	682	27	0.041	0.148
ACTA1	uc001htm.1	0.000	81	298	0.000	0.007

Table S4.1

 \mathbf{R}_{s} values for a list of genes whose proteins have been detected in modern hair shaft.

We selected original gene accessions from Lee et al. [2006], filtering for genes showing sufficient sequence coverage.

Gene	UCSC	Cov(G	Cov(PM)	Rs
MAP2K3 CAPN9	uc002avu.1 uc009xfg.1.uc001hua.1.uc001htz.1	1029 697	95 85	0.708 0.707
IGSF9B	uc001qqy.1	747	132	0.707
MYH3 RAB3II 1	ucuu2gmq.1 uc001nsp.1	433 784	36	0.707
NFIC	uc002lxq.1	1471	225	0.706
KIAA0802	uc010dkw.1	95 335	268	0.705
ATP6V1B1	uc002shi.1,uc010fdv.1,uc002shj.1	265	35	0.704
FHAD1	uc001awe.1	560	58	0.704
CTBP1	ucuusmkx.1,ucuusmku.1 uc003act.1	269 1674	102	0.704
ACTN4	uc002ojb.1	173	152	0.703
PDE6B SLC11A1	uc003gao.2,uc003gap.1 uc002vbw.1 uc002vby.1	1649 428	132 41	0.703
PLEKHA7	uc001mmn.1	363	34	0.702
	uc001lxj.1,uc009ydv.1	231	146	0.702
SCN3B	uc001pzb.1,uc001pza.1	190	213	0.701
TNR	uc001gkp.1	458	37	0.700
GRIK4	ucoozwik. i	3407	86	0.698
BNPI	uc002pno.1	216	181	0.698
KIAA1855	ucuu2ypw.2 uc003our.2	147	57 133	0.698
CNGA3	uc010fij.1	323	25	0.697
ZNF714 SIGLEC1	uc002npl.2,uc010ecp.1,uc002npo.2	106 537	59 103	0.696
MB	uc003anz.1,uc003aoa.1	159	63	0.693
PRG-3	uc010mtc.1	150	26	0.693
DKFZp762	uc004cet.1	686	104	0.693
NFKBID	uc002och.1	104	108	0.692
vWF-CP	ucuuseqe.1,ucuuseqt.1 uc004cdp.2	294 1536	37 43	0.692
UPK1B	uc003ecc.1	113	26	0.690
SLC38A10 SLC22A2	uc002jzy.1 uc003ate 1 uc003atf 1	2553 272	165 125	0.689
GPR56	uc002emg.2	585	52	0.689
	uc002xbi.1	1234	25	0.689
ESF1	ucoosvsa. 1,ucoo Ibin.2,ucoosvsa. 1,ucoo Ibin.2 ucoo2woj.1	213	110	0.689
AACS	uc009zyi.1	1081	60	0.688
TMPRSS13	uc010bx2.1 uc009vzr.1.uc001prt.1	288	41 54	0.688
FAM60A	uc001rke.1,uc001rkd.1,uc001rkb.1	193	398	0.687
RAB19 FAM125B	uc010lni.1,uc003vvr.1 uc010mxd 1	163 2603	28 49	0.687
ZNF57	uc002lwr.1	151	166	0.687
TDRD12	uc002ntr.2,uc002ntq.2	483	300	0.686
FAS/ER	uc000kit.1	341	26	0.686
FAM107B	uc001ina.1,uc001imx.1	1750	45	0.686
PSMA8	uc002kvr.1,uc002kvp.1,uc002kvp.1	168	69	0.685
ZFHX2	uc010akg.1	104	61	0.684
XHRIP110	uc003/mi.2 uc003mff.1	169	32 33	0.684
DLGAP3	uc001byc.1	734	76	0.683
C1orf177		221	338	0.683
ZNF506	uc002noh.2,uc010eci.1	152	83	0.683
SLC39A4	uc003zcq.1,uc003zco.1,uc003zcp.1	334	243 50	0.682
C11orf52	uc001pmh.1	113	77	0.681
SMURF1	uc003upt.1 uc002rob 1 uc002roa 1	364	62 62	0.681
AGT	uc009xff.1,uc001hty.2,uc009xfe.1	279	95	0.681
TGM3	uc002wfx.2	441	25	0.680
PHLDB3	ucoo nex. 1,ucoo nex. 1,ucoo new. 1,ucoo new. 1,ucoo neu. 1 uco10eit.1	287	20 65	0.679
CEP192	uc002krw.1	318	27	0.679
hBSSP-4	ucousings. i ucousings. i	178	143	0.679
PRSS22	uc002cry.1	178	161	0.678
KIAA1486 CDH26	uc002vor.1 uc002vbg.1.uc002vbf.1	326 155	182 35	0.678
DNMT3L	uc002zeg.1,uc002zeh.1	676	109	0.677
ZC3H14 PRICKLE4	uc001xwz.1 uc003ore 1	193 263	28 89	0.677 0.677
ABCB9	uc001udr.2	461	120	0.677
FLJ00133	uc002wak.2	437 343	180 87	0.677
RNF17	uc001ups.1,uc001upr.1	302	84	0.675
C17orf64	uc002iyq.1	120	45	0.675
LZTS1	ucoosgcm. i ucoosgcm. i	276	121	0.675
MYT1	uc002yih.2,uc002yij.1	1943	66	0.674
PLXNB3	ucuuzwyi.1,ucuuzwyK.1 uc004fii.1	95 668	32 72	0.674
CACNA1H	uc002ckv.1,uc010brj.1,uc002cku.1	1058	344	0.674
BMI1	ucuungpw.n,ucuungpv.n,ucuungpu.n uc001irh.1	201 112	377	0.673

OBSL1	uc010fwl.1	485
AZU1	uc002lpz.1	124
HNMT SLC34A1		97
BRD1	ucoostiv.2	2446
STK17B	uc010fsh.1	417
GRHL3	uc001biz.1,uc001biy.1	286
DKFZp586B		423
CALCOCO2	uc002iof.1	169
EIF2C1	uc001bzk.1	399
DKEZn686E		225
C4orf44	uc003ggs.1,uc003ggt.1	266
TMPRSS4	uc001psd.2,uc009yzt.1	418
PHKG1 BC042092		257
IL34	uc002ezi.1,uc002ezh.1	262
ARRDC2	uc002nhv.1	115
		5621
ADRA1A	uc010lum.1,uc003xfe.1,uc010lu.1,uc003xfc.1,uc003xfn.1,uc003xfg.1	338
AK096230	uc001ukg.1	136
NAV2	uc009yhz.1	407
INPP5D	uc002vtw.1.uc002vtv.1	2060
KIAA1075	uc001sbo.1,uc001sbm.2	178
FBXO46		513
BC132948	uc002vvm.1	214
ALS2CL	uc003cpx.1	139
TP73	uc001aks.2,uc009Vk.1,uc001akr.2	1315
WDR5		507 876
UNC84B	uc010gxs.1	546
EIF2S3	uc004dbc.1	97
		88
HSPA12B	uc002wje.1,uc002wjd.1	781
ITGA2B	uc002igu.1	186
PKD11 2	uc0101ru.1 uc002fai 1 uc002fab 1	719
SV2B	ucO2bqv.1	426
GRK1		190
NTN3	ucou rygv. r,ucou rygu. r ucou2cai.1	369 140
PRPF40B	uc001rus.1,uc001rur.1	118
CEL	uc010naa.1 uc002voi 1 uc010ckc 1	265
P2RX1	uc002yei. 1,uc002yei. 1,uc010gkc. 1 uc002fww.1	425
ATXN7L1	uc003vdg.2,uc003vdf.1	187
PGLYRP2	uc002nbg.2,uc002nbf.2	313
ACRC	uc004eae.1	134
VRL	uc002gpz.1	280
I RPV2		280
TEX14	uc010dda.1,uc010dcz.1,uc002iws.1,uc002iwr.1	1079
SERPINA3	uc001ydo.2	286
CAV3		1828
EIF4G1	uc003fnx.2	292
S100Z	uc003keq.2,uc003kep.1	291
SUT-1	ucuusiww.1 uc003vtc 1	326 239
RGS14	uc003mgf.1,uc003mgi.1,uc003mgh.1	609
DAGK1		124
EYA2	ucousvuk.2 uc002xsn.1.uc002xso.1	1765
FLJ00369	uc002osc.2	1461
OAT4L	uc009ypt.1	154
TRXR2A		211
SLC27A5	uc002qtc.1,uc010eus.1	295
RHBDL2		368
WISP1	uc010med.1.uc010mec.1.uc010meb.1.uc003vuc.1.uc003vub.1	448
TRAPPC9	uc010mel.1	6978
ALDH1L1	uc003eio.2,uc003ein.1	445 1115
PTPRE	uc001lkd.1	819
SHANK1	uc002psx.1,uc002psw.1	2072
ENIL2 ARC41	ucuu2pcq.1 uc003uge 1	802 495
C14orf4	uc001xsy.1	239
WFIKKN2	uc002isv.2,uc010dbu.1	238
FBXO47	ucu03xff.1 uc002hrc.1	118
CRYBB1	uc003acy.1	205
	uc001lbf.2,uc001lbi.1,uc001lbh.1,uc001lbg.1	254
TSSC6	ucou ripj. r,ucou ripi. r,ucou ripi. r,ucou ripg. r uc009vdk.1	336
PABPN1	uc001wjh.2	327
FLJ00059	uc003uwv.1	221 1/0
ARPC4	uc003btb.1,uc003bta.1,uc003bsz.1	126

 $\frac{14744}{102} \\ \frac{1613}{112} \\ \frac{1253}{122} \\ \frac{1253}{33} \\ \frac{13448}{132} \\ \frac{1202}{33} \\ \frac{1153}{112} \\ \frac{1253}{312} \\ \frac{1202}{33} \\ \frac{1153}{112} \\ \frac{1202}{122} \\ \frac{120$

0.671 0.671 0.670 0.669 0.669 0.669 0.669 0.669 0.665 0.655 0.654

SERPINA6	uc001ycv.1	146	28	0.639
CES7	uc002eip.1,uc002eio.1	155	33	0.639
NLRP1 PPFIA1	uc010clh.1,uc002gcg.1,uc002gch.2,uc002gcl.1,uc002gcj.1,uc002gck.1,uc002gci.1 uc001opr.1	337 207	43 44	0.638
RHBDF2	uc002jrs.1	255	133	0.637
BTBD2	uc0012m2.1,uc0012my.1 uc002luo.1	290	29 231	0.637
NBR1 WNK4	uc002idj.1,uc002idk.1	113 117	72 205	0.637
SSH1	uc001tno.1	201	48	0.637
SERPINE2 KLK15	uc002vnt.1 uc002pto.1.uc002ptn.1.uc002ptn.1.uc002ptl.1	438 170	44 48	0.636
MUC17	uc003uxp.1	255	54	0.635
IL2RA	uc0010pk.1 uc009xih.1,uc001iiz.1	358	31	0.635
RAPGEF1 CERCAM	uc010mzm.1,uc010mzl.1,uc010mzs.1,uc010mzr.1,uc010mzg.1,uc010mzp.1,uc010mzo.1,uc010mzn.1,uc004cbc.1	540 706	75 56	0.635
C21orf56	uc002zii.1	763	175	0.634
DPEP2	ucu02010.1 uc002eve.1	232	64 49	0.634
INPP5A ZNE236	uc001llq.1	8836 1840	173	0.633
HYDIN2	uc002ers.1	294	143	0.632
BC061638 TJP3	uc009zrx.1 uc002lvk.1	399 1065	63 82	0.632
CAMLG	uc003kzu.1,uc003kzt.1	99	125	0.631
KIAA1618	uc002pdu.1 uc002jyg.1	831	121	0.631
SMCR7L ZEHX4	uc003axw.2 uc003yaw 1	333 434	45 57	0.631
MYOC	uc001ghu.1	108	34	0.630
ZMIZ2 TMEM111	uc003tis.1,uc003tit.1 uc003buo.2,uc003bun.1	365 97	53 61	0.629
GPR77	uc010ela.1	210	48	0.629
TINAGL1	uc001bta.1	228	91	0.627
TGFBRAP1	uc010fjc.1 uc001yaz 1 uc001yaz 1	287 83	36 52	0.627
ELFN1	uc010ksg.1	224	421	0.626
DKFZp434 KIAA2034	uc002kdy.2 uc010env.1	895 623	58 65	0.626
ZBTB46	uc002ygv.1	2998	113	0.626
F7	uc001vsw.1,uc010agp.1	1037	108	0.625
PPP1R16A ILDR1	uc003zdf.1,uc003zdd.1 uc003eeg.1	429 152	119 43	0.624
CDH23	uc001jsc.1,uc001jsi.2,uc001jsh.2,uc001jsg.2	617	36	0.622
MYL6B	uc0020kw.2 uc001sjt.1,uc001sjs.1	423	374 69	0.622
FAM30A FHOD3	uc001ysr.2,uc001ysg.2 uc010dmz 1	656 1242	94 34	0.621
TFAP2C	uc002xya.1	139	431	0.620
CUX2 KIAA1669	uc001tsb.1 uc003bka.1	2520 283	62 193	0.620
UNC84A	uc003sji.1	836	88	0.620
TRIM63	uc001bli.1	92	38	0.620
FLJ00074 hSK4	uc001bbd.1 uc010eiz 1	143 160	59 173	0.619
KCNE3	uc001ovc.1	75	139	0.618
GIP	uc002vtr.1 uc002iol.1	291 81	30	0.617
	uc002sup.1	2094	34 35	0.617
C5orf38	uc003idc.1	132	244	0.616
SRCRB4D AK128833	uc003ufb.1 uc001acx.1	607 190	34 156	0.616 0.616
LRIT1	uc001kcz.1	156	32	0.615
GMEB2	uc002yfp.1	1133	114	0.613
DBN1 FGFR2	uc003mgz.1 uc009xzs.1 uc009xzr.1	297 809	238 55	0.613
C11orf41	uc001mun.1,uc001mup.2	352	38	0.612
LRRC24	uc010Kxr.1 uc003zdn.1,uc003zdm.1	495	74 528	0.612
C19orf54	uc002oou.1,uc002ooy.1	186	34	0.609
KCNIP2	uc009xwv.1,uc009xwu.1,uc001kuf.1,uc001kue.1,uc001kud.1,uc001kuc.1,uc001kub.1,uc001kug.1	479	136	0.608
KCHIP2 KLF17	uc001kua.1 uc001clp.1	479 102	136 62	0.608 0.608
RPL3L	uc002cnh.1	264	126	0.607
BC026998	uc002gte.2	179	31	0.606
SERPINB6 ACTG2	uc003muk.1 uc010fev.1.uc002siw 1	245 132	27 40	0.606
MED1	uc002hru.2,uc002hrv.2	352	64	0.606
FOSL1	ucuuyzxp.1 uc001ogg.1	85	∠5 309	0.606
SLC22A6	uc001nwm.1,uc001nwl.1,uc001nwj.1,uc001nwk.1	97	47	0.606
UROC1	uc010hsi.1,uc003eiz.1	814	56	0.605
LOC645545 KLHL30	uc010ddb.1 uc002vxr.1	333 566	72 224	0.605 0.604
AMBP	uc004bie.2	149	72	0.604
GLISS	ucuuszny, i,ucuuszic, i,ucuuszitz, i	102	217	0.004

GCNT7	uc002xxw.2	418	28 104	0.603
PLOD3	uc010lhs.1	269	162	0.602
FOXRED1 HAL	uc001qdk.1 uc001tem.1	123 131	37 90	0.602
GALNTL2	uc003car.2	366	33 115	0.601
P/OKcl.13	uc010mrd.1	429	106	0.600
C1orf167 MUC20	uc001ata.2 uc010bzo 1	170 316	51 54	0.600
NOD27	uc002ekp.1	328	49	0.598
BARHL1	uco10mm.1,uco10mm.1,uco02smq.1,uco02smp.1 uc004cbp.1	375 447	64 160	0.597
NLGN3	uc004dzd.1	114 150	51 161	0.596
SOHLH2	uc001uvj.1	108	161	0.596
KIAA0999 EML1	uc001ppw.1 uc010avt.1	143 2293	31 135	0.596
FLJ00009	uc003uwu.1,uc003uws.1	292	87	0.596
NLRP5	uc002qmi.1	698	43	0.596
GLTSCR1 GABRB3	uc002phi.2 uc001zbb.1	1133 3074	428 29	0.595
TET3	uc002skb.2,uc010fez.1	691	112	0.594
C12orf34	uc001tpf.1	702	20 61	0.594
SYDE1 HIP1R	uc002naj.1 uc001udk 1	221 252	223 184	0.594
SLC41A3	uc003eii.1	492	56	0.592
P2X	ucousego.1 uco10ckm.1	1301	56 82	0.592
PLA2G2F	uc009vpp.1	142	36 106	0.592
PCDHGA5	uc003lju.1	3468	144	0.591
ABCA13 STK24	uc010kyu.1 uc001vnm.1	430 1201	186 59	0.590 0.590
DHX33	uc002gbz.1	190	42	0.589
SERPIND1	ucoostan i ucoo2ztc.1	84	33	0.589
DKFZp313K ATXN7L3	uc002jew.2 uc002jga 1 uc002ifz 1	280 87	33 64	0.589
DUSP22	uc003msy.1	1047	88	0.588
TBL1X	ucouzagc.1 uc004css.1	398	54 26	0.588
FLJ00010 PCDHGB7	uc003uwr.1	326 1961	87 157	0.587
KIAA1274	uc001jre.2	486	95	0.586
TORBV2S1 TNFRSF18	uc003vzp.2 uc001add.1,uc001adb.1	765 95	46 167	0.586
HRIHFB200	uc001avr.2	1494	35	0.586
SIX3	uc002run.1	103	241	0.585
TCL6 KIAA0447	uc001yev.1,uc001yeu.1 uc001abe 2	106 118	31 69	0.585
SLC24A6	uc001tvb.1	344	86	0.583
RDH10	uc001uid.1 uc003xzj.1	944 103	55 30	0.583
BV13S6J2. NSUN4	uc010lnu.1 uc009vvg 1 uc009vvf 1 uc001cpr 1	103 146	30 85	0.583
SP6	uc002img.1	232	135	0.582
ADCY6	uc003yue.1 uc001rsh.2	252	48 213	0.582
SLC23A1	uc003leh.1,uc003leg.1	269 161	78 35	0.580
MID1	uc004cub.1,uc010ndy.1,uc004ctw.2,uc004ctv.2,uc004ctu.2,uc004ctt.2,uc004ctt.1,uc004ctn.1,uc004ctm.1	88	34	0.580
SPDYA WDR85	uc002rmk.1,uc002rmj.1 uc004cnj.1,uc010ncl.1	99 188	172 79	0.579
C10orf71	uc001jhp.1	395 156	57 120	0.577
RLTPR	uc002etn.1,uc010cel.1	831	213	0.577
GALN16 SULT4A1	uc001ryk.1 uc003bed.1	336 642	31 167	0.577
	uc002lvh.2	288	199 182	0.576
KCTD1	uc002kvv.1	1706	40	0.574
RNF165 CLDN10	uc002lby.1 uc001vma.1	1805 921	37 48	0.574
FXY	uc004cty.2	89	34	0.573
R3HDML	uc004cent 1 uc002xls.1	110	63	0.573
PHAX PRSS35	uc003kua.1 uc010kbm.1.uc003piz.1	154 77	147 44	0.573
ITIH3	uc003dfv.2	217	62	0.571
FGF6	uco Toeor. 1, ucoo2pux. 1 uc001qmr.1	103	98	0.571
OVCH1 CACNA1A	uc001rix.1 uc002mwx.2	137 1480	65 41	0.569
CHRNE	uc002fzk.1	295	195	0.567
CCDC144N	ucou rocs.1 uc002gyf.1	106	328 36	0.566
GPRC5C DYRK4	uc002jkt.2,uc002jks.1 uc009zeh 1	244 277	113 57	0.566
RHOBTB2	uc003xcp.1	792	28	0.566
C8orf82	ucuu2orr.1,ucuu2ors.1 uc003zdq.1	87 163	131 674	0.565
MGC70857	uc003zdp.1	163	674	0.564

uc001asi 1
4000 1401.1
uc003jby.1
uc010clb.1
uc002gbh.2,uc002gbi.2
uc010jpr.1
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PLA2G5 EBI3 IQCA1 GTF2A1L MX2 POLR1A NRG1 TUSC5 TM9SF4 AMFR PAX1 TM4SF5 TKT ATP2C2 ABHD12B UBE20 ANKS6 GCLC DKFZp686 DUS1L SLC25A21 URFN4 BEST2 TRAF3IP3 C11orf9 KIAA1234 TNPO2 JMJD5 SASH3 TOP3A ZHX3 PCDHA5 SASH3 TOP3A ZHX3 PCDHA5 SASH3 TOP3A ZHX3 PCDHA5 SASH3 TOP3A ZHX3 PCDHA5 SASH3 TOP3A ZHX3 PCDHA5 SASH3 TOP3A ZHX3 PCDHA5 SASH3 TOP3A ZHX3 PCDHA5 SASH3 TOP3A ZHX3 PCDHA5 SASH3 TOP3A ZHX3 PCDH45 SASH3 SASH3 TOP3A ZHX3 PCDH45 SASH3 SASH3 TOP3A ZHX3 PCDH45 SASH3 SASH3 TOP3A ZHX3 PCDH45 SASH3 SASH
PLA2G5 EBI3 IQCA1 GTF2A1L MX2 POLR1A NRG1 TUSC5 TM9SF4 AMFR PAX1 TM4SF5 TKT ATP2C2 ABHD12B UBE20 ANKS6 GCLC DKFZp686 DUS1L SLC25A31 LRFN4 BEST2 TRAF3IP3 C110rf9 KIAA1234 TNP02 JMJD5 SASH3 TOP3A ZHX3 PCDHA5 SAMM50 SERPINF1 DA0 CAPN8 SLC25A24 VESPR LOC91948 KIAA1835 AK125726 KCNQ1 SPNS3 ZNF564 CAPZB
PLA2G5 EBI3 IQCA1 GTF2A1L MX2 POLR1A NRG1 TUSC5 TM9SF4 AMFR PAX1 TM4SF5 TKT ATP2C2 ABHD12B UBE20 ANKS6 GCLC DKFZp686 DUS1L SLC25A31 LRFN4 BEST2 TRAF3IP3 C110f9 KIAA1234 TNPO2 JMJD5 SASH3 TOP3A ZHX3 PCDHA5 SAMM50 SERPINF1 DA0 CAPN8 SLC25A24 VESPR LOC91948 KIAA1835 AK125726 KCNQ1 SPNS3 ZNF564 CAPZB TSGA10IP

1143 179 1449

 $\begin{array}{c} 235\\ 792\\ 209\\ 181\\ 1778\\ 595\\ 153\\ 121\\ 212\\ 99\\ 1120\\ 1014\\ 92\\ 1949\\ 660\\ 1629\\ 77\\ 6450\\ 212\\ 237\\ 118\\ 358\\ 361\\ 114\\ 141\\ 257\\ 6100\\ 470\\ 316 \end{array}$

 $\begin{array}{c} 1766\\ 444\\ 29664\\ 3822\\ 7592\\ 232\\ 6855\\ 495\\ 497\\ 682\\ 117\\ 754\\ 117\\ 288\\ 9109\\ 438\\ 766\\ 094\\ 3766\\ 092\\ 312\\ \end{array}$

0.562 0.561 0.561 0.561 0.561 0.551 0.554 0.558 0.555 0.5554 0.555 0.5554 0.555 0.552 0.55
AK310432	uc009vII.1	259	96	0.519
CMIP	uc002fgg.1	4964	43 63	0.518
SPTBN5	uc001zos.1	1589	95	0.518
NKX6-3 FLJ00275	ucu1uixa.1 uc002gfa.1	3067	62 61	0.518
CD4	uc009zfc.1,uc001qqv.1,uc009zez.1	260	48	0.517
AMI 2	uc002a0i.1 uc009vrl 1	89 946	276	0.517
TMEM108	uc003epm.1	91	94	0.516
KIAA0702	uc002apg.1,uc010bhm.1	194	25 47	0.515
SLC26A9	uc001hdo.1	198	68	0.515
C7orf27	uc003smh.2	188	172	0.515
UHRF1BP1	ucuoziro.z	273	26	0.514
PEBP4	uc003xcn.1	2579	32	0.513
PHOX2A RASGRP4	uc001osn.2 uc010ept 1 uc002oir 2	285 293	219 50	0.512
SLC18A1	uc003/3/2012	127	26	0.512
PARP14		127	26 76	0.512
FKSG71		88	30	0.512
PLA2G12B		88	30	0.511
NLRP7 VGLL2	ucu10esk.1,ucu02qin.2,ucu02qin.2,ucu02qig.2 uc003axo.1	356 270	39 138	0.511
AFAP1L1	uc003lqi.1	184	47	0.511
C4orf29	uc010inz.1,uc003ifu.1,uc003ifu.2,uc003ifu.2,uc003ifu.2,uc003ifu.2,uc003ifu.2,uc003ifu.2,uc010aig.1,uc010aig	94 300	32 181	0.511
OTOLE		500	101	0.511
FAM107A	uc003dko.1	635	54	0.510
TRIM2	uc003inh.1	961	28	0.510
CD226	uc002lkm.2	365	31	0.510
BPI	uc002xib.2	242	46	0.507
PCDHGA1 ASAP3	uc003) .1 uc001bbb 2	4289 150	54 57	0.507
SPATA16	uc003fin.2	545	46	0.506
MEGF2	uc010hkt.1	177	64	0.506
AK094715	uc003pta.1	126	53	0.505
BIN2	uc001ryh.1.uc009zlz.1.uc001ryg.1	248	143	0.505
LILRB5	uco Tocky 1 ucoO2aey, 1,ucO02aey, 1	119	35 60	0.505
GATA4	uc003wub.1	2101	45	0.503
PRSS16 NCF4	uc003njb.1,uc003nja.1 uc003anz 2,uc003anz 2	163 281	41 77	0.503
FCGBP	uc002omp.2	1130	36	0.502
NKX6-1	uc003hpa.1	75	301	0.502
DKFZp434B		224	159	0.502
KIAA0467	uc001cjk,1,uc001cjl.1	499	40	0.501
MS4A10		263 135	94 45	0.500
PTPRU	uc001brx.1	88	44	0.500
NM 001001 FPAS1	uc002yxh.1 uc002ynw 1	390 78	30 39	0.500
BCR-ABL	uc010gtx.1	900	88	0.500
BSPH1		107	32	0.498
IFNGR2	ucoupsc. 1/2004 pt.2.2 ucoupsc.2004 pt.2.2	259	43	0.498
ZNF718	uc003fzt.2	621	581	0.497
MEFV	ucuusjow.z ucQuzeun.1	236	65	0.496
COLQ	uc003bzv.1	353	25	0.496
KCNN4 SLC25A19	uc0020x1.1 uc0020is 2 uc010dge 1	386 231	51 52	0.495
SPATA20	uc002irf.1,uc002ire.1,uc002ire.1	248	135	0.495
	uc001sep.1	330	136	0.495
CEACAM20	uc010ejg.1,uc010ejp.1,uc010ejp.1,uc010ejn.1	151	32	0.495
CASS4	uc002xxp.1,uc002xxr.1,uc002xxq.2	506	30	0.494
XKR9	uc00101ze_1.uc00101zd_1.uc003xvg.1	141	139	0.493
BC111724	uc002cuf.1	141	139	0.493
ZNF717	ucuu <u>s</u> gaw.1 uc010hoa.1	207	34	0.493
CACNA1B	uc004coh.1	2334	60	0.493
PROZ		326 470	111 33	0.492 0.401
CNGB1	uc010cdh.1,uc002emt.1	1276	44	0.491
KIAA1123	uc002isj.2	149	122	0.491
FRG1	ucoo3izs.1	220 89	240	0.491
DOCK4	uc003vfv.1	204	25	0.490
ZNF232 BOC	uc002gat.1 uc003eac 2	403 121	158 83	0.490 0.490
DKFZp547P	uc001zbi.1	212	135	0.490
SLC7A9	uc002ntw.2,uc002ntu.2,uc002ntv.2	564 804	48	0.489
PPP1R14A	uc010efv.1,uc002ohq.1	80	234	0.488
MAN1B1	uc004clf.1,uc004cle.1	382	62	0.487
SPG20	ucuuzwap.z uc001uvo.1	305	83 37	0.486
C20orf107	uc002xxy.1	99	32	0.485
ARRDC5	uc002mbm.1	181	39	0.485

DYNC2LI1	uc002rtl.1,uc002rtk.1,uc002rtj.1	95	46	0.484
dJ402G11.5	uc010hap.1,uc003bjy.1	537	195	0.484
GRM2	uc003dbp.1	204	69	0.483
PEAR1 HOXD3	ucuu110.1 uc002.044 1	546 232	111	0.483
CD3001 F		140	27	0.403
PRKAG2	uc003wkj.1	5405	75	0.482
FRMD4A	uc001imu.1	2489	38	0.481
_IL7	uc003ybg.1	131	126	0.481
ZFAT	uc010meh.1,uc003yuo.1,uc003yun.1,uc003yur.1	1968	187	0.480
		313	50 36	0.479
NPHS2		82	157	0.478
LDB3	uc009xsv.1.uc001kds.1.uc001kdr.1.uc001kdv.1.uc001kdv.1	604	51	0.478
LIG4	uc001vqp.1	136	195	0.478
SNCG	uc001keb.1	76	121	0.478
HEFL	uc010gio.1	506	29	0.478
NKX2-5		181	259	0.477
		039 455	39 50	0.470
PTPRS		216	57	0.475
TXNDC3	uc003tfn.1	93	44	0.473
RNB6	uc010avu.1	1598	28	0.473
LOC201175	uc002iiz.1	444	105	0.473
ZNF284	uc0020vg.1	86	61	0.473
		730	95	0.473
TAF15		218	103	0.472
PVALB	uc003apx.1	392	37	0.472
DDC	uc010kza.1	677	69	0.471
ACSBG1	uc002bdh.1	821	43	0.471
KIAA1569		397	28	0.470
GL125D1 C10orf110		3/5	47	0.470
PCDHGB5	uc003kf 1	2529	138	0.408
CCDC79	uc002edd.1	109	102	0.468
BAG4	uc003xkz.1,uc003xky.1	202	157	0.466
UBE3B	uc001tos.1	530	38	0.466
CLCN2	uc003foh.2	260	69	0.464
	ucou2mtr.1	96	89	0.464
ZC3H12D	uccosnint. i, uccosnins. z, uccosnint. i, uccosnint. i uccosnint.	02 247	30 88	0.403
EPB42	uc001zg.2	81	75	0.463
ABCD1	uc004fig.1	128	79	0.463
NINJ2	uc001qil.1	1395	28	0.462
PECAM1	uc002ieg.1	585	40	0.462
SGULZ SAD120		78	72	0.462
CD300A		254	91	0.401
TH	uc001lyt 1.uc001lys 1.uc001lyr 1.uc001lyr 1.uc001lyr 1.uc001lyr 1.	365	96	0.460
SFTPB	uc002sqj.1,uc002sqi.1,uc002sqh.1	174	32	0.460
ZNF595	uc003fzv.1	158	581	0.460
RAB11FIP1	uc003xkp.1	99	26	0.460
	uc0003tp.2	1371	40	0.460
LRRC37B		102	35	0.458
CAPN3		125	80	0.457
CPN2	uc003fts.1	259	93	0.457
IL22RA2	uc003qhn.1,uc003qhm.1,uc003qhl.1	124	34	0.457
CR617046	uc001myp.2	148	146	0.455
IEPP SSD2		346	35	0.455
RTN1		397	219	0.450
PIP5KL1	uc004bsu.1	175	159	0.454
GPIHBP1	uc003yxu.1	116	171	0.454
C7orf51	uc003uve.1	222	181	0.453
ACVRL1		217	86	0.453
PIK3R6		400 640	35	0.452
SELO	ucoo3biz.1	292	117	0.451
ATP1A4	uc001fve.2	234	35	0.449
CYP27C1	uc002tod.2	284	100	0.448
MST091	uc002yqq.2	186	50	0.448
KIAA1132		4245	98 47	0.448
C20orf197		226	56	0.448
CRHR2	uc003tbp.2	655	46	0.445
PPP1R1B	uc002hsc.1,uc002hsb.1	90	32	0.444
MRP6	uc002dem.1	90	60	0.444
IUBB3	uc00265.2	1043	240	0.444
KIF25		504 686	/ 1 85	0.443
DUSP18	ucoodani - jucoodani - j	194	143	0.442
LPIN2	uc002klo.1	643	42	0.441
ASZ1	uc003vjb.2	113	149	0.440
TOP3b	uc002zvr.2.uc010gtm.1	332	31	0.436
PRSS36		405	197	0.432
RDS6KA1		02 315	62	0.432
arh7V		164	101	0.431
EDC4	uc002eus.1	202	87	0.431
CELSR3	uc010hkg.1	405	58	0.430
KAT2A	uc002hyx.2	270	116	0.430
KIAA1501	uc0021qc.1	599	147	0.429
NK4A1		331	/1	0.429
LIFUL	ucoosaun. i	109	29	0.428

KIAA1787	uc002ggc.1	173	84	0.425
ADAMTS17	uc002bvx.1	3868	33	0.424
NM 001127	uc010nmy.1	319	45	0.423
	uc002wlg.1	188	159	0.423
C1orf186	uc001hdt.1	278	47	0.423
MCM5	uc003anw.1	354	52 25	0.422
BOLL	uc004eay.2,uc004eax.2 uc002uut.2,uc002uus.2,uc002uuu.1,uc002uur.2	91	460	0.421
C6orf146	uc003mvy.1	270	31	0.421
CNTN2	uc001hbs.1	486	71	0.421
AX721097	uc002fji.2	86	48	0.419
C21orf63	uc002ypu.1	558	29 35	0.418
SCML4	uc003pry.2	192	37	0.418
KLK8	uc002e0.1	92	134	0.416
FAM83E	uc002pjn.2	481	153 84	0.416
ALPL	uc001beu.2	453	25	0.414
RBM47		790	28	0.414
HSF5	uc002iwi.1	324	401	0.413
DNAJC14	uc001shu.1	461 306	57 56	0.412
WDR59	uc003yde.1	447	35	0.411
NUP210L	uc001fdw.1	849 186	74 87	0.411
EXOC3L	uc002erx.1	348	79	0.409
PCDH8	uc001vhj.1,uc001vhi.1	285	582	0.408
ETV6	uc001raa.1	1387	49	0.406
CCDC64B	uc002ctf.2,uc002cte.2	386 494	67 25	0.405
TCF4	uc002lgb.1	267	36	0.404
VAX1	uc009xyx.1,uc001ldb.1	220 312	400	0.404
ARX	uc004dbp.2	413	83	0.402
ST3GAL2 MYL5	uc002eyw.2 uc003gay 1	627 196	84 105	0.402
graf-2	uc003ili.1	337	27	0.401
ADAM2 akin	uc003xnl.1,uc003xnk.1,uc003xnl.1 uc001oee 1	162 245	54 49	0.400
XAF1	uc002gdo.1,uc002gdn.1,uc002gdr.1,uc002gdq.1,uc002gdp.1	85	68	0.400
CGB8	ucu1uncc.1 uc002pmc.1	317 89	38 105	0.400
CETP	uc002eki.2,uc002eki.2	372	64	0.393
FOXN1	ucu10iwq.1,ucu03xmn.2 uc002hbj.1	208	37 109	0.393 0.393
CHRFAM7A	uc010baj.1,uc010bak.1	229	30	0.393
PCDHGB1	uc003ljo.1	3925	90 85	0.393
KIAA1218	uc003vdh.2	786	41	0.391
SEPX1	uc002cng.1	118	184	0.391
LYST FAM13C	uc001hxi.1	308	40 57	0.390
KIAA0148	uc001pp.1	207	46	0.389
WNT5B	uc009zdg.1	1291 76	35	0.389
LAX1	uc001haa.1	80	31	0.388
RIMKLB MYST1	uc001quu.2	792 90	34 52	0.386
AGAP3	uc003wiji.1	417	117	0.383
HOM-TES- SDC3	uc001gpg.2 uc001bsd 2	159 131	73 60	0.383 0.382
ABCC3	uc002isn.1	228	26	0.380
C10orf108	ucuu3pqn.2,ucu10kcw.1,ucuu3pqp.2,ucuu3pqo.2 uc001ifr.2	734 211	89	0.380
ZNF580	uc002qlm.1	506	64	0.379
CA5A	uc002evs.1 uc002fkn.1	1016	69 81	0.379
CYP4F12	uc002nbl.2	111	30	0.378
DUXA	uc002nev.2 uc002goa.1	116	28 29	0.377
TP53	uc002gih.1,uc002gig.1	98	55	0.374
SPAG4L	ucuu3qrw.1 uc002wyi.1	242	38 45	0.373
SLC6A12	uc001qhy.2,uc001qhx.2	147	41	0.372
TNNT1	uc002qjd.2,uc002qjc.2,uc002qjb.2,uc002qjf.2	400	299	0.372
FAM151A	uc001cxn.1	158	35	0.369
OGT	uc004eau.2 uc004eac.2	95	28 28	0.368
MIOX SNX20	uc003bln.1,uc003bln.1,uc003bll.1	125	138	0.368
ADAT1	uc002egn.1,uc002egn.2 uc002fep.1,uc002fe0.1	164	120	0.366
BEAN	uc002eog.1	241 343	80 48	0.365
PIWIL2	uc010ltv.1,uc003xbn.1	440	160	0.364
SLC5A11 CATSPER4	uc010bxt.1,uc002dmu.1,uc002dmt.1,uc002dms.1	473 110	43 40	0.364 0.364
GSG1	uc001rbp.1,uc001rb0.1,uc001rbn.1,uc001rbq.1	235	32	0.363
VSTM1	uc002qcx.2,uc002qcw.2	144	87	0.362

EPX	uc002ivq.2
DISC1 C9	uc001hvc.2
MRĂP	uc002ypl.1
ZBTB7B	uc001fgl.2
FAT1	uc010isn.1,uc003ize.1
COL8A1	uc003dth.1,uc003dtg.1
RASAL3	uc0030qk.1 uc002nbe.2.uc010eaa.1
TMEFF1	uc004bay.1
FLJ00251 PPP1R7	uc001mdw.2
OGDH	uc003tlp.1,uc003tlo.1
PILRA	uc003uuq.1,uc003uup.1,uc003uuo.1
SYNGAP1	ucu10jvt.1,ucu10jvs.1,ucu030kk.1 uc010iuz 1
CLCNKA	uc001axv.1,uc001axu.1
ARSA	uc003bmz.2,uc003bnd.2,uc003bnc.2,uc003bnb.2,uc003bna.2
PRMT2	ucoo2nen.2,ucoo2nen.2,ucoo2nen.1
CACNA1S	uc001gvv.1
UNQ385 SAMD14	uc003gpb.1 uc002ige 1
ERCC6L	uc004eap.1,uc004eaq.1
AP2M1	uc003fmy.1,uc010hxu.1
ARRDC3	ucoo izci. i,ucoo izci. i,uco i oazj. i ucoo3kiz.1
C3orf22	uc003ejb.1
ARID3C SERINC3	UC003ZUV.1 UC010ggs 1 UC002xmf 1 UC002xme 1
SCIN	uc003sso.2
CCDC81	uc001pbx.1,uc001pbw.1
GRIP2 NEUROD2	UCUU3Dyt.1 uc002hrv 1
SAG	uc002vuh.2
KIAA0613	
ACCN3	ucouswip.2,ucouswi0.2,ucouswin.2 uco01toc.1
HTR3A	uc001pon.1,uc001pom.1
LGALS12 CAPN11	uc001nxc.1,uc001nxb.1,uc001nxa.1 uc003owt 1
ZNF687	uc009wmp.1
TMEM82	uc001axc.1
limkain	
LIM2	uc002pwm.1,uc002pwl.1
CPT1A CTNND1	uc009ysj.1
TBC1D24	uc001111.1
SLC9A3R1	uc002ilp.1
POU2F3 KLK14	uc001pxe.1 uc002pys 1
NKX2-3	uc009xwj.1
SYTL1 C19orf51	uc009vsv.1 uc002ail 1 uc002ail 1 uc002aii 1 uc002aii 1
FUT3	uc002mdl.2,uc002mdj.2,uc002mdm.2,uc002mdk.2
KPL1	uc009ytr.1
PAOR6	uc0016a0.1 uc001fnv 1 uc001fnv 1 uc001fnv 1 uc001fnv 1 uc001fnu 1
DQ866763	uc001vue.2
DPH1	uc002ftu.1
OPN4	uc002pwc.2 uc001kdg.1,uc001kdp.1
FBXO32	uc003yqq.1
MICAL2PV2 MICAL2PV1	uc001mkc.2
GHDC	uc002hzd.1
MEOX1	uc002iea.1,uc002idz.1,uc002ieb.1
SCAND2	uco101cl: 1,0co101ce: 1,0co0212w.2,0c00212x.2
BTNL9	uc003mmt.1
HCCA2	uc002utb.1 uc001lto.1
COL8A2	uc001bzw.1,uc001bzv.1
NM 207423	uc001ijx.1
IRF1	uc003kxb.1
SNRPA	uc002opa.1
ATG9B	ucuu2qge.1 uc010lpv.1
NRBP2	uc003yzw.1
1PSG1 FL 100180	uc002ckw.1
TRIM61	uc003igw.1
ATP13A3	uc003ftx.2
AK308580 TBC1D4	UCU1UeKI.1 uc010aer 1
CYP8B1	uc010hif.1
TEKT4	uc002stw.1
DHH	ucou2nds.1 uc001rtf.1
C15orf39	uc002azp.2
AQP4 KANK4	uc002kwa.1
DGUOK	uc002sjy.1,uc002sjx.1

70 111	0.360
31	0.359
154 126	0.359 0.357
81	0.357
30	0.356
126 75	0.355
53	0.354
60 26	0.353
28 64	0.352
39	0.351
47 41	0.351 0.350
280	0.350
52	0.349
40 216	0.348
48	0.346
76 50	0.345
32 62	0.345
55	0.343
43 87	0.343
25	0.342
28	0.342
297 38	0.341
51	0.340
244 33	0.340
34	0.337
30	0.334
71 121	0.333 0.332
42	0.331
94 60	0.330
60 148	0.329
232	0.326
97 48	0.323
46 143	0.323
56	0.321
31	0.321
30 49	0.319
78	0.318
167 51	0.318 0.316
98 50	0.316
34	0.315
51 51	0.313
75	0.313
72	0.311
78 45	0.308
57	0.306
68	0.304
281 79	0.302
47	0.298
33 39	0.298 0.298
48	0.297
128	0.295
33 41	0.295
26	0.290
36	0.290
57 134	0.287
39	0.286
93 42	0.285 0.285
29	0.284
∠8 69	0.284 0.283

 $\begin{array}{c} 162\\ 95\\ 115\\ 200\\ 141\\ 142\\ 101\\ 337\\ 213\\ 364\\ 799\\ 389\\ 394\\ 446\\ 243\\ 361\\ 295\\ 117\\ 80\\ 154\\ 491\\ 1019\\ 256\\ 491\\ 1019\\ 256\\ 312\\ 110\\ \end{array}$

uc003mgo.2 uc002iog.1.uc002iop.1 uc004bah.1 uc002xel.1 uc002jkn.1 uc002xjz.1 uc001urt.2 uc002bql.1,uc002bqk.1,uc002bqj.1 uc001ova.1 uc010eag.1,uc010eae.1,uc002nds.1 uc002fhs.1 uc009yeb.1,uc001lye.1 uc010ecc.1 uc010dxu 1 uc004brg.1 uc004brg.1 uc001lcq.1,uc009xyt.1 uc001aru.1 uc003amm.2 uc001uax.1 uc001ocf.1 uc002xau.1 uc002lga.1 uc010els 1 uc002zcy.1 uc002axd.1,uc010bjf.1,uc002axe.1,uc002axf.1 uc001bwh.1 uc001iiu.1 uc002orc.1 uc001mpw.1 uc001ohs.1 uc001qcw.1 uc002nvc.1 uc010hqs.1,uc003ebz.1,uc003eby.1 uc003ooe.1 uc003svp.1 uc002gho.1 uc003xnf.1.uc003xne.1 uc001nwn.1 uc009zvh.1 uc003ief.1,uc010inr.1 uc004buc.1,uc004bub.1 uc001fkk.2,uc001fkj.2 uc003ssl.1 uc002ama 1 uc002ojq.1 uc002rta.1 uc003itl.2 uc002wrj.1 uc001sbb.1 uc003uvo.2 uc001efy.1,uc009wgz.1 uc004fjo.1 uc002fal.1,uc010cgc.1,uc002fam.1 uc010bcl.1 uc002pes.2 uc010dbg.1,uc010dbf.1,uc002inn.1 uc002ior.1 uc009vwr.1.uc001civ.2 uc010fvx.1 uc001fwl.2 uc003lsr.2,uc003lss.2 uc003lsr.2,uc003lss.2 uc002mde.1 uc002gfz.1 uc001rvn.1 uc001wib.1 uc002uke.1 uc001tua.1 uc003uns.1 uc002etm.1 uc003edl 1 uc002pon.2 uc002xav.1 uc004bzl.2 uc002prt.1 uc001git.1 uc002oim.1 uc004dxe.2,uc004dxd.2 uc003zcc.1,uc003zce.1,uc003zcd.1 uc003xop.1,uc003xoo.1 uc003xjy.1 uc003uss.1 uc003pog.2 uc003pul.1,uc003puk.1 uc003pfq.1 uc003ndi.1 uc003kxv.1 uc010izj.1,uc003kdi.1,uc003kdh.1 uc003mbh.1,uc003mbj.1,uc003mbi.1 uc003ksh.1 uc003ith.1 uc003dct 1 uc003cld.1 uc003avb.1 uc002rqx.1 uc002avi.1 uc002pkg.1

F12 ABI3 NR4A3 HCA58 BTBD17 KIAA1335 PDX1 RCCD1 CAPN5 CIB3 AK123582 ART1 NCAN IDIR UNQ6496 PNLIPRP2 MASP2 C22orf28 MORN3 BATF2 TP53INP2 PRTN3 DKFZp686B U2AF1 ISLR2 HPCA IL15RA UNQ3098 DBX1 SLC29A2 HYLS1 FXYD1 IGSF11 DNAH8 AX747263 EIF4A1 tMDC AB209692 MYO1H TRPC3 DNM1 RUSC1 BC075797 NI RP13 RINL ABCG8 NBI A00301 C20orf78 KRT79 PCOLCE NHLH2 RENBP ZNF19 MAPKBP1 PNMAL2 HOXB3 PHOSPHO MPL TTLL4 SLAMF1 SLAMF1 MYOZ3 NRTN GPS2 AQP2 C14orf93 EVX2 KIAA0985 PON1 CTCF **NR112** PRRG2 ZNF335 hCG 31249 KCNC3 SERPINC1 SPRED3 EFNB1 GPR172A AP3M2 ZNF703 PP838 EPHA7 AMD1 C6orf57 SOX4 SHROOM1 GFM2 NPM1 TNFAIP8 HAND2 DUSP7 CCK RPS19BP1 HNRPLL SOX11 FLJ36070

518 244

304 145

396 172 209

288

75 123

89

126 136

91

179

223

161 130

402 149

304 287

206

250 666

199 114

263 144 97

297

134

667 776

224 347 81

84 469

76 492 342

192

213

284 258

196 130

124 231 272

101 94

352 280

1207

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201

174 77 130

82 179

94

269 364

83 311 106

76 76

130 94

94 148 89

83 112

186

85 93

110 83

110 95 83

87

245

268

36 45

111 33 167

27 45 72

69 38

229

72

25 248

69

35 125

81 28

187

45 55

38 39

62 92 81

99 70

36 405

39

149 27

28

103 51

247 26

60 55

43 61

54

165 195

30 68

173

339 243

458 174

293

323

83 336

306

101 193

88 332 325

139 126

209

293

132

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0.270 0.273 0.272 0.270 0.270

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0.248 0.246 0.243 0.243 0.238 0.237

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0.184 0.181 0.178 0.177 0.176

0.173

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0.163 0.159 0.148 0.146 0.142 0.140

0.137

0.120

0.111 0.106

0 105

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KISS1R	uc002lgk.2	262	352	0.000
TOB1	uc010dbv.1	126	295	0.000
HOXB13	uc002ioa.1	85	128	0.000
GALR2	uc002iam.1	161	401	0.000
HEXIM1	uc002iia.1	122	153	0.000
BCL6B	uc002geg.2.uc010clt.1	100	179	0.000
IRX5	uc002eby 1	131	422	0,000
TMEM159	uc002dih.2.uc002dif.2	137	122	0.000
IMP3	uc002bat 2	219	221	0,000
AFN	uc010bnl 1	75	142	0,000
ZEP36I 1	uc001xki 1 uc001xkh 1	83	102	0,000
ATP6V1D	uc001xif.1	107	143	0.000
KLF5	uc001vie.1	95	404	0.000
C13orf36	uc001uvt.2	83	240	0.000
GPN3	uc001tas.1.uc001tar.1	88	82	0.000
SILV	uc001sig.1.uc001sip.1	75	27	0.000
ELA1	uc001rvi.1	89	32	0.000
PRPH	uc001rtu.1	100	357	0.000
WEE1	uc001mhs.1	95	404	0.000
AL157440	uc001kec.1	89	147	0.000
C10orf114	uc001ian.2	98	312	0.000
KIAA0335	uc001klr.1.uc001kla.1	94	121	0.000
ZNF518A	uc001klo.1.uc001klp.1	94	121	0.000
ACTA1	uc001htm.1	81	298	0.000
UBE2U	uc001dbn.1	76	42	0.000

Table S4.2

List of candidate genes whose expression was down-regulated in the Saqqaq hairs.

This list corresponds to the first 1,000 non-redundant genes showing the lowest R_s values (transcripts are collapsed into a single Gene name, providing the maximal R_s value observed in this quartile). R_s values and respective coverage in PM and RE regions are provided.

Gene	UCSC	Cov(GB)	Cov(PM)	Rs
UTY	uc004fsv.1.uc004fsx.1	400	49	Inf
MPP1		86	103	Inf
FAM3A	uc004flu.1,uc004flt.1,uc004flw.1,uc004fls.1	161	98	Inf
DNASE1L1	uc004fkw.1.uc004fkv.1.uc004fku.1	127	140	Inf
		192	164	Inf
HCFC1	ucou4iju. 1, ucou4iju. 1, ucou4iji. 1 ucou4iju 1, ucou4iju. 1	710	148	Inf
ARD1A	uc004fjn.1,uc04fjm.1	89	141	Inf
L1CAM	uc004fje.1,uc010nuo.1,uc004fjc.1,uc004fjb.1	213	156	Inf
UCHL5IP	uc004fb0.1.uc004fbn.1	341	95	Inf
CD99L2		83 564	123	Inf
IDS	uc004fcv.2,uc004fct.2	94	59	Inf
ZNF75D	uc004eyp.1	124	51	Inf
MOSPD1	uc004evb.1	103	66	Inf
API N		78	40 152	Inf
SMARCA1	uc004eup.2,uc004eun.2	101	74	Inf
CUL4B	uc004esw.1	149	30	Inf
FAM70A	uc010ngo.1,uc004esp.2,uc004eso.2	144	99	Inf
KI HI 13		182	95	Inf
AMOT	uc004eps.1	158	56	Inf
TRPC5	uc004epm.1,uc004epl.1	400	36	Inf
AMMECR1	uc004eop.1.uc004eop.1	110	103	Inf
NUP62CI		128	102	Inf
MORC4	uc004emv.2,uc004emv.2,uc004emu.2	77	124	Inf
MCART6	uc004elu.1	209	39	Inf
BTK		152	38	Inf
	ucou4egq, 1,ucou4egq, 1,ucou4egt, 1,ucou4egt, 1 ucou4egq, 1,ucou4egt, 1,ucou4egt, 1	144	39	Inf
CXorf34	uco04eqs.1	144	39	Inf
HDX	uc004eel.1,uc004eek.1	103	35	Inf
RPS6KA6		120	92	Inf
ZDHHC15	ucou4edu.z,ucou4edu.z,ucou4edu.z,ucou4edu.z,ucou4edu.z,ucou4edu.z,ucou4edu.z,ucou4edu.z,ucou4edu.z,ucou4edu.z	194	50 41	Ini
ZMYM3	uc004dzi,1,uc004dzi,1,uc004dzh,1	178	76	Inf
LAS1L	uc004dwd.1,uc004dwc.1,uc004dwa.1	80	44	Inf
KIAA0424		117	48	Inf
PHF8	uco04dvm1.ruc004dv12 uco04dsv2 uco04dsv2 uco04dsv2 uco04dsv2	307	48	Inf
HUWE1	uc004dsp.1	522	71	Inf
JARID1C	uc004dsa.1,uc004drz.1	196	75	Inf
SYP	uc004dna.1,uc004dmz.1	79	89	Inf
WDR45 DKF7p761 118		190	6U 92	INT
TFE3	uc004dmb.2,uc024dmd.1	99	92	Inf
SLC35A2	uc004dlo.1	76	54	Inf
ELK1	uc010nhv.1,uc010nhv.1,uc004dik.2	107	64	Inf
5YN1 7NF41		3/1	135	INT
SLC9A7	uc004dgv.1.uc004dgu.1	146	140	Inf
BCOR	uc004deq.2,uc004deo.2,uc004dep.2,uc004dem.2,uc004den.2	571	236	Inf
SRPX	uc004ddz.1.uc004ddy.1	118	80	Inf
		190	27	Ini
DKFZp434M1	uco10nfd.1	247	144	Inf
SCML2	uc004cyl.2	250	144	Inf
	uc004cyg.1,uc004cyh.2,uc010nta.1	191	135	Inf
MID1	ucou4cun. i uco04cta 2	400 320	44	Inf
HDHD1A	uc010ndl.1,uc004crv.1	403	166	Inf
NLGN4X	uc010ndh.1,uc010ndj.1,uc010ndi.1,uc004crr.1,uc004crq.1	600	110	Inf
MXRA5 BPCC3		158	158 114	Int Inf
TAZ	uc004flc.2.uc010nuv.1.uc004flb.1.uc004fla.1.uc004fkv.1.uc004fkx.1	93	133	Inf
SLC6A8	uc004fic.2,uc004fib.2	114	125	Inf
BGN	uc004fhr.1	273	75	Inf
NSDHL MTMR1	UCUU4Tgs.1,UCUU4Tgt.1 uc004feb 1 uc004fei 1	312	70 97	INT Inf
FHL1	uc010nrz.1,uc004ezl.1	198	172	Inf
FAM122C	uc010nru.1	86	64	Inf
HPRT1		84	117	Inf
FL 130058	ucou4exi.2,ucou4exi.1,ucou4exi.1 uco04ewa 1 uco04evz 1	238	54 75	Inf
XPNPEP2	uc004eut.1	169	31	Inf
XIAP	uc010nav.1.uc010nau.1.uc004etx.1	227	73	Inf
GRIA3	uc004etr.2,uc004etq.2	301	67	Inf
LONRF3	ucoo4eax.1.uc004eaw 1	100	103	Inf
IL13RA1	uc004eqt.1,uc004eqs.1	214	126	Inf
PLS3	uc010nqf.1,uc010nqg.1,uc004eqd.1	198	84	Inf
ZCCHC16		392	30	Inf
ATG4A	uco10npu. 1,uco10npu. 1,uco04eo2.2 uc004ens.1.uc004ent.1.uc004enr 1	76	52	Inf
IL1RAPL2	uc004elz.1	896	106	Inf
DACH2	uc010nmg.1,uc004eex.1,uc004eew.1	459	63	Inf
CHIC1		92 142	66 53	Inf
NLGN3	ucou+eao.,,ucou+eaa., uco10nlb.1,uco04dzc.1,uco04dzb.1	169	42	Inf
MED12	uc004dyz.1,uc004dyy.1	166	41	Inf
DLG3	uc004dyi.1,uc004dyi.1	192	35	Inf
KIF4A	ucu04ayr.1,ucu10nkw.1,ucu04ayg.1	180	43	int

EDA	uc004dxm.1.uc004dxn.1.uc004dxa.1.uc004dxp.1.uc004dxs.1.uc004dxr.1	223	98	Inf
STARD8	uc004dxc.2	235	85	Inf
MSN GNI 3I	uc004dwt.1	162 178	58 47	Inf
CCNB3	uc004dox.2	140	33	Inf
CLCN5	uc004dog,1,uc004dor.1	203	92	Inf
ZUCCCCC22 ZNE81	uc004ana.1 uc010nby.1	158 75	31 26	Inf
DKFZp434P0	uc004dhe.1	328	67	Inf
RBM10	uc004dhi.1,uc010nhq.1,uc004dhi.1,uc004dhg.1,uc004dhf.1	500	60	Inf
TSPAN7		121 154	74 96	Inf
LANCL3	uc004ddp.1	89	55	Inf
PRRG1	uc004ddo.1.uc004ddn.1	128	94	Inf
PRGP1 PDK3	uc004dbm.2	113 185	94 93	Inf
PTCHD1	uc010nfu.1,uc004dal.2	85	163	Inf
YY2	uc010nfq.1	129	60	Inf
MBTPS2		177	60 100	Inf
REPS2	uc004cxv.1,uc004cxv.1	404	75	Inf
SYAP1	uc004cxp.1_uc004cxp.1	165	58	Inf
OFD1	ucuu4cxc.1 uc004cxv.2 uc004cvu.2 uc010pen 1 uc004cvg.2 uc004cvg.2	97	54 63	Inf
PRPS2	uc004cvb.1,uc004cva.1	99	144	Inf
FRMPD4	uc004cuz.1	769	101	Inf
KIAA1280 WWC3		991 991	171 171	Inf
GYG2	uc004cgw.1	257	50	Inf
SURF6	uc004cdb.2	288	149	Inf
CRAT		174	153 49	Inf
ANGPTL2	uc010mxg.1,uc004bgr.1	447	118	Inf
PPP6C	uc010mww.1,uc004bpg.2,uc010mwv.1	232	220	Inf
RBM18	ucuu4bma.1 uc004bly.1	109	106	Inf
ASTN2	uc004bjv.1,uc004bjp.1,uc004bjg.1,uc004bjt.1,uc004bjs.1,uc004bjr.1	150	111	Inf
DFNB31	uc004bix.1	424	59	Inf
FKBP15 KIAA0674	ucuu4pgt.2.ucuu4pgs.2 uc010mut 1	144	99	Inf
C9orf80	uc004bgg.1	80	91	Inf
PTPN3	uc010mtv.1	344	34	Inf
KIAA0573		408	225	Inf
ERP44	uc004bam.1	350	131	Inf
ANKS6		484	41	Inf
CDC14B		446	120	Ini
hfrc	uc004awe.1,uc004awd.1	263	163	Inf
SLC35D2	uc010msf.1,uc010mse.1,uc004awc.1	488	163	Inf
IPPK	uc004atl.2 uc004asl 1	144	258 175	Inf
CCRK	uc004apu.1,uc004apt.1,uc004aps.1,uc004apr.1	113	90	Inf
C9orf64	uc004anc.1,uc004anb.1	145	141	Inf
UBQLN1		175	217	Inf
PRUNE2	uc004akn.1,uc010mpk.1	343	146	Inf
TRPM6 FAM108B1	uc004ajn.1,uc010mpe.1,uc010mpd.1,uc010mpc.1,uc004ajk.1,uc004ajl.1 uc004aji 1	235	106 270	Inf
MCART1		81	194	Inf
FBXO10	uc004aab.1	584	163	Inf
PAX5 RNE38	uc010mls.1,uc010mlr.1,uc010mlp.1,uc010mlp.1,uc003zzo.1 uc003zzi 1 uc003zzi 1 uc003zzi 1 uc003zzm 1 uc003zzi 1	2969	136	Inf
KIAA1539	uc003zw1,10000zw1,10000zw1.1	102	84	Inf
FLJ00135	uc003zwg.1	113	104	Inf
FLJ00350 PIGO	ucu03zwc 1 uc003zwf 1	113 113	104 104	Inf
VCP	uc010mki.1,uc0010mkh.1,uc003zvy.2	95	128	Inf
UBAP2	uc003ztr.1,uc003ztr.1,uc003ztr.1	862	89	Inf
NDUFB6	ucouszti. 1,ucouszti. 1,ucousz	107	53	Ini
TOPORS	uc003zrc.1,uc003zrb.1	82	217	Inf
DDX58		154	58	Inf
CDKN2A		165	123	Inf
PSIP1	uc003zlz.2	136	139	Inf
TTC39B	uc010mirt.1.uc010mire.1.uc003zIr.1	466	171	Inf
ZDHHC21		218	142	Inf
NFIB	uc003zlf.1,uc003zle.1	507	248	Inf
RIAD2026	ucuu3zkn.1,ucuu3zko.1,ucuu3zkm.1,ucuu3zkl.1,ucuu3zkl.1,ucuu3zkl.1	3789	75 205	Inf
ERMP1	uc003zjn.1,uc010mhs.1,uc003zjm.1	80	194	Inf
C9orf68	uc003zim.2,uc010mhj.1	223	47	Inf
кгаз С90rf7	uco romne.1,uco10mna.1,uco03zns.1,uc003znr.1 uc004cec 1 uc010nan 1 uc004ced 1	427 332	140 194	Inf Inf
GTF3C5	uc010mzz.1,uc004cci.2,uc004ccj.2	419	149	Inf
GFI1B	uc004ccq.1	348	51	Inf
EXOSC2	ucuu4can.∠ uc004bzu 2	77	30 67	Inf Inf
USP20	uc004byt.1,uc004byr.2,uc004bys.2	1285	192	Inf
DKFZp781M1	uc010myr.1	568	198	Inf
PPP2R4 PHYHD1	ucuu4dxo.1,ucuu4dxn.1,ucuu4dxl.1,ucuU4dxl.1 uc004hwn 2	568 221	198 80	Inf Inf
SET	uc004bvt.2	622	28	Inf
SPTAN1	uc004bvn.2,uc004bvn.2,uc004bvl.2	1282	311	Inf
URM1	ucoo+bvc. i,ucoo+bvc. i,ucoo+bvb. i,ucoo+bvd.z uc004buv.1	319	78	Inf
			-	

SLC25A25	uc004btb.1	837	187	In
STXBP1 MRRF	uc004brl.1,uc004brk.1 uc010mwa 1 uc004bmc 1 uc004bmb 1	822 201	298 108	ln In
TRIM32	uc004bjw.2,uc004bjx.2	108	122	İn
ZNF618 RGS3	uc004bib.1,uc004bid.1,uc004bic.1 uc010muz 1 uc010mva 1 uc004bbw 1	1395 399	206 69	ln In
SLC31A1	uc004bgv.2,uc004bgu.1	192	101	In
HSDL2 DKFZp666L09	uc004bqc.1,uc004bqa.1 uc004bab.1	399 399	190 190	In In
AKAP2	uc004bem.1	650	162	In
SMC2	uc004bbu.1,uc004bbw.1,uc004bbv.1,uc004bbx.1	80	74 94	in In
RNF20	uc004bbn.1	89	60	In
INVS	uc004bbb.1 uc010mta.1,uc010mtb.1,uc004bar.1,uc004baq.1,uc004bap.1,uc004bao.1	505	114	In
NR4A3	uc004bae.1,uc004bai.1,uc004bag.1,uc004baf.1	439	288	In
ANP32B	uc004aya.1	191	631	In
TDRD7 HABP4	uc004axj.1	197 351	294 290	ln In
SYK	uc004arc.1	664	36	In
CR613032	uc004aqe.1 uc004aoh.1	267 82	174 35	in In
GCNT1	uc004akf.2,uc010mph.1	247	141	In
C9orf61	uc014aix.2,uc004aiw.2,uc004aiv.2 uc010moo.1,uc010mon.1	102	34	In
RG9MTD3	uc004aak.1,uc004aai.1	113	69 195	In
UNC13B	uc010mkl.1	612	128	In
UNC13B KIAA1045	uc003zwr.1,uc003zwq.1 uc003zvg 1	799 79	128 238	ln In
IL11RA	uc003zvi.1	108	54	In
NFX1 MTAP	ucu1umjr.1,ucuu3zsr.1,ucuu3zso.1,ucuu3zsp.1,ucuu3zsg.1 uc003zpi.1,uc003zph.1	303 1683	90 134	In In
JMJD2C	uc010mhv.1,uc003zkg.1,uc003zkh.1	390	277	In
KANK1	uc003zgs.1	304	96	In
TRAPPC9	uc003yvi.1	9100 265	86 143	In
TATDN1	uc010mdm.1,uc003yrf.1,uc003yrd.1	219	103	In
DERL1 MRPL13	uc003ypn.1,uc003ypm.1,uc003ypl.1 uc003yp.1	126 89	155 51	ln In
SAMD12	uc003yom.2,uc010mda.1	606	112	In
CSMD3	uc003ynx.2,uc010mcx.1,uc003ynv.1,uc003ynu.1	290 396	48 55	In
NUDCD1	uc010mci.1,uc003ynb.2	224	111	In
PABPC1	uc003yjt.1,uc003yjs.1	100	275	In
SNX31 ANKRD46	uc003yjr.1 uc003yim 2 uc003yin 1 uc003yin 1 uc003yin 1	360 166	146 140	ln In
STK3	uc003yio.1	1837	300	In
KIAA1429	uc010mbk.1 uc003yap.1,uc003yao.1	82 199	81 69	In In
FAM82B	uc003ydu.1	117	103	In
MRPS28	uc003ybo.1,uc003ybp.1	232	106	In
TPD52 MSC	uc003ybt.1,uc003ybs.1	510 124	193 197	ln In
EYA1	uc003xyv.1,uc003xyu.1	438	39	In
COPS5 PDE7A	uc003xxd.1,uc003xxt.1,uc003xxe.1 uc003xva.1,uc003xvr.1	86 322	37 141	In In
ARMC1	uc003xvl.1	190	85	In
TCEA1	uco10iyi.1,uco10iyi.1,uco03xsi.2 uc003xrv.1,uc003xru.1	234 248	153	In
	uc003xot.2,uc003xos.2	581 1324	37 56	In
FLJ43582	uc003xlx.1	142	33	İn
FGFR1 DUSP26	uc003xlt.2 uc003xig.1.uc003xip.1	566 110	31 93	ln In
C8orf41	uc003xjl.2,uc003xjm.2	75	40	In
INTS9	uc003xld.1 uc003xhb.1,uc003xha.1	363	51	in In
PNMA2	uc003xez.2	77 241	186 158	In
CSGALNACT	uc003wzf.1	1289	25	İn
ASAH1 MTMR7	uc003wym.2,uc003wyl.2,uc003wyo.2,uc003wyn.2 uc003wxm 1	137 575	155 120	In In
SGCZ	uc003wwg.1	2273	171	In
LOC649305	uc003wwc.2	223	67	in In
PINX1	uc003wti.2,uc003wth.2	630 3104	119	In
ZNF707	uc003yzh.2,uc003yzf.2,uc010mfi.1,uc010mfh.1,uc003yze.2	468	109	In
PHF20L1 EFR3A	uc003yts.1,uc003ytt.1 uc003vte.1	76 166	159 276	ln In
MTBP	uc003ypc.1	153	39	In
TM7SF4	ucuusymw.1 uc003ylx.1	91	134 29	in In
C8orf47	uc003yih.1	115	153	In
WWP1	uc003ydt.1	394	221	in In
CHMP4C PKIA	uc003ycl.1	83 120	75 140	In
CRISPLD1	uc003yan.1	83	131	İn
SULF1	uc003xz1.2 uc003xyg.2,uc003xyf.2	237 771	43 48	In In
C8orf46		253	33	In
	40000AUZ. 1,40000AVA. 1,400 10195. 1	144	101	

YTHDF3 RI BP1I 1	uc003xuy.1 uc003xug 1 uc010lyo 1 uc003xub 1	144 185	200 63	Inf Inf
RP1	uc003xsc.1	249	30	Inf
KIAA0146	uc010ixz.1,uc003xqs.1 uc010lxs.1,uc003xqe.1,uc003xqd.1	2158	29 186	Inf
FNTA POLB	uc003xpu.1,uc003xpt.1,uc003xps.1	126	228	Inf
LETM2	uc003xlo.1,uc003xln.1,uc003xln.1	122	124	Inf
DCTN6 HMBOX1	uc003xhy.1 uc003xhc 2 uc010lyd 1 uc003xhd 2 uc003xhe 1	122 421	76 114	Inf Inf
ELP3	uc003xg0.2	347	54	Inf
DKFZp762K0	uc003xtq.1,uc003xtp.1 uc003xea.1	1017 175	95 116	Inf
CDCA2	uc003xep.1	175	116	Inf
XPO7	uc003xaa.2,uc010lti.1	328	255	Inf
ATP6V1B2 INTS10	uc003wzp.1	87 182	124 119	Inf Inf
PCM1	uc003wyg.2,uc003wyh.2,uc010lta.1,uc003wyi.2	82	212	Inf
FDFT1	uc003wwr.1,uc003wwy.1,uc003wws.1,uc003www.1,uc003wwv.1,uc003wwu.1,uc003wwt.1 uc003wuh.1	429 481	129 29	Inf
MTMR9	uc010lrx.1,uc003wtm.1	118	128	Inf
CLN8	uc003wp0.2	422	194	Inf
FBXO25 FAM62B	uc003woz.1,uc003woy.1,uc003wox.1 uc003woc.1,uc003wob.1,uc003wod.1	264 1117	97 31	Inf
SMARCD3	uc003wju.1	1228	124	Inf
FAM115A SLC37A3	uc003wdp.1,uc003wdo.1 uc010lnh.1,uc003vvp.1	253 543	109 152	Inf
CREB3L2	uc003vty.2,uc003vtx.1,uc003vtw.1	487	116	Inf
DKFZp586B0	uco10lmg.1	287 251	32 26	Inf
WDR91	uc003vsp.1	289 280	214 184	Inf
UBE2H	uc003vpg.1,uc003vpf.1	683	149	Inf
TNPO3 TRN-SR	uc010llz.1,uc010lly.1,uc003vol.1 uc003vom.1	372 372	90 90	Inf Inf
RBM28	uc003vmp.2	181	78	Inf
IQUB	ucousvia.1,ucousvia.1 ucousvko.1	100	69	Inf
CTTNBP2	uc003vif.1	478	333 187	Inf
COG5	uc003vee.1,uc003ved.1,uc003vec.1	1066	106	Inf
SLC26A5 ZRF1	uc003vbv.1,uc003vbu.1,uc003vbz.1,uc003vbt.1 uc003vbp.1	349 190	193 106	Inf Inf
DNAJC2	uc010lix.1,uc003vbo.1	190	106	Inf
NAPEPLD	uc003vbd.2,uc003vbc.2	180	173	Inf
ALKBH4	uc003uzm.1,uc003uzl.1	231 90	200	Inf
LRCH4	uc003uvj.1	558	147	Inf
STAG3OS DKFZp761F1	uc003uua.2,uc003uuc.1 uc010lgg.1	723 121	96 56	Inf Inf
GAL3ST4	uc003utu.1	121	56	Inf
TECPR1	uc003upg.1	958	158	Inf
CALCR	uc003umw.2,uc003umv.1,uc003umu.1	179 485	96 253	Inf Inf
PEX1	uc010ley.1,uc003uly.1	106	184	Inf
SEMA3E	uc003ulq.1 uc003uhv.1	135 502	40 26	Inf
STYXL1	uc003uek.2,uc010ldh.1,uc003uej.2,uc003uem.1,uc003uel.1	516	132	Inf
HIP1	uc003uds.1	2401	163	Inf
POM121C MI XIPI	uc003tvn 1 uc003tvn 1 uc003tvn 1 uc003tvl 1 uc003tvk 1	192 586	26 251	Inf Inf
BAZ1B	uc003tyc.1	525	266	Inf
LOC401365 TNS3	ucu10lac.1 uc010kyo.1,uc003tnv.1	858 2054	177 40	Inf
DDX56		86 102	119	Inf
POLR2J2	uc003tjd.1	153	40	Inf
C7ort44 DPY19L2P1	uc003tip.1,uc003tiq.1,uc003tio.1,uc003tin.1 uc003tea.1	311 124	91 57	Inf
CRH2R	uc010kvy.1	288	213	Inf
KIAA0644	uc003szt.1	200	378	Inf
SKAP2 IGF2BP3	uc003syc.1 uc003swa.1	324 879	69 265	Inf Inf
DRCTNNB1A	uc003svn.2	100	158	Inf
MGC87042	uc003svh.2	197	30	Inf
DKFZp761E0	uc003sty.2	150 164	152 152	Inf
SNX13		344	152	Inf
ETV1 THSD7A	uc003ssw.2 uc003ssf.2	264 686	33 99	Inf Inf
	uc003srs.1,uc003srp.2,uc003srr.1,uc003sro.2,uc003srn.2,uc003srg.1,uc003srm.1	281	202	Inf
KDELR2	uc003sqf.2,uc003sqe.2	182	194	Inf
RNF216 C7orf27	uc003soz.1,uc003soy.1,uc003sox.1 uc003smi 1 uc003smi 1	1255 298	162 157	Inf Inf
KIAA0010	uc003wni.2	627	30	Inf
NUB1 DKFZp761E2	ucuu3wjx.1,ucuu3wjw.1 uc003wjk.1	497 167	161 74	Inf Inf
ZNF282	uc003wfm.1	910 1050	230 184	Inf
EPHB6	uc003wbt.1	322	29	Inf

CHRM2	uc003vtm.1,uc003vto.1,uc003vtl.1,uc003vtg.1,uc003vtn.1,uc003vtk.1,uc003vtj.1,uc003vtf.1,uc003vti.1	316	262	Inf
CALD1	uc003vsd.1,uc003vsc.1	375	36	Inf
EXOC4	uc003vri,2,uc003vri,1,uc003vrk,1	605	25	Inf
FAM40B	uc003vow.1,uc003vox.1	181	198	Inf
CCDC136	Uc003vny.1	114	27 145	Inf
METTL2B		132	46	Inf
SND1	uc003vmi.1	2821	178	Inf
CEIR HBP1	uc003vjd.1 uc003vjd.1	331	77 187	Inf
RINT1	uc010lij.1,uc003vda.1	260	96	Inf
DKFZp586I12	uc010liv.1,uc010liv.1,uc003vbm.1	77	93	Inf
PMPCB PRKRIP1		76 434	93 72	Inf
SERPINE1	uc003uxt.2	175	52	Inf
TRIM56	uc003uxr.2,uc003uxr.1	317	86	Inf
FBXO24	ucc03uvl.1uc003uvm.1	154	178	Inf
ZNF789	uc010lfw_1,uc003uqq_1	122	94	Inf
MYH16 I MTK2	uc003upv.1 uc003upd 1	361	27 183	Inf
KIAA1861	uc003µmp.1	145	38	Inf
CCDC132	uc003umo.1	145	38	Inf
OTPBP10		1274	294 64	Inf
ZNF804B	uc003uju.1	791	160	Inf
ADAM22	uc003uji.1,uc003uji.1,uc003uji.1,uc003uji.1,uc003uji.1,uc003uji.1,uc003uji.1,uc003uji.1,uc003uji.1,uc003uji.1	387	194	Inf
MDH2	uc003ue0.1,uc003ue0.1	420	125	Inf
RHBDD2	uc003udw.1.uc003udv.1	155	90	Inf
ELN POM121	uc003tzq.1,uc003tzp.1,uc003tzo.1,uc003tzz.1,uc003tzv.1,uc003tzx.1,uc003tzv.1,uc003tzv.1,uc003tzv.1,uc003tzu.1,uc003tz	757	115 25	Inf
AUTS2	uc003tvr.2,uc003tvr.2	1754	258	Inf
STAG3L4	uc003tvt.2	89	151	Inf
I YW1 C7orf42	uc003tvn.1 uc003tvl 1 uc003tvk 1	715 367	29 32	Inf Inf
TPST1	uc010laa.1,uc010lazz.1,uc003tuw.1	548	197	Inf
VKORC1L1		352	160	Inf
ZNF273 ZNF138	uc003tto.1,uc003tto.1	85 96	53 52	Inf
ZNF107	uc003tte.1,uc003ttd.1	223	41	Inf
DKFZP566I10	uc003trt 1 uc003trc 1 uc003trc 1 uc003trc 1	190	101	Inf
CMAP		171	117	Inf
DBNL	uc003tip.2,uc003tiq.2,uc003tio.2	238	117	Inf
BMPER	ucoustna.1 uc003tdw 1	261 828	232	Inf
BBS9	uc003tdq.1,uc003tdp.1,uc003tdo.1,uc003tdn.1	784	74	Inf
ADCYAP1R1	uc003tcf.1	125	29	Inf
FAPP2	ucoostax.1	226	197	Inf
PLEKHA8	uc003tam.1,uc003tap.1,uc003tan.1	115	197	Inf
TAX1BP1 CCDC126	uc003szL1,uc003szk.1	161 145	127 57	Inf
KLHL7	uco03svs.2,uc003svr.2	234	122	Inf
TSPAN13		105	168	Inf
JTV1		138	135	Inf
EIF3B	uc003slz.1,uc003sly.1,uc003sly.1	109	225	Inf
		386	265 78	Inf
UNC84A	ucotoks,ucotosk	1388	46	Inf
THBS2	uc003qwt.1	1388	57	Inf
PARK2 RSPH3	ucu10kke.1.ucu03dtz.2.ucu03dtv.2.ucu03dtx.2	7523	196 165	Inf
EZR	uc003qrt.2	607	137	Inf
SERAC1	uc003grc.1	303	180	Inf
SYNE1	uc010kiy.1	102	75	Inf
ZC3H12D	uc010kid.1.uc003gmm.2	915	56	Inf
HIVEP2 AX747618	uc003qia1	856 226	211	Inf
MAP	uc010kgv.1	585	26	Inf
	uc010kgr.1,uc010kgs.1,uc010kgt.1,uc010kgt.1,uc003qha.1,uc003ggz.1	664 357	26	Inf
SGK1	uc003ge0.2	1386	43	Inf
SLC2A12	uc003gem.1	271	43	Inf
SIX/ ARHGAP18	ucUU3qq22	160 440	103	Inf
C6orf174	uc003qbg.2,uc003qbf.1	246	61	Inf
ECHDC1	uc010kez.1.uc003qay.2.uc010key.1.uc003qaz.2	87	115	Inf
LAMA4	uc003ptw.2.uc003pt.2.uc003pt.2	102	113	Inf
TRAF3IP2	uc010kdx.1,uc010kdw.1,uc003pvg.1,uc003pvf.1,uc003pve.1	220	34	Inf
	uc003pud.1,uc003puc.1	280	25 80	Inf Inf
ASCC3	uc003pgl.2,uc003pgk.1	298	153	Inf
USP45	uc003ppz.1,uc003pqa.1,uc010kcq.1,uc003ppx.1	139	168	Inf
AKIRIN2	ucuU3ppq.2,ucuU3ppp.2,ucuU3pp0.2 uc003pmk.1	135	55 200	Inf
KIAA1009	uc003pkk.2,uc003pkj.2,uc010kbp.1	85	61	Inf
		370	84	Inf
KIAA1417	ucouspin. r.ucouspin. r uc010kbi.1	200 246	116	Inf
LCA5	uc003pix.1,uc003piy.1	92	80	Inf
SLC17A5 C6orf150	ucuu3pnn.2 uc003pax.1	393 128	163 191	Inf Inf
	· · · · · · · · · · · · · · · · · · ·	-		

B3GA12	uc003ptv.2,uc003ptv.1	318	408	Inf
COL9A1		217	34	INT
		200	64 70	INT
C6orf138		253	102	Inf
GPR116		166	42	Inf
RCAN2	uc003oyb.1	195	28	Inf
NFKBIE	uc003oxe.1	77	152	Inf
TRERF1	uc003osc.1,uc003osb.1,uc003ose.1,uc003osd.1	1689	34	Inf
USP49	uc003ori.1	620	161	Inf
FRS3		171	143	Inf
MOCS1	ucuu3ope.2,ucuu3opc.2,ucuu3opb.2,ucuu3opa.2,ucuu3opd.2	300	74	Inf
		444	207	IIII Inf
Cforf120		237	140	IIII
TMFM217	uc003onm 2 uc010iws 1 uc010iwr 1 uc003onl 1	169	78	Inf
PPIL1	uc003omu.1	122	77	Inf
STK38	uc003omi.1,uc003omh.1	258	97	Inf
AK309286	uc010jgw.1	226	63	Inf
DCDC2	uc003ndx.1	424	87	Inf
DINBP1	uc010jph.1,uc003nbp.1,uc003nbm.1,uc003nbl.1	579	226	Inf
CEOD1		100	249	INT
		126	40	IIII
MAK		435	102	Inf
F13A1		530	57	Inf
TBP	uc003axu.1.uc003axt.1	77	86	Inf
PACRG	uc003quc.1,uc003qub.1,uc003qua.1	4487	137	Inf
GTF2H5	uc003qrd.1	109	183	Inf
OPRM1	uc010kjf.1,uc003qpq.1,uc003qpt.1,uc003qpo.1,uc003qpr.1,uc003qpn.1	235	70	Inf
C6orf97		594	219	Inf
C60ff211		89	171	INT
DREG		475	250	Inf
GPR126	$\mu c 010 khf 1 \mu c 010 khc 1 \mu c 010 khc 1$	230	250	Inf
VTA1	uc003giw.1	148	69	Inf
TBPL1	uc003gel.1	112	137	Inf
ENPP1	uc003qcx.2	267	203	Inf
ENPP3	uc003qcu.2	295	73	Inf
LAMA2	uc010kfe_1,uc003qbo_1,uc003qbn.1	1397	53	Inf
RSP03	uc003qas.1,uc003qar.1	106	336	Inf
	ucc10kev.1,ucc003gam.1	97	95	Inf
		273	207	IIII
NKAIN2		1378	248	Inf
TCBA1	uc010kec_1.uc003pzp.1	1624	248	Inf
PKIB	uc003pyz.1	758	79	Inf
CDC40	uc003pua.1	92	50	Inf
FIG4	uc003ptt.2	370	74	Inf
QRSL1	uc003prl.1,uc003prm.1	119	80	Inf
AIM1		260	317	Inf
		202	163	Ini
KLILSZ KIAA1000		570	163	Inf
ORC3I	uc003pmi 1 uc003pmg 1	274	95	Inf
DOPEY1	uco03pis.1	195	144	Inf
CD109	uc010kaz.1,uc010kba.1,uc003phg.1,uc003php.1	331	161	Inf
KCNQ5	uc003pgi.2,uc010kat.1,uc003pgk.1	936	285	Inf
SMAP1	uc010kap.1	557	79	Inf
FAM135A	uc003pth.2,uc003pth.2	394	189	Inf
DAIS		029	70	Ini
7NF451	uc003pdt 1 uc003pdt 1 uc003pdn 1 uc003pdm 1	146	88	Inf
EFHC1	uccospan, had been ha	132	95	Inf
PAQR8	uc003pao.2	239	246	Inf
CENPQ	uc003ozh.1	90	54	Inf
CDC5L	uc003oxl.1	142	76	Inf
DKFZp547K1	uc010jyy.1	111	184	Inf
		75	100	IIII
TTBK1		1329	130	Inf
PARC	uc003oui.1	105	90	Inf
KIAA0708	uc010jyk.1	595	90	Inf
CUL9	uc003oul.1,uc003ouk.1	595	90	Inf
PTK7	uc010jyj.1	427	55	Inf
		84	93	INT
		102	120	Ini
CNPY3	ucoo3ata 2	102	120	Inf
UBR2	uc003osf,2,uc003osg,1	159	67	Inf
BYSL	uc003orl.1	123	92	Inf
RNF8	uc003onr.2.uc003ong.2	117	143	Inf
FBC1D22B	uc003onn.1	356	74	Inf
FLJ00276	uc003ong.2	432	27	Inf
FGDZ C6orf80		432	21	Int
AK125083		203 158	41	Inf
MAPK14	uc003olo,1,uc003olr,1,uc003ola,1,uc003olo,1	216	160	Inf
ZKSCAN3	uc003nlf.2,uc010jrc.1,uc003nle.2	105	40	Inf
SCGN	uc010ipz.1,uc003nfb.1	123	49	Inf
LRRC16A	uc010jpy.1,uc010jpx.1	971	177	Inf
	UCUTUIDV.1,UCUU3NEK.1	129	131	Inf
F2F3	1.pqUU 000 Licharda 2 Licharda 2	00 430	364	III Inf
RNF144B	uc003ncs.1	192	42	Inf
GMPR	uc003nbs.1	593	152	Inf
JARID2	uc003nbj.1	2373	104	Inf
HIVEP1	uc003nac.1	662	199	Inf

RRFB1	uc003mxe 1	1483	78	Inf
PRPF4B	uc003mvv.1	143	103	Inf
BPHL	uc003mva.1,uc003muy.1	421	197	Inf
AX748230	ucousmuw.z,ucousmuv.z,ucousmux.i ucousmut.z,ucousmux.i	252	43	Ini
MGAT1	uc003mmi.2,uc010jlg.1,uc010jlf.1,uc003mmh.2,uc003mmg.2,uc010jlh.1	855	29	Inf
HK3	uc003mfa.1	476	32	Inf
GABRB2	uc010iiu.1.uc003lvr.1.uc003lvr.1.uc003lvs.1	442	182	Inf
RNF145	uc010iiq.1,uc003lxp.2	113	267	Inf
EBF1	uc003lxl.2,uc010jip.1	1588	138	Inf
DKFZp434C1	uccolinu.1	289	100	Inf
CCDC69	uc003ltg.1	289	100	Inf
DCTN4 CD74	uc003lsu.1,uc010jhi.1,uc003lsv.1 uc003lst 1,uc003lse 1,uc003lsd 1,uc003lsc 1	186 174	34	Int
ARSI		190	162	Inf
CSF1R	uc003lrm.1	699	32	Inf
PCDH1	ucou3ipo.1,ucou3ipK.1,ucou3ipi.1,ucou3ipi.1,ucou3ipi.1,ucou3ipi.1,ucou3ipi.1, uc003ilio 1,ucou3ipi.1	263	143 281	Inf
HARS	uc010jfu.1,uc003jgw.1	116	93	Inf
PFDN1	uc003lff.1	250	31	Inf
DNAJC18		153	223	Inf
SIL1	uc003ldp.1.uc003ldo.1	1357	98	Inf
CDC25C	uc003lcs.1	364	124	Inf
BRD8	uc03l0i.1	89	25	Inf
FAM13B	uc003lca.2,uc003lcb.2,uc003lbz.2	332	213	Inf
KLHL3	uc010jem.1,uc010jek.1	318	25	Int
SKP1	uc010jdw.1	90	199	Inf
VDAC1	uc003kyr.1,uc003kyg.1,uc003kyp.1	221	446	Inf
ZCCHC10	uc003kyh.1,uc003kyg.1 uc003kyf 2 uc003kye 1 uc003kyd 1	180 305	85 202	Int
KIF3A	uc003kxp.1,uc003kxo.1	141	124	Inf
ZNF608	uc003kts.1,uc003ktg.1	579	37	Inf
PGGT1B P.IA2	ucu10jcn.1,ucu03kqx.2,ucu03kqw.2 uc003kqs 2	85 192	84 135	Inf
GIN1	uc003koc.1,uc003kob.1,uc003koa.1	84	42	Inf
SLCO4C1	uc003knm.1	95	121	Inf
ELLZ FAM172A		191 463	258 75	Inf
TMEM161B	uc003kjc.1	142	46	Inf
DHFR	uc003kgx.1,uc003kgy.1	126	159	Inf
C5orf37	uc003kih.1 uc003keh.2	194	83	Ini
CENPK	uc003jtu.1,uc003jtt.1,uc003jts.1	152	80	Inf
ADAMTS6		715	27	Inf
ERCC8	uc003jtk. 1 uc003ism.2.uc003isl.2	89 161	62 49	Ini
PDE4D	uc003jsc.2,uc003jrz.2,uc003jry.2	956	39	Inf
SLC38A9	uc010ivy.1,uc003jqf.1	218	68 20	Inf
C5orf28	uc003jny.1,uc003jny.2,uc003jnx.1	93	115	Inf
OXCT1	uc003jmn.1	205	233	Inf
NUP155	uc010iuz.1,uc003jku.1,uc003jkt.1	507	55 135	Int
MTMR12	uc010iul.1,uc010iuk.1,uc003jhg.1	511	235	Inf
CDH12	uc003jgk.1	1715	26	Inf
CDH18 FAM134B		777	44 187	Inf
FAM173B	uc003jeo.2	151	95	Inf
FLJ33360	uc003jdn.1	157	25	Inf
CCDC127	ucuu3ipn.2 uc003iam 1	1242 140	141 121	Inf
RMND5B	uc003miq.1	119	43	Inf
PRR7	uc003mgw.1	671	47	Inf
C5orf25	uc003mdt.2.uc010ika.1	429	26	Ini
HMP19	uc003mcx.1	548	32	Inf
BNIP1	uc003mcl.2,uc003mck.2,uc003mcj.2,uc003mci.2	193	93	Inf
ATP6V0E1	ucoo3mcd.1	418	117	Inf
ERGIC1	uc003mbw.2	2172	159	Inf
	uc003map.1	167 3485	25	Inf
RARS	uc003lzx.1	137	78	Inf
HMMR	uc003jzh.1,uc003jzg.1,uc003jzf.1	113	58	Inf
GABRG2 FARP6	uc003lyz.2,uc003lyy.2,uc010ljc.1 uc003lya 1	122	26	Inf
CNOT8	uc003lvu.1	109	190	Inf
GRIA1	uc003luz.2,uc003lva.2,uc003luy.2	955	25	Inf
SIVIZA SLC6A7	ucuusiii.2 uc003lrr.1	135 414	143	Inf
SLC26A2	uc003lrh.1	85	218	Inf
FLJ41603		421	123	Inf
FBXO38	uc003lph.2,uc003lpg.1,uc003lpf.1	149	50	Inf
STK32A	uc003lom.2,uc003lol.2,uc010jgn.1	370	95	Inf
SH3RF2 KCTD16	uc003Int.1	500 667	92 60	Inf
PCDHGB2	ucoo3iiii.1	3625	114	Inf
PCDHGC3	uc003lkw.1,uc003lkv.1	954	179	Inf
PAIP2 FAM53C		204 147	154 111	Inf Inf
STSG4523	uc010jev.1	147	111	Inf
SEC24A	uc003kzs.1	641	160	Inf

UBE2B	uc003kzh.1	80	169	Inf
TCF7 CDC42SE2	uc003kzb.1,uc003kyt.1 uc003kvb 2 uc003kvi 1 uc003kvi 1	85 546	49 249	Inf Inf
CSNK1G3	uc003kto.1,uc003ktn.1,uc003ktn.1	163	244	Inf
SNX24 SNCAIP	uc010jcy.1,uc003ktt.1,uc003ktg.1 uc010jct 1 uc003ksz 1 uc003ksy 1 uc003ksy 1 uc003ksy 1	558 287	172 252	Inf Inf
SRFBP1	uc003kst.1	112	48	Inf
DCP2 ZRSR1	uc003kgh.1 uc003kgd 2 uc010icb 1	137 151	127 74	Inf Inf
APC	uc010jbz.1,uc003kpy.2,uc003kpz.2	325	54	Inf
FER HISPPD1	uc003koq.1,uc003kop.1 uc010ibo 1 uc003kod 2	289 117	205 53	Inf Inf
CAST	uc003klt.1,uc003kly.1,uc003klx.1,uc003klw.1,uc003klv.1,uc003klu.1	453	283	Inf
RHOBTB3 XRCC4	uc003kim.1 uc003kia 1 uc003kie 1 uc003kie 1 uc003kid 1 uc003kib 1	225 449	280 53	Inf Inf
MSH3	uc003kgz.1	636	160	Inf
FAM151B ZEYVE16	uc003kgv.1 uc003kgs 2 uc003kgs 2 uc003kgs 2 uc003kgr 2	193 94	179 179	Inf Inf
THBS4	uc003kgh.1	324	164	Inf
CMYA5 PAPD4	uc003kgc.1 uc003kfz 2 uc003kga 2 uc010iaf 1 uc003kgb 2 uc010iae 1	371 234	104 42	Inf Inf
JMY	uc003kfv.1	358	114	Inf
HMGCR TNPO1	uc003kdg.1,uc003kdp.1 uc003kck.2	88 459	163 187	Inf
PTCD2	uc003kcc.1,uc003kcb.1	123	73	Inf
MAP1B CENPH	uc010iyy.1 uc010ixc 1 uc003ivp 1	167 116	49 78	Inf Inf
SLC30A5	uc003jvh.1	142	150	Inf
SFRS12	uc003juv.1 uc010iwy.1.uc003jup.1	311 105	50 201	Inf
C5orf44	uc010iwv.1.uc003juc.2.uc003jua.2.uc003jtz.2	140	62	Inf
RGS7BP	uc003jtm.2,uc010iwt.1,uc003jtn.1 uc003iti.1	98 198	56 146	Inf
IPO11	uc003itc.1	781	127	Inf
NDUFAF2 GPBP1	uc003jsp.2 uc003jri,2.uc003jrh,2.uc010jwg,1	294 479	98 178	Inf
NDUFS4	uc003jpe.2	241	33	Inf
ITGA1	ucuu3jos.1 ucu03jou.1	117 394	81 81	Inf
NNT	uc003joe.1,uc003jof.1	248	141	Inf
MGC42105 GHR	ucu03jno.1 uc003imt.1	380 498	177 206	Inf
DKFZp547K2	uc010iva.1	858	38	Inf
DNAJC21	uc003jkv.1 uc003jic.1.uc003jib.1	975 105	38 211	Inf
KIAA1334	uc010ius.1	538	28	Inf
TARS	uc010iur.1,uc003jir.1 uc003ihz.1.uc010jup.1.uc003ihv.1	742 84	103	Inf
PDZD2	uc003jhm.1	2714	36	Inf
GUSBP1	uc003jnd.1,uc003jne.1 uc010jub.1	321 99	136 25	Inf
FBXL7	uc003jfn.1	1405	219	Inf
KIAA1234	ucousicy. 1 uco10isv.1	292 3954	25	Inf
AHRR	uc003jaw.1,uc003jav.1	3954	25	Inf
ACSL1	uc003ixy.2,uc003ixw.2 uc003iwt.1	565	29	Inf
MLF1IP	uc003iwr.1,uc003iwg.1	152	179	Inf
ENPP6	uc003iwg.1	1542	27	Inf
GPM6A	uc003iuh.1,uc003iug.1	977 361	150	Inf
FBXO8	uc003itg.1,uc003itg.1	84	68	Inf
C4orf27	uc003isl.2	96 907	116	Inf
FSTL5	uco03in.2,uc003in.1,uc003in.1	1572	33	Inf
C4orf18 PDGEC	uc003ipp.2 uc003ini 1 uc003ini 1	82 457	83 196	Inf
PET112L	uc003imm.2,uc003iml.1	323	36	Inf
SH3D19 I RBA	uc003imc.2 uc003ilu 2	318 2468	29 104	Inf Inf
DKFZp761C0	uc003ild.1	161	138	Inf
LOC90826 OTUD4	uc003ilc.1 uc003ikb.2.uc003iiz.2.uc003ika.2	161 116	138 76	Inf
ANAPC10	uc003ijv.2,uc003iju.2,uc003ijw.2	233	96	Inf
SCLT1	uc003iat.2.uc003iar.2.uc010iob.1.uc003iaa.2.uc003iap.2	390 86	97 138	Inf
BBS7	uc003iee.1,uc003ied.1	122	91	Inf
PDE5A PRSS12	ucuosidg. 1,ucuosidi. 1 uc003ica.1	206	289	Inf
PITX2	uc003iaf.1	1181	31	Inf
PAPSS1	uco10inc.1,uco03n2c.1 uc003hyk.1	226	232	Inf
	uc003hye.1,uc010ilv.1,uc003hyf.1,uc003hyc.1	472	78	Inf
CENPE	uc003hxc.1,uc003hxb.1	135	82	Inf
NHEDC1	uc010ilm.1,uc003hwu.1,uc003hww.1	242	71	Inf
MANBA	ucoosniwi. r.ucoosniwi. r ucooshwg.1	398	141	Inf
PPP3CA	uc010ilk.1,uc010ilj.1,uc003hvu.1,uc003hvs.1,uc003hvv.1	528 528	236	Inf
SNCA	uc003hsq.1,uc003hsp.1	247	170	Inf
HEL308	uc010ikb.1,uc003hom.1	234 182	61 125	Inf
KIAA2037	uc010iju.1,uc003hny.2	398	78	Inf
LIN54 SEC314	uc010ijt.1,uc003hnz.2,uc003hnz.2 uc003hno 2 uc003hno 1 uc003hno 1 uc003hno 1 uc003hno 1 uc003hno 1 uc003hno 1 uc003hno 1 uc003hno 1 uc003hno 1	398 166	78 167	Inf
SCARB2	uc003hju.1	207	234	Inf

NUP54 SDAD1 SDAD1 PPEF-2 G3BP2 CDKL2 BTC RASSF6 ANKRD17 ADAMTS3 ADAMTS3 CENPC1 EPHA5 PPAT CLOCK SCFD2 TEC CRN CORIN GABRA2 PHOY28 PHOX2B RBM47 C4orf34 RFC1 KIAA0746 KIAA0746 PPARGC1A KCNIP4 LCORL FBXL5 BC035722 RAB28 ACOX3 AFAP1 JAKMIP1 STX18 STX18 WHSC2 FAM53A FAM53A FLJ14297 ZNF721 NEIL3 CLCN3 PALLD ETFDH TMEM144 TRIM2 ARFIP1 IL15 MGST2 INTU SPATA5 FGF2 KIAA1627 NDST3 C4orf16 SCYE1 GSTCD CISD2 KIAA1122 SMARCAD1 ARHGAP24 COPS4 C4orf22 C40ff22 ANXA3 MRPL1 AK310457 CR627383 FAM47D MOBKL1A ARL9 KIAA1211 FIP1L1 PDGFRA PDGFRA SPATA18 SLAIN2 GABRB1 LIMCH1 PGM2 TBC1D19 ZCCHC4 CD38 C40of23 XIAA0232 KIAA0232 DKFZp762M1 KFZp762M STK32B AK056081 RGS12 C4orf8 ZNF141 IQCG RNF168 PCYT1A MUC4 FGF12 BCL6 EHHADH EHHADH ABCC5 PARL MCC-B MCCC1 ZMAT3

uc003hit.1,uc010ije.1,uc003his.1
uc003bit 1 uc003bis 1 uc003bir 1
uc003hig.2
uc010iil.1,uc010iik.1,uc003hhd.1,uc003hhc.1
uc003hqr.1,uc003hqq.1,uc003hqp.1
uc003hgk.1
uc010ihm.1,uc003hdd.1
uc003hda.1,uc003hcx.1,uc003hcz.1,uc003hcy.1
uc003hbr.1
ucuu3haz.1
uc003gwt 2
uc003gvc 2
uc003quo.1
uc003gtz.1,uc003gty.1,uc003gtx.1
uc003gru.2
uc003gqs.1
uc003qqe.1
uc003gpq.1
uc010idy.1,uc003god.1,uc010idx.1,uc003goc.1,uc003gob.1
uc003gmw.1
uc003gmu.1,uc003gmt.1
ucousgen. 1,ucousgen. 1
uconount, ucoogae.2,ucoogae.2,ucoogae.2,ucoogae.2
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uc003iev.1
uc003icg.1,uc003icf.1
uc003ibx.1
uc010imm.1,uc003iak.2,uc003iaj.2
uc003hyg.1,uc003hyh.1
uc003hxx.2,uc010ils.1,uc003hxy.2
uc003hwt.2
uc003htb.2,uc003htc.2,uc003htd.2
ucuu3npi.1,ucuu3npi.2,ucuu3npk.1
uc010iha.1
uc003gzx.2.uc003hab.1.uc003gzz.1.uc003gzv.1
uc003haa.1
uc003qzk.1,uc003qzl.1
uc003qya.2
uc003gxh.1
uc003gwd.2,uc003gwc.2
uc003gta.1
uc010iew.1,uc003gsf.2
uc003arl.2
uc003goj.1,uc003gol.1
uc003glf.1
uc003glq.2,uc003glr.2
uc003fzz 2 uc003gab 2 uc003gab 2
uc003fva.2.uc003fvn.1.uc003fvo.1
uc010jah 1 uc003fwg 1
uc003fwh.1.uc003fwg.1
uc010hzu.1
uc010hzu.1 uc003fsx.1
uc010hzu.1 uc003fsx.1 uc003frp.2,uc010hza.1
uc010hzu.1 uc003fsx.1 uc003frp.2,uc010hza.1 uc003fpf.1
uc010hzu.1 uc003fsx.1 uc003frp.2,uc010hza.1 uc003frf.1 uc010hxo.1,uc010hxn.1,uc003fmh.1,uc003fmg.1
uc010hzu.1 uc003ftsx.1 uc003ftr.2,uc010hza.1 uc003ftrf.1 uc010hxo.1,uc003ftnf.1,uc003ftng.1 uc0003ftme.1,uc003ftnd.1
uc010hzu.1 uc003fsx.1 uc003fp.2,uc010hza.1 uc003fpf.1 uc010hxo.1,uc010hxn.1,uc003fml.1,uc003fmg.1 uc003fme.1,uc003fml.1 uc003flf.1
uc010hzu.1 uc003fsx.1 uc003fp.2,uc010hza.1 uc003fpf.1 uc010hxo.1,uc010hxn.1,uc003fml.1,uc003fmg.1 uc003fme.1,uc003fml.1 uc003fle.1
uc010hzu.1 uc003fsx.1 uc003fp.2,uc010hza.1 uc003fpf.1 uc010hxo.1,uc010hxn.1,uc003fmh.1,uc003fmg.1 uc003ff.1 uc003ff.1 uc003ff.1 uc003ff.1

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AADACL1	uc003fig.1	429	80 95	Inf
PDCD10	uc003fez.1,uc003fex.1,uc003fey.1	97	103	Inf
ZBBX KPNA4	uc003fdp.1 uc003fdn.1	235 213	60 358	Inf
TRIM59 SHOX2	uc003fdm.1	81 261	170 189	Inf
DHX36	uc003ezz.2.uc003ezy.2.uc010hvq.1	118	46	Inf
WWTR1 HLTF	uc003exe.1,uc003exf.1 uc003ews.1.uc010hye.1.uc003ewr.1.uc003ewa.1	666 198	165 124	Inf Inf
PLSCR4	uc003evu.2,uc010hva.1,uc010huy.1,uc003evt.2,uc010huz.1	96 154	147	Inf
AMOTL2	uc003eqh.1,uc003eqg.1	452	230	Inf
TOPBP1 ACAD11	uc003eps.1 uc003eov.2	163 94	113 114	Inf Inf
CPNE4	uc003eok.1	819	114	Inf
C3orf25	ucuosemx.2 uc003emg.1	324	83	Inf
KIAA1160 RAB43	uc010hsy.1	300 300	250 250	Inf Inf
KIAA1257	uc003elg.1,uc003elj.2	653	135	Inf
ZNF148	uc003ekr.1 uc003ehy.2,uc003eia.2,uc010hsa.1,uc003ehz.2,uc003ehx.2	259 521	196 157	Inf Inf
SLC12A8	uc010hrz.1,uc003ehv.2	1002	154	Inf
FSTL1	uc0004ee.2 uc010hrb.1,uc003eds.1	151	252	Inf
GSK3B LSAMP	uc003edo.1,uc003edn.1 uc003ebt.1	753 1207	53 59	Inf Inf
ZBTB20	uc003ebn.1	1722	121	Inf
CCDC52	uc003eaq.2	185	76 72	Inf
CBLB ZBTB11	uc003dwe.1,uc003dwd.1,uc003dwc.1	369 136	165 205	Inf Inf
PROS1	uc003drb.2	258	122	Inf
PDZRN3	ucuu3apq.2,ucuu3app.2 uc003dpl.1	91 1247	45 388	Inf
FRMD4B	uc003dny.2,uc003dny.2 uc003dmr 2 uc003dmp 1 uc003dmp 1 uc003dmp 1	320 2118	117	Inf
THOC7	uc003dlu.2,uc003dlt.2	85	250	Inf
FHIT DNASE1L3	uc010hnn.1,uc003dky.2,uc003dkx.2 uc003dio.1	3032 130	100 29	Inf Inf
FAM116A	uc003dia.1	232	160	Inf
IL17RD	uc003dik.1	373	25	Inf
ARHGEF3 WNT5A	uc003dif.2,uc003dih.2 uc003dbm 1 uc010bmw 1	234 208	27 35	Inf Inf
NEK4	uc003dfr.2,uc003dfq.2	144	101	Inf
TWF2	uc010nmc.1 uc003ddd.1	282	194 194	Inf
WDR51A DKF7p434C2	uc003dcw.1,uc003dcu.1	878 878	113 113	Inf
PCBP4	uc003dcl.1	92	75	Inf
RASSF1	uc003db0.1 uc003dab.1,uc003dac.2	159 81	106 238	Inf
HYAL3	uc003czg.1,uc003czf.1,uc003cze.1,uc003czd.1	119	114	Inf
CAMKV	uc003cxv.1,uc003cxv.1	123	138	Inf
IHPK1 IP6K1	uc003cxo.1 uc003cxn.1.uc003cxm.1	444 500	142 142	Inf Inf
RHOA	uc010hku.1	439	236	Inf
SLC25A20	uc0003cwp.1 uc010hkj.1,uc003cva.2	225 263	25 72	Inf
COL7A1 SHISA5	uc003ctz.2 uc003cto 1 uc003cto 1	900 338	159 123	Inf Inf
DKFZp547H1	uc003crw.1	297	25	Inf
MAP4 KIF9	uc003cst.2,uc003csc.2,uc003csg.1,uc003csp.1 uc010hjp.1,uc003cqz.1,uc003cqy.1,uc003cqx.1	839 347	171 165	Inf
LTF FYCO1	uc010hjh.1,uc003cpq.1	104 502	103 108	Inf
ZDHHC3	uc003cog.1,uc003cof.1	352	158	Inf
ZNF445 SEC22C	uc003cnf.2 uc003cli.1	193 364	106 141	Inf Inf
ULK4	uc003ckx.1,uc003ckw.2,uc003ckv.2	395	113	Inf
FLJ45032	ucooscii. 1,ucooscii. 1,ucooscii. 1 ucooscik.1	84	159	Inf
SBC2 SI C4A7	uc010hfm.1 uc003cdu 2	330 419	304 304	Inf Inf
NGLY1	uc003cdm.1,uc010hfg.1,uc003cdl.1	136	129	Inf
NKIRAS1	ucuu3caa.2,ucuu3cae.1,ucuu3cac.2,ucuu3ccz.2,ucuu3ccz.2,ucuu3ccx.2,ucuu3ccx.2,ucu10nte.1 uc003ccm.1,uc003ccl.1,uc003cck.1,uc003ccj.1	587 105	204 173	Inf
ZNF385D TBC1D5	uc003cce.1	520 1197	53 29	Inf Inf
SH3BP5	uc003bzp.1	585	217	Inf
RAF1 C3orf31	uc003bxf.2 uc003bwj.1.uc003bwh.1	363 238	150 74	Inf Inf
SEC13	uc003bvq.1,uc003bvm.1,uc003bvp.1,uc003bvo.1,uc003bvn.1	166	73	Inf
KIAA0411	uc003brk.2	1342	48	Inf
SRGAP3 RAD18	uc003brg.1,uc003brf.1 uc003brd 1	2039 154	48 56	Inf Inf
SUMF1	uc010hby.1,uc003bpz.1	315	84	Inf
LMLN OPA1	ucuuuas.1,ucuu3tyu.1,ucuu3tyt.1,ucuu10iar.1,ucu03tyt.1 uc003ftn.1,uc003ftm.1,uc003ftl.1,uc003fti.1,uc003fti.1,uc003fti.1,uc003fti.1,uc003ft	468 319	151 112	Inf Inf
IL1RAP	uc010hzf.1,uc003fsl.1,uc003fso.1,uc010hzg.1,uc003fsq.2,uc003fsk.1,uc003fsm.1	375	198	Inf
MAP3K13	ucousiis.i uco10hyf.1	895	25 86	Inf
VPS8 FAM131A	uc010hyd.1,uc003fpb.1 uc003foq.1	781 125	96 51	Inf Inf

PSMD2	uc003fnn.1	97	165	Inf
ECE2	uc003fni.2	1132	107	Inf
FXR1	uc003fkr.1.uc003fkg.1.uc003fkp.1	370	100	Inf
NDUFB5	uc003fke.1.uc003fkc.1	97	80	Inf
NI GN1	uc010bww 1 uc003fin 1	1309	52	Inf
ENDC3B	uc003fbz 2 uc010bwt 1	1593	241	Inf
SERPINI1		178	53	Inf
SCHIP1		535	208	Inf
GEM1		112	230	Inf
	ucoolici.2, ucoolici.1	112	111	Inf
KONADA		002	100	li li
KUNABI		728	120	Ini
MME		206	132	int
MED12L	uc003eyo.2,uc003eyn.2,uc003eyp.1,uc003eym.1	428	232	Inf
SELT	uc003evf.1	189	143	Inf
FAM62C	uc003esk.1	435	283	Inf
ARMC8	uc003esb.1	376	30	Inf
BFSP2	uc003epn.1	535	52	Inf
DNAJC13	uc010htg.1.uc003eor.1	131	129	Inf
NEK11	uc003enw.1.uc003eoa.1.uc003enx.1.uc003env.1	201	84	Inf
nek11l	uc003enz 1	544	84	Inf
C3orf37	uc003elu 1 uc003elw 1 uc003elv 1 uc003elt 1	137	234	Inf
		372	134	Inf
PAR7		622	170	Inf
		022	170	

Table S4.3

List of candidate genes whose expression was up-regulated in the Saqqaq hairs.

This list corresponds to the top 1,000 non-redundant genes showing the highest R_s values (transcripts are collapsed into a single Gene name, providing the maximal R_s value observed in this quartile). R_s values and respective coverage in PM and RE regions are provided.

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