Supplemental Materials Molecular Biology of the Cell

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SUPPLEMENTAL FIGURE LEGENDS

Supplemental Figure 1: Aim44p-GFP is a functional fusion protein. The fusion of a C-terminal GFP tag to Aim44p does not produce a multibudded phenotype in cells in late-log phase liquid culture (SC), indicating that the tag does not interrupt its normal function in promoting cell division. The percentage of multibudded cells in the Aim44p-GFP expressing cells is not statistically different from wild-type cells (p = 0.9, Student's t-test). *aim44* Δ cells are multibudded compared to wild-type and Aim44p-GFP expressing cells (p = .0085 and p = 0.01, respectively, Student's t-test). Error bars represent standard deviation of 3 independent experiments. $n \ge 100$ cells for each strain in each independent experiment.

Supplemental Figure 2: Localization of Cdc3p-mCherry and Myo1p-GFP during cell division in wild-type cells. Cells expressing *MYO1* tagged with GFP at its chromosomal locus and plasmidborne Cdc3p-mCherry were grown to mid-log phase and imaged by fluorescence microscopy. Deconvolved projections of representative images of Cdc3p-mCherry (red) and Myo1p-GFP (green) at the bud neck of cells with small, medium, and large buds (**a-c**, respectively) and separated mother and daughter cells (**d**). (**a**) Myo1p co-localizes with a single septin ring in a small-budded cell. (**b-c**) In cells bearing medium to large buds, the septin ring thickens and forms 2 rings that surround the single Myo1p ring. (**d**) After contractile ring closure and cell separation, mother and daughter cell each have a single septin ring. Scale bar = 0.3 μm.

Supplemental Figure 3: (A) *aim44* Δ yeast expressing a plasmid-borne septin, Cdc3p-mCherry (red), were grown to mid-log phase and imaged by fluorescence microscopy. Cdc3p-mCherry forms single and double rings in *aim44* Δ cells. (B) Myo1p is not required for localization of Aim44p to a ring at the bud neck. *myo1* Δ cells in which *AIM44* was tagged with GFP at its chromosomal locus were grown in SC to mid-log phase and imaged by fluorescence microscopy. Deletion of *MYO1* did not prevent localization of Aim44p-GFP to the bud neck. Scale bar = 1 µm.

Supplemental Figure 4: The lower-mobility band containing Hof1p-13Myc is due to phosphorylation of the protein. Wild-type and *aim44* Δ cells expressing Hof1p tagged with 13Myc at its chromosomal locus were synchronized as for Figure 1. An aliquot was removed 90 min after release from G₁ arrest for extraction of protein. Half of the protein sample was left untreated and the other half was treated with 10 units of calf intestinal alkaline phosphatase (CIP) for 1 hr at 37°C. Western blots are shown in which Hof1p was detected using anti-Myc antibody and the load control, hexokinase, was detected using a polyclonal anti-hexokinase antibody. *, lower mobility band(s) of Hof1p-13Myc. The lower mobility Hof1p-13Myc band(s) is phosphorylated, as measured by sensitivity to CIP treatment.

Supplemental Figure 5: Deletion of *AIM44* inhibits cytokinesis but does not affect rates of cell cycle progression. Cell cycle progression was analyzed by quantifying spindle length every 15 min for 90 min after release from pheromone-induced arrest in G₁ phase in wild-type and *aim44* Δ cells expressing tubulin tagged at its C-terminus with GFP. Cells were fixed using paraformaldehyde and stained with the DNA-binding dye DAPI and imaged. These data are pooled from two independent experiments and measurements average n ≥ 100 cells per strain. Error bars represent standard error of the mean.

Supplemental Table 1: Yeast strains used in this study were constructed as described in the Methods section. All yeast strains were created in the *BY4741* wild-type background except for the temperature-sensitive septin strain (*YEF743 cdc12-6*) and the wild-type control (*YEF6866 CDC12*). The two *YEF* strains were kindly provided by Dr. Erfei Bi.

Strain	Genotype	Source
BY4741	MAT a his3∆1 leu2∆0 met15∆0 ura3∆0	Open Biosystems (Huntsville, AL)
7394	MAT a aim44∆::kanMX6 his3∆1 leu2∆0 met15∆0 ura3∆0	Open Biosystems
ISY007	MAT a cbk1∆:HIS3 leu2∆0 met15∆0 ura3∆0	(García-Rodríguez <i>et</i> <i>al</i> ., 2009)
THY158	MAT a MYO1-GFP::HIS3 leu2∆0 met15∆0 ura3∆0	(Huckaba <i>et al.</i> , 2006)
RHY045	MAT a his3∆1 myo1∆::LEU2 met15∆0 ura3∆0	(Higuchi <i>et al</i> ., 2013)
DAY001	<i>MATa aim44∆::kanMX6 his3∆1 leu2∆0 met15∆0 ura3∆0</i> [pYIP128-CDC3-mCherry::LEU2]	This study
DAY002	MAT a MYO1-GFP:HIS3 aim44∆::kanMX6 leu2∆0 met15∆0 ura3∆0	This study
DAY003	MAT a AIM44-GFP::HIS3 leu2∆0 met15∆0 ura3∆0	This study
DAY004	<i>MATa AIM44-GFP::HIS3 leu2∆0 met15∆0 ura3∆0</i> [pYIP128-CDC3-mCherry::LEU2]	This study
DAY005	MAT a AIM44-GFP::HIS3 myo1∆::LEU2 met15∆0 ura3∆0	This study
DAY006	MAT a AIM44-GFP::HIS3 MYO1-mCherry::hphMX4 leu2∆0 met15∆0 ura3∆0	This study
DAY007	MAT a MYO1-GFP::HIS3 CDC3-mCherry::hphMX4 leu2∆0 met15∆0 ura3∆0	This study
DAY008	MAT a HOF1-13Myc::HIS3 leu2∆0 met15∆0 ura3∆0	This study
DAY009	MAT a HOF1-13Myc::HIS3 aim44∆::LEU2 met15∆0 ura3∆0	This study
LPY057-1	MAT a HOF1-GFP::HIS3 leu2Δ0 met15Δ0 ura3Δ0	This study
DAY010	MAT a aim44Δ::KANMX6 HOF1-GFP::HIS3 met15Δ0 ura3Δ0	This study
DAY011	CDC12+ AIM44-GFP::KanMX6 ura3 leu2∆0 HIS3+ TRP1+ [pYIP128-CDC3-mCherry::LEU2]	This study

Strain	Genotype	Source
DAY012	<i>cdc12-6 AIM44-GFP::KanMX6 ura3 leu2∆0 HIS3+ TRP1+</i> [pYIP128-CDC3-mCherry::LEU2]	This study
DAY013	MAT a his3∆1 TUB1-GFP::LEU2 met15∆0 ura3∆0	This study
DAY014	MAT a aim44∆::kanMX6 his3∆1 TUB1-GFP::LEU2 met15∆0 ura3∆0	This study
DAY015	<i>MATa his3∆1 leu2∆0 met15∆0 ura3∆0 [</i> pGB1805- <i>DBF2::URA3]</i>	This study
DAY016	<i>MATa aim44∆::kanMX6 his3∆1 leu2∆0 met15∆0 ura3∆0</i> [pGB1805- <i>DBF2::URA3</i>]	This study
DAY017	MAT a AIM44-3HA∷kanMX6 his3∆1 leu2∆0 met15∆0 ura3∆0	This study
DAY018	MAT a AIM44-3HA::kanMX6 HOF1-13Myc::HIS3 leu2∆0 met15∆0 ura3∆0	This study



Cdc3p-mCherry Myo1p-GFP

Merge





WT 90 min after release from arrest CIP Hof1p-13Myc

Hexokinase



