

Figure S1. Experimental strategy for 3D alginate culture for induction of chondrogenesis of rat MSC. BM-MSC were isolated from the tibia and femurs of Dark Agouti (DA) and Lewis (LEW) rats and expanded in vitro. Chondrogenic differentiation was induced in 3D alginate culture by incubation with TGF β -3 and BMP-2 for up to 21 days (21d). Differentiated MSC refer to MSC that were cultured in 3D alginate layers and which displayed morphological and gene expression changes consistent with a chondrocyte-like phenotype.

Figure S2. Lewis chondrogenic differentiated MSC do not suppress allogeneic T-cell proliferation. (a) CFSE-labeled allogeneic DA T cells were stimulated with anti-CD3/CD28 beads in the presence of undifferentiated or differentiated Lewis MSC at the indicated ratios. On day 4, cells were harvested and the percentage proliferation was analysed by flow cytometry. (b) Gating strategy used for analysis is shown. CD4⁺ and CD8⁺ T cells were identified using anti-CD4-APC and anti-CD8-PE antibodies and analysed by flow cytometry. (c). CFSE dilution was analysed in gated CD4⁺ and CD8⁺ cells and the percentage proliferation was calculated, and graphed as shown. Representative results of two independent experiments are shown \pm SEM. . *p<0.05, **p<0.01, ***p<0.001

Figure S3. Chondrogenic differentiated human MSC do not suppress allogeneic T-cell proliferation. (a) CFSE-labeled allogeneic PBMCs were stimulated with anti-CD3/CD28 soluble antibodies in the presence of undifferentiated or differentiated MSC at indicated ratios for 4 days. (b) The corresponding CFSE dilution histograms are shown to indicate the percentage of CD4⁺ and CD4⁻ lymphocyte proliferation. CD4⁺ and CD4⁻ proliferation was determined similarly to that outlines in the gating strategy in Figure S2b. Data from three independent MSC donors are shown. *p<0.05, **p<0.01, ***p<0.001

Figure S4. Experimental strategy for the generation of cytotoxic T cells DA-specific allogeneic cytotoxic T cells (CTLs) were generated in a one-way mixed lymphocyte culture of LEW and DA T cells as outlined. Undifferentiated and differentiated syngeneic and allogeneic MSC were incubated with allo-antigen specific CTLs in an effector to target ratio of 100:1, for 4 hours, following which time the fluorescence of the supernatants were analysed by a fluorescence plate reader. Specific lysis was calculated from mean fluorescence of replicates as follows: $(F[\text{sample}] - F[\text{spontaneous release}] / (F[\text{maximum lysis}] - F[\text{spontaneous release}]) \cdot 100 = \% \text{ specific lysis}$.

Figure S5. Subcutaneous Implantation experimental strategy and morphological evidence for chondrogenic differentiation in syngeneic differentiated and allogeneic differentiated alginate layers. (a) The experimental strategy for subcutaneous implantation is outlined here. Syngeneic and allogeneic undifferentiated and differentiated MSC were implanted subcutaneously in LEW rats. Allogeneic splenocytes were implanted as an immunogenicity positive control. 6 weeks later, the area surrounding the implant was removed, formalin fixed and paraffin embedded. The number of CD3⁺ and CD68⁺ cells within the implants were analysed by immunohistochemistry (b) Representative histological sections, at 20X magnification (upper row) and 40X magnification (lower row) illustrating H&E staining of syngeneic differentiated, allogeneic undifferentiated and allogeneic differentiated MSC implants. 40X magnification (lower row) clearly illustrates the morphological characteristics of chondrocytes within the syngeneic and allogeneic differentiated alginate layers.

Figure S6. Experimental strategy for the *in vivo* analysis of injection of unencapsulated and implantation of alginate encapsulated differentiated and undifferentiated MSC syngeneic

and allogeneic MSC. The experimental strategy for subcutaneous injection and implantation is outlined here. Syngeneic and allogeneic undifferentiated and differentiated MSC were either injected or implanted subcutaneously in LEW rats. Allogeneic splenocytes were also implanted as an immunogenicity positive control. 6 weeks later, the draining blood, lymph nodes and spleen were removed from each animal. An ex vivo re-stimulation assay was used to analyze the allo-specific and third party memory T cell responses, both locally and systemically, in the recipient animals. Anti-donor antibody responses were analysed by flow cytometry of donor strain (DA) cells incubated with recipient sera from the various groups.

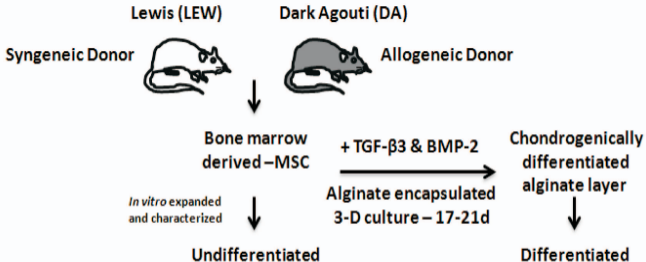


Figure S1

a

Dark Agouti (DA) → Lewis (LEW)



Lewis (LEW) → Dark Agouti (DA)

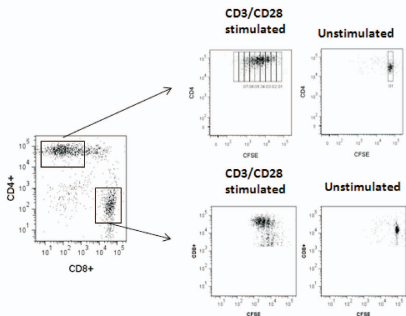
CFSE labeled
Mixed lymphocytesUndifferentiated MSC
&
Differentiated MSC

CD28/CD3 Stimulation



MSC-mixed lymphocyte co-culture

MSCs - 1:1000-1:10

b**c**

LEW MSC → DA T-cells

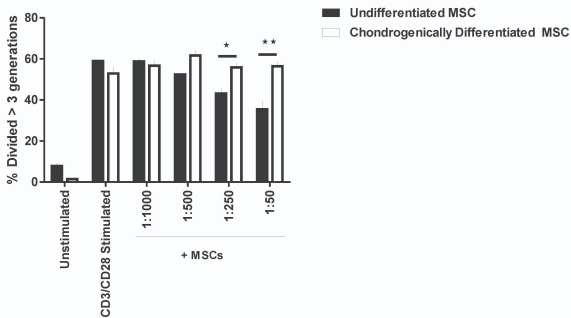
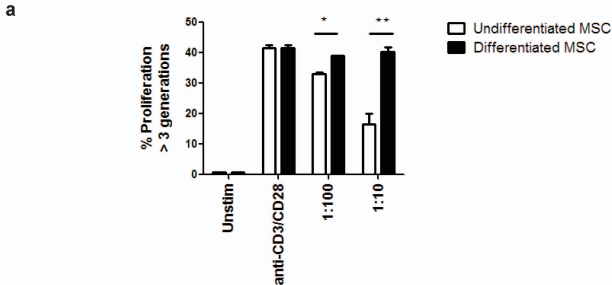


Figure S2



b

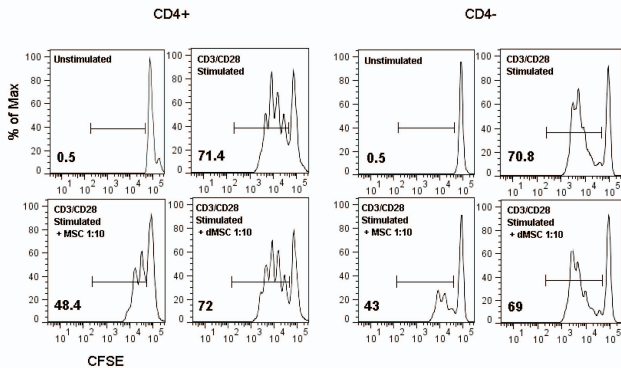


Figure S3

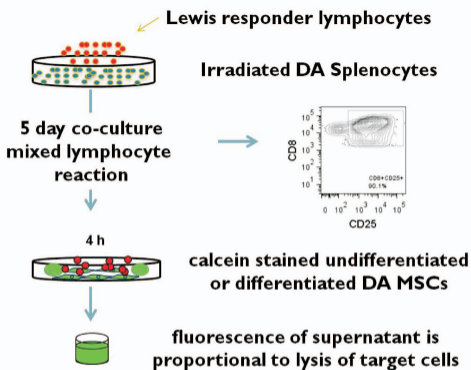


Figure S4

a**Subcutaneous implantation
(alginate encapsulated cells)****1 x 10⁶ cells
in alginate layer****Lewis (LEW)
Recipient**

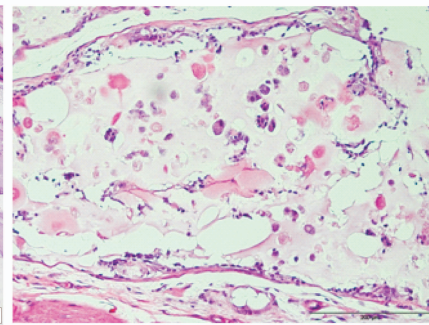
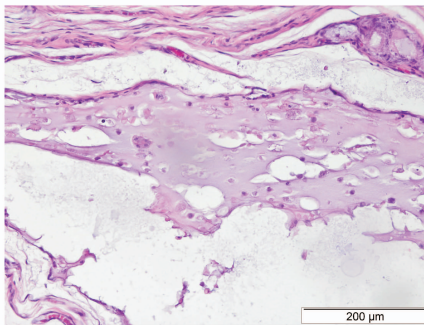
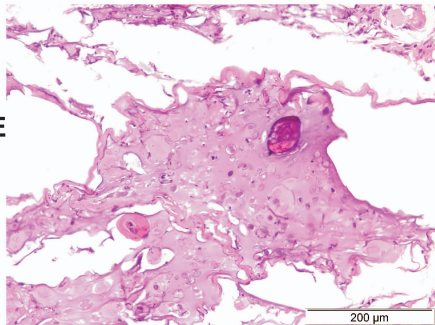
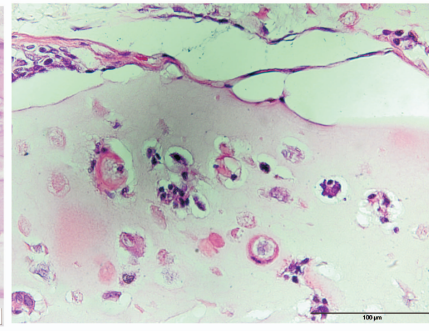
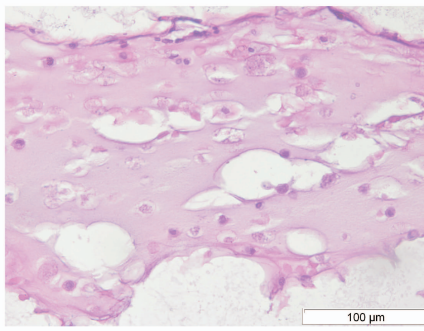
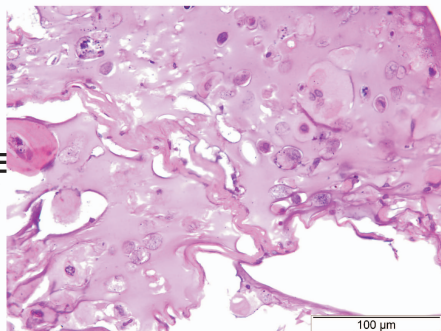
- Syngeneic undifferentiated MSC
- Syngeneic Differentiated MSC
- Allogeneic undifferentiated MSC
- Allogeneic Differentiated MSC
- Allogeneic Splenocytes00

d42 (6 weeks)**Isolation of subcutaneous implant site****Histological and Immunohistochemical analysis
(H&E, CD68, CD3)****b**

Syngeneic differentiated MSC

Allogeneic undifferentiated MSC

Allogeneic differentiated MSC

H&E
20XH&E
40X**Figure S5**

Subcutaneous injection
(alginate released cells)

Subcutaneous implantation
(alginate encapsulated cells)

1×10^6
single cell suspension

1×10^6 cells
in alginate layer

- Syngeneic undifferentiated MSC
- Syngeneic Differentiated MSC
- Allogeneic undifferentiated MSC
- Allogeneic Differentiated MSC
- Allogeneic Splenocytes



Lewis (LEW)
Recipient



- Syngeneic undifferentiated MSC
- Syngeneic Differentiated MSC
- Allogeneic undifferentiated MSC
- Allogeneic Differentiated MSC
- Allogeneic Splenocytes



d42 (6 weeks)



Isolated serum
- Detection of allo-antibodies

Isolated draining lymph nodes
and spleen

Isolated serum
- Detection of allo-antibodies

4d

Ex vivo re-stimulation assay

DA OX62* (Donor)

Wistar Furth (WF) OX62* (Third Party)

Proliferation and Activation

Figure S6