

***JAK2* or *CALR* mutation status defines subtypes of essential thrombocythemia with substantially different clinical course and outcomes**

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Supplemental Methods and Tables

Supplemental Methods

Diagnosis of *JAK2* mutated polycythemia vera according to the criteria of the British Committee for Standards in Hematology

Diagnosis of *JAK2* mutated polycythemia vera requires both of the following 2 criteria to be present: 1) high hematocrit (>0.52 in men, >0.48 in women) or raised red cell mass (>25% above predicted); 2) mutation in *JAK2*, i.e., *JAK2* (V617F) or mutation of exon 12. These criteria are very similar to the major criteria of the WHO classification, which considers hemoglobin level (>18.5 g/dL in men, >16.5 g/dL in women, or other evidence of increased red cell mass) instead of hematocrit. The only difference is that the WHO classification requires, in addition to the two major criteria, at least one minor criterion (bone marrow biopsy showing hypercellularity, serum erythropoietin below the normal range, or endogenous erythroid colony formation in vitro).

In order to assess polycythemic transformation of essential thrombocythemia, we adopted the criteria of the British Committee for Standards in Hematology since sequential evaluations of bone marrow biopsy were not available in most patients.

References

1. McMullin MF, Bareford D, Campbell P, et al. Guidelines for the diagnosis, investigation and management of polycythaemia/erythrocytosis. *Br J Haematol*. 2005;130(2):174-195.
2. McMullin MF, Reilly JT, Campbell P, et al. Amendment to the guideline for diagnosis and investigation of polycythaemia/erythrocytosis. *Br J Haematol*. 2007;138(6):821-822.

Sanger sequencing of CALR exon 9

The following M13-tagged primers were used for PCR

- Forward 5'-tgtaaacgacggccagtAAGCAAGGGCTATCGGGTAT-3'

- Reverse 5'-caggaaacagctatgaccAACCAAATCCACCCCAAAT-3'

and for sequencing

- Forward 5'-TGTA AACGACGGCCAGT-3'.

Supplemental Table 1. Patients studied in the present work in the analyses of genotype/phenotype relationships and clinical outcomes.

Myeloproliferative neoplasm	JAK2 (V617F) mutated (%)	CALR exon 9 mutated (%)	JAK2/MPL/CALR wild-type (%)	All genotypes
<i>Patients belonging to the initial patient population</i>				
Essential thrombocythemia	466	176	75	717
Polycythemia vera	468	-	-	468
All patients with PV or ET				1185
<i>Patients with secondary myelofibrosis evaluated for mutant allele burden</i>				
Post-ET myelofibrosis	18	10		28
Post-PV myelofibrosis	55			55
All patients with secondary myelofibrosis				83
<i>Total no.</i>				<i>1268</i>

Supplemental Table 2. Type and frequency of *CALR* mutations in patients with essential thrombocythemia with nonmutated *JAK2* and *MPL*. The nomenclature adopted here is that of Klampfl et al [N Engl J Med. 2013 Dec 19;369(25):2379-90. Epub 2013 Dec 10].

<i>CALR</i> mutation	Nucleotide	Change	Frequency	
			No.	%
Type 1	c.1092_1143del	AAEKQMKDKQDEEQRTRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	81	46.0
Type 2	c.1154_1155insTTGTC	AAEKQMKDKQDEEQRLKEEEDKRRKEEEEAE DN CRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	67	38.1
Type 3	c.1095_1140del	AAEKQMKDKQDEEQRQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	2	1.1
Type 4	c.1102_1135del	AAEKQMKDKQDEEQRLRRRQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	4	2.3
Type 5	c.1091_1142del	AAEKQMKDKQDEGQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	2	1.1
Type 6	c.1094_1139del	AAEKQMKDKQDEERRQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 8	c.1104_1137del	AAEKQMKDKQDEEQRLKRRQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 9	c.1140del	AAEKQMKDKQDEEQRLKEEEDKRRKEEERQTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	2	1.1
Type 10	c.1154delinsTGTGTC	AAEKQMKDKQDEEQRLKEEEDKRRKEEEEAE DM CRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 13	c.1100_1134delinsA	AAEKQMKDKQDEEQRQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 16	c.1102_1137delinsCA	AAEKQMKDKQDEEQRLQRRQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 18	c.1104_1155del	AAEKQMKDKQDEEQRLKRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 20	c.1118_1136del	AAEKQMKDKQDEEQRLKEEEDGRRQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 21	c.1118_1145delinsCGTTTA	AAEKQMKDKQDEEQRLKEEEDALRGQGG-	1	0.6
Type 22	c.1120_1123del	AAEKQMKDKQDEEQRLKEEEDNAKRRRRQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 28	c.1131_1152del	AAEKQMKDKQDEEQRLKEEEDKRRKRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 32	c.1153_1154delinsTGTC	AAEKQMKDKQDEEQRLKEEEDKRRKEEEEAE DC RRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 34	c.1154_delinsCTTGTC	AAEKQMKDKQDEEQRLKEEEDKRRKEEEEAE TC RRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 36	c.1155_1156insTGTCG	AAEKQMKDKQDEEQRLKEEEDKRRKEEEEAE DK CRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	2	1.1
Type 37 [§]	c.1091-1124del	AAEKQMKDKQDEDAKRRRRQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 38 [§]	c.1101-1152del	AAEKQMKDKQDEEQRLRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 39 [§]	c.1109-1145del	AAEKQMKDKQDEEQRLKEERTRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Type 40 [§]	c.1113A>C;1122-1140del	AAEKQMKDKQDEEQRLKEEDEDKRRQRTRRRMMRTKMRMRMRRTRRKMRRKMSPARPRTSCREACLQGWTEA-	1	0.6
Total			176	100
<i>Wild-type reference sequence</i>		AAEKQMKDKQDEEQRLKEEEDKRRKEEEEAE DK EDEDKDEDEDEEDKEEDEEDVPGQAKDEL-		

* The aminoacid sequence starting from codon A352 is reported. Acid and basic residues are in red and blue, respectively.

[§] Novel *CALR* mutations with respect to *CALR* abnormalities described in the original report by Klampfl et al [N Engl J Med. 2013 Dec 19;369(25):2379-90. Epub 2013 Dec 10].