

Targeting Extracellular DNA to Deliver IGF-1 to the Injured Heart

Raffay S. Khan¹, Mario D. Martinez², Jay C. Sy², Karl D. Pendergrass¹, Pao-lin Che², Milton E. Brown^{1, 2}, E. Bernadette Cabigas², Madhuri Dasari², Niren Murthy^{2,3}, Michael E. Davis^{1,2,3,4,*}

¹ Division of Cardiology, Emory University, Atlanta, Georgia 30322, USA

² Wallace H. Coulter Department of Biomedical Engineering, Emory University and Georgia Institute of Technology, Atlanta, Georgia 30322, USA.

³ Department of Bioengineering, University of California at Berkeley, Berkeley, CA, 94720, USA

⁴ Emory+Children's Center for Cardiovascular Biology, Children's Healthcare of Atlanta, Atlanta, GA 30332, USA

* To whom correspondence should be addressed:

Michael E. Davis, PhD

The Wallace H. Coulter Department of Biomedical Engineering

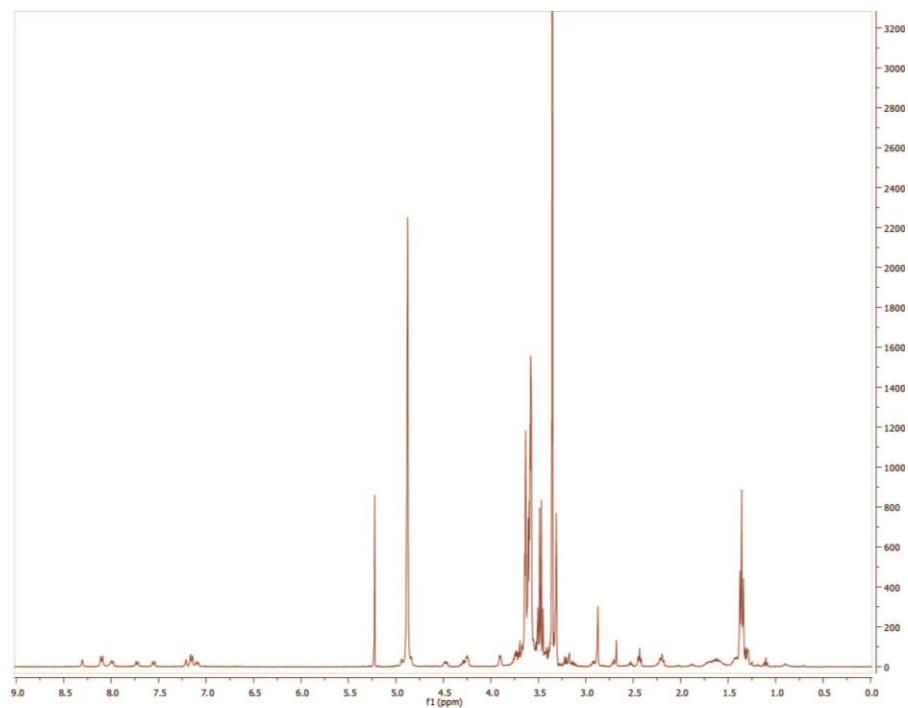
Emory University and Georgia Institute of Technology

101 Woodruff Circle, Suite 2001, Atlanta, GA 30322, USA

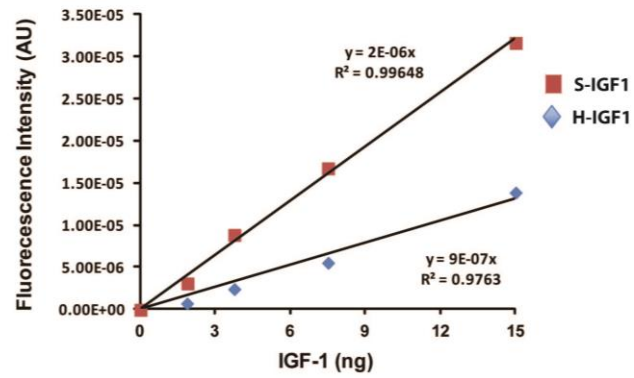
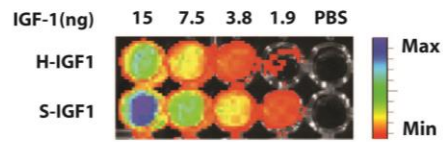
E-mail: michael.davis@bme.emory.edu

Supplemental Data

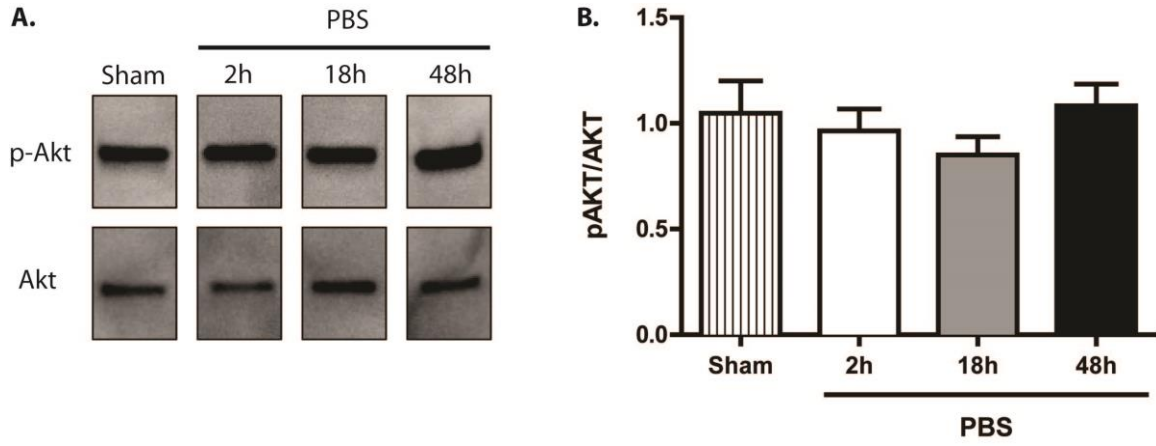
Supplemental Figure 1



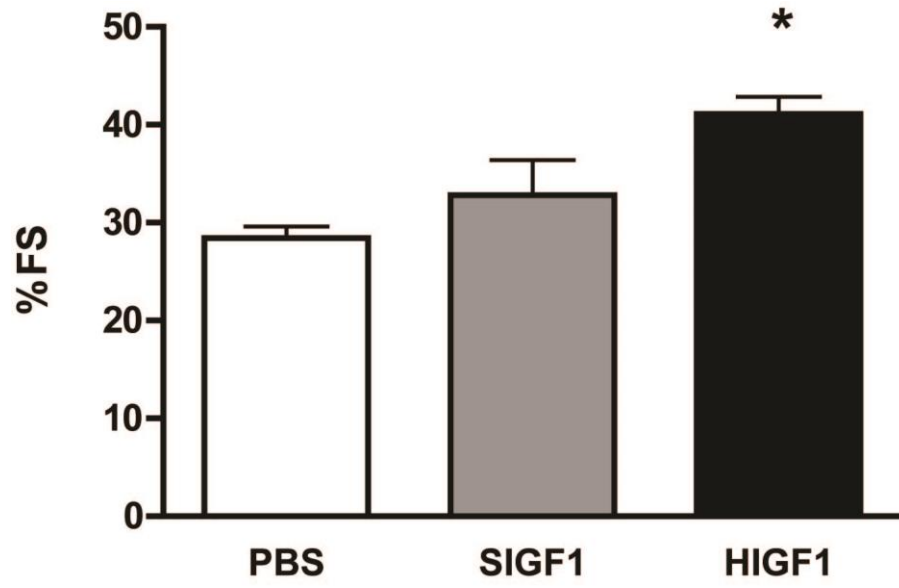
Supplemental Figure 2



Supplemental Figure 3



Supplemental Figure 4



Supplemental Figure Legends

Figure 1: **Hoechst-Biotin Nuclear Magnetic Resonance (NMR) spectra.**

Hoechst-biotin was characterized by $^1\text{H-NMR}$ (400 MHz, CD_3OD) by tracking aromatic Hoechst Protons (δ 8.10, d, 8.68 Hz, 2H; δ 7.99, d, 9.5 Hz, 2H; δ 7.73, d, 8.68 Hz, 1H; δ 7.55, d, 9.5 Hz, 1H; δ 7.15, m, 4H) to Biotin protons (δ 4.48, m, 1H; δ 4.30, m, 1H; δ 2.68, m, 1H; δ 2.20, t, 8.74 Hz, 2H).

Figure 2: **Hoechst-IGF-1 (H-IGF1) and Streptavidin-IGF-1 (S-IGF1) standard curves.**

Differences in fluorescence efficiencies between H-IGF1 and S-IGF1 were accounted for by generating standard curves. Known amounts of H-IGF1 and S-IGF1, as measured by ELISA were added to a 96-well plate and imaged using a fluorescence-imaging camera (Xenogen IVIS 200 camera) with the same settings used to image harvested tissues. Fluorescence efficiency was measured and plotted against IGF-1 to generate standard curves for both H-IGF1 and S-IGF1. Compared to equal amounts of H-IGF1, S-IGF1 had considerably greater fluorescence efficiency.

Figure 3: **No changes in Akt phosphorylation between sham and PBS-**

treated mice following ischemia-reperfusion. Proteins were extracted from animals subject to sham surgery or ischemia-reperfusion for the indicated time and Western analysis was performed for both phospho- and total Akt. Bands were quantified and expressed as a ratio of phospho-to-total (p-Akt/Akt).

Representative blots and grouped data demonstrate no difference in Akt phosphorylation in any control group (n=4 per group).

Figure 4: **H-IGF1 preserves cardiac function two weeks following ischemia-reperfusion.** Echocardiography was used to measure cardiac contractility in PBS, S-IGF1 and H-IGF1 treated mice. Percent fractional shortening (%FS), an index of systolic function, was significantly greater in H-IGF1 treated mice compared to S-IGF1 and PBS treated mice (n=7-11 per group, *, $p < 0.05$ vs. S-IGF1, one-way ANOVA followed by Tukey's post-test).