

# Supporting Information

## Expedient Synthesis of Highly Potent Antagonists of Inhibitor of Apoptosis Protein (IAPs) with Unique Selectivity for ML-IAP

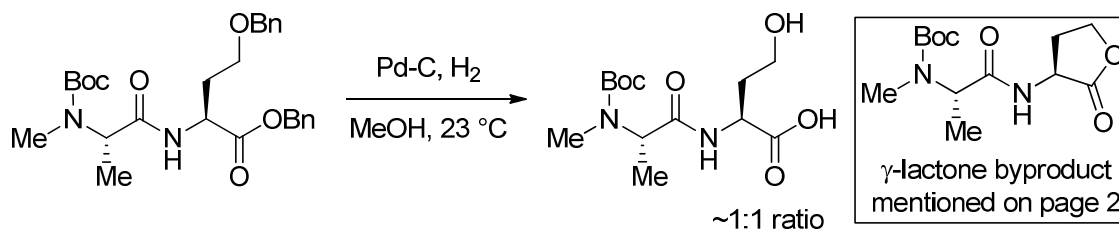
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**Figure SI-1:  $\gamma$ -Lactone Byproduct (mentioned on Page 2 in text)**



**Table SI-1: Fluorescence Polarization Competition Assay Data with Error Values**

Cmpd <sup>[a]</sup>	XIAP		clAP1	clAP2	ML-IAP
	BIR2 $K_i$ ( $\mu$ M)	BIR3 $K_i$ ( $\mu$ M)	BIR3 $K_i$ ( $\mu$ M)	BIR3 $K_i$ ( $\mu$ M)	$K_i$ ( $\mu$ M)
<b>1a</b>	>56.0	>39.1	8.98±0.67	28.90±2.03	>25
<b>1b</b>	>56.0	13.3±0.4	1.18±0.16	3.31±0.38	8.337 ± 1.059
<b>1c</b>	>56.0	6.87±0.28	1.05±0.13	2.70±0.14	5.070 ± 3.934
<b>1d</b>	51.0±8.5	>39.1	5.36±0.35	9.71±0.55	>7.5
<b>1e</b>	>56.0	28.3±6.3	1.49±0.09	3.81±0.10	>7.5
(S)- <b>10a</b>	>56.0	0.44±0.02	0.061±0.001	0.149±0.013	0.344±0.137
(S)- <b>10b</b>	17.2±2.0	9.23±0.82	0.411±0.032	0.799±0.064	4.708±0.048
(S)- <b>10c</b>	>56.0	0.61±0.02	0.130±0.001	0.260±0.027	0.043±0.036
<b>10d</b>	>56.0	0.930±0.139	0.034±0.009	0.056±0.012	0.233±0.134
(S)- <b>10e</b>	9.64±0.71	0.100±0.014	0.022±0.003	0.047±0.009	0.0021±0.00019
<b>10f</b>	15.62±2.52	0.270±0.011	0.085±0.008	0.116±0.026	0.0046±0.0038
(S)- <b>12</b>	>56.0	32.64±3.17	6.97±0.80	9.72±1.60	>25
(S)- <b>15</b>	>56.0	0.350±0.016	<0.010	0.071±0.008	0.294±0.121
(R)- <b>15</b>	>56.0	>39.1	12.72±0.63	27.26±1.56	ND
GDC-0152 <sup>[b]</sup>	29.8±4.6 (0.112)	0.258±0.015 (0.028)	0.102±0.012 (0.017)	0.144±0.014 (0.043)	0.041±0.006 (0.014)

[a] Compounds **1a-1e**, **10d** and **10f** are diastereomixtures; otherwise major stereoisomer tested is indicated.

[b] Measured values; reported values in parentheses, see structure above (Scheme 1A).

### XIAP Fluorescence Polarization Assay Conditions

Binding of compounds to BIR1/2 of XIAP or BIR3 of XIAP was determined by fluorescence polarization. Assay buffer was 25 mM Hepes at pH 7.5/1 mM TCEP (Tris(2-carboxyethyl)phosphine hydrochloride)/0.005% Tween 20/ 20 nM AVPIAQK-rhodamine. BIR1/2 was present at 1  $\mu$ M while BIR3 was present at 200 nM. Compound ranged from 100  $\mu$ M to 0.0061  $\mu$ M using 2 fold dilutions from the highest concentration. Assays were run in 384 well black plates read in an Analyst in fluorescence polarization mode with excitation at 530 nm, emission at 580 nm and a dichroic mirror at 565 nm. Data was fit to a nonlinear regression curve in Prism to determine the  $IC_{50}$  values of the compounds.  $K_i$ 's were then calculated from the  $IC_{50}$  values.<sup>1</sup>

### clAP1 and clAP2 Fluorescence Polarization Assay Conditions

Binding assays with clAP1 BIR3 or clAP2 BIR3 were done with Hepes at 25 mM, Tween 20 at 0.005% and AVPIAQK-rhodamine at 20 nM. clAP1 BIR3 was at 0.095  $\mu$ M while clAP2 BIR3 was at 0.123  $\mu$ M. Compound

ranged from 100 to 0.0061  $\mu\text{M}$  by 2 fold dilutions. Assays were run in 384 well black plates read in an Analyst in fluorescence polarization mode with excitation at 530 nm, emission at 580 nm and a dichroic mirror at 565 nm. Data was fit to a nonlinear regression curve in Prism to determine the  $\text{IC}_{50}$  values of the compounds.  $K_i$ 's were then calculated from the  $\text{IC}_{50}$  values.

#### **ML-IAP Fluorescence Polarization Assay Conditions**

Binding of compounds to ML-IAP was determined by fluorescence polarization. Assay buffer was 25 mM Hepes at pH 7.5/1 mM TCEP (Tris(2-carboxyethyl)phosphine hydrochloride)/0.005% Tween 20/ 20 nM AVPIAQK-rhodamine. ML-IAP was present at 20 nM. Compound ranged from 200  $\mu\text{M}$  to 0.00056  $\mu\text{M}$  using 3 fold dilutions from the highest concentration. Assays were run in 384 well black plates read in a BMG POLARstar in fluorescence polarization mode with excitation at 544 nm and emission at 590 nm. Data was fit to a nonlinear regression curve in Prism to determine the  $\text{IC}_{50}$  values of the compounds.  $K_i$ 's were then calculated from the  $\text{IC}_{50}$  values.

#### **Caspase Derepression and Competition Fluorescence Polarization Assay Conditions**

Purified recombinant human caspases 3 [0.1 nM], -7 [1 nM] or -9 [2.2  $\mu\text{M}$ ] were used in combination with XIAP BIR2 [1 nM] and BIR3 [2.1  $\mu\text{M}$ ]. During the derepression assay, caspases and the BIRs were allowed to form a complex for 30 minutes before the addition of increasing concentrations of the compound **10e**. For the competition assay, increasing concentrations of compound **10e** were added at the same time as the enzyme and inhibitor. The ratio of inhibited to uninhibited caspase activity ( $V_i/V_o$ ) was obtained by monitoring the release of fluorescence from the fluorogenic substrates Ac-DEVD-afc for caspases 3 and -7 or Ac-LEHD-afc for caspase 9.

#### **Caspase 3/7 Activity Assay**

Caspase 3/7 activity assays in MDA-MB-231 cells pretreated with vehicle (0.1% DMSO) or 5  $\mu\text{M}$  of **10e** or **10f** before treatment with 0 or 100 ng/mL TRAIL for 4 h. Activity is normalized to that of vehicle +TRAIL values (in the cases where TRAIL was used) or vehicle -TRAIL (in the cases where TRAIL was not used) and all assays are carried out in triplicate as least twice. Caspase 3/7 activity was assessed utilizing CaspaseGlo® 3/7 Assay (Promega Corp., Madison, WI). Cells are seeded as for CellTiterGlo and treated as described. The assays were carried out exactly as per manufacturer's instructions before being read on a FlexStation3 plate reader utilizing SoftTek software.

#### **TNF ELISA Assay Conditions**

MDA-MB-231 and SKVO3 cells were cultured as described below before treatment with vehicle (0.1% DMSO) or 5  $\mu\text{M}$  of **10e** or **10f**. 1 mL of media was removed after 6 h (6 h data not shown) and 24h and cellular debris pelleted at 16 k g for 2 min. 100  $\mu\text{L}$  of supernatant was removed per sample in duplicate and TNF levels were assessed using a BD OptEIA™ Human TNF ELISA Kit II (BD Biosciences, San Jose, CA) exactly as per

manufacturer's instructions. The data was processed and graphed in excel and is expressed as averages +/- s.e.m. Error values are present, but small, thus making the error bars difficult to see.

### **Western Blot Analysis**

Immunoblot analysis of cIAP1/2 and XIAP expression in MDA-MB-231 cells treated with vehicle (0.1% DMSO) or 5  $\mu$ M of **10e** or **10f** for 24 h. Tot. Erk 1/2 and  $\beta$ -actin are shown as equal loading controls. Antibodies used are anti-pan-cIAP1/2 (Clone 315301, 1:1000, R&D Systems, Inc., Minneapolis, MN); anti-XIAP (#2042, 1:2000) and anti-Total Erk 1/2 (#9102, 1:5000) (both from Cell Signaling Technologies Inc. Beverly, MA); or anti- $\beta$ -Actin (A5441, 1:10000, Sigma-Aldrich, St. Louis, MO).

### **MDA-MB-231 Cell Viability Assay Conditions**

Five thousand cells per well were plated in a 96 well plate and incubated overnight at 37 °C / 5% CO<sub>2</sub>. The following day, compounds in 2 fold dilutions were added to the plates and the plates returned to the incubator for 4 hrs. TRAIL at a final concentration of 5 ng/mL was then added to half the plate while an equivalent volume of culture media was added to the other half of the plate as a control. Twenty-three hrs later, plates were removed to the bench and 25  $\mu$ L CellTiter Glo was added to each well and the plates were read for luminescence on a Luminoskan Ascent. Data was fit in Prism to a nonlinear regression curve to determine the LD<sub>50</sub> of the compounds.

### **MDA-MB-231 Cell Viability Assay Conditions (TRAIL Dose-Response Assay)**

Dose response cell viability assay of MDA-MB-231 cells cultured in the presence of vehicle (0.1% DMSO) or 5  $\mu$ M of **10e** or **10f** for 4 h before addition of a dose of TRAIL for a further 20 h as described. Viability is assessed by ATP content using CellTiter Glo® (Promega Corp., Madison, WI) as per manufacturer's instructions. Values are normalized to that of vehicle/ drug alone and all experiments are carried out in at least triplicate at least three times. Error values are small and thus the error bars on the graph are difficult to discern, yet still present.

### **SKOV3 Cell Viability Assay Conditions**

One thousand seven hundred cells per well were plated in a 384 well plate and incubated overnight at 37 °C / 5% CO<sub>2</sub>. The following day, compounds in 10 fold dilutions were added to the plates and the plates returned to the incubator for 4 hrs. TRAIL at a final concentration of 100 ng/mL was then added to half the plate while an equivalent volume of culture media was added to the other half of the plate as a control. Twenty-three hrs later, plates were removed to the bench and 20  $\mu$ L ATPlite was added to each well and the plates were read for luminescence on a BMG Labtech POLARstar. Data was fit in Prism to a nonlinear regression curve to determine the LD<sub>50</sub> of the compounds.

### **HFF Cell Viability Assay Conditions**

Normal human fibroblasts were maintained in DMEM with 10% (v/v) FBS, and penicillin/streptomycin/L-Glutamine (Omega Scientific Inc, Tarzana, CA). 5000 cells/well of a 96-well dish in 50  $\mu$ L complete medium were seeded

and allowed to attach overnight. 40  $\mu$ L of fresh media containing the specified drug at the concentrations described is added before re-incubation of the cells at 37 °C for 4 h. TRAIL to a final concentration of 100 ng/mL is added and the cells are again incubated at 37 °C for 20 h. Plates are removed to room temperature before addition of one half volume of freshly prepared CellTiterGlo reagent and luminescence is read on a Biotek Synergy 2 plate reader.

### **Molecular Modelling Specifications**

The *ChemSketch* software (ACD Labs) was used to generate a 2D chemical structure of the ligand that was subsequently converted to 3D coordinate set using the Dictionary interface in the *MIFit* software (<http://code.google.com/p/mifit/>).

Modeling the protein:ligand complex was performed by generating a chain of low energy conformers from this starting model and overlaying selected common atoms onto the structurally related compound, (3S,6S,7R,9aS)-6-(((2S)-2-aminobutanoyl)amino)-7-(2-aminoethyl)-N-(diphenylmethyl)-5-oxooctahydro-1H-pyrrolo[1,2-a]azepine-3-carboxamide, from the protein crystal structure of the cIAP1-BIR3 domain (entity SMK in PDB entry 3MUP).

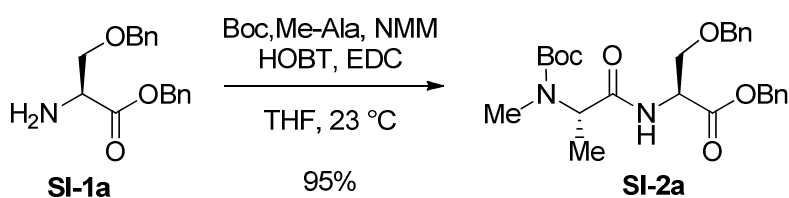
Low energy ligand conformers were generated using programs in the OpenBabel suite<sup>2</sup> by rotating atomic groups about randomly selected torsion angles with subsequent energy minimization. Target atoms for overlaying low energy conformers onto the docked pose of the known protein:ligand complex included the carbonyl and adjoining atoms in the central conjugated ring system. For each trial pose, the docked ligand was evaluated for the formation of acceptable non-bonded interactions with the protein binding site. Minor steric conflicts were relieved by relaxation of the conformation of the bound ligand. In parallel calculations the extent of the volume overlap with the known ligand was also assessed. All docking calculations and bound ligand evaluations were managed via the SDsearch interface.

Several docking runs of 1000 conformers each were performed in order to assess the available space in the protein binding site and the potential diversity of docking solutions. No solutions were obtained with ring puckers different from the related reference structure. When poses were scored so as to maximize volume overlap with the conformation of the known ligand the extent of the docked conformational ensemble was somewhat reduced and all of the allowed docking solutions were tightly clustered into two marginally distinct subsets.

Overall, these docking studies provided a relatively unique prediction for the protein:ligand complex in which the bound ligand adopts a low-energy conformation, makes plausible non-bonded interactions with the protein site and is consistent with previously determined cocrystal structures.

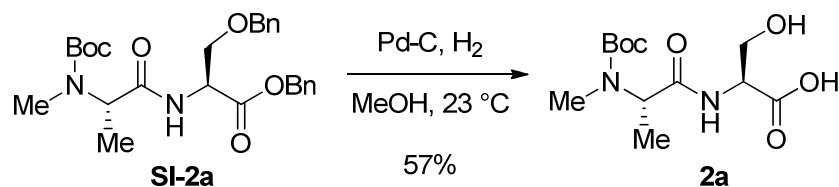
## Experimental Procedures

**General.** All solvents were used as purchased from commercial sources or dried over 4Å molecular sieves prior to use in the case of moisture sensitive reactions. Reactions conducted under microwave irradiation were performed in a CEM Discover microwave reactor using either CEM 10 mL reaction vessels or a ChemGlass heavy wall pressure vessel (100 mL, 38 mm x 190 mm). Reaction progress was monitored by reverse-phase HPLC and/or thin-layer chromatography (TLC). High resolution mass spectrometry was performed using ESI-TOFMS, EI-MS (reference: perfluorokerosene) and APCI-MS. TLC was performed using silica gel 60 F254 pre-coated plates (0.25 mm). Flash chromatography was performed using silica gel (32-63 μm particle size) or aluminum oxide (activated, basic, ~150 mesh size). All products were purified to homogeneity by TLC analysis (single spot, unless stated otherwise), using a UV lamp and/or iodine and/or CAM or basic KMnO<sub>4</sub> for detection purposes. NMR spectra were recorded on 400 MHz and 500 MHz spectrometers at ambient temperature. <sup>1</sup>H and <sup>13</sup>C NMR chemical shifts are reported as δ using residual solvent as an internal standard; CDCl<sub>3</sub>: 7.26, 77.16 ppm; CD<sub>3</sub>OD: 3.31, 49.00 ppm; DMSO-d<sub>6</sub>: 2.50, 39.52 ppm, CD<sub>3</sub>CN: 1.94 (<sup>1</sup>H), 1.32 (<sup>13</sup>C) ppm. Abbreviations used: alanine (Ala), *t*-butoxycarbonyl (Boc), 1-hydroxybenzotriazole (HOBT), *N*-methylmorpholine (NMM), *N*-(3-dimethylaminopropyl)-*N'*-ethylcarbodiimide (EDC), palladium on carbon (Pd-C), dichloromethane (DCM), diethyl ether (Et<sub>2</sub>O), ethyl acetate (EtOAc), 2,2,2-trifluoroethanol (TFE), methanol (MeOH), homoserine (HSer), homocysteine (HCys), triphenylmethyl or trityl (Trt), trifluoroacetic acid (TFA). Isocyanide **4e** was synthesized by the previously established route.<sup>3</sup>

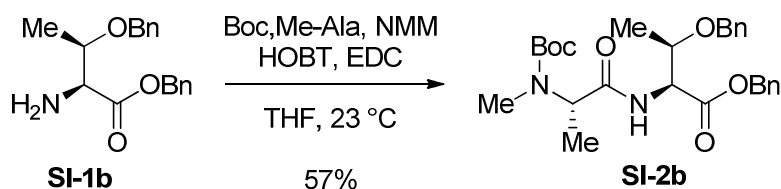


### (S)-Benzyl 3-(benzyloxy)-2-((S)-2-((*tert*-butoxycarbonyl)(methyl)amino)propanamido)propanoate (**SI-2a**).

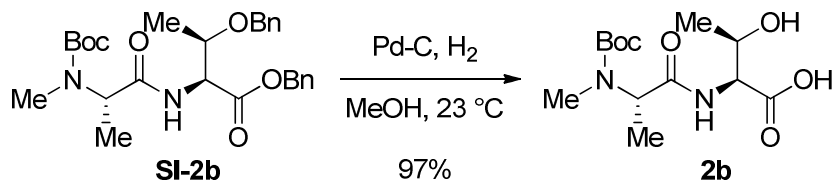
To a solution of **SI-1a** (1.74 g, 3.80 mmol, 1.0 equiv), Boc-*N*-Me-Ala-OH (773 mg, 3.80 mmol, 1.0 equiv), HOBT·xH<sub>2</sub>O (641 mg, 4.18 mmol, 1.1 equiv) and NMM (1.25 mL, 11.4 mmol, 3 equiv) in THF (45 mL) at 0 °C was added EDC·HCl (766 mg, 3.99 mmol, 1.05 equiv). After 30 min the cold bath was removed. The solution stirred for 14 h and then was quenched with saturated aqueous NaHCO<sub>3</sub> (50 mL), extracted with EtOAc (2 x 40 mL), dried over sodium sulfate and then concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (5:1→4:1→3:1 hexanes/EtOAc) to yield **SI-2a** (1.70 g, 95%). *R<sub>f</sub>* = 0.20 (5:1 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 7.34-7.27 (m, 8H), 7.19 (dd, 2H, *J* = 2.0, 8.0 Hz), 5.18 (q, 2H, *J* = 12.0 Hz), 4.79-4.74 (m, 1H), 4.45 (q, 2H, *J* = 12.0 Hz), 3.89 (dd, 1H, *J* = 3.2, 9.6 Hz), 3.66 (dd, 1H, *J* = 3.2, 9.6 Hz), 2.75 (s, 3H), 1.45 (s, 9H), 1.34 (t, 3H, *J* = 7.2 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 171.6, 170.0, 137.5, 135.4, 128.7, 128.5, 128.5, 128.3, 127.9, 127.7, 73.4, 69.8, 67.4, 52.9, 30.0, 28.4, 13.9; HRMS calcd for C<sub>26</sub>H<sub>34</sub>N<sub>2</sub>O<sub>6</sub>Na: 493.23091, found 493.23211.



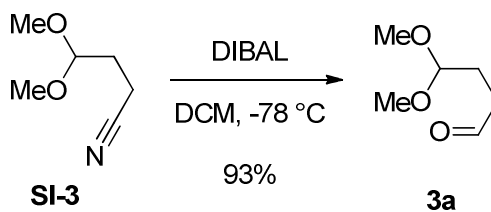
**(S)-2-((S)-2-((Tert-butoxycarbonyl)(methyl)amino)propanamido)-3-hydroxypropanoic acid (2a).** To a solution of **SI-2a** (1.70 g, 3.61 mmol, 1.0 equiv) in methanol (25 mL) was added 10 wt% Pd-C (100 mg). A balloon of H<sub>2</sub> was applied for 16 h, then the mixture was filtered through Celite with DCM and concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (1:1 hexanes/EtOAc→100% DCM→5% MeOH/DCM) to yield **2a** (591 mg, 57%). *R<sub>f</sub>* = 0.12 (7% MeOH/DCM). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ: 4.41 (t, 1H, *J* = 3.6 Hz), 3.91 (dd, 1H, *J* = 4.4, 10.8 Hz), 3.83 (dd, 1H, *J* = 4.0, 11.2 Hz), 3.35-3.34 (m, 1H), 2.86 (s, 3H), 1.47 (s, 9H), 1.38 (d, 3H, *J* = 6.8 Hz); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>CN) δ: 207.9, 173.1, 172.4, 81.0, 62.6, 55.4, 30.9, 28.5. HRMS calcd for C<sub>12</sub>H<sub>22</sub>N<sub>2</sub>O<sub>6</sub>Na: 313.1370, found 313.1371.



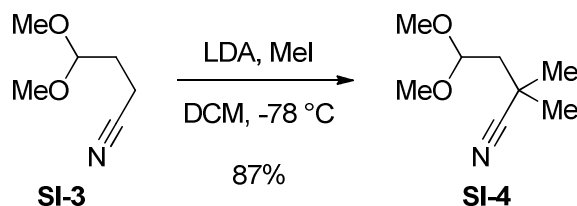
**(2S,3R)-Benzyl 3-(benzyloxy)-2-((S)-2-((tert-butoxycarbonyl)(methyl)amino)propanamido)butanoate (SI-2b).** Same procedure as above (**SI-2a**) using **SI-1b** (4.65 g, 11.9 mmol, 1.0 equiv), Boc-*N*-Me-Ala-OH (2.43 g, 11.9 mmol, 1.0 equiv), HOBT·xH<sub>2</sub>O (2.19 g, 14.3 mmol, 1.1 equiv), NMM (3.94 mL, 35.8 mmol, 3 equiv) and EDC·HCl (2.75 g, 14.3 mmol, 1.05 equiv) in THF (100 mL). The resultant oil was purified by flash chromatography on silica gel (5:1→4:1→2:1 hexanes/EtOAc) to yield **SI-2b** (3.32 g, 57%). *R<sub>f</sub>* = 0.26 (5:1 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 7.31-7.25 (m, 8H), 7.17-7.15 (m, 2H), 5.14 (d, 1H, *J* = 6.0 Hz), 5.06 (d, 1H, *J* = 6.0 Hz), 4.67 (dd, 1H, *J* = 2.4, 9.2 Hz), 4.48 (d, 1H, *J* = 12.0 Hz), 4.27 (d, 1H, *J* = 12.0 Hz), 4.15 (qd, 1H, *J* = 2.0, 6.0 Hz), 2.79 (s, 3H), 1.60 (s, 1H), 1.42 (s, 9H), 1.35 (d, 3H, *J* = 7.2 Hz), 1.16 (d, 3H, 6.4 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 172.2, 170.4, 135.5, 128.7, 128.7, 128.5, 128.5, 128.5, 128.4, 127.8, 127.8, 74.3, 70.9, 67.3, 56.8, 28.4, 16.4. HRMS calcd for C<sub>27</sub>H<sub>36</sub>N<sub>2</sub>O<sub>6</sub>Na: 507.2466, found 507.2468.



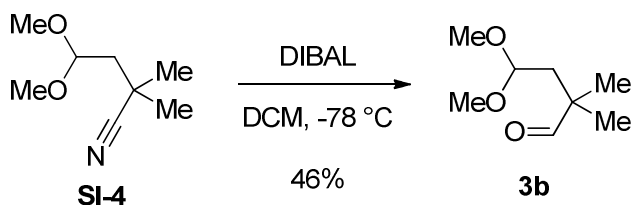
**(2S,3R)-2-((S)-2-((Tert-butoxycarbonyl)(methyl)amino)propanamido)-3-hydroxybutanoic acid (2b).** Same procedure as above (**2a**) using **SI-2b** (3.306 g, 6.82 mmol, 1.0 equiv) and 10 wt% Pd-C (150 mg) in methanol (50 mL). The resultant oil was sufficiently pure as a crude product, **2b** (2.01 g, 97%). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ: 7.44 (bs, 1H), 4.70 (bs, 1H), 4.40-4.36 (m, 1H), 4.33 (dd, 1H, *J* = 2.8, 6.4 Hz), 2.87 (s, 3H), 1.48 (s, 9H), 1.39 (d, 3H, *J* = 7.2 Hz), 1.18 (d, 3H, *J* = 6.4 Hz); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD) δ: 174.7, 173.7, 157.5, 81.9, 68.2, 59.0, 55.7, 30.9, 28.6, 20.7, 14.9. HRMS calcd for C<sub>13</sub>H<sub>24</sub>N<sub>2</sub>O<sub>6</sub>Na: 327.15266, found 327.15236.



**4,4-Dimethoxybutanal (3a).** To a solution of **SI-3** (1.2 g, 9.29 mmol, 1.0 equiv) in DCM (75 mL) at -78 °C under N<sub>2</sub> was added 1.1 M DIBAL in cyclohexane (23.23 mL, 10.2 mmol, 1.1 equiv). After 3 h at -78 °C, the mixture was slowly warmed to r.t. and quenched with sat. aq. NH<sub>4</sub>Cl (25 mL) and Rochelle salt (25 mL). Reaction progress was monitored by TLC (vanillin stain). After stirring for 1 h, the mixture was extracted with DCM (3 x 20 mL). The organics were then washed with brine (30 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo to yield a colorless, relatively volatile liquid **3a** (1.14 g, 93%) which was sufficiently pure to use without further purification. The analytical data match those previously reported.<sup>4</sup>

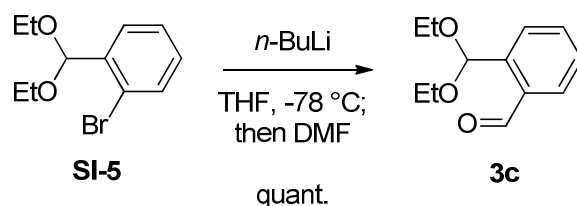


**4,4-Dimethoxy-2,2-dimethylbutanenitrile (SI-4).** To a solution of diisopropylamine (4.77 mL, 34.1 mmol, 2.2 equiv) in THF (50 mL) at -10 °C under N<sub>2</sub> was added 1.5 M *n*-BuLi in hexanes (22.7 mL, 34.1 mmol, 2.2 equiv). After 30 min the mixture was cooled to -78 °C and a solution of **SI-3** (2.0 g, 15.5 mmol, 1.0 equiv) in THF (10 mL) was added. After 1 h iodomethane (2.12 mL, 34.1 mmol, 2.2 equiv) was added. The mixture was slowly warmed to 0 °C and kept there for 14 h, at which time it was quenched with sat. aq. NH<sub>4</sub>Cl (40 mL) and extracted with EtOAc (3 x 20 mL). The organics were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (5:1→3:1 hexanes/EtOAc) to yield **SI-4** (2.105 g, 87%) as a yellow oil. R<sub>f</sub> = 0.49 (3:1 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 4.60 (t, 1H, J = 5.6 Hz), 3.37 (s, 6H), 1.83 (d, 2H, J = 4.4 Hz), 1.39 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 124.7, 102.4, 53.3, 43.0, 30.0, 27.5

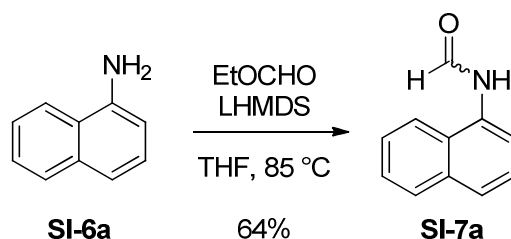


**4,4-Dimethoxy-2,2-dimethylbutanal (3b).** Same procedure as above (**3a**) using **SI-4** (500 mg, 3.18 mmol, 1.0 equiv) in DCM (25 mL) and 1.1 M DIBAL in cyclohexane (3.18 mL, 10.2 mmol, 1.1 equiv). The resultant oil was purified by flash chromatography on silica gel (DCM) to yield **3b** (232 mg, 46%) as a colorless, relatively volatile oil. Yield is highly variable and dependent upon extent of drying as the compound is fairly volatile. R<sub>f</sub> = 0.39 (7:1 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 9.39 (s, 1H), 4.36 (t, 1H, J = 6.0 Hz), 3.30 (s, 6H), 1.85 (d, 2H, J = 5.6 Hz), 1.07 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 204.7, 102.7, 53.7, 43.9, 41.6, 21.9.

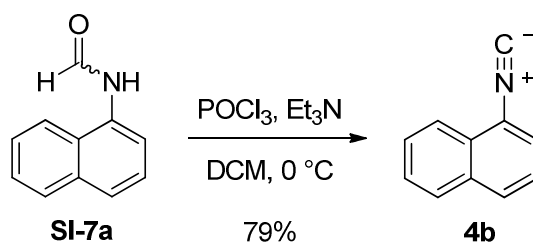




**2-(Diethoxymethyl)benzaldehyde (3c).** To a solution of **SI-5** (1.94 g, 7.49 mmol, 1.0 equiv) in THF (20 mL) at -78 °C under N<sub>2</sub> was added 1.5 M *n*-BuLi in hexanes (7.49 mL, 11.2 mmol, 1.5 equiv). After 30 min DMF (869 μL, 11.2 mmol, 1.5 equiv) was added. The mixture was slowly warmed to r.t. over 4 h, at which time it was quenched with sat. aq. NH<sub>4</sub>Cl (40 mL) and extracted with EtOAc (3 x 20 mL). The organics were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (95:4:1 hexanes/EtOAc/Et<sub>3</sub>N) to yield **3c** (1.34 g, quantitative) as a yellow oil. R<sub>f</sub> = 0.46 (3:1 hexanes/EtOAc). The analytical data match those previously reported.<sup>5</sup>

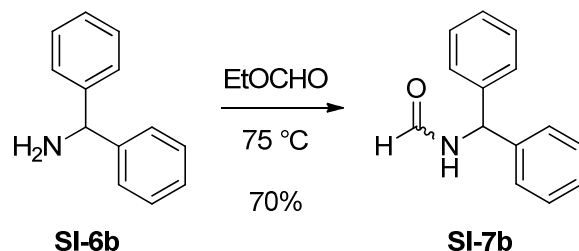


**N-(Naphthalen-1-yl)formamide (SI-7a).** To a mixture of 1-naphthylamine (**SI-6a**, 6.0 g, 41.9 mmol, 1.0 equiv) and ethyl formate (6.74 mL, 83.8 mmol, 2 equiv) in THF (200 mL) was added 1 M LHMDs in THF (75.4 mL, 75.4 mmol, 1.8 equiv). The mixture was heated to 85 °C for 14 h and then concentrated. The resulting solid was filtered and rinsed with hexanes to yield the first batch of **SI-7a**. The filtrate was concentrated and the filtration procedure was repeated for a second batch of product to finally yield **SI-7a** (3.05 g, 64%) as a brown solid and a 2:1 mixture of rotational isomers. R<sub>f</sub> = 0.10 (5:1 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 8.65-8.61 (m, 2H), 8.45 (bs, 1H), 8.04-7.99 (m, 2H), 7.92-7.85 (m, 2H), 7.80 (d, 1H, *J* = 8.4 Hz), 7.73 (d, 1H, *J* = 8.0 Hz), 7.63-7.51 (m, 3H), 7.50-7.44 (m, 2H), 7.32 (d, 1H, *J* = 7.6 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 164.1, 159.7, 134.4, 134.2, 132.2, 131.1, 129.0, 128.7, 127.9, 127.2, 127.2, 127.2, 127.0, 126.7, 126.4, 126.3, 125.9, 125.7, 121.4, 121.0, 120.5, 119.3. HRMS calcd for C<sub>11</sub>H<sub>9</sub>NO: 171.0679, found 171.0681.

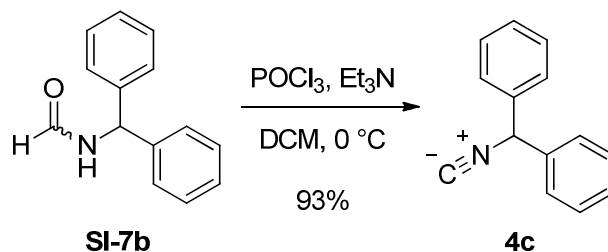


**1-Isocyanonaphthalene (4b).** To a solution of **SI-7a** (1.048 g, 6.12 mmol, 1.0 equiv) in DCM (20 mL) at 0 °C was added Et<sub>3</sub>N (4.33 mL, 31.2 mmol, 5.1 equiv) followed by phosphorus oxychloride (841 μL, 9.18 mmol, 1.5 equiv). The mixture was warmed to 23 °C and stirred for 2 h, at which time it was poured into a mixture of saturated NaHCO<sub>3</sub> (40 mL) and 1 M NaOH (20 mL) and extracted with DCM (3 x 20 mL). The organics were dried over

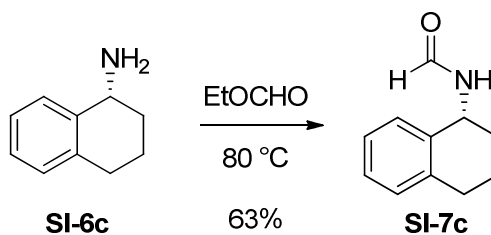
Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (1:1 hexanes/DCM) to yield **4b** (740 mg, 79%) as a brown oil which was stored at 0 °C. *R<sub>f</sub>* = 0.72 (3:1 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 8.19 (d, 1H, *J* = 8.4 Hz), 7.90 (d, 2H, *J* = 8.0 Hz), 7.68 (t, 1H, *J* = 7.6 Hz), 7.61 (t, 2H, *J* = 7.2 Hz), 7.45 (td, 1H, *J* = 2.4, 8.4 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 167.3, 133.7, 129.9, 128.5, 128.2, 128.1, 127.6, 125.1, 124.7, 123.1. HRMS calcd for C<sub>11</sub>H<sub>8</sub>N: 154.06513, found 154.06671.



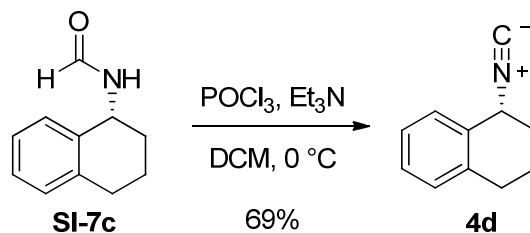
**N-Benzhydrylformamide (SI-7b).** A mixture of benzhydrylamine (**SI-6b**, 4.0 g, 21.8 mmol, 1.0 equiv) and ethyl formate (2.0 mL, 24.9 mmol, 1.14 equiv) was heated to 75 °C for 14 h. EtOAc was added and the mixture was triturated by sonication, then filtered and rinsed with Et<sub>2</sub>O to yield **SI-7b** (3.24g, 70%) as a white solid. The compound exists as a mixture of rotational isomers. *R<sub>f</sub>* = 0.29 (3:1 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 8.15 (s, 1H), 7.34-7.19 (m, 10H), 6.69 (d, 1H, *J* = 6.0 Hz), 6.27 (d, 1H, *J* = 8.4 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 160.4, 141.0, 128.8, 127.7, 127.5, 55.7. HRMS calcd for C<sub>14</sub>H<sub>14</sub>NO: 212.10699, found 212.100748.



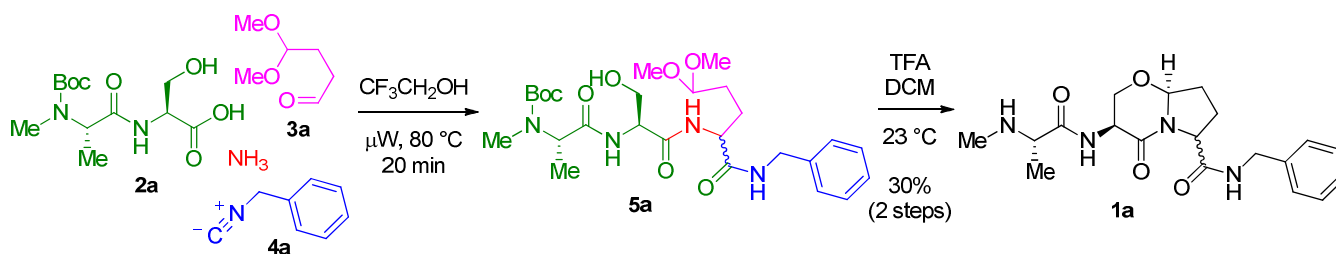
**(Isocyanomethylene)dibenzene (4c).** To a solution of **SI-7b** (1.727 g, 8.17 mmol, 1.0 equiv) in DCM (35 mL) at 0 °C was added Et<sub>3</sub>N (5.79 mL, 41.7 mmol, 5.1 equiv) followed by phosphorus oxychloride (1.12 mL, 12.3 mmol, 1.5 equiv). The mixture was warmed to 23 °C and stirred for 18 h, at which time it was poured into a mixture of saturated NaHCO<sub>3</sub> (50 mL) and 1 M NaOH (20 mL) and extracted with DCM (3 x 30 mL). The organics were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (DCM→5:1 DCM/EtOAc) to yield **4c** (1.467 g, 93%) as an orange solid which was stored at 0 °C. *R<sub>f</sub>* = 0.73 (7:1 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 7.41-7.33 (m, 10H), 5.92 (s, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 158.5, 137.7, 129.1, 128.6, 126.7, 77.2, 62.1. HRMS calcd for C<sub>14</sub>H<sub>11</sub>NNa: 216.07837, found 216.07971.



**(R)-N-(1,2,3,4-Tetrahydronaphthalen-1-yl)formamide (SI-7c).** A mixture of (*R*)-(-)-1,2,3,4-tetrahydro-1-naphthylamine (**SI-6c**, 10.0 g, 67.9 mmol, 1 equiv) and ethyl formate (6.23 mL, 77.4 mmol, 1.14 equiv) was heated to 80 °C for 14 h. Hexanes was added and the mixture was triturated by sonication, then filtered and rinsed with hexanes to yield **SI-7c** (7.44 g, 63%) as a tan solid.  $R_f = 0.22$  (3:1 hexanes/EtOAc).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$ : 8.23 (s, 1H), 7.29-7.25 (m, 1H), 7.23-7.16 (m, 2H), 7.13-7.08 (m, 1H), 5.82 (bs, 1H), 5.28 (dd, 1H,  $J = 5.2, 14.0$  Hz), 2.85-2.73 (m, 2H), 2.15-2.03 (m, 1H), 1.88-1.81 (m, 3H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 160.5, 137.7, 136.1, 129.4, 128.8, 127.6, 126.5, 46.4, 30.3, 29.3, 20.0. HRMS calcd for  $\text{C}_{11}\text{H}_{13}\text{NONa}$ : 198.0889, found 198.0890.

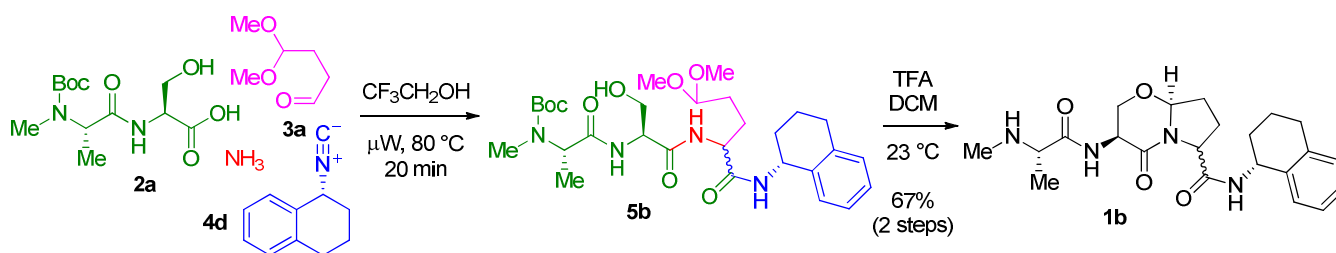


**(R)-1-Isocyano-1,2,3,4-tetrahydronaphthalene (4d).** To a solution of **SI-7c** (2.85 g, 16.3 mmol, 1.0 equiv) in DCM (40 mL) at 0 °C was added  $\text{Et}_3\text{N}$  (11.51 mL, 82.9 mmol, 5.1 equiv) followed by phosphorus oxychloride (2.23 mL, 24.4 mmol, 1.5 equiv). The mixture was warmed to 23 °C and stirred for 2 h, at which time it was poured into saturated  $\text{NaHCO}_3$  (200 mL) and extracted with DCM (2 x 100 mL). The organics were dried over  $\text{Na}_2\text{SO}_4$ , filtered and concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (3:1→1:1 hexanes/DCM) to yield **4d** (1.76 g, 69%) as a brown oil which was stored at 0 °C.  $R_f = 0.59$  (5:1 hexanes/EtOAc).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.45-7.43 (m, 1H), 7.26-7.23 (m, 2H), 7.14-7.11 (m, 1H), 4.83 (app. s, 1H), 2.92-2.84 (m, 1H), 2.80-2.72 (m, 1H), 2.18-2.12 (m, 2H), 2.11-2.01 (m, 1H), 1.87-1.78 (m, 1H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 155.2, 136.5, 132.1, 129.5, 128.6, 128.6, 126.7, 52.6, 30.7, 28.6, 19.4. HRMS calcd for  $\text{C}_{11}\text{H}_{12}\text{N}$ : 158.0964, found 158.0966.

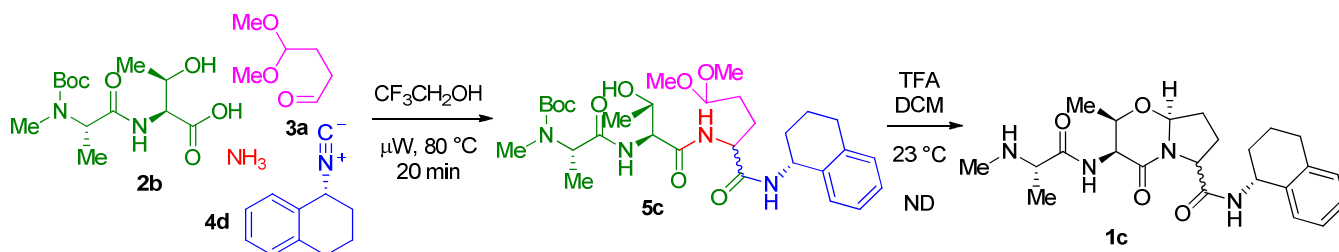


**(3S,8aS)-N-Benzyl-3-((S)-2-(methylamino)propanamido)-4-oxohexahydro-2H-pyrrolo[2,1-b][1,3]oxazine-6-carboxamide (1a).** A mixture of carboxylic acid **2a** (93 mg, 0.320 mmol, 1.0 equiv), aldehyde **3a** (44 mg, 0.336 mmol, 1.05 equiv), benzyl isocyanide (**4a**) (38 mg, 0.320 mmol, 1.0 equiv) and 7 M ammonia in MeOH (92  $\mu\text{L}$ , 0.641 mmol, 2.0 equiv) in TFE (3 mL) was stirred under microwave irradiation at a set temperature of 80 °C for 20 min. The mixture was then transferred to a round bottom flask and concentrated in vacuo, then 1 M NaOH (15 mL) was added and the mixture was extracted with DCM (3 x 7 mL). The organics were dried over  $\text{Na}_2\text{SO}_4$ , filtered and concentrated in vacuo. The resultant oil **5a** was combined with TFA (147  $\mu\text{L}$ , 1.92 mmol, 6 equiv) in DCM (5 mL) and stirred at 23 °C for 14 h. The mixture was concentrated in vacuo and the product purified by

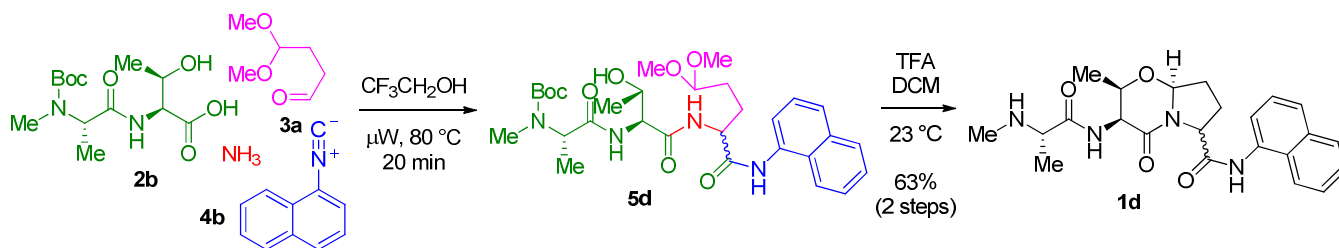
flash chromatography on basic alumina (3:1 hexanes/EtOAc→DCM→7% MeOH/DCM) to yield **1a** as a 1:1 diastereomixture of the the free base (36 mg, 30% over 2 steps). Some of the material was further purified by preparative scale HPLC for use in biological assays. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ: 8.51 (bs, 1H), 7.33-7.28 (m, 8H), 7.26-7.21 (m, 2H), 5.23 (t, 1H, *J* = 5.2 Hz), 5.16 (dd, 1H, *J* = 5.2, 8.4 Hz), 4.68 (dd, 1H, *J* = 3.2, 6.4 Hz), 4.61-4.56 (m, 2H), 4.48 (d, 1H, *J* = 15.2 Hz), 4.42-4.33 (m, 4H), 4.28 (dd, 1H, *J* = 6.4, 11.6 Hz), 4.24 (dd, 1H, *J* = 6.0, 11.6 Hz), 4.01 (dd, 1H, *J* = 3.2, 11.6 Hz), 3.92 (dd, 1H, *J* = 3.2, 11.6 Hz), 3.69 (q, 2H, *J* = 6.8 Hz), 2.61 (s, 3H), 2.60 (s, 3H), 2.41-2.29 (m, 2H), 2.26-2.16 (m, 2H), 1.94-1.82 (m, 2H), 1.49 (d, 3H, *J* = 7.2 Hz), 1.47 (d, 3H, *J* = 7.2 Hz); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD) δ: 173.6, 173.4, 167.9, 167.1, 139.7, 139.7, 129.5, 129.5, 128.4, 128.4, 128.2, 128.2, 91.1, 90.9, 71.7, 70.8, 60.7, 59.8, 58.8, 44.2, 44.0, 32.3, 32.2, 32.2, 31.2, 27.2, 26.7, 16.8, 16.7. HRMS calcd for C<sub>19</sub>H<sub>27</sub>N<sub>4</sub>O<sub>4</sub>: 375.2027, found 375.2028.



**(3*S*,8*aS*)-3-((*S*)-2-(Methylamino)propanamido)-4-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)hexahydro-2*H*-pyrrolo[2,1-*b*][1,3]oxazine-6-carboxamide (**1b**).** Same procedure as above (**1a**) with carboxylic acid **2a** (97 mg, 0.334 mmol, 1.0 equiv), aldehyde **3a** (46 mg, 0.351 mmol, 1.05 equiv), isocyanide **4d** (53 mg, 0.334 mmol, 1.0 equiv) and 7 M ammonia in MeOH (97 μL, 0.668 mmol, 2.0 equiv) in TFE (3 mL). The resultant oil **5b** was combined with TFA (177 μL, 1.55 mmol, 6 equiv) in DCM (5 mL) and stirred at 23 °C for 14 h. The mixture was concentrated in vacuo and the product purified by flash chromatography on basic alumina (3:1 hexanes/EtOAc→DCM→7% MeOH/DCM) to yield **1b** as the free base (72 mg, 67% over 2 steps). Some of the material was further purified by preparative scale HPLC for use in biological assays. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 8.51 (s, 1H), 7.40-7.36 (m, 1H), 7.17-7.06 (m, 7H), 5.24 (dd, 1H, *J* = 4.8, 6.4 Hz), 5.17 (dd, 1H, *J* = 4.8, 8.0 Hz), 5.10-5.04 (m, 2H), 4.66-4.62 (m, 2H), 4.57 (t, *J* = 8.0 Hz), 4.35 (d, 1H, *J* = 7.6 Hz), 4.27 (dd, 1H, *J* = 6.4, 11.6 Hz), 4.24 (dd, 1H, *J* = 6.0, 12.0 Hz), 3.76-3.65 (m, 2H), 2.87-2.72 (m, 4H), 2.63 (s, 3H), 2.61 (s, 3H), 2.43-2.31 (m, 2H), 2.28-2.17 (m, 2H), 2.05-1.88 (m, 7H), 1.86-1.74 (m, 5H), 1.52 (d, 3H, *J* = 7.2 Hz), 1.49 (d, 3H, *J* = 7.2 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 206.6, 172.9, 172.8, 171.7, 167.7, 167.0, 138.7, 138.5, 137.6, 137.6, 130.1, 129.9, 129.7, 129.3, 128.2, 128.1, 127.2, 127.2, 127.1, 91.1, 91.0, 71.6, 70.9, 60.8, 59.8, 58.9, 58.8, 58.8, 32.3, 32.3, 32.2, 31.3, 31.3, 31.2, 30.2, 30.2, 27.2, 26.8, 21.8, 21.5, 16.8, 16.7. HRMS calcd for C<sub>22</sub>H<sub>30</sub>N<sub>4</sub>O<sub>4</sub>Na: 437.21593, found 437.20535.

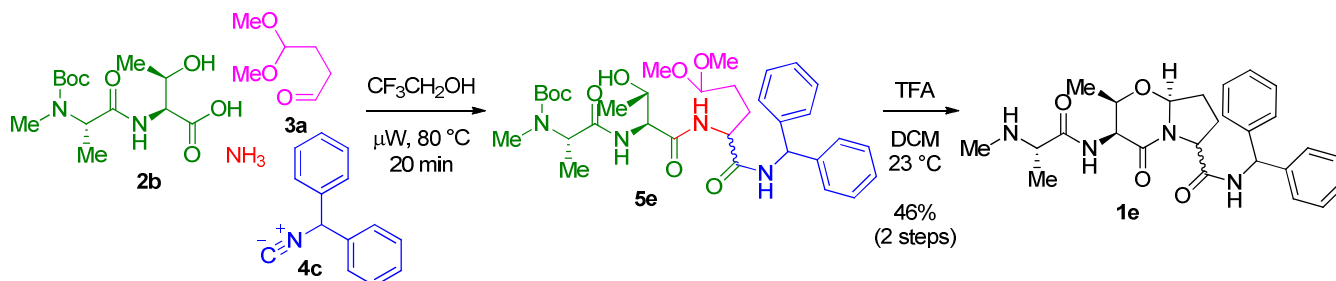


**(2*R*,3*S*,8*aS*)-2-Methyl-3-((*S*)-2-(methylamino)propanamido)-4-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)hexahydro-2*H*-pyrrolo[2,1-*b*][1,3]oxazine-6-carboxamide (1c).** Same procedure as above (1a) with carboxylic acid **2b** (85 mg, 0.279 mmol, 1.0 equiv), aldehyde **3a** (39 mg, 0.293 mmol, 1.05 equiv), isocyanide **4d** (44 mg, 0.279 mmol, 1.0 equiv) and 7 M ammonia in MeOH (80  $\mu$ L, 0.559 mmol, 2.0 equiv) in TFE (3 mL). The resultant oil **5c** was combined with TFA (128  $\mu$ L, 1.67 mmol, 6 equiv) in DCM (5 mL) and stirred at 23 °C for 14 h. The mixture was concentrated in vacuo and the product purified by flash chromatography on basic alumina (3:1 hexanes/EtOAc $\rightarrow$ DCM $\rightarrow$ 7% MeOH/DCM) to yield **1c** as a slightly impure free base (94 mg, 79% semi-pure). Some of the material was further purified by preparative scale HPLC for use in biological assays and data collection.  $^1\text{H}$  NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$ : 8.28 (d, 1H,  $J$  = 8.8 Hz), 8.26 (d, 1H,  $J$  = 8.8 Hz), 8.24 (s, 2H), 8.18 (d, 1H,  $J$  = 8.8 Hz), 8.00 (d, 1H,  $J$  = 8.8 Hz), 7.28 (d, 1H,  $J$  = 7.2 Hz), 7.16-7.06 (m, 6H), 5.25 (t, 1H,  $J$  = 5.6 Hz), 5.20 (dd, 1H,  $J$  = 5.2, 7.6 Hz), 4.99-4.92 (m, 2H), 4.60 (dd, 1H,  $J$  = 5.6, 8.4 Hz), 4.50 (dd, 1H,  $J$  = 5.2, 8.4 Hz), 4.46 (t, 1H,  $J$  = 7.2 Hz), 4.34-4.27 (m, 2H), 4.24 (t, 2H,  $J$  = 8.4 Hz), 3.13 (q, 1H,  $J$  = 6.8 Hz), 3.09 (q, 1H,  $J$  = 6.8 Hz), 2.76-2.70 (m, 3H), 2.24 (s, 3H), 2.22 (s, 3H), 1.93-1.80 (m, 6H), 1.80-1.60 (m, 6H), 1.17 (d, 3H,  $J$  = 6.8 Hz), 1.15 (d, 3H,  $J$  = 6.8 Hz), 1.07 (d, 3H,  $J$  = 6.4 Hz), 1.00 (d, 3H,  $J$  = 6.4 Hz);  $^{13}\text{C}$  NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$ : 174.1, 170.3, 169.9, 165.7, 165.2, 137.6, 137.4, 137.0, 136.9, 128.7, 128.5, 128.3, 127.7, 126.7, 126.6, 125.8, 125.7, 99.5, 87.7, 87.6, 73.4, 72.6, 59.2, 58.8, 58.7, 57.9, 50.5, 50.3, 46.6, 46.5, 34.0, 33.7, 30.7, 30.0, 29.9, 28.8, 28.8, 26.0, 25.7, 20.5, 18.9, 18.7, 16.5. HRMS calcd for C<sub>23</sub>H<sub>32</sub>N<sub>4</sub>O<sub>4</sub>Na: 451.23158, found 451.23286.

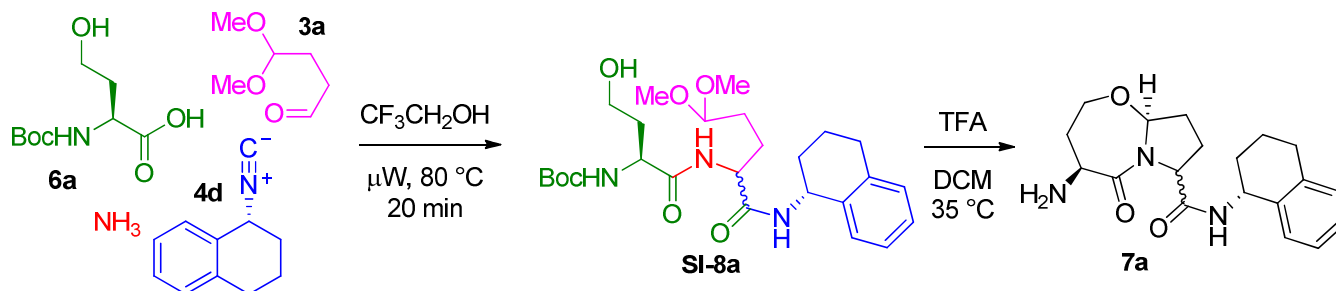


**(2*R*,3*S*,8*aS*)-2-Methyl-3-((*S*)-2-(methylamino)propanamido)-*N*-(naphthalen-1-yl)-4-oxohexahydro-2*H*-pyrrolo[2,1-*b*][1,3]oxazine-6-carboxamide (1d).** Same procedure as above (1a) with carboxylic acid **2b** (100 mg, 0.328 mmol, 1.0 equiv), aldehyde **3a** (46 mg, 0.344 mmol, 1.05 equiv), isocyanide **4b** (50 mg, 0.328 mmol, 1.0 equiv) and 7 M ammonia in MeOH (94  $\mu$ L, 0.657 mmol, 2.0 equiv) in TFE (3 mL). The resultant oil **5d** was combined with TFA (151  $\mu$ L, 1.97 mmol, 6 equiv) in DCM (5 mL) and stirred at 23 °C for 14 h. The mixture was concentrated in vacuo and the product purified by flash chromatography on basic alumina (1:1 hexanes/EtOAc $\rightarrow$ DCM $\rightarrow$ 7% MeOH/DCM) to yield **1d** as the free base (88 mg, 63% over 2 steps). Some of the material was further purified by preparative scale HPLC for use in biological assays.  $^1\text{H}$  NMR (400 MHz, CD<sub>3</sub>OD)

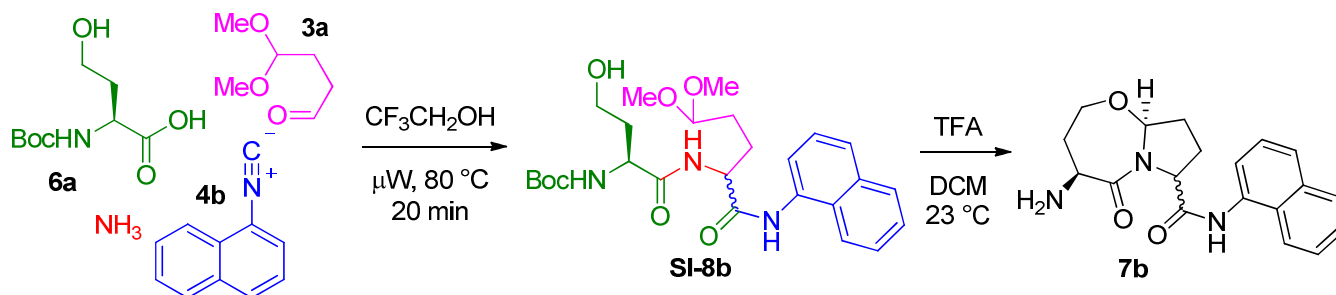
$\delta$ : 8.53 (bs, 1H), 8.13-8.09 (m, 1H), 8.05 (d, 1H,  $J = 6.8$  Hz), 7.92-7.87 (m, 2H), 7.81 (t, 2H,  $J = 6.4$  Hz), 7.56-7.46 (m, 8H), 5.29 (dd, 1H,  $J = 5.2, 8.0$  Hz), 5.21 (dd, 1H,  $J = 4.8, 8.4$  Hz), 4.83 (t, 1H,  $J = 8.4$  Hz), 4.71-4.67 (m, 2H), 4.62 (d, 1H,  $J = 4.0$  Hz), 4.36-4.24 (m, 2H), 3.70 (q, 1H,  $J = 6.8$  Hz), 3.65 (q, 1H,  $J = 6.8$  Hz), 2.59 (s, 3H), 2.52 (s, 3H), 2.42-2.32 (m, 2H), 2.31-2.18 (m, 2H), 2.16-2.02 (m, 2H), 1.96-1.85 (m, 1H), 1.50 (d, 3H,  $J = 6.8$  Hz), 1.38 (d, 3H,  $J = 7.2$  Hz), 1.24 (t, 6H,  $J = 6.8$  Hz);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 173.3, 173.2, 172.7, 172.4, 168.3, 167.5, 135.7, 135.7, 134.0, 133.7, 130.6, 130.6, 129.3, 129.2, 128.2, 128.1, 127.5, 127.4, 127.3, 127.2, 126.4, 126.4, 124.6, 124.3, 124.1, 123.8, 91.0, 90.9, 76.3, 75.7, 60.6, 59.5, 59.1, 58.9, 52.8, 52.5, 32.6, 32.4, 32.2, 31.2, 26.7, 26.2, 17.3, 17.2, 16.7. HRMS calcd for  $\text{C}_{23}\text{H}_{29}\text{N}_4\text{O}_4$ : 425.2183, found 425.2181.



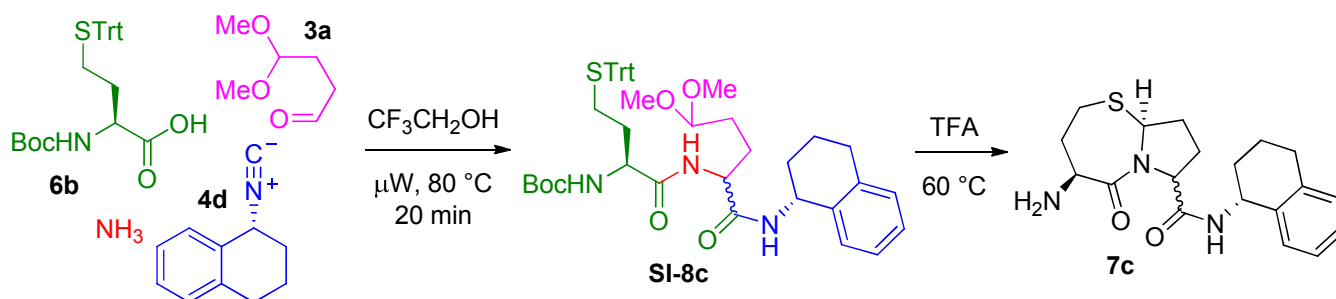
**(2R,3S,8aS)-N-Benzhydryl-2-methyl-3-((S)-2-(methylamino)propanamido)-4-oxohexahydro-2H-pyrrolo[2,1-b][1,3]oxazine-6-carboxamide (1e).** Same procedure as above (**1a**) with carboxylic acid **2b** (105 mg, 0.345 mmol, 1.0 equiv), aldehyde **3a** (47 mg, 0.362 mmol, 1.05 equiv), isocyanide **4c** (67 mg, 0.345 mmol, 1.0 equiv) and 7 M ammonia in MeOH (99  $\mu\text{L}$ , 0.690 mmol, 2.0 equiv) in TFE (3 mL). The resultant oil **5e** was combined with TFA (159  $\mu\text{L}$ , 2.07 mmol, 6 equiv) in DCM (5 mL) and stirred at 23 °C for 14 h. The mixture was concentrated in vacuo and the product purified by flash chromatography on basic alumina (1:1 hexanes/EtOAc $\rightarrow$ DCM $\rightarrow$ 7% MeOH/DCM) to yield **1e** as the free base (73 mg, 46% over 2 steps).  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 7.38-7.18 (m, 20H), 6.17 (s, 1H), 6.15 (s, 1H), 5.21 (dd, 1H,  $J = 5.2, 8.0$  Hz), 5.13 (dd, 1H,  $J = 4.8, 8.8$  Hz), 4.66 (t, 1H,  $J = 7.6$  Hz), 4.62 (d, 1H,  $J = 4.4$  Hz), 4.56 (d, 1H,  $J = 4.4$  Hz), 4.47 (d, 1H,  $J = 8.4$  Hz), 4.28 (dd, 1H,  $J = 4.4, 6.4$  Hz), 4.22 (dd, 1H,  $J = 4.4, 6.4$  Hz), 3.25 (q, 1H,  $J = 6.8$  Hz), 3.21 (q, 1H,  $J = 6.8$  Hz), 2.36 (s, 3H), 2.30 (s, 3H), 2.39-2.25 (m, 2H), 2.19-2.12 (m, 2H), 2.06-1.86 (m, 4H), 1.85-1.79 (m, 2H), 1.31 (d, 3H,  $J = 6.8$  Hz), 1.26 (d, 3H,  $J = 7.2$  Hz), 1.20 (d, 3H,  $J = 6.4$  Hz), 1.18 (d, 3H,  $J = 6.4$  Hz);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 176.8, 176.6, 173.0, 172.7, 168.0, 167.6, 143.0, 142.8, 142.6, 142.6, 129.7, 129.6, 129.5, 129.4, 128.9, 128.8, 128.7, 128.6, 128.5, 128.5, 128.3, 128.1, 90.7, 90.6, 76.3, 75.5, 60.4, 60.3, 60.0, 59.2, 58.5, 58.4, 52.5, 52.2, 34.2, 34.1, 32.1, 31.1, 29.5, 26.7, 26.1, 19.2, 19.1, 16.8, 16.7. HRMS calcd for  $\text{C}_{26}\text{H}_{32}\text{N}_4\text{O}_4\text{Na}$ : 487.23158, found 487.23308.



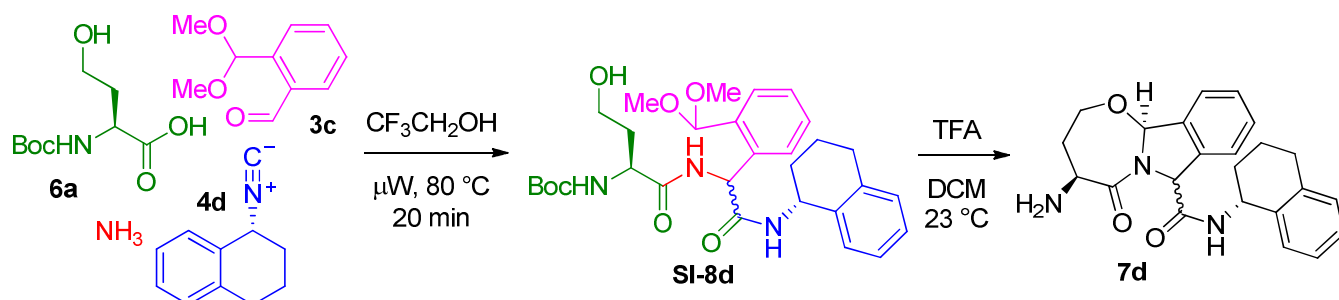
**(4*S*,9*aS*)-4-Amino-5-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)octahydropyrrolo[2,1-*b*][1,3]oxazepine-7-carboxamide (7a).** A mixture of Boc-*N*-HSer-OH (**6a**) (318 mg, 1.45 mmol, 1.0 equiv), aldehyde **3a** (201 mg, 1.52 mmol, 1.05 equiv), isocyanide **4d** (228 mg, 1.45 mmol, 1.0 equiv) and 7 M ammonia in MeOH (414  $\mu$ L, 2.90 mmol, 2.0 equiv) in TFE (5 mL) was stirred under microwave irradiation at a set temperature of 80  $^{\circ}$ C for 20 min. The mixture was then transferred to a round bottom flask and concentrated in vacuo, then 1 M NaOH (15 mL) was added and the mixture was extracted with DCM (3 x 7 mL). The organics were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo. The resultant oil **SI-8a** was combined with TFA (834  $\mu$ L, 10.9 mmol, 8 equiv) in DCM (5 mL) and stirred at 35  $^{\circ}$ C for 14 h. The mixture was concentrated in vacuo and **7a** used without further purification in the next step.



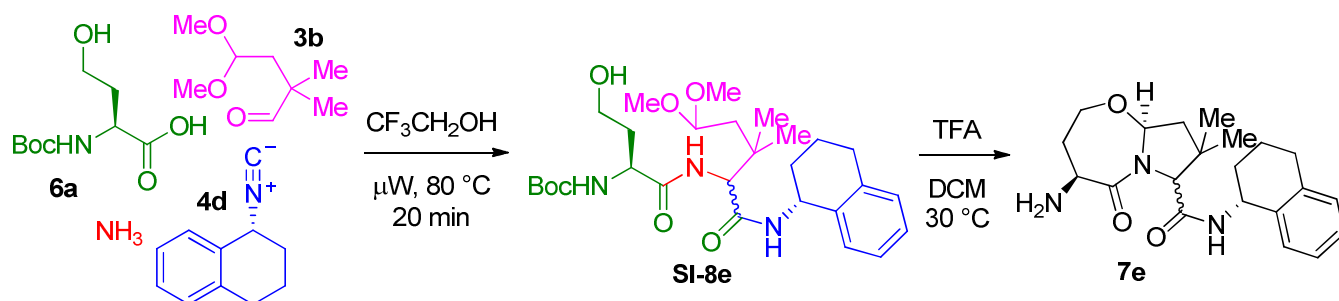
**(4*S*,9*aS*)-4-Amino-*N*-(naphthalen-1-yl)-5-oxooctahydropyrrolo[2,1-*b*][1,3]oxazepine-7-carboxamide (7b).** Same procedure as above (**7a**) with Boc-*N*-HSer-OH (**6a**) (150 mg, 0.684 mmol, 1.0 equiv), aldehyde **3a** (95 mg, 0.718 mmol, 1.05 equiv), isocyanide **4b** (105 mg, 0.684 mmol, 1.0 equiv) and 7 M ammonia in MeOH (195  $\mu$ L, 1.37 mmol, 2.0 equiv) in TFE (4 mL). The resultant oil **SI-8b** was combined with TFA (314  $\mu$ L, 4.10 mmol, 6 equiv) in DCM (5 mL) and stirred at 23  $^{\circ}$ C for 14 h. The mixture was concentrated in vacuo and **7b** used without further purification in the next step.



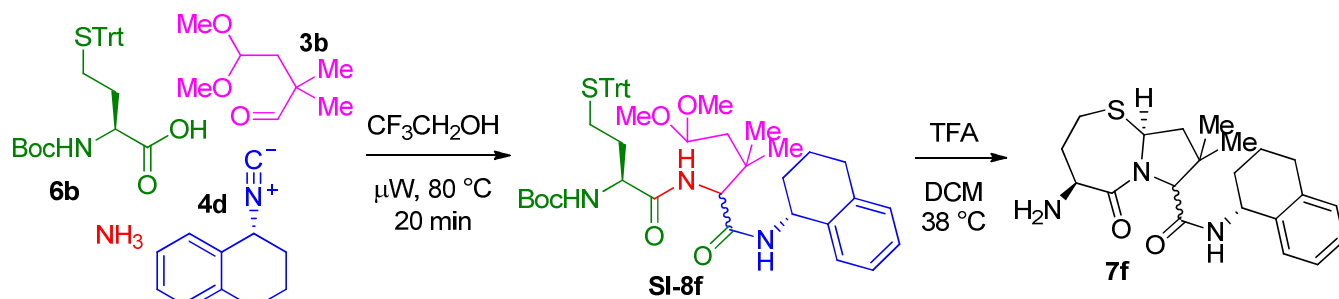
**(4*S*,7*S*,9*aS*)-4-Amino-5-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)octahydropyrrolo[2,1-*b*][1,3]thiazepine-7-carboxamide (7c).** Same procedure as above (**7a**) with Boc-*N*-HCys(Trt)-OH (**6b**) (665 mg, 1.39 mmol, 1.0 equiv), aldehyde **3a** (193 mg, 1.46 mmol, 1.05 equiv), isocyanide **4d** (219 mg, 1.39 mmol, 1.0 equiv) and 7 M ammonia in MeOH (398  $\mu$ L, 2.78 mmol, 2.0 equiv) in TFE (5 mL). The resultant oil **SI-8c** was combined with TFA (1.07 mL, 13.9 mmol, 10 equiv) in DCM (5 mL) and stirred at 60  $^{\circ}$ C for 6 h. The mixture was concentrated in vacuo, then partially purified (trityl byproduct removed and more polar product(s) collected) by flash chromatography on basic alumina (3:1 hexanes/EtOAc $\rightarrow$ DCM $\rightarrow$ 7% MeOH/DCM) to yield semi-pure **7c**.



**(4*S*,11*bS*)-4-Amino-5-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)-2,3,4,5,7,11*b*-hexahydro-[1,3]oxazepino[2,3-*a*]isoindole-7-carboxamide (**7d**). Same procedure as above (**7a**) with Boc-*N*-HSer-OH (**6a**) (175 mg, 0.800 mmol, 1.0 equiv), aldehyde **3c** (144 mg, 0.800 mmol, 1.0 equiv), isocyanide **4d** (126 mg, 0.800 mmol, 1.0 equiv) and 7 M ammonia in MeOH (229  $\mu\text{L}$ , 1.60 mmol, 2.0 equiv) in TFE (4 mL). The resultant oil **SI-8d** was combined with TFA (490  $\mu\text{L}$ , 6.40 mmol, 8 equiv) in DCM (3 mL) and stirred at  $23\text{ }^\circ\text{C}$  for 14 h. The mixture was concentrated in vacuo and **7d** used without further purification in the next step.**



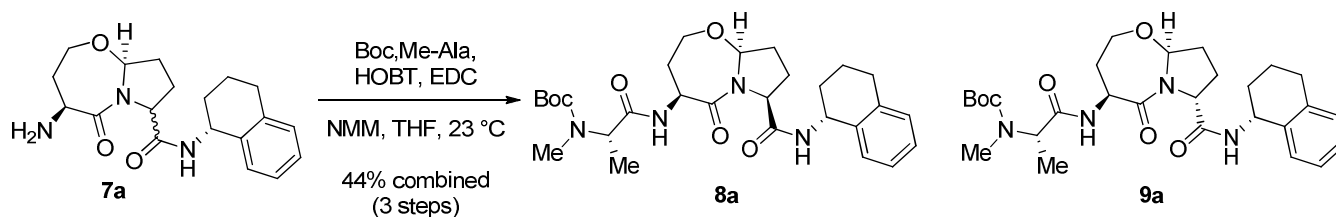
**(4*S*,9*aS*)-4-Amino-8,8-dimethyl-5-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)octahydropyrrolo[2,1-*b*][1,3]oxazepine-7-carboxamide (**7e**). Same procedure as above (**7a**) with Boc-*N*-HSer-OH (**6a**) (157 mg, 0.718 mmol, 1.0 equiv), aldehyde **3b** (144 mg, 0.718 mmol, 1.0 equiv), isocyanide **4d** (113 mg, 0.718 mmol, 1.0 equiv) and 7 M ammonia in MeOH (205  $\mu\text{L}$ , 1.44 mmol, 2.0 equiv) in TFE (4 mL). The resultant oil **SI-8e** was combined with TFA (473  $\mu\text{L}$ , 7.18 mmol, 10 equiv) in DCM (4 mL) and stirred at  $30\text{ }^\circ\text{C}$  for 14 h. The mixture was concentrated in vacuo and **7e** used without further purification in the next step.**



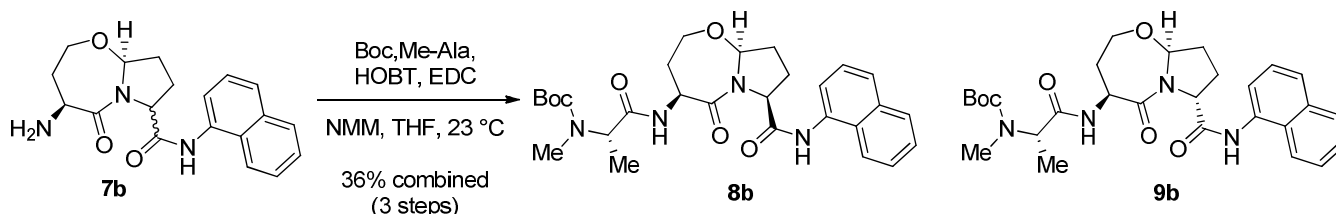
**(4*S*,7*S*,9*aS*)-4-Amino-8,8-dimethyl-5-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)octahydropyrrolo[2,1-*b*][1,3]thiazepine-7-carboxamide (**7f**). Same procedure as above (**7a**) with Boc-*N*-HCys(Trt)-OH (**6b**) (500 mg, 1.05 mmol, 1.0 equiv), aldehyde **3b** (176 mg, 1.10 mmol, 1.05 equiv), isocyanide **4d** (165 mg, 1.05 mmol, 1.0 equiv) and 7 M ammonia in MeOH (299  $\mu\text{L}$ , 2.09 mmol, 2.0 equiv) in TFE (5 mL). The resultant oil **SI-8f** was**



combined with TFA (804  $\mu\text{L}$ , 10.5 mmol, 10 equiv) in DCM (5 mL) and stirred at 38  $^{\circ}\text{C}$  for 14 h. The mixture was concentrated in vacuo, then partially purified (trityl byproduct removed and more polar product(s) collected) by flash chromatography on basic alumina (3:1 hexanes/EtOAc $\rightarrow$ DCM $\rightarrow$ 7% MeOH/DCM) to yield semi-pure **7f**.

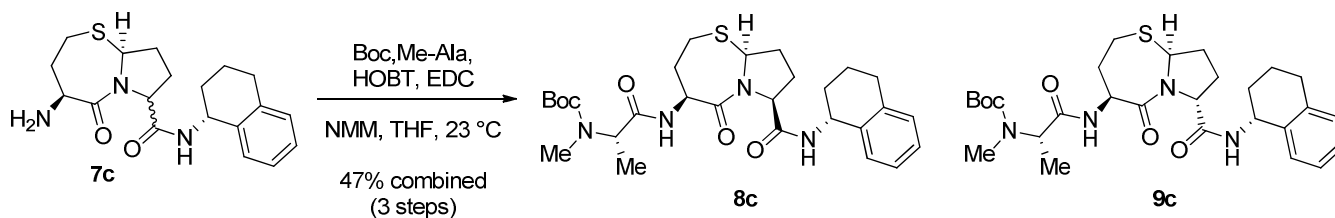


**tert-Butyl methyl((2S)-1-oxo-1-(((4S,9aS)-5-oxo-7-((R)-1,2,3,4-tetrahydronaphthalen-1-yl)carbamoyl)octahydropyrrolo[2,1-b][1,3]oxazepin-4-yl)amino)propan-2-yl)carbamate (**8a**) and (**9a**).** To a solution of crude **7a**·TFA (622 mg, 1.36 mmol, 1.0 equiv), Boc-*N*-Me-Ala-OH (276 mg, 1.36 mmol, 1.0 equiv), HOBT·xH<sub>2</sub>O (229 mg, 1.50 mmol, 1.1 equiv) and NMM (598  $\mu\text{L}$ , 5.44 mmol, 4 equiv) in THF (15 mL) at 0  $^{\circ}\text{C}$  was added EDC·HCl (274 mg, 1.43 mmol, 1.05 equiv). After 30 min the cold bath was removed. The solution stirred for 14 h and then was quenched with saturated aqueous NaHCO<sub>3</sub> (25 mL), extracted with EtOAc (2 x 20 mL), dried over sodium sulfate and then concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (2:1 $\rightarrow$ 1:1 $\rightarrow$ 1:3 hexanes/EtOAc) to yield, after 3 steps, partially separated diastereomers **8a** (30 mg, 4%, ~3:1 d.r.) and **9a** (40 mg, 5%, ~3:1 d.r.), along with unseparated **8a+9a** (267 mg, 35%). Data for **8a**:  $R_f$  = 0.40 (1:3 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.23-7.05 (m, 4H), 6.84 (d, 1H,  $J$  = 8.0 Hz), 5.22 (t, 1H,  $J$  = 6.4 Hz), 5.18-5.08 (m, 1H), 4.69 (dd, 1H,  $J$  = 5.6, 10.8 Hz), 4.62 (d, 1H,  $J$  = 7.6 Hz), 4.13-4.03 (m, 1H), 3.95 (q, 1H,  $J$  = 12.8 Hz), 2.75 (s, 3H), 2.80-2.74 (m, 1H), 2.47-2.37 (m, 1H), 2.17-1.89 (m, 4H), 1.88-1.69 (m, 5H), 1.43 (s, 9H), 1.32 (d, 3H,  $J$  = 7.2 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 171.4, 169.9, 169.8, 137.6, 137.3, 136.9, 136.7, 129.3, 129.2, 128.6, 128.3, 27.4, 127.3, 126.4, 126.2, 90.3, 90.0, 70.7, 70.6, 61.1, 60.6, 53.1, 52.6, 47.7, 47.7, 33.3, 32.7, 32.5, 30.2, 30.1, 29.8, 29.3, 29.3, 28.4, 28.4, 25.9, 20.5, 20.1. Data for **9a**:  $R_f$  = 0.55 (1:3 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.24-7.11 (m, 4H), 7.11-7.05 (m, 1H), 6.69 (bs, 1H), 5.21 (d, 1H,  $J$  = 5.6 Hz), 5.10 (q, 1H,  $J$  = 6.8 Hz), 4.75 (dd, 1H,  $J$  = 7.6, 11.6 Hz), 4.55 (d, 1H,  $J$  = 8.0 Hz), 4.47 (t, 1H,  $J$  = 8.8 Hz), 4.01 (d, 1H,  $J$  = 12.8 Hz), 3.97 (t, 1H,  $J$  = 12.4 Hz), 2.82-2.75 (m, 2H), 2.77 (s, 3H), 2.45-2.33 (m, 1H), 2.32-2.24 (m, 1H), 2.24-2.13 (m, 2H), 2.06-1.93 (m, 3H), 1.84-1.76 (m, 2H), 1.74-1.64 (m, 5H), 1.45 (s, 9H), 1.34 (d, 3H,  $J$  = 6.8 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 172.1, 171.0, 169.8, 137.6, 136.7, 129.3, 128.6, 127.4, 126.4, 90.0, 70.6, 66.0, 61.2, 53.2, 47.8, 33.3, 32.7, 30.2, 29.3, 28.5, 25.8, 20.2, 14.0. HRMS calcd for C<sub>28</sub>H<sub>40</sub>N<sub>4</sub>O<sub>6</sub>: 551.2840, found 551.2838.



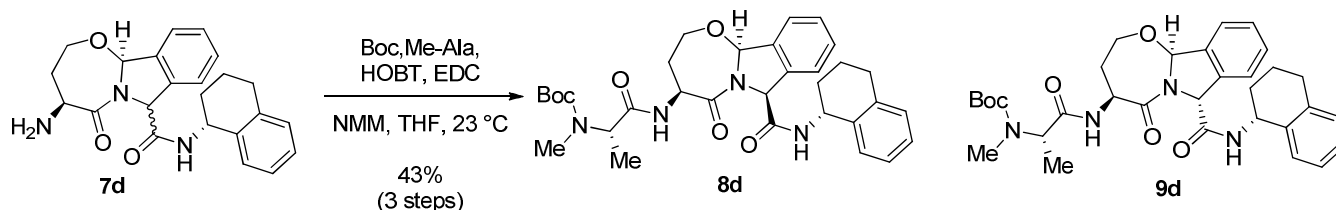
**tert-Butyl methyl((2S)-1-(((4S,9aS)-7-(naphthalen-1-ylcarbamoyl)-5-oxooctahydropyrrolo[2,1-b][1,3]oxazepin-4-yl)amino)-1-oxopropan-2-yl)carbamate (**8b**) and (**9b**).** Same procedure as above (**8a**) using

crude **7b**•TFA (209 mg, 0.615 mmol, 1.0 equiv), Boc-*N*-Me-Ala-OH (125 mg, 0.615 mmol, 1.0 equiv), HOBT·xH<sub>2</sub>O (104 mg, 0.677 mmol, 1.1 equiv), NMM (338 μL, 3.08 mmol, 5 equiv [to soak up xs TFA]) and EDC·HCl (124 mg, 0.646 mmol, 1.05 equiv) in THF (10 mL). The resultant oil was purified by flash chromatography on silica gel (2:1→1:1→1:3 hexanes/EtOAc) to yield, after 3 steps, separated diastereomers **8b** (43 mg, 12%, ~6:1 d.r.) and **9b** (37 mg, 10%, ~6:1 d.r.), along with unseparated **8b+9b** (49 mg, 14%). Data for **8b**: R<sub>f</sub> = 0.33 (1:3 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 9.11 (s, 1H), 8.10 (d, 1H, *J* = 7.6 Hz), 7.98 (d, 1H, *J* = 8.8 Hz), 7.86 (d, 1H, *J* = 8.0 Hz), 7.67 (d, 1H, *J* = 8.4 Hz), 7.56-7.44 (m, 3H), 7.32 (s, 1H), 5.33 (t, 1H, *J* = 6.4 Hz), 4.90 (d, 1H, *J* = 6.4 Hz), 4.81 (dd, 1H, *J* = 5.2, 10.4 Hz), 4.19 (dt, 1H, *J* = 2.8, 12.8 Hz), 4.07-3.98 (m, 1H), 2.78 (s, 3H), 2.59-2.46 (m, 2H), 2.32-2.21 (m, 1H), 2.01-1.91 (m, 3H), 1.85-1.74 (m, 1H), 1.44 (s, 9H), 1.36 (d, 3H, *J* = 6.8 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 172.1, 171.4, 169.1, 168.5, 134.1, 132.7, 128.9, 126.6, 126.5, 126.0, 125.9, 125.5, 120.7, 119.8, 90.7, 70.8, 61.2, 52.8, 32.8, 32.6, 30.2, 28.5, 28.4, 25.6. Data for **9b**: R<sub>f</sub> = 0.42 (1:3 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 9.47 (s, 1H), 8.03 (d, 1H, *J* = 7.2 Hz), 7.94 (d, 1H, *J* = 7.6 Hz), 7.82 (d, 1H, *J* = 7.6 Hz), 7.63 (d, 1H, *J* = 8.0 Hz), 7.55 (s, 1H), 7.50-7.40 (m, 2H), 7.31 (s, 1H), 5.21 (s, 1H), 4.96 (d, 1H, *J* = 7.6 Hz), 4.85-4.78 (m, 1H), 4.43 (t, 1H, *J* = 8.8 Hz), 4.14 (d, 1H, *J* = 12.8 Hz), 3.99 (t, 1H, *J* = 12.0 Hz), 2.79 (s, 3H), 2.61-2.53 (m, 1H), 2.26-2.14 (m, 1H), 2.11-1.97 (m, 2H), 1.90-1.78 (m, 1H), 1.46 (s, 9H), 1.34 (d, 3H, *J* = 7.2 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 175.0, 173.3, 172.4, 168.5, 134.1, 132.8, 128.7, 126.5, 126.1, 125.8, 125.4, 121.0, 119.5, 90.3, 70.6, 65.9, 61.6, 53.2, 49.2, 33.6, 32.5, 30.3, 30.3, 28.5, 28.5, 28.4, 24.6. HRMS calcd for C<sub>28</sub>H<sub>36</sub>N<sub>4</sub>O<sub>6</sub>Na: 547.25271, found 547.25362.



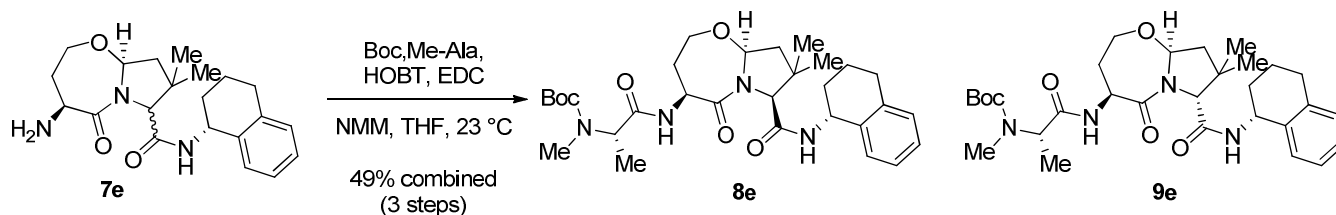
**tert-Butyl methyl((2*S*)-1-oxo-1-(((4*S*,9*aS*)-5-oxo-7-(((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)carbamoyl)octahydropyrrolo[2,1-*b*][1,3]thiazepin-4-yl)amino)propan-2-yl)carbamate (**8c**) and (**9c**). Same procedure as above (**8a**) using crude **7c**•TFA (658 mg, 1.39 mmol, 1.0 equiv), Boc-*N*-Me-Ala-OH (282 mg, 1.39 mmol, 1.0 equiv), HOBT·xH<sub>2</sub>O (234 mg, 1.39 mmol, 1.1 equiv), NMM (917 μL, 8.34 mmol, 6 equiv [to soak up xs TFA]) and EDC·HCl (280 mg, 1.46 mmol, 1.05 equiv) in THF (18 mL). The resultant oil was purified by flash chromatography on silica gel (1:1→1:2→1:3 hexanes/EtOAc) to yield, after 3 steps, separated diastereomers **8c** (121 mg, 16%, ~3:1 d.r.) and **9c** (100 mg, 13%, ~3:1 d.r.) along with unseparated **8c+9c** (136 mg, 18%). Data for **8c**: R<sub>f</sub> = 0.27 (1:3 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 7.32 (d, 1H, *J* = 7.6 Hz), 7.25-7.21 (m, 1H), 7.16-7.04 (m, 4H), 5.17 (q, 1H, *J* = 7.2 Hz), 5.08 (t, 1H, *J* = 7.2 Hz), 4.74 (d, 1H, *J* = 8.0 Hz), 4.53 (dd, 1H, *J* = 6.0, 10.8 Hz), 3.35-3.22 (m, 1H), 2.76 (s, 3H), 2.63-2.46 (m, 1H), 2.20 (d, 1H, *J* = 12.8 Hz), 2.12-1.98 (m, 2H), 1.92-1.71 (m, 5H), 1.59 (q, 1H, *J* = 12.4 Hz), 1.43 (s, 9H), 1.31 (d, 3H, *J* = 7.2 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 171.3, 169.6, 169.3, 137.3, 129.2, 129.1, 128.8, 127.2, 126.1, 62.3, 61.8, 52.8, 47.6, 33.0, 32.1, 30.4, 30.2, 29.3, 28.4, 28.4, 26.5, 20.5. Data for **9c**: R<sub>f</sub> = 0.44 (1:3 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 7.22-7.12 (m, 4H), 7.09-7.04 (m, 1H), 6.62 (bs, 1H), 5.28 (d, 1H, *J* = 7.6 Hz), 5.09 (d, 1H, *J* = 6.4 Hz), 4.66-4.56 (m, 2H), 3.32 (t,**

1H,  $J = 12.0$  Hz), 2.87-2.68 (m, 3H), 2.75 (s, 3H), 2.35-2.19 (m, 3H), 2.08-1.96 (m, 2H), 1.85-1.69 (m, 5H), 1.45 (s, 9H), 1.29 (d, 3H,  $J = 7.2$  Hz);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 170.9, 169.6, 137.6, 136.6, 129.3, 128.6, 127.4, 126.3, 63.8, 61.3, 53.5, 47.7, 33.7, 31.7, 30.1, 29.3, 28.5, 28.4, 20.1. HRMS calcd for  $\text{C}_{28}\text{H}_{40}\text{N}_4\text{O}_5\text{SNa}$ : 567.26116, found 567.26151.



**tert-Butyl methyl((2S)-1-oxo-1-(((4S,11bS)-5-oxo-7-(((R)-1,2,3,4-tetrahydronaphthalen-1-yl)carbamoyl)-2,3,4,5,7,11b-hexahydro-[1,3]oxazepino[2,3-a]isoindol-4-yl)amino)propan-2-yl)carbamate (8d) and (9d).**

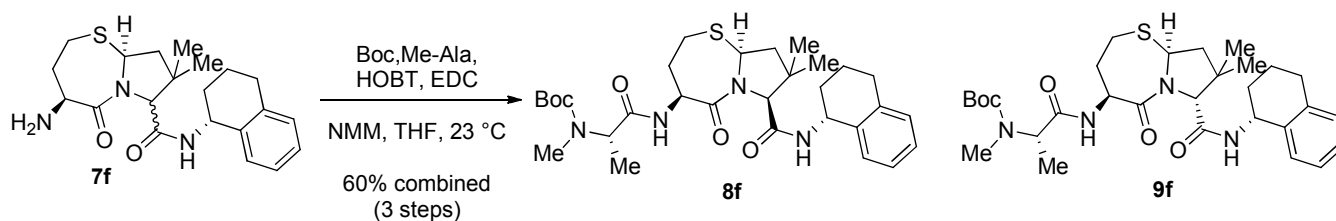
Same procedure as above (**8a**) using crude **7d**•TFA (323 mg, 0.640 mmol, 1.0 equiv), Boc-*N*-Me-Ala-OH (130 mg, 0.640 mmol, 1.0 equiv), HOBT•xH<sub>2</sub>O (108 mg, 0.704 mmol, 1.1 equiv), NMM (281  $\mu\text{L}$ , 2.56 mmol, 4 equiv) and EDC•HCl (129 mg, 0.672 mmol, 1.05 equiv) in THF (12 mL). The resultant oil was purified by flash chromatography on silica gel (3:1→1:1→1:2 hexanes/EtOAc) to yield, after 3 steps, the unseparated diastereomixture **8d**+**9d** (200 mg, 43%). By NMR, one of the diastereomers seems to exist as a pair of rotational isomers.  $R_f = 0.18$  (1:1 hexanes/EtOAc).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.56 (d, 1H,  $J = 7.6$  Hz), 7.47 (q, 1H,  $J = 4.4$  Hz), 7.44-7.39 (m, 5H), 7.38-7.34 (m, 1H), 7.32-7.27 (m, 1H), 7.18-7.14 (m, 3H), 7.10-7.06 (m, 1H), 7.03 (d, 1H,  $J = 7.2$  Hz), 6.90 (d, 1H,  $J = 7.2$  Hz), 6.74 (d, 1H,  $J = 7.6$  Hz), 6.44-6.36 (m, 3H), 6.21 (s, 1H), 5.50 (bs, 2H), 5.17-5.10 (m, 1H), 5.03 (dd, 1H,  $J = 8.0, 14.4$  Hz), 4.88-4.80 (m, 2H), 4.72-4.66 (m, 1H), 4.44 (td, 2H,  $J = 8.8$  Hz), 4.31-4.15 (m, 5H), 2.80 (s, 3H), 2.79 (s, 3H), 2.77 (s, 3H), 2.71 (t, 4H,  $J = 6.4$  Hz), 2.22-2.08 (m, 3H), 2.06-1.98 (m, 2H), 1.86-1.73 (m, 5H), 1.71-1.61 (m, 2H), 1.48 (s, 9H), 1.46 (s, 9H), 1.35 (d, 3H,  $J = 7.2$  Hz), 1.34 (d, 3H,  $J = 7.2$  Hz), 1.33 (d, 3H,  $J = 7.2$  Hz);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 175.0, 172.4, 170.5, 168.3, 168.1, 137.7, 137.2, 136.8, 136.5, 136.5, 135.9, 135.7, 135.7, 135.2, 130.7, 130.5, 129.4, 129.3, 129.0, 128.7, 127.8, 127.4, 127.2, 126.4, 126.2, 125.0, 125.0, 122.9, 122.3, 122.3, 92.0, 91.5, 71.4, 71.4, 66.7, 66.5, 65.9, 53.3, 52.8, 49.2, 47.9, 47.7, 30.3, 29.3, 29.2, 28.5, 28.4, 28.4, 20.4, 20.2. HRMS calcd for  $\text{C}_{32}\text{H}_{41}\text{N}_4\text{O}_6\text{Na}$ : 599.28401, found 599.28561.



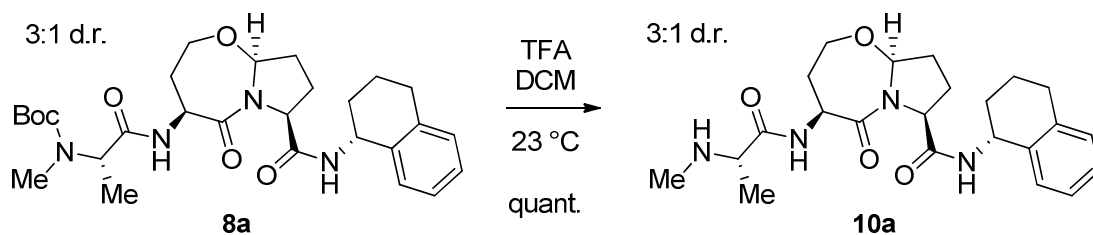
**tert-Butyl ((S)-1-(((4S,7S,9aS)-8,8-dimethyl-5-oxo-7-(((R)-1,2,3,4-tetrahydronaphthalen-1-yl)carbamoyl)octahydropyrrolo[2,1-b][1,3]oxazepin-4-yl)amino)-1-oxopropan-2-yl)(methyl)carbamate (8e) and (9e).**

Same procedure as above (**8a**) using crude **7e**•TFA (270 mg, 0.555 mmol, 1.0 equiv), Boc-*N*-Me-Ala-OH (113 mg, 0.555 mmol, 1.0 equiv), HOBT•xH<sub>2</sub>O (93 mg, 0.610 mmol, 1.1 equiv), NMM (366  $\mu\text{L}$ , 3.33 mmol, 6 equiv [to soak up xs TFA]) and EDC•HCl (112 mg, 0.582 mmol, 1.05 equiv) in THF (10 mL). The resultant oil was purified by flash chromatography on silica gel (3:1→1:1→1:3 hexanes/EtOAc) to yield, after 3 steps, separated

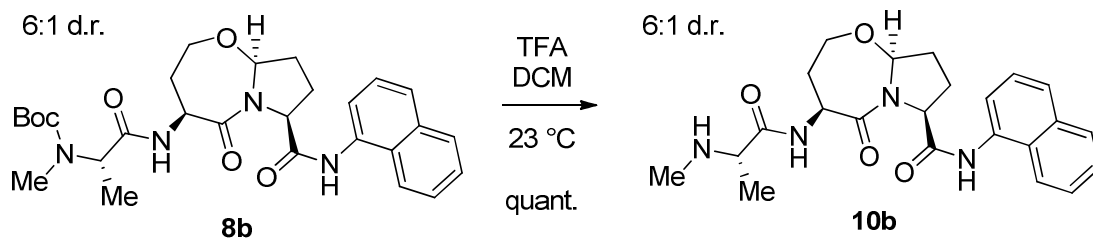
diastereomer **8e** (29 mg, 7%, >10:1 d.r.) along with unseparated **8e+9e** (168 mg, 42%). Data for **8e**:  $R_f = 0.30$  (1:1 hexanes/EtOAc).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.29-7.25 (m, 2H), 7.17-7.12 (m, 2H), 7.09-7.05 (m, 1H), 6.72 (d, 1H,  $J = 8.0$  Hz), 5.24 (t, 1H,  $J = 5.6$  Hz), 5.16 (dd, 1H,  $J = 5.6, 6.8$  Hz), 4.70 (dd, 1H,  $J = 5.6, 11.2$  Hz), 4.16 (s, 1H), 4.05-3.98 (m, 1H), 3.93 (q, 1H,  $J = 12.4$  Hz), 2.79 (s, 3H), 2.78-2.73 (m, 2H), 2.19 (dd, 1H,  $J = 6.8, 14.0$  Hz), 2.06-1.96 (m, 2H), 1.88 (dd, 1H,  $J = 6.0, 14.0$  Hz), 1.87-1.69 (m, 5H), 1.66-1.60 (m, 1H), 1.47 (s, 9H), 1.34 (d, 3H,  $J = 7.2$  Hz), 1.18 (s, 3H), 1.07 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 170.7, 168.8, 137.3, 136.7, 136.6, 129.2, 128.9, 127.4, 126.4, 89.3, 89.2, 70.9, 70.7, 52.6, 47.5, 46.1, 39.6, 30.2, 29.7, 29.2, 28.5, 28.4, 23.8, 21.2, 19.9, 14.3, 14.0. Data for **9e**:  $R_f = 0.39$  (1:3 hexanes/EtOAc). HRMS calcd for  $\text{C}_{30}\text{H}_{44}\text{N}_4\text{O}_6\text{Na}$ : 579.3153, found 579.3155.



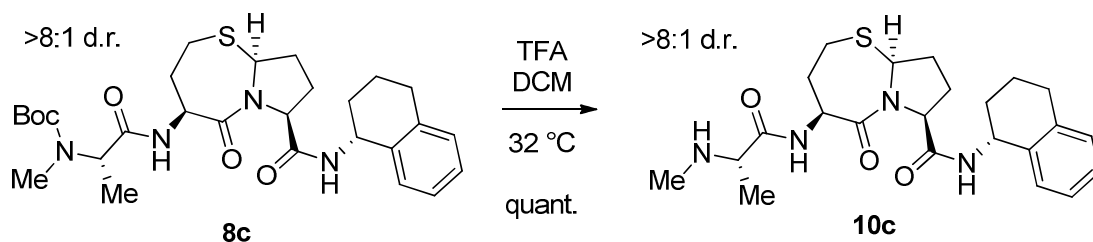
**tert-Butyl ((2S)-1-(((4S,9aS)-8,8-dimethyl-5-oxo-7-((R)-1,2,3,4-tetrahydronaphthalen-1-yl)carbamoyl)octahydropyrrolo[2,1-b][1,3]thiazepin-4-yl)amino)-1-oxopropan-2-yl)(methyl)carbamate (8f) and (9f).** Same procedure as above (**8a**) using crude **7f**•TFA (387 mg, 0.998 mmol, 1.0 equiv), Boc-*N*-Me-Ala-OH (202 mg, 0.998 mmol, 1.0 equiv), HOBT·xH<sub>2</sub>O (168 mg, 1.10 mmol, 1.1 equiv), NMM (329  $\mu\text{L}$ , 2.99 mmol, 3 equiv) and EDC·HCl (201 mg, 1.05 mmol, 1.05 equiv) in THF (10 mL). The resultant oil was purified by flash chromatography on silica gel (3:1→1:1→1:3 hexanes/EtOAc) to yield, after 3 steps, separated diastereomer **8f** (12 mg, 2%) and **9f** (47 mg, 8%), along with unseparated **8f+9f** (300 mg, 50%) and unreacted Boc-**7f** (59 mg, 12%) left over from the previous reaction. Data for diastereomixture:  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 7.32-7.28 (m, 1H), 7.18-7.11 (m, 6H), 7.10-7.06 (m, 2H), 5.49 (d, 1H,  $J = 9.2$  Hz), 5.41 (q, 1H,  $J = 8.0$  Hz), 5.09 (t, 1H,  $J = 6.0$  Hz), 5.03 (t, 1H,  $J = 6.0$  Hz), 5.03 (t, 1H,  $J = 12.0$  Hz), 4.69-4.57 (m, 4H), 4.24 (d, 1H,  $J = 12.4$  Hz), 4.19-4.16 (m, 1H), 3.31 (d, 2H,  $J = 2.0$  Hz), 3.29-3.21 (m, 2H), 2.86 (s, 6H), 2.81 (s, 3H), 2.80-2.75 (m, 2H), 2.68-2.56 (m, 1H), 2.31-2.20 (m, 3H), 2.02-1.75 (m, 13H), 1.48 (s, 18H), 1.37 (d, 3H,  $J = 7.6$  Hz), 1.32 (d, 3H,  $J = 7.2$  Hz), 1.15 (s, 3H), 1.13 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 172.7, 171.8, 171.4, 138.8, 138.5, 137.4, 137.4, 130.2, 130.1, 130.0, 130.0, 129.8, 129.8, 128.5, 128.3, 128.2, 127.2, 73.3, 73.3, 63.9, 61.9, 61.7, 54.8, 54.2, 54.1, 47.6, 47.2, 40.9, 40.9, 40.8, 33.8, 33.2, 32.2, 31.3, 31.2, 31.1, 30.8, 30.2, 30.1, 28.7, 28.7, 28.7, 25.3, 23.9, 21.3, 21.0. Data for **8f**:  $R_f = 0.24$  (1:1 hexanes/EtOAc). Data for **9f**:  $R_f = 0.38$  (1:1 hexanes/EtOAc).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.34-7.29 (m, 1H), 7.20-7.12 (m, 3H), 7.08 (d, 1H,  $J = 7.2$  Hz), 6.00 (d, 1H,  $J = 8.8$  Hz), 5.33 (d, 1H,  $J = 8.8$  Hz), 5.14-5.07 (m, 1H), 4.57-4.47 (m, 1H), 4.06-4.02 (m, 1H), 3.28 (t, 1H,  $J = 12.8$  Hz), 2.85-2.79 (m, 2H), 2.76 (s, 3H), 2.34-2.26 (m, 1H), 2.01-1.90 (m, 2H), 1.87-1.73 (m, 6H), 1.47 (s, 9H), 1.35 (s, 3H), 1.30 (d, 3H,  $J = 7.2$  Hz), 1.25-1.20 (m, 1H), 1.15 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 171.5, 170.6, 170.6, 169.3, 137.9, 136.3, 129.4, 129.1, 129.0, 127.5, 126.3, 73.0, 62.8, 53.9, 47.8, 46.5, 39.9, 39.8, 33.3, 32.7, 30.6, 30.1, 29.3, 28.5, 28.5, 24.6, 19.8. HRMS calcd for  $\text{C}_{30}\text{H}_{44}\text{N}_4\text{O}_5\text{S}$ : 595.2925, found 595.2922.



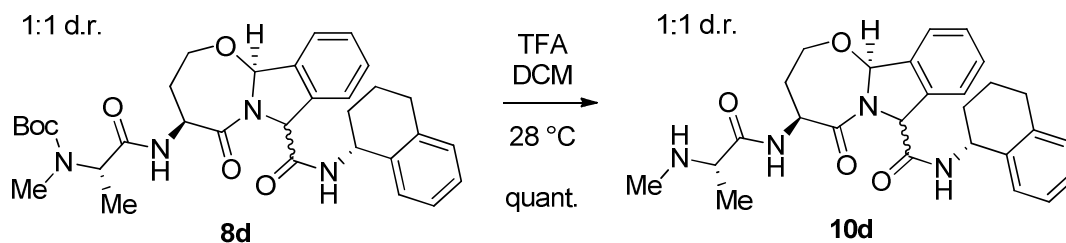
**(4*S*,7*S*,9*aS*)-4-((*S*)-2-(Methylamino)propanamido)-5-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)octahydropyrrolo[2,1-*b*][1,3]oxazepine-7-carboxamide (10a).** To a solution of **8a** (30 mg, 0.057 mmol, 1 equiv, ~3:1 d.r.) in DCM (2 mL) was added TFA (35  $\mu$ L, 0.454 mmol, 8 equiv). After stirring for 20 h at 23 °C, the solution was concentrated. The product was eluted through a short plug (~400 mg) of Silicycle<sup>®</sup> TMA-chloride ion exchange resin with MeOH to yield **10a**•HCl (26 mg, quantitative) as the major diastereomer (~3:1). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$ : 7.38-7.35 (m, 1H), 7.15-7.06 (m, 3H), 5.41-5.38 (m, 1H), 5.09-5.03 (m, 1H), 4.42 (t, 1H,  $J$  = 6.4 Hz), 4.15 (dt, 1H,  $J$  = 2.8, 12.8 Hz), 4.04-3.96 (m, 1H), 3.95-3.89 (m, 1H), 2.86-2.71 (m, 2H), 2.67 (s, 3H), 2.32-2.25 (m, 1H), 2.12 (q, 2H,  $J$  = 7.2 Hz), 2.06-1.96 (m, 2H), 1.94-1.85 (m, 1H), 1.85-1.74 (m, 2H), 1.58 (d, 3H,  $J$  = 7.2 Hz); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD)  $\delta$ : 173.4, 172.7, 172.7, 172.2, 169.6, 169.3, 138.6, 138.5, 137.8, 137.7, 130.0, 130.0, 129.6, 129.2, 128.2, 128.1, 127.1, 91.0, 71.3, 71.2, 62.4, 62.4, 58.4, 58.3, 54.4, 54.2, 34.0, 33.6, 33.3, 33.2, 31.8, 31.3, 31.2, 30.2, 30.2, 28.2, 28.0, 21.7, 21.6, 16.4, 16.4. HRMS calcd for C<sub>23</sub>H<sub>33</sub>N<sub>4</sub>O<sub>4</sub>: 429.2496, found 429.2495.



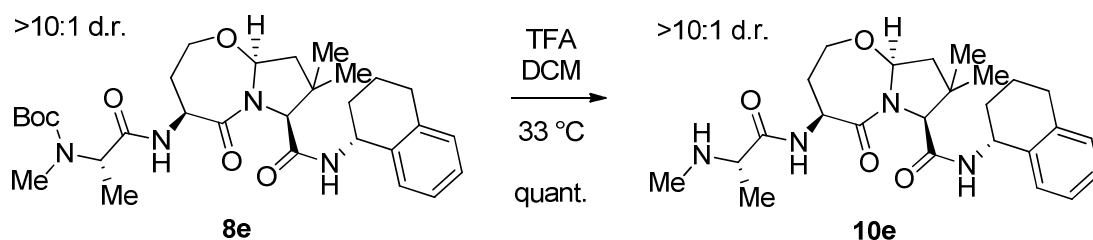
**(4*S*,7*S*,9*aS*)-4-((*S*)-2-(Methylamino)propanamido)-*N*-(naphthalen-1-yl)-5-oxooctahydropyrrolo[2,1-*b*][1,3]oxazepine-7-carboxamide (10b).** To a solution of **8b** (12 mg, 0.023 mmol, 1 equiv, ~6:1 d.r.) in DCM (1 mL) was added TFA (14  $\mu$ L, 0.183 mmol, 8 equiv). After stirring for 20 h at 23 °C, the solution was concentrated to yield **10b**•TFA (12 mg, quantitative) as the major diastereomer (~6:1). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$ : 8.12-8.08 (m, 1H), 7.92-7.88 (m, 1H), 7.79 (d, 1H,  $J$  = 8.4 Hz), 7.67 (dd, 1H,  $J$  = 1.2, 7.2 Hz), 7.56-7.45 (m, 3H), 5.48 (q, 1H,  $J$  = 2.8 Hz), 4.99 (d, 1H,  $J$  = 12.0 Hz), 4.75 (t, 2H,  $J$  = 6.8 Hz), 4.21 (dt, 1H,  $J$  = 2.8, 12.4 Hz), 4.10-4.00 (m, 1H), 3.96-3.87 (m, 1H), 2.68 (s, 3H), 2.44-2.29 (m, 2H), 2.22-2.07 (m, 2H), 1.83 (dd, 1H,  $J$  = 2.0, 14.4 Hz), 1.60 (d, 3H,  $J$  = 6.8 Hz); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD)  $\delta$ : 172.9, 172.5, , 169.7, 135.7, 134.0, 129.9, 127.3, 127.2, 126.5, 123.5, 91.1, 71.4, 62.8, 58.4, 54.3, 49.0, 33.8, 33.4, 31.8, 28.1, 16.4. HRMS calcd for C<sub>23</sub>H<sub>28</sub>N<sub>4</sub>O<sub>4</sub>Na: 447.20028, found 447.20189.



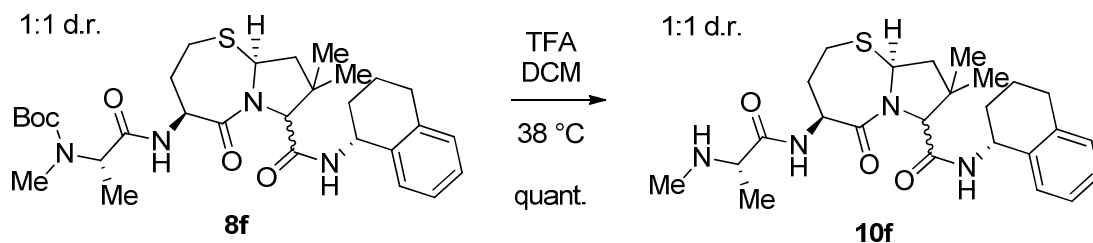
**(4*S*,7*S*,9*aS*)-4-((*S*)-2-(Methylamino)propanamido)-5-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)octahydropyrrolo[2,1-*b*][1,3]thiazepine-7-carboxamide (10c).** Same procedure as above (**10a**) using **8c** (90 mg, 0.165 mmol, 1 equiv, >8:1 d.r.) and TFA (126  $\mu\text{L}$ , 1.65 mmol, 10 equiv) in DCM (4 mL). After stirring for 20 h at 32  $^\circ\text{C}$ , the solution was concentrated. The product was eluted through a short plug (~500 mg) of Silicyle<sup>®</sup> TMA-chloride ion exchange resin with MeOH to yield **10c**•HCl (79 mg, quantitative) as the major diastereomer (>8:1 d.r.). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$ : 7.39-7.34 (m, 1H), 7.17-7.05 (m, 4H), 5.46-5.39 (m, 1H), 5.07 (t, 1H,  $J = 6.8$  Hz), 4.77 (dd, 1H,  $J = 2.0, 11.2$  Hz), 4.57 (dd, 1H,  $J = 5.2, 7.6$  Hz), 3.94-3.87 (m, 1H), 3.29-3.21 (m, 1H), 3.02 (ddd, 1H,  $J = 2.8, 6.0, 14.4$  Hz), 2.82-2.75 (m, 2H), 2.66 (s, 3H), 2.60-2.49 (m, 1H), 2.25-2.17 (m, 2H), 2.15-2.09 (m, 1H), 2.05-1.95 (m, 2H), 1.95-1.74 (m, 4H), 1.53 (d, 3H,  $J = 7.2$  Hz); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD)  $\delta$ : 172.3, 171.9, 169.5, 138.7, 138.5, 137.6, 137.3, 130.2, 130.0, 129.9, 129.5, 128.4, 128.3, 127.2, 127.1, 63.9, 63.4, 63.1, 58.4, 58.3, 55.1, 54.2, 54.1, 34.1, 33.3, 31.8, 31.8, 31.3, 31.0, 30.1, 30.1, 28.8, 28.5, 21.5, 21.1, 16.4, 16.3. HRMS calcd for C<sub>23</sub>H<sub>33</sub>N<sub>4</sub>O<sub>3</sub>S: 445.2268, found 445.2267.



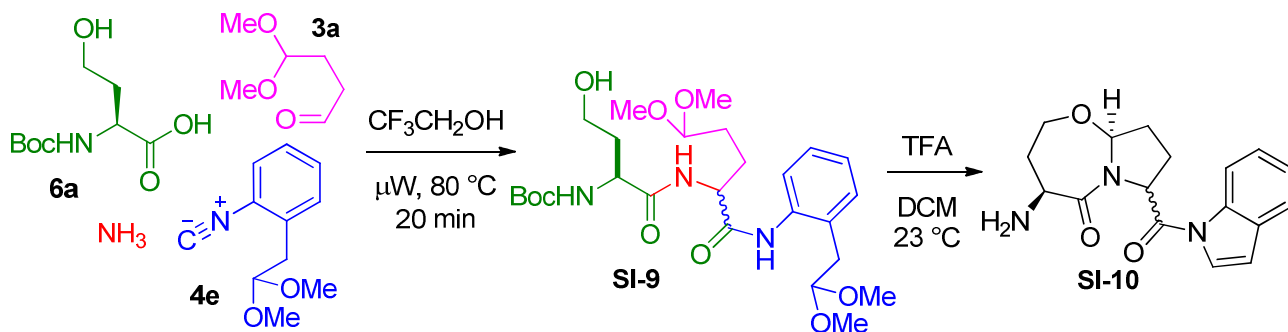
**(4*S*,11*bS*)-4-((*S*)-2-(Methylamino)propanamido)-5-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)-2,3,4,5,7,11*b*-hexahydro-[1,3]oxazepino[2,3-*a*]isoindole-7-carboxamide (10d).** Same procedure as above (**10b**) using **8d** (38 mg, 0.066 mmol, 1 equiv, 1:1 d.r.) and TFA (40  $\mu\text{L}$ , 0.527 mmol, 8 equiv) in DCM (2 mL). After stirring for 20 h at 28  $^\circ\text{C}$ , the solution was concentrated to yield **10d**•TFA (38 mg, quantitative) as a 1:1 diastereomixture. Data for the 1:1 diastereomixture: <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$ : 7.53-7.45 (m, 7H), 7.39-7.35 (m, 1H), 7.26 (d, 1H,  $J = 7.2$  Hz), 7.16-7.07 (d, 2H,  $J = 2.0$  Hz), 6.53 (d, 1H,  $J = 1.6$  Hz), 6.47 (s, 1H), 5.57 (d, 1H,  $J = 1.6$  Hz), 5.47 (s, 1H), 5.11-5.03 (m, 3H), 4.66 (dd, 1H,  $J = 9.2, 11.2$  Hz), 4.46 (td, 1H,  $J = 2.0, 9.2$  Hz), 4.35-4.27 (m, 4H), 3.97 (q, 1H,  $J = 6.8$  Hz), 3.88 (q, 1H,  $J = 7.2$  Hz), 2.89-2.74 (m, 3H), 2.70 (s, 3H), 2.69 (s, 3H), 2.62-2.54 (m, 1H), 2.33 (tt, 1H,  $J = 1.6, 10.8$  Hz), 2.02-1.92 (m, 6H), 1.85-1.76 (m, 3H), 1.62 (d, 3H,  $J = 7.2$  Hz), 1.55 (d, 3H,  $J = 7.2$  Hz), 1.54 (d, 3H,  $J = 7.2$  Hz); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD)  $\delta$ : 176.8, 172.2, 172.0, 171.0, 170.5, 170.4, 169.7, 169.2, 162.8, 162.4, 138.7, 138.6, 138.5, 138.4, 137.7, 137.5, 136.9, 136.6, 131.3, 131.3, 130.3, 130.2, 130.1, 130.0, 129.7, 129.5, 128.2, 128.2, 127.1, 126.3, 123.2, 123.2, 101.3, 93.2, 92.4, 72.1, 72.0, 67.3, 67.2, 66.9, 58.4, 58.4, 58.2, 54.6, 54.4, 50.2, 34.2, 33.5, 31.8, 31.8, 31.4, 31.0, 30.2, 30.2, 29.2, 21.6, 21.4, 16.4, 16.4, 16.2. HRMS calcd for C<sub>27</sub>H<sub>33</sub>N<sub>4</sub>O<sub>4</sub>: 477.2496, found 477.2493.



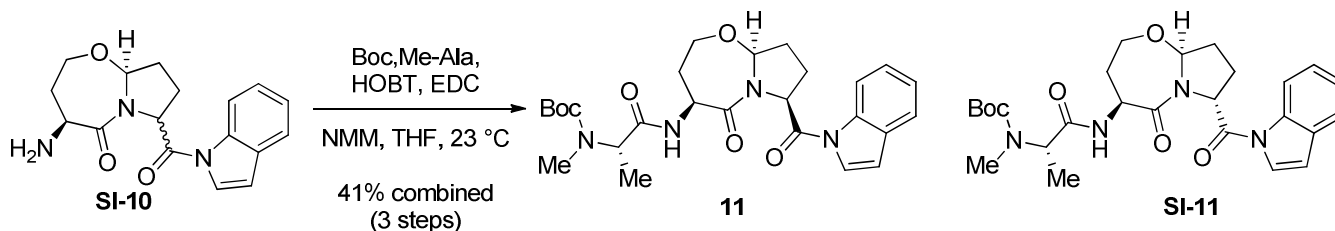
**(4*S*,7*S*,9*aS*)-8,8-Dimethyl-4-((*S*)-2-(methylamino)propanamido)-5-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)octahydropyrrolo[2,1-*b*][1,3]oxazepine-7-carboxamide (10e).** Same procedure as above (10b) using **8e** (25 mg, 0.045 mmol, 1 equiv, 10:1 d.r.) and TFA (35  $\mu$ L, 0.449 mmol, 10 equiv) in DCM (1 mL). After stirring for 20 h at 33  $^\circ$ C, the solution was concentrated to yield **10e**•TFA (25 mg, quantitative) as the major diastereomer (10:1 d.r.).  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 8.15 (d, 1H,  $J = 8.4$  Hz), 7.32 (d, 1H,  $J = 6.4$  Hz), 7.17-7.07 (m, 3H), 5.44 (t, 1H,  $J = 6.4$  Hz), 5.10 (q, 1H,  $J = 6.8$  Hz), 4.14 (dt, 1H,  $J = 3.2, 12.0$  Hz), 4.08 (s, 1H), 3.99-3.91 (m, 2H), 2.80 (p, 2H,  $J = 6.0$  Hz), 2.68 (s, 3H), 2.20 (dd, 1H,  $J = 6.4, 13.2$  Hz), 2.08-1.96 (m, 3H), 1.89-1.77 (m, 4H), 1.58 (d, 3H,  $J = 7.2$  Hz);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 172.2, 171.5, 169.6, 138.5, 137.6, 130.1, 129.7, 128.3, 127.1, 117.5, 114.6, 90.5, 71.7, 71.3, 58.4, 54.2, 47.0, 40.1, 33.2, 31.8, 31.4, 30.1, 29.3, 24.2, 21.4, 16.3. HRMS calcd for  $\text{C}_{25}\text{H}_{37}\text{N}_4\text{O}_4$ : 457.2809, found 457.2811.



**(4*S*,9*aS*)-8,8-Dimethyl-4-((*S*)-2-(methylamino)propanamido)-5-oxo-*N*-((*R*)-1,2,3,4-tetrahydronaphthalen-1-yl)octahydropyrrolo[2,1-*b*][1,3]thiazepine-7-carboxamide (10f).** Same procedure as above (10a) using **8f** (62 mg, 0.108 mmol, 1 equiv, 1:1 d.r.) and TFA (66  $\mu$ L, 0.866 mmol, 8 equiv) in DCM (3 mL). After stirring for 20 h at 38  $^\circ$ C, the solution was concentrated. The product was eluted through a short plug (~500 mg) of Silicyle<sup>®</sup> TMA-chloride ion exchange resin with MeOH to yield **10f**•HCl (54 mg, quantitative) as a 1:1 diastereomixture.  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 7.34-7.27 (m, 2H), 7.18-7.06 (m, 7H), 5.54-5.45 (m, 1H), 5.41 (t, 1H,  $J = 8.0$  Hz), 5.11-5.06 (m, 1H), 5.06-5.01 (m, 1H), 4.77-4.71 (m, 2H), 4.23 (s, 1H), 4.16 (s, 1H), 3.97-3.89 (m, 2H), 3.29-3.19 (m, 2H), 2.93-2.84 (m, 2H), 2.78 (dd, 4H,  $J = 6.4, 12.8$  Hz), 2.68 (s, 6H), 2.32-2.21 (m, 3H), 2.01-1.75 (m, 12H), 1.55 (d, 3H,  $J = 7.2$  Hz), 1.54-1.50 (m, 2H), 1.47 (d, 3H,  $J = 6.8$  Hz), 1.40-1.37 (m, 2H), 1.16 (s, 6H), 1.14 (s, 3H), 1.13 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 172.4, 172.3, 171.8, 171.4, 169.3, 168.9, 138.8, 138.5, 137.4, 137.4, 130.1, 130.1, 129.8, 128.3, 127.1, 127.0, 73.4, 63.8, 61.8, 58.3, 55.1, 54.4, 40.9, 40.9, 40.7, 33.6, 32.1, 31.8, 31.7, 31.3, 31.1, 30.9, 30.2, 30.1, 28.7, 23.9, 21.3, 21.0, 16.3, 16.2. HRMS calcd for  $\text{C}_{25}\text{H}_{37}\text{N}_4\text{O}_3\text{S}$ : 473.2581, found 473.2579.



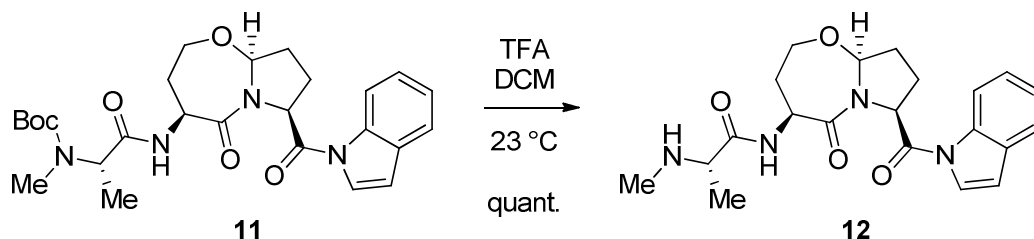
**(4*S*,9*aS*)-4-Amino-7-(1*H*-indole-1-carbonyl)hexahydropyrrolo[2,1-*b*][1,3]oxazepin-5(2*H*)-one (SI-10).** Same procedure as above (**7a**) with Boc-*N*-HSer-OH (**6a**) (313 mg, 1.43 mmol, 1.0 equiv), aldehyde **3a** (198 mg, 1.50 mmol, 1.05 equiv), isocyanide **4e** (273 mg, 1.43 mmol, 1.0 equiv) and 7 M ammonia in MeOH (408  $\mu$ L, 2.85 mmol, 2.0 equiv) in TFE (5 mL). The resultant oil **SI-9** was combined with TFA (1.09 mL, 14.3 mmol, 10 equiv) in DCM (5 mL) and stirred at 23 °C for 14 h. The mixture was concentrated in vacuo and **SI-10** used without further purification in the next step.



**tert-Butyl ((2*S*)-1-(((4*S*,9*aS*)-7-(1*H*-indole-1-carbonyl)-5-oxooctahydropyrrolo[2,1-*b*][1,3]oxazepin-4-yl)amino)-1-oxopropan-2-yl)(methyl)carbamate (**11**) and (**SI-11**).** Same procedure as above (**8a**) using crude **SI-10**•TFA (611 mg, 1.43 mmol, 1.0 equiv), Boc-*N*-Me-Ala-OH (291 mg, 1.43 mmol, 1.0 equiv), HOBT·xH<sub>2</sub>O (241 mg, 1.57 mmol, 1.1 equiv), NMM (786  $\mu$ L, 7.15 mmol, 5 equiv [to soak up xs TFA]) and EDC·HCl (288 mg, 1.50 mmol, 1.05 equiv) in THF (15 mL). The resultant oil was purified by flash chromatography on silica gel (2:1→1:1→1:4 hexanes/EtOAc) to yield, after 3 steps, separated diastereomers **11** (150 mg, 21%) and **SI-11** (144 mg, 20%). Data for **11**:  $R_f$  = 0.27 (1:3 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.51 (d, 1H,  $J$  = 8.4 Hz), 7.57 (d, 1H,  $J$  = 8.0 Hz), 7.50 (d, 1H,  $J$  = 4.0 Hz), 7.35 (t, 1H,  $J$  = 8.4 Hz), 7.28 (t, 1H,  $J$  = 7.6 Hz), 7.16 (s, 1H), 6.69 (d, 1H,  $J$  = 3.6 Hz), 5.35 (dd, 1H,  $J$  = 3.6, 6.4 Hz), 5.28 (dd, 1H,  $J$  = 4.8, 8.0 Hz), 4.80 (dd, 1H,  $J$  = 6.0, 10.8 Hz), 4.75-4.65 (m, 1H), 4.31 (dt, 1H,  $J$  = 3.2, 12.8 Hz), 4.12 (q, 1H,  $J$  = 7.2 Hz), 4.05 (t, 1H,  $J$  = 13.2 Hz), 2.76 (s, 3H), 2.44-2.31 (m, 2H), 2.30-2.19 (m, 2H), 2.05-1.98 (m, 2H), 1.42 (s, 9H), 1.34 (d, 3H,  $J$  = 7.2 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 171.1, 170.8, 168.8, 135.9, 130.2, 125.3, 124.0, 124.0, 120.8, 117.0, 110.0, 89.7, 80.6, 80.6, 77.2, 70.8, 64.3, 60.4, 59.7, 53.0, 32.6, 30.3, 28.3, 28.3, 28.3, 27.2, 21.0. Data for **SI-11**:  $R_f$  = 0.50 (1:3 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.38 (s, 1H), 7.57 (d, 1H,  $J$  = 7.6 Hz), 7.49 (d, 1H,  $J$  = 4.0 Hz), 7.35 (t, 1H,  $J$  = 7.2 Hz), 7.28 (d, 1H,  $J$  = 7.6 Hz), 7.18 (s, 1H), 6.71 (d, 1H,  $J$  = 3.6 Hz), 5.44-5.39 (m, 2H), 4.88 (dd, 1H,  $J$  = 5.6, 11.2 Hz), 4.75-4.69 (m, 1H), 4.47 (t, 2H,  $J$  = 8.8 Hz), 4.31-4.24 (m, 1H), 4.17 (dt, 1H,  $J$  = 3.2, 12.8 Hz), 4.13-4.04 (m, 1H), 3.72-3.66 (m, 1H), 3.56-3.48 (m, 1H), 2.79 (s, 3H), 2.66-2.54 (m, 1H), 2.37 (sept, 1H,  $J$  = 6.8 Hz), 2.21-2.06 (m, 4H), 1.81 (qd, 1H,  $J$  = 3.6, 14.0 Hz), 1.67-1.58 (m, 1H), 1.43 (s, 9H), 1.33 (d, 3H,  $J$  =

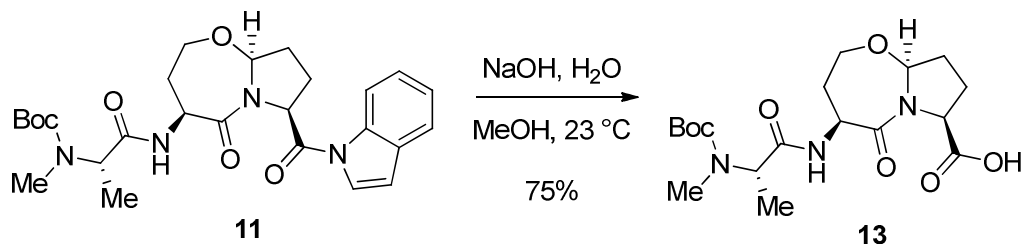


7.2 Hz);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 172.3, 171.1, 168.7, 135.8, 130.2, 125.4, 124.1, 123.8, 121.0, 116.7, 110.3, 89.6, 70.7, 65.8, 60.0, 53.0, 49.1, 33.1, 32.2, 30.4, 28.4, 28.4, 28.4, 26.9. Also included below are 1D-NOESY and DEPT-135 spectra for **11** and the HSQC spectra for both **11** and **SI-11**. HRMS calcd for  $\text{C}_{26}\text{H}_{34}\text{N}_4\text{O}_6\text{Na}$ : 521.2371, found 521.2372.



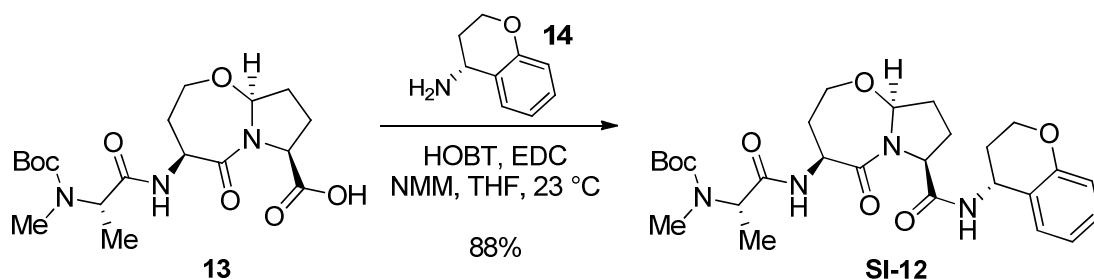
**(S)-N-((4S,7S,9aS)-7-(1H-Indole-1-carbonyl)-5-oxooctahydropyrrolo[2,1-b][1,3]oxazepin-4-yl)-2-**

**(methylamino)propanamide (12).** Same procedure as above (**10b**) using **11** (52 mg, 0.104 mmol, 1 equiv) and TFA (64  $\mu\text{L}$ , 0.834 mmol, 8 equiv) in DCM (2 mL). After stirring for 20 h at 23  $^\circ\text{C}$ , the solution was concentrated to yield **12**·TFA (53 mg, quantitative) as a single diastereomer.  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 8.39 (d, 1H,  $J$  = 8.0 Hz), 7.84 (d, 1H,  $J$  = 4.0 Hz), 7.58 (d, 1H,  $J$  = 7.2 Hz), 7.33-7.24 (m, 2H), 6.73 (d, 1H,  $J$  = 4.0 Hz), 5.50-5.46 (m, 1H), 5.34 (t, 1H,  $J$  = 6.8 Hz), 4.26 (dt, 1H,  $J$  = 3.2, 12.4 Hz), 4.10-4.02 (m, 1H), 3.92-3.84 (m, 2H), 2.65 (s, 3H), 2.28 (qd, 1H,  $J$  = 3.6, 12.4 Hz), 2.19-2.05 (m, 2H), 1.86 (d, 1H,  $J$  = 14.0 Hz), 1.55 (dd, 2H,  $J$  = 4.0, 7.2 Hz), 1.50 (d, 3H,  $J$  = 7.2 Hz);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$ : 172.2, 171.0, 169.6, 137.2, 131.9, 126.0, 126.0, 125.0, 122.0, 117.4, 110.7, 90.9, 71.4, 61.5, 58.3, 54.4, 49.0, 33.7, 33.2, 31.7, 28.3, 16.3. HRMS calcd for  $\text{C}_{21}\text{H}_{26}\text{N}_4\text{O}_4\text{Na}$ : 421.18463, found 421.18593.

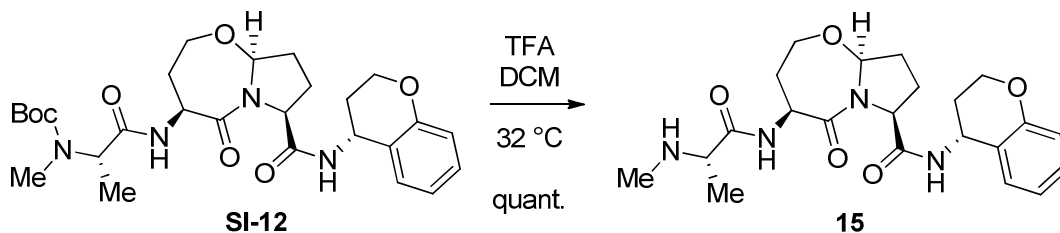


**(4S,7S,9aS)-4-((S)-2-((tert-Butoxycarbonyl)(methyl)amino)propanamido)-5-oxooctahydropyrrolo[2,1-**

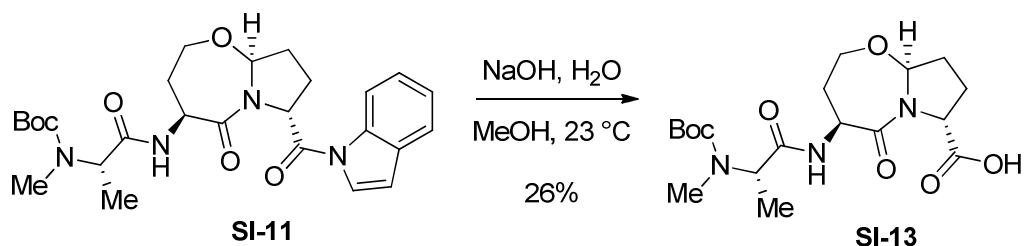
**b][1,3]oxazepine-7-carboxylic acid (13).** To a solution of **11** (142 mg, 0.285 mmol, 1.0 equiv) in MeOH (6 mL) was added 1M NaOH (1 mL). After stirring for 3 h, the methanol was removed in vacuo. Then EtOAc (10 mL) and 1 M NaOH (8 mL) were added and an extraction was performed, with the organic layer being discarded. The aqueous layer was acidified with 3M HCl to  $\text{pH} \leq 2$  and then extracted with DCM (3 x 5 mL). The organics were dried over sodium sulfate, filtered and concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (1:3 hexanes/EtOAc  $\rightarrow$  DCM  $\rightarrow$  5% MeOH/DCM) to yield **13** as a colorless oil (85 mg, 75%) as a single diastereomer.  $R_f$  = 0.17 (7% MeOH/DCM).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.30 (bs, 1H), 5.22 (m, 1H), 4.77 (t, 1H,  $J$  = 8.0 Hz), 4.52-4.46 (m, 1H), 4.14 (d, 1H,  $J$  = 12.8 Hz), 3.95 (t, 1H,  $J$  = 12.0 Hz), 2.78 (s, 3H), 2.32-2.18 (m, 2H), 2.13-2.02 (m, 2H), 2.00-1.85 (m, 2H), 1.44 (s, 9H), 1.33 (d, 3H,  $J$  = 7.2 Hz);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 171.5, 171.5, 156.2, 156.1, 89.8, 80.8, 80.7, 70.7, 59.7, 52.9, 32.7, 30.4, 30.4, 28.4, 28.4, 26.5, 26.5, 14.2. HRMS calcd for  $\text{C}_{18}\text{H}_{29}\text{N}_3\text{O}_7\text{Na}$ : 422.18977, found 422.19015.



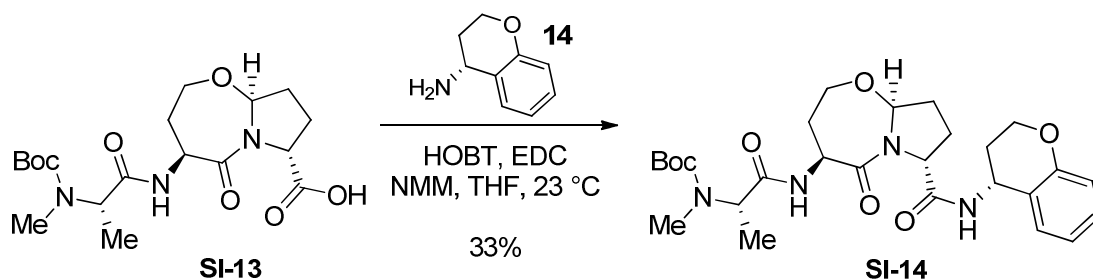
**tert-Butyl ((S)-1-(((4S,7S,9aS)-7-((R)-chroman-4-ylcarbamoyl)-5-oxooctahydropyrrolo[2,1-b][1,3]oxazepin-4-yl)amino)-1-oxopropan-2-yl)(methyl)carbamate (SI-12).** To a solution of **13** (50 mg, 0.125 mmol, 1.0 equiv), (*R*)-chroman-4-ylamine·HCl (**14**, 23 mg, 0.125 mmol, 1.0 equiv), HOBT·xH<sub>2</sub>O (21 mg, 0.138 mmol, 1.1 equiv) and NMM (41 μL, 0.376 mmol, 3 equiv) in THF (5 mL) at 0 °C was added EDC·HCl (25 mg, 0.131 mmol, 1.05 equiv). After 30 min the cold bath was removed. The solution stirred for 14 h and then was quenched with saturated aqueous NaHCO<sub>3</sub> (15 mL), extracted with EtOAc (2 x 10 mL), dried over sodium sulfate and then concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (1:3 hexanes/EtOAc) to yield **SI-12** (58 mg, 88%) as a single diastereomer. *R<sub>f</sub>* = 0.11 (1:2 hexanes/EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 7.16-7.10 (m, 3H), 6.91 (d, 1H, *J* = 7.2 Hz), 6.86-6.77 (m, 2H), 5.22 (t, 1H, *J* = 6.0 Hz), 5.12 (q, 1H, *J* = 6.8 Hz), 4.68 (dd, 1H, *J* = 6.0, 11.2 Hz), 4.59 (d, 1H, *J* = 7.2 Hz), 4.22 (td, 1H, *J* = 2.8, 7.2 Hz), 4.15-4.08 (m, 1H), 4.06-4.01 (m, 1H), 3.92 (t, 1H, *J* = 12.4 Hz), 2.74 (s, 3H), 2.41-2.37 (m, 2H), 2.25-2.17 (m, 1H), 2.16-2.07 (m, 1H), 2.02 (pd, 1H, *J* = 2.8, 7.2 Hz), 1.95-1.84 (m, 2H), 1.61-1.45 (m, 1H), 1.42 (s, 9H), 1.31 (d, 3H, *J* = 7.2 Hz); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 171.5, 170.1, 155.0, 129.3, 128.9, 122.3, 120.7, 117.2, 90.2, 77.2, 70.6, 63.6, 60.5, 52.6, 43.8, 32.7, 32.5, 30.2, 29.0, 28.4, 25.9. HRMS calcd for C<sub>27</sub>H<sub>38</sub>N<sub>4</sub>O<sub>7</sub>Na: 553.26327, found 553.26399.



**(4S,7S,9aS)-N-((R)-Chroman-4-yl)-4-((S)-2-(methylamino)propanamido)-5-oxooctahydropyrrolo[2,1-b][1,3]oxazepine-7-carboxamide (15).** To a solution of **SI-12** (58 mg, 0.109 mmol, 1 equiv) in DCM (2 mL) was added TFA (83 μL, 1.09 mmol, 10 equiv). After stirring for 20 h at 32 °C, the solution was concentrated. The product was eluted through a short plug (~500 mg) of Silicycle® TMA-chloride ion exchange resin with MeOH to yield **15**·HCl (51 mg, quantitative) as a single diastereomer. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ: 7.33 (d, 1H, *J* = 7.6 Hz), 7.13 (t, 1H, *J* = 8.4 Hz), 6.86 (t, 1H, *J* = 7.2 Hz), 6.76 (d, 1H, *J* = 8.0 Hz), 5.39 (dd, 1H, *J* = 3.6, 6.8 Hz), 5.08 (t, 1H, *J* = 6.0 Hz), 4.40 (d, 1H, *J* = 6.8 Hz), 4.26-4.12 (m, 3H), 4.03-3.89 (m, 2H), 2.67 (s, 3H), 2.33-2.24 (m, 1H), 2.14-1.97 (m, 6H), 1.81 (dd, 1H, *J* = 2.0, 14.0 Hz), 1.58 (d, 3H, *J* = 6.8 Hz); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD) δ: 172.9, 172.2, 169.6, 156.4, 130.5, 130.0, 123.5, 121.6, 117.8, 91.0, 71.3, 64.6, 62.3, 58.4, 54.2, 49.0, 44.9, 33.6, 33.3, 31.8, 30.2, 28.0, 16.4. HRMS calcd for C<sub>22</sub>H<sub>31</sub>N<sub>4</sub>O<sub>5</sub>: 431.2289, found 431.2286.

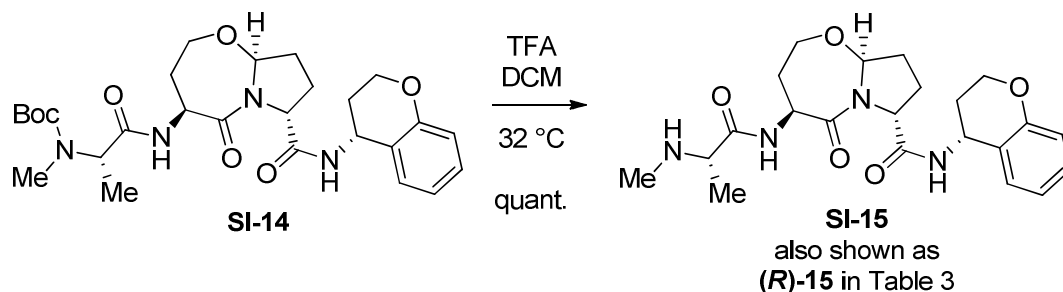


**(4*S*,7*R*,9*aS*)-4-((*S*)-2-((*tert*-butoxycarbonyl)(methyl)amino)propanamido)-5-oxooctahydropyrrolo[2,1-*b*][1,3]oxazepine-7-carboxylic acid (SI-13).** To a solution of **SI-11** (105 mg, 0.211 mmol, 1.0 equiv) in MeOH (4 mL) was added 1M NaOH (1 mL). After stirring for 3 h, the methanol was removed in vacuo. HPLC analysis of the crude reaction mixture revealed that diastereomer **SI-11** didn't react as cleanly as **11**. Then DCM (10 mL) and 1 M NaOH (8 mL) were added and an extraction was performed, with the organic layer being discarded. The aqueous layer was acidified with 3M HCl to pH  $\leq$  2 and then extracted with DCM (3 x 5 mL). The organics were dried over sodium sulfate, filtered and concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (1:3 hexanes/EtOAc $\rightarrow$ DCM $\rightarrow$ 5% MeOH/DCM) to yield **SI-13** as a colorless oil (22 mg, 26%) as a single diastereomer.  $R_f = 0.14$  (7% MeOH/DCM).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$ : 5.25 (d, 1H,  $J = 6.8$  Hz), 4.83 (dd, 1H,  $J = 5.6, 9.6$  Hz), 4.65 (d, 1H,  $J = 8.8$  Hz), 4.14-4.09 (m, 1H), 4.0 (t, 1H,  $J = 12.0$  Hz), 2.79 (s, 3H), 2.41-2.31 (m, 1H), 2.27-2.11 (m, 2H), 2.06-1.96 (m, 1H), 1.78 (qd, 1H,  $J = 3.6, 12.0$  Hz), 1.46 (s, 9H), 1.33 (d, 3H,  $J = 7.6$  Hz);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 173.9, 172.0, 171.3, 89.6, 80.9, 70.7, 60.6, 59.8, 53.1, 33.0, 32.5, 30.5, 28.5, 26.1, 21.2, 14.3, 14.1. HRMS calcd for  $\text{C}_{18}\text{H}_{29}\text{N}_3\text{O}_7\text{Na}$ : 422.18977, found 422.19015.



***tert*-Butyl ((*S*)-1-(((4*S*,7*R*,9*aS*)-7-((*R*)-chroman-4-ylcarbamoyl)-5-oxooctahydropyrrolo[2,1-*b*][1,3]oxazepin-4-yl)amino)-1-oxopropan-2-yl)(methyl)carbamate (SI-14).** To a solution of **SI-13** (21 mg, 0.0053 mmol, 1.0 equiv), (*R*)-chroman-4-ylamine $\cdot$ HCl (**14**, 10 mg, 0.0053 mmol, 1.0 equiv), HOBT $\cdot$ xH<sub>2</sub>O (9 mg, 0.0058 mmol, 1.1 equiv) and NMM (17  $\mu\text{L}$ , 0.0158 mmol, 3 equiv) in THF (3 mL) at 0  $^\circ\text{C}$  was added EDC $\cdot$ HCl (11 mg, 0.0055 mmol, 1.05 equiv). After 30 min the cold bath was removed. The solution stirred for 14 h and then was quenched with saturated aqueous  $\text{NaHCO}_3$  (10 mL), extracted with EtOAc (2 x 10 mL), dried over sodium sulfate and then concentrated in vacuo. The resultant oil was purified by flash chromatography on silica gel (1:1 $\rightarrow$ 1:3 hexanes/EtOAc) to yield **SI-14** (9 mg, 33%) as a single diastereomer.  $R_f = 0.51$  (1:3 hexanes/EtOAc).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.20-7.12 (m, 3H), 6.89 (t, 1H,  $J = 7.6$  Hz), 6.82 (d, 1H,  $J = 8.4$  Hz), 5.23-5.19 (m, 1H), 5.12-5.05 (m, 1H), 4.79-4.71 (m, 1H), 4.55 (d, 1H,  $J = 8.0$  Hz), 4.26-4.19 (m, 1H), 4.15-4.06 (m, 2H), 3.97 (t, 1H,  $J = 12.0$  Hz), 2.77 (s, 3H), 2.39-2.26 (m, 1H), 2.24-2.13 (m, 2H), 2.07-2.00 (m, 1H), 1.99-1.91 (m, 2H), 1.80-1.70 (m, 2H), 1.44 (s, 9H), 1.34 (d, 3H,  $J = 7.2$  Hz);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 172.3, 171.1, 169.9, 155.2, 129.4,

129.3, 122.0, 120.9, 117.3, 90.1, 70.6, 63.4, 61.1, 53.1, 43.8, 33.4, 32.7, 32.1, 29.8, 29.1, 28.5, 25.6, 22.8, 14.3.  
HRMS calcd for C<sub>27</sub>H<sub>38</sub>N<sub>4</sub>O<sub>7</sub>Na: 553.26327, found 553.26399.



**(4S,7R,9aS)-N-((R)-chroman-4-yl)-4-((S)-2-(methylamino)propanamido)-5-oxooctahydropyrrolo[2,1-b][1,3]oxazepine-7-carboxamide (SI-15).** To a solution of **SI-14** (58 mg, 0.109 mmol, 1 equiv) in DCM (2 mL) was added TFA (83  $\mu$ L, 1.09 mmol, 10 equiv). After stirring for 20 h at 32 °C, the solution was concentrated to yield **SI-15**•TFA (51 mg, quantitative) as a single diastereomer. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$ : 8.43 (d, 1H, *J* = 8.0 Hz), 7.15-7.09 (m, 2H), 6.85 (t, 1H, *J* = 8.0 Hz), 6.78-6.73 (m, 1H), 5.40 (d, 2H, *J* = 5.6 Hz), 5.10-5.04 (m, 1H), 4.99 (dd, 1H, *J* = 2.4, 11.2 Hz), 4.53 (d, 1H, *J* = 9.2 Hz), 4.21 (t, 2H, *J* = 5.2 Hz), 4.14 (dt, 1H, *J* = 3.2, 13.2 Hz), 4.05-3.96 (m, 1H), 3.91 (q, 1H, *J* = 7.2 Hz), 2.67 (s, 3H), 2.44-2.31 (m, 1H), 2.30-2.18 (m, 1H), 2.16-2.07 (m, 1H), 2.04-1.95 (m, 3H), 1.93-1.81 (m, 2H), 1.52 (d, 3H, *J* = 6.8 Hz); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD)  $\delta$ : 173.5, 172.7, 169.3, 156.5, 130.2, 129.9, 123.5, 121.6, 117.9, 91.1, 71.2, 64.6, 62.3, 58.3, 54.4, 44.9, 34.0, 33.3, 31.8, 30.1, 28.2, 16.4. HRMS calcd for C<sub>22</sub>H<sub>30</sub>N<sub>4</sub>O<sub>5</sub>Na: 453.21084, found 453.21280.

## References

- (1) Nikolovska-Coleska, Z.; Wang, R.; Fang, X.; Pan, H.; Tomita, Y.; Li, P.; Roller, P. P.; Krajewski, K.; Saito, N. G.; Stuckey, J. A.; Wang, S. *Analytical Biochemistry* **2004**, *332*, 261.
- (2) O'Boyle, N.; Banck, M.; James, C.; Morley, C.; Vandermeersch, T.; Hutchison, G. *Journal of Cheminformatics* **2011**, *3*, 33.
- (3) Gilley, C. B.; Buller, M. J.; Kobayashi, Y. *Org. Lett.* **2007**, *9*, 3631.
- (4) Griesbaum, K.; Jung, I. C.; Mertens, H. *J. Org. Chem.* **1990**, *55*, 6024.
- (5) Ueda, M.; Kawai, S.; Hayashi, M.; Naito, T.; Miyata, O. *J. Org. Chem.* **2010**, *75*, 914.

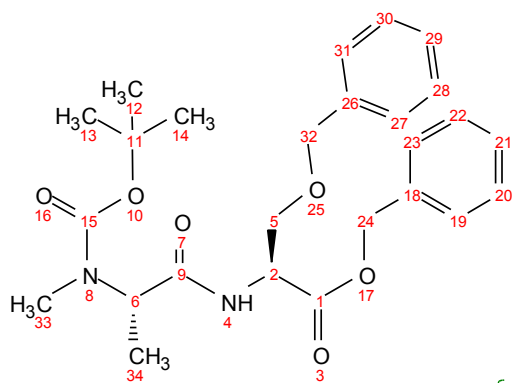
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7.202  
7.187

5.227  
5.196  
5.170  
5.140  
4.776  
4.755  
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4.431  
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4.377  
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3.678  
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3.654  
3.646

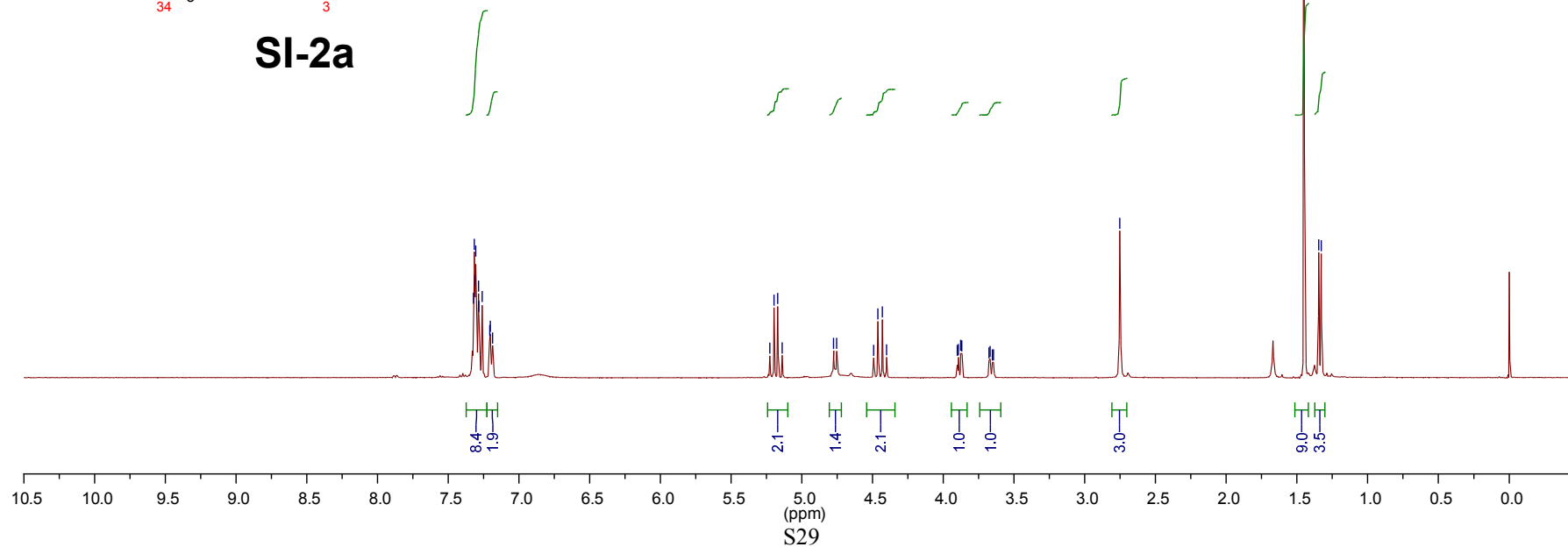
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1.329

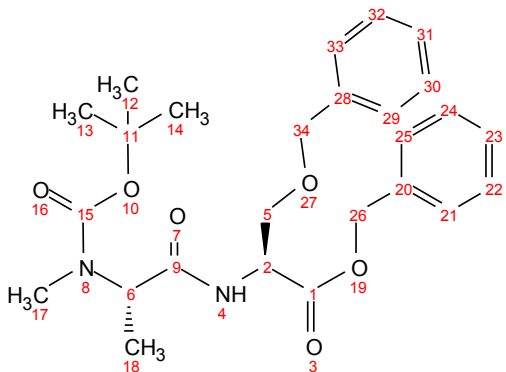
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Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



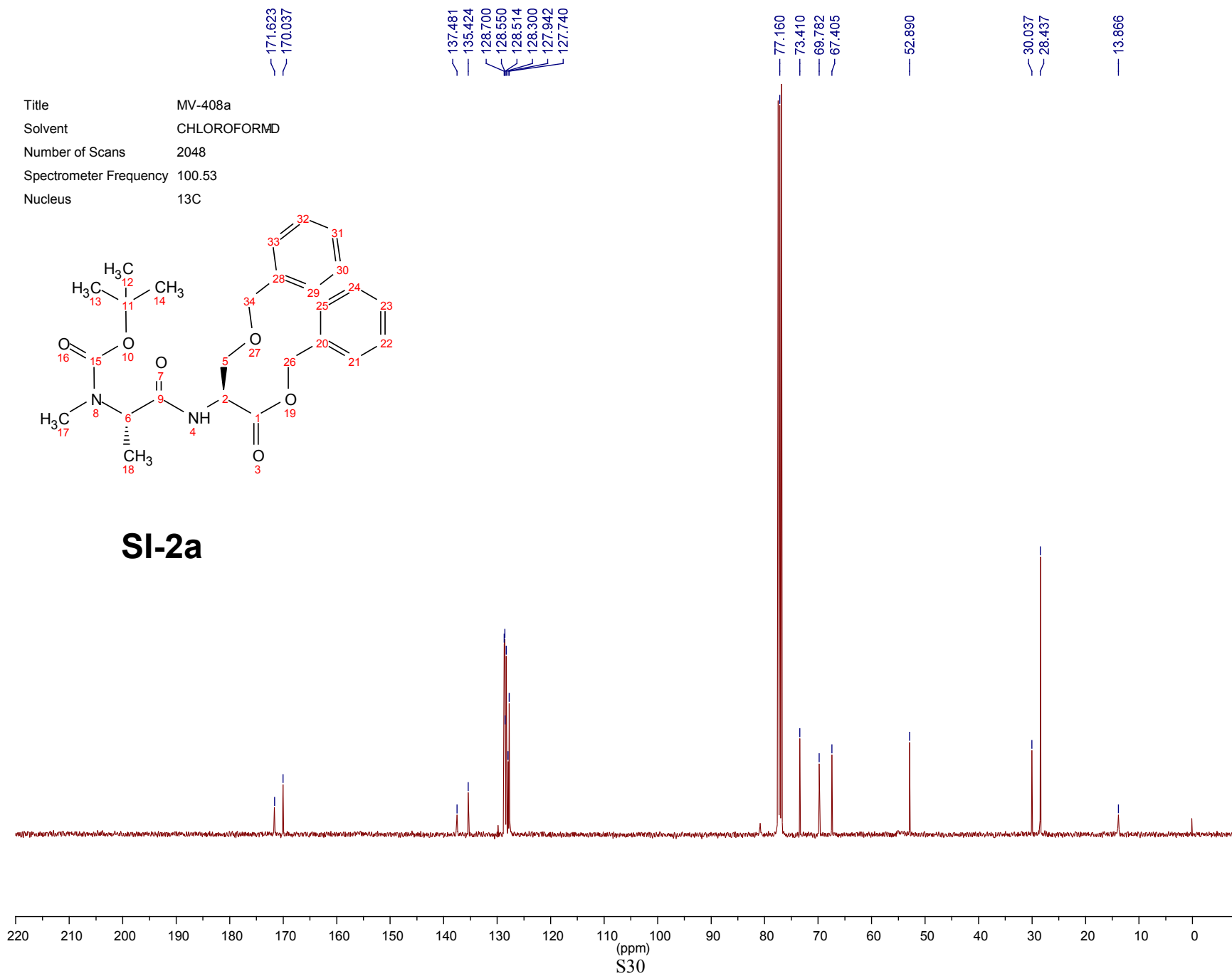
SI-2a



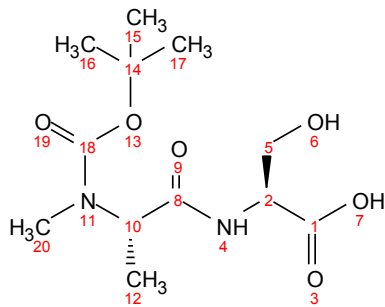
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Nucleus <sup>13</sup>C



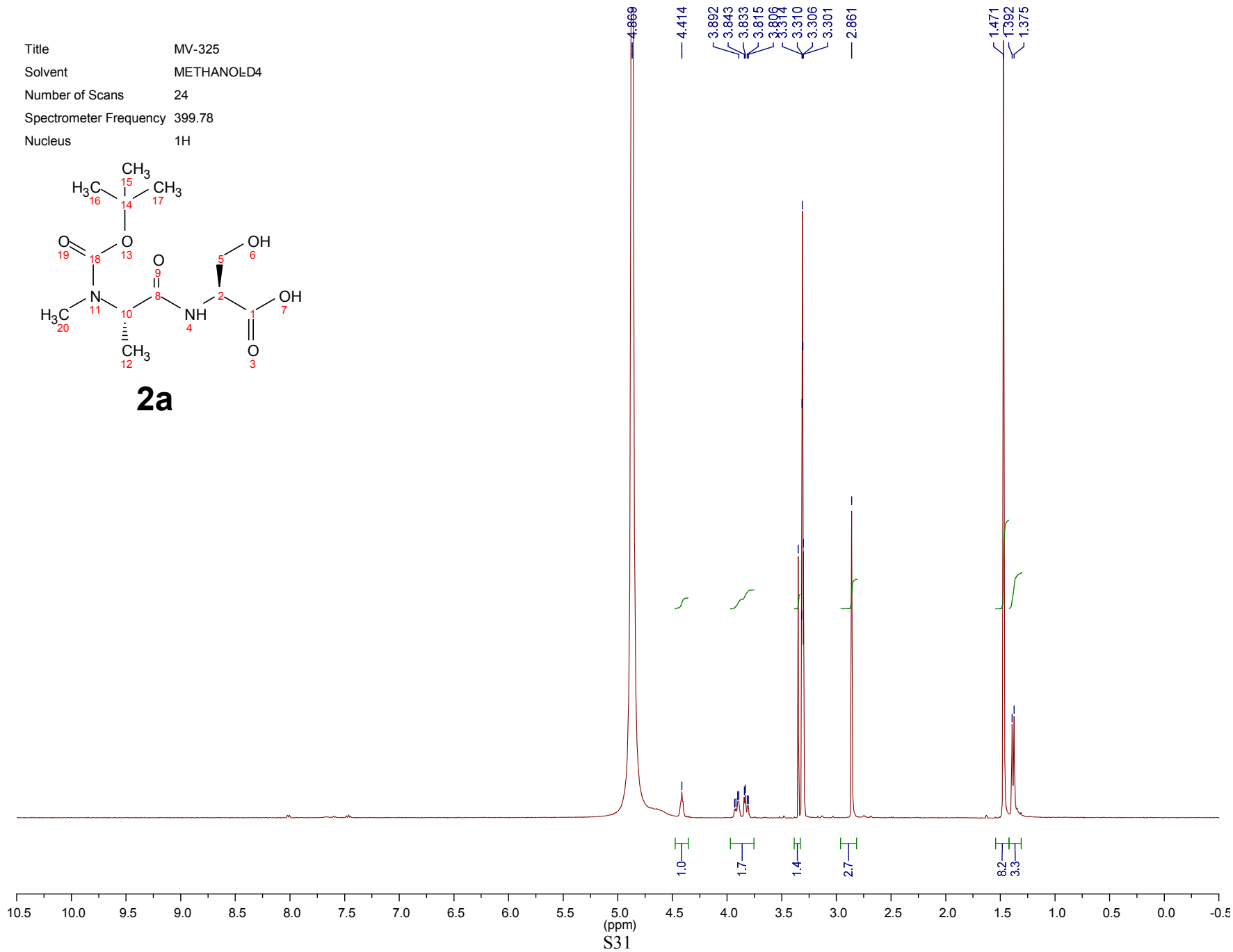
SI-2a



Title MV-325  
Solvent METHANOL-D4  
Number of Scans 24  
Spectrometer Frequency 399.78  
Nucleus 1H



**2a**



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81.029

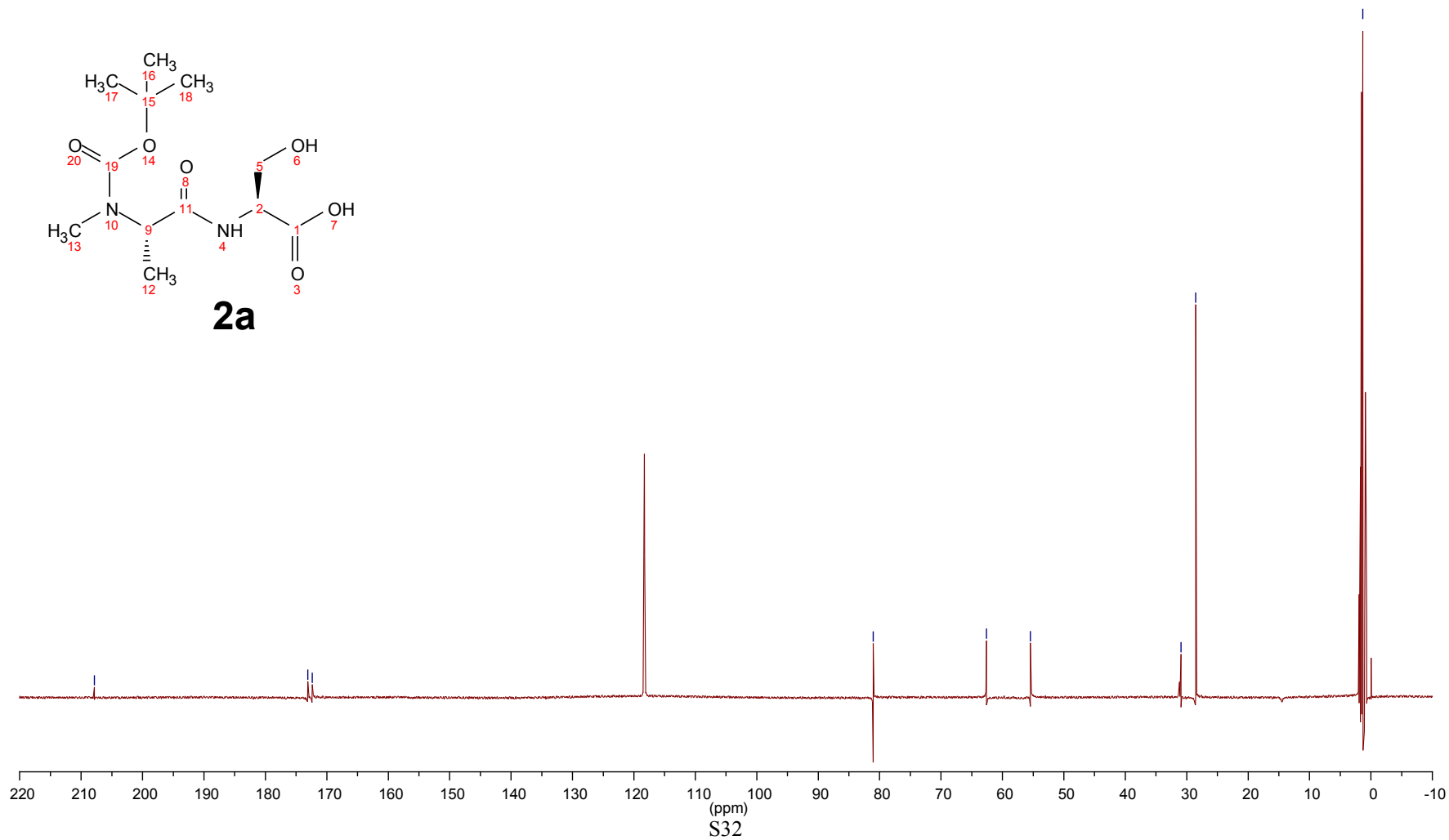
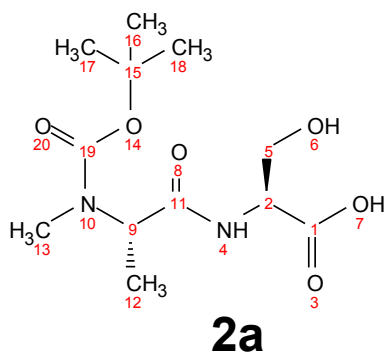
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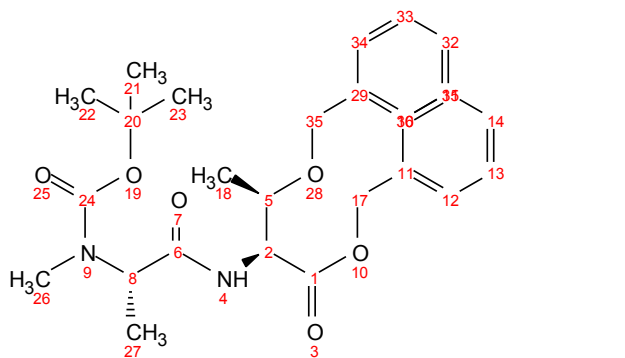
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Title MV-409dcarbon  
Solvent ACETONITRILED3  
Number of Scans 1600  
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Nucleus 13C

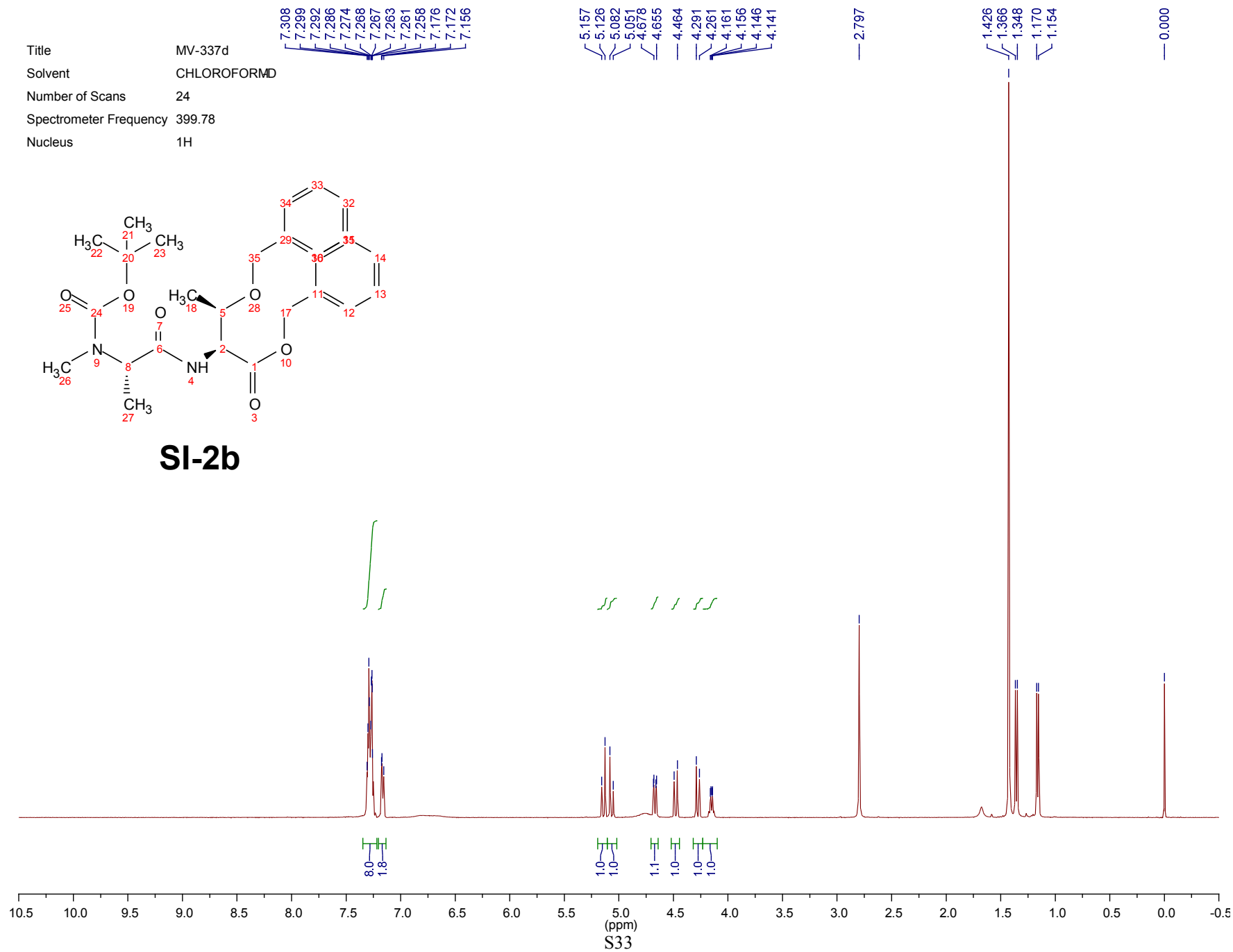




Title MV-337d  
Solvent CHLOROFORMD  
Number of Scans 24  
Spectrometer Frequency 399.78  
Nucleus 1H



SI-2b



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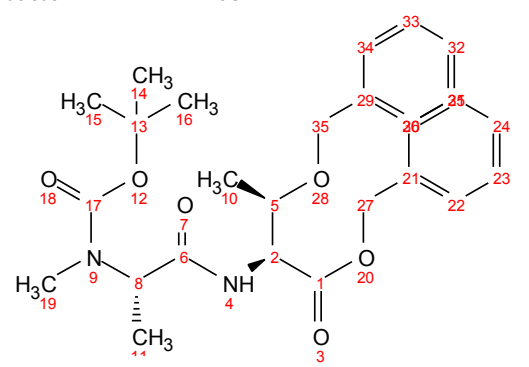
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56.783

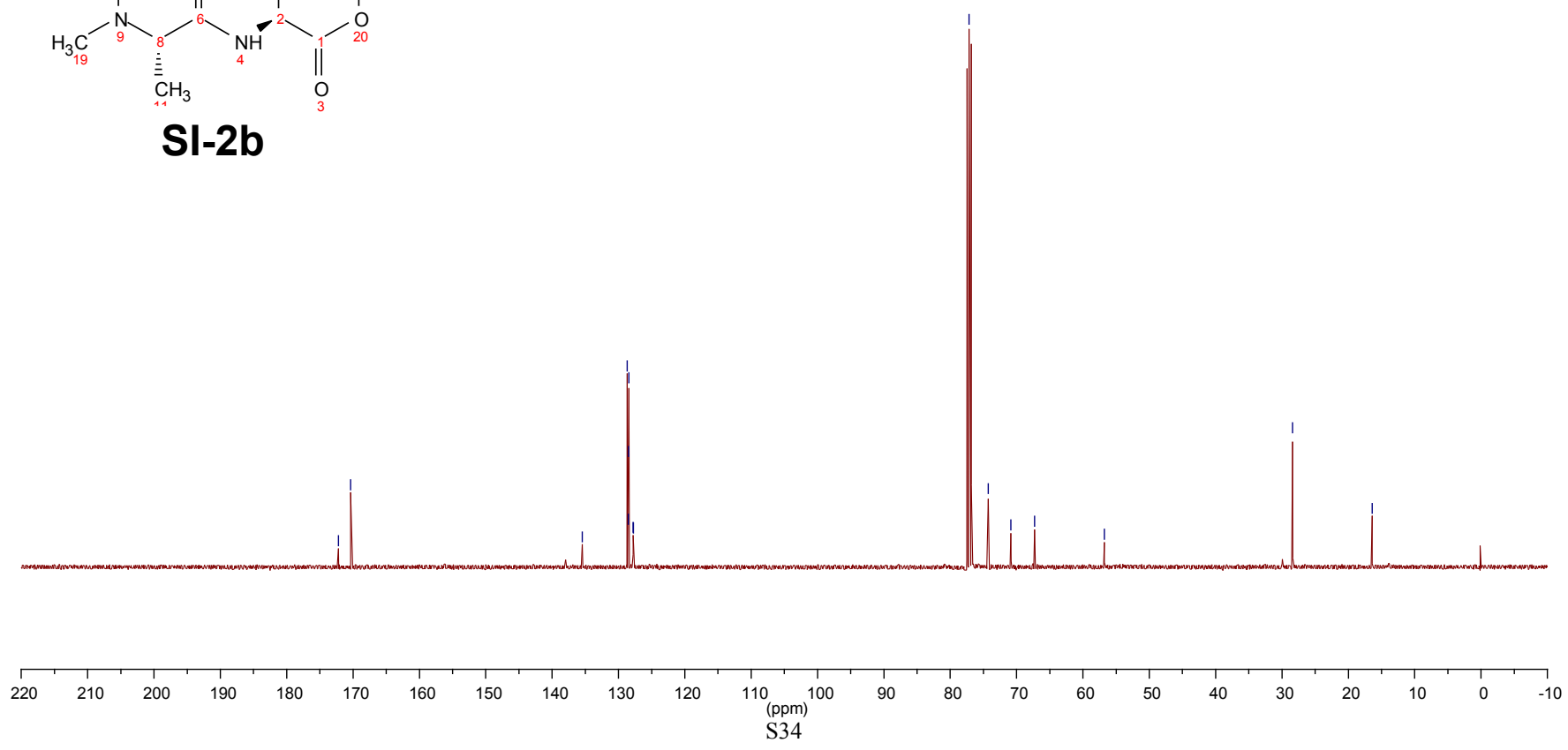
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16.420

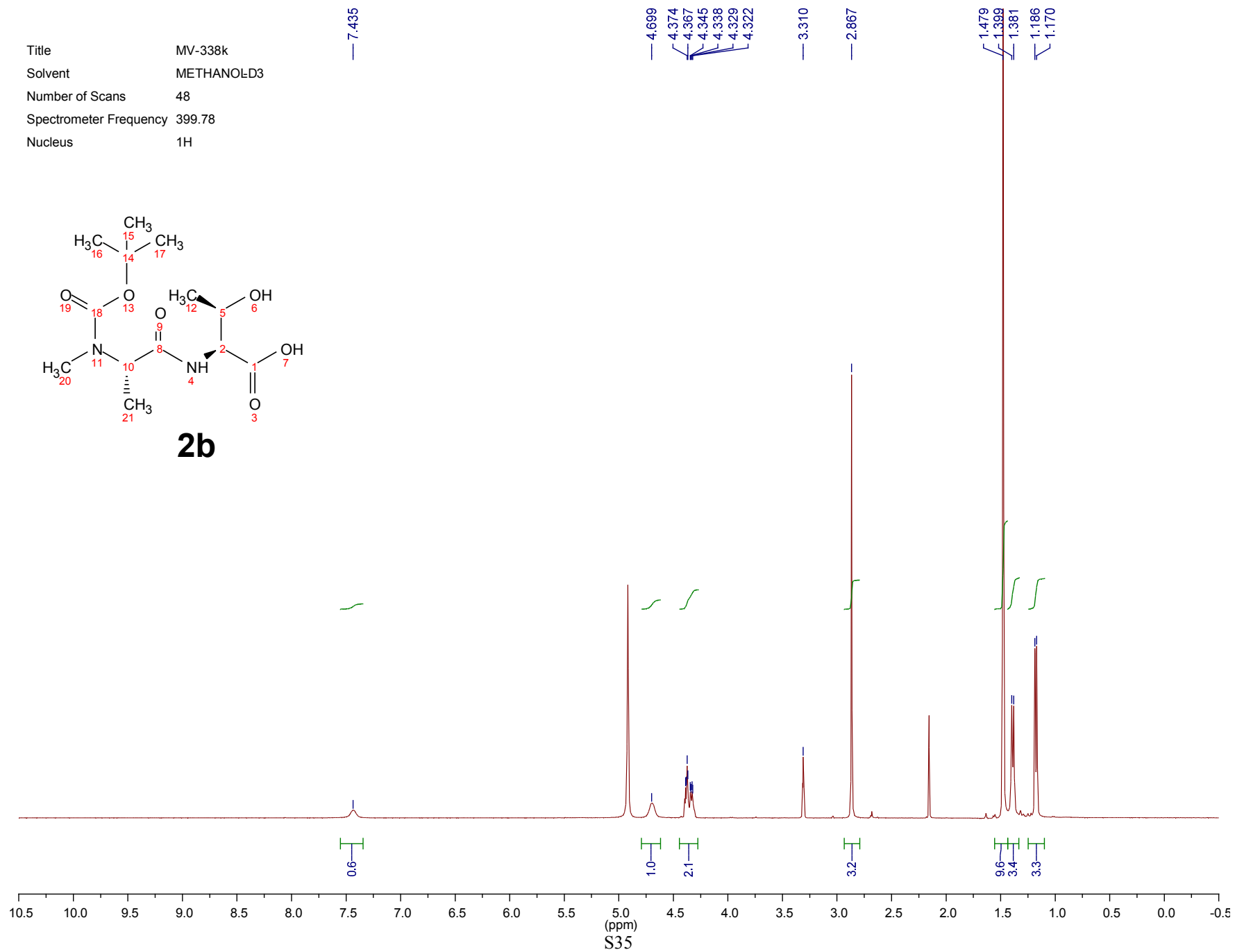
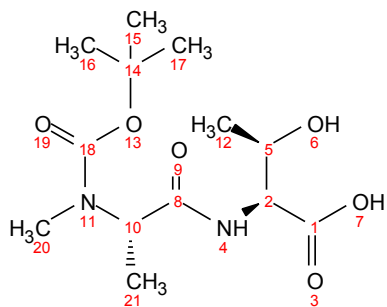
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Number of Scans 6120  
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Nucleus 13C



**SI-2b**

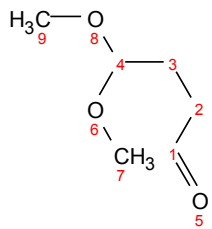


Title MV-338k  
Solvent METHANOL-D3  
Number of Scans 48  
Spectrometer Frequency 399.78  
Nucleus 1H

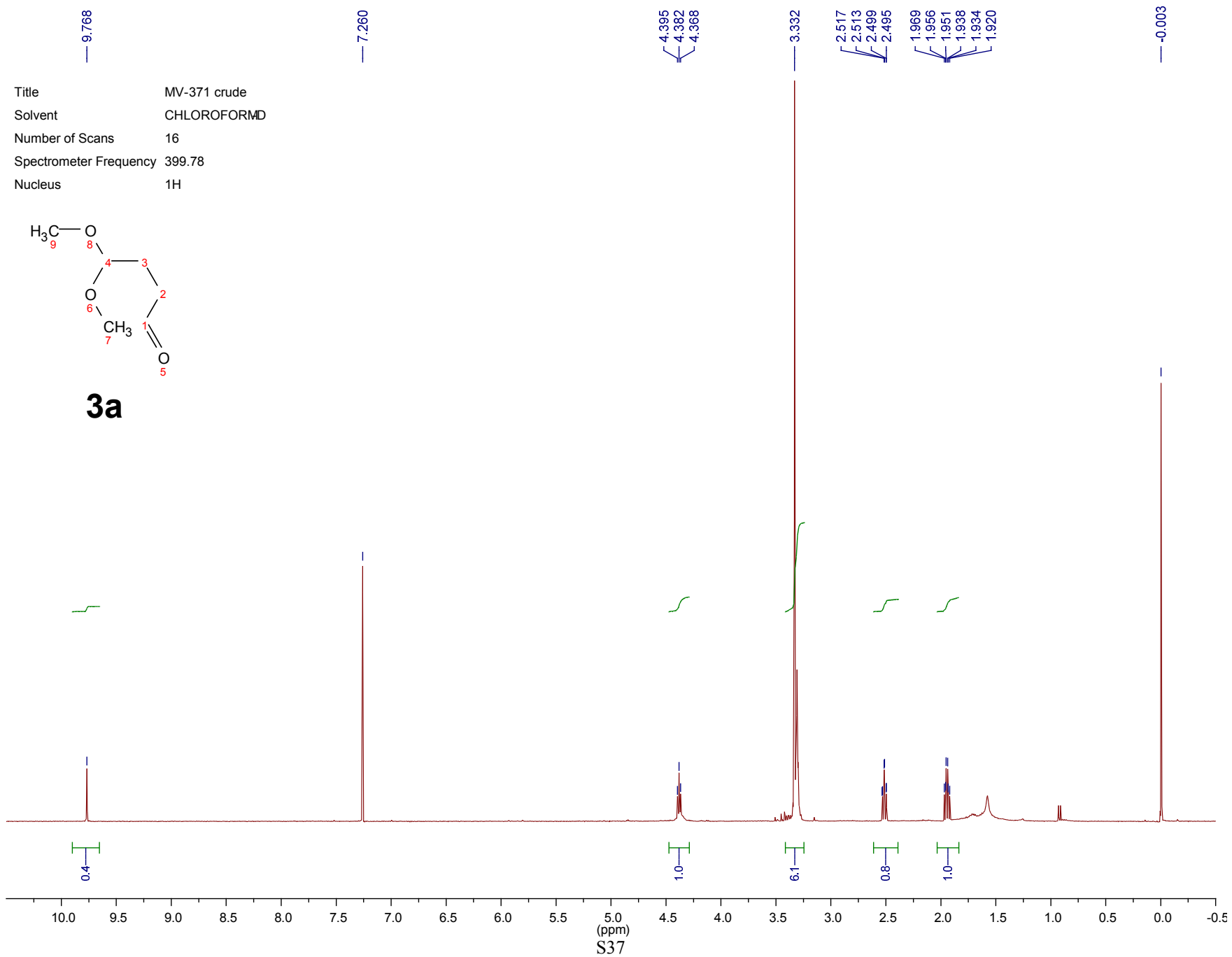




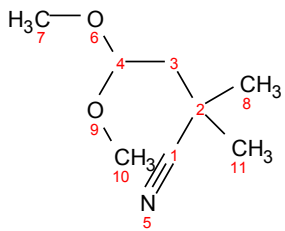
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Nucleus 1H



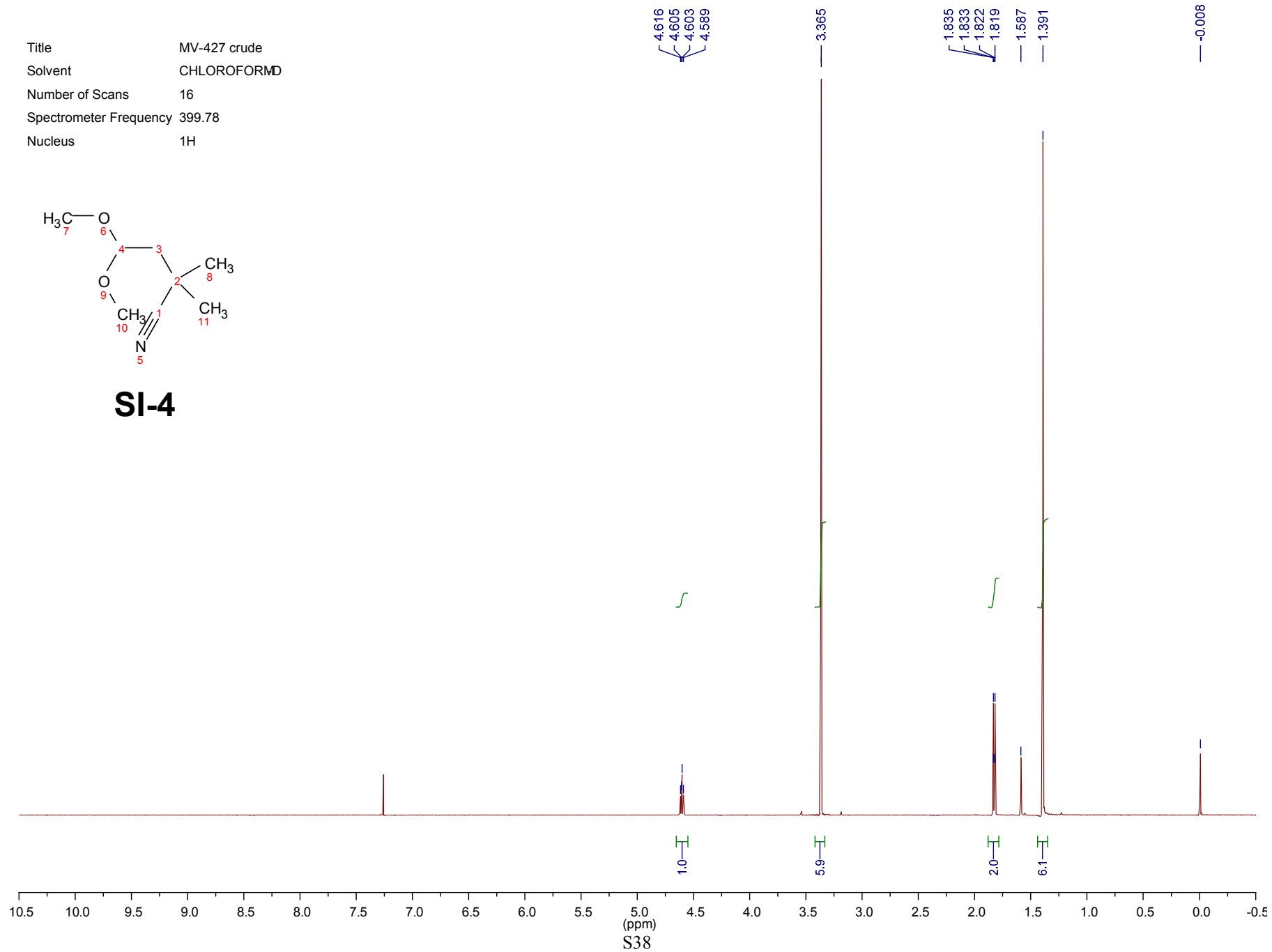
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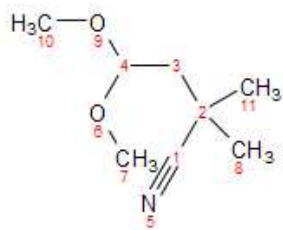
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Nucleus 1H



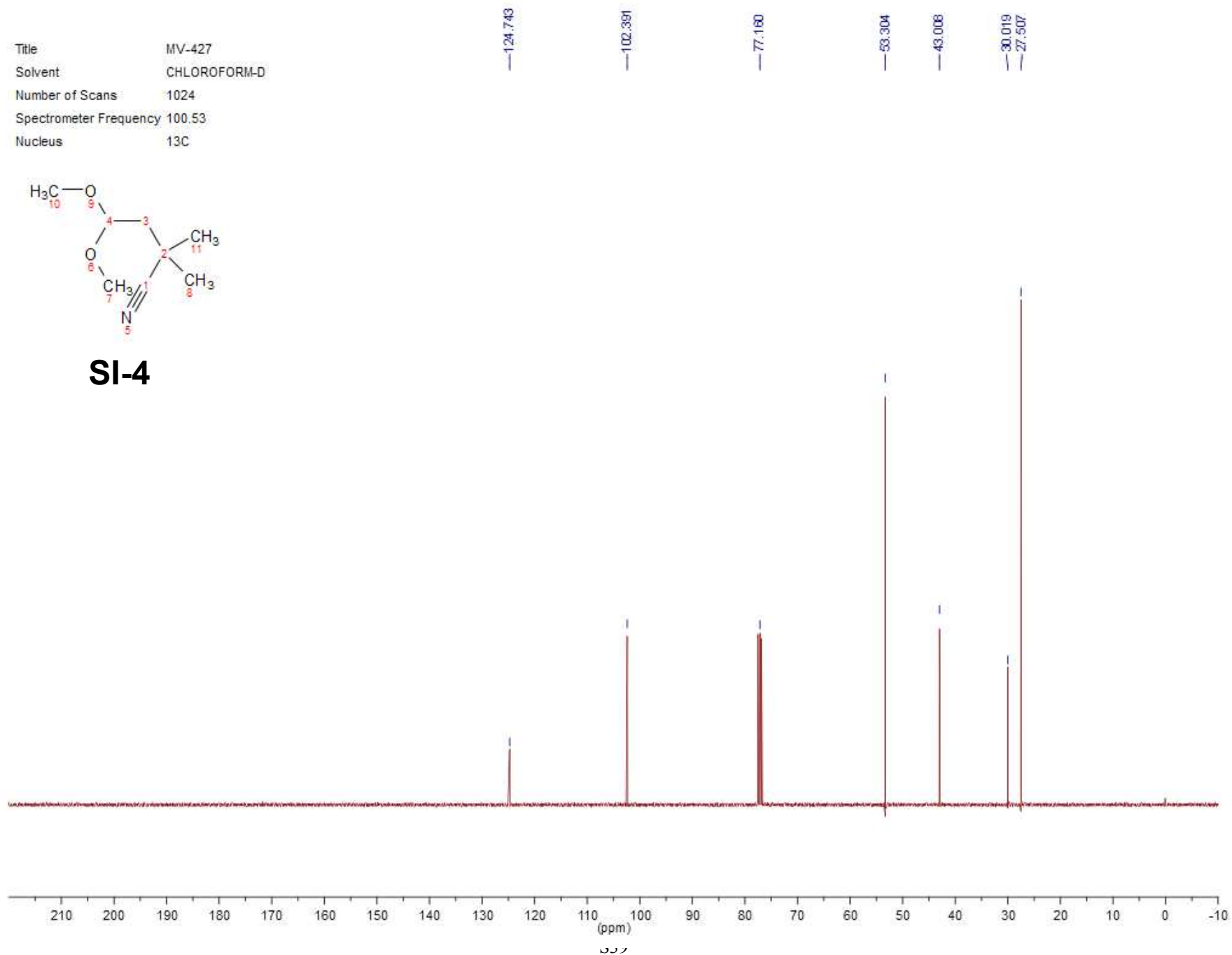
SI-4

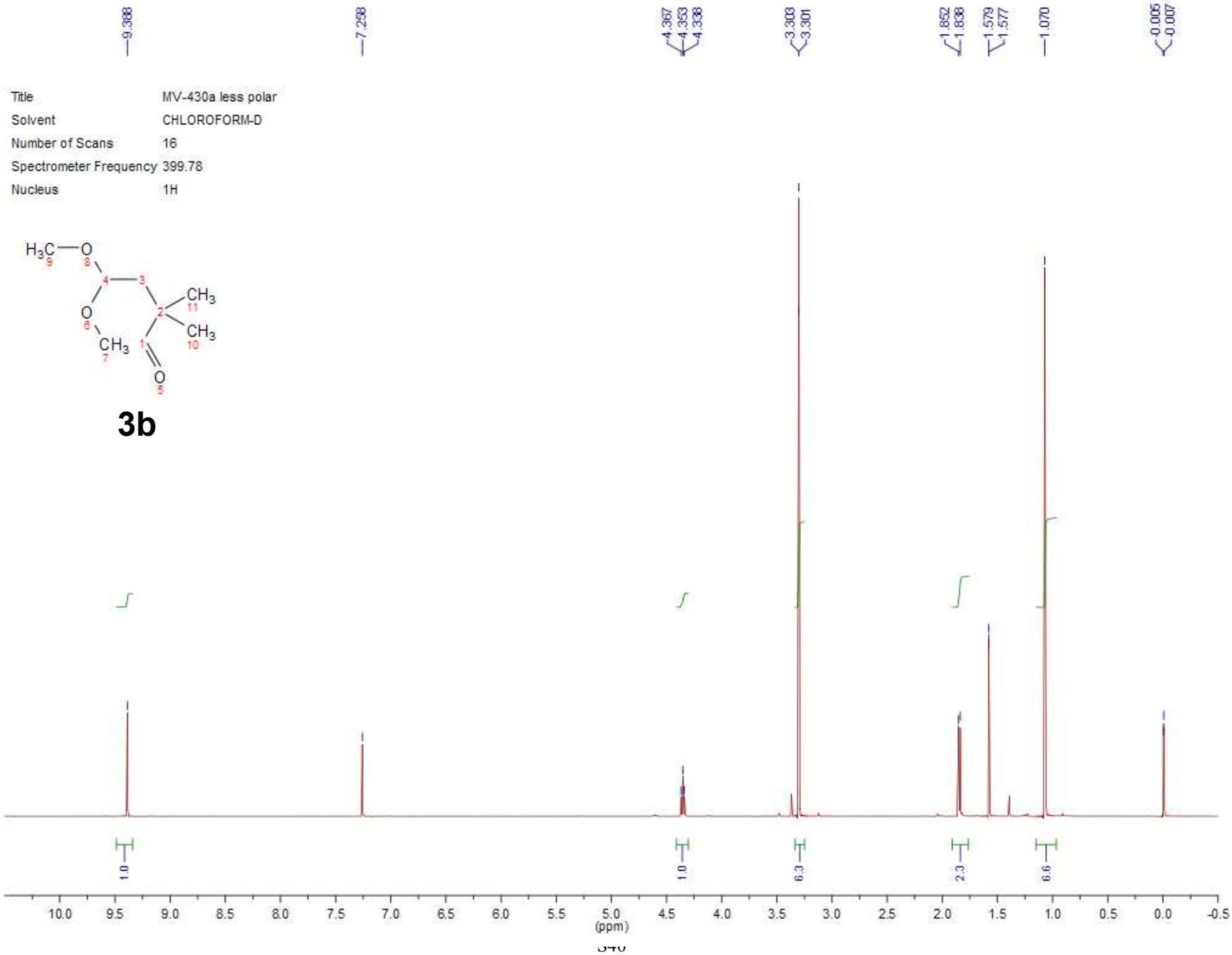


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Nucleus <sup>13</sup>C



SI-4







204.677

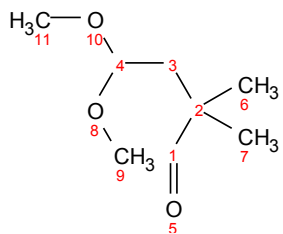
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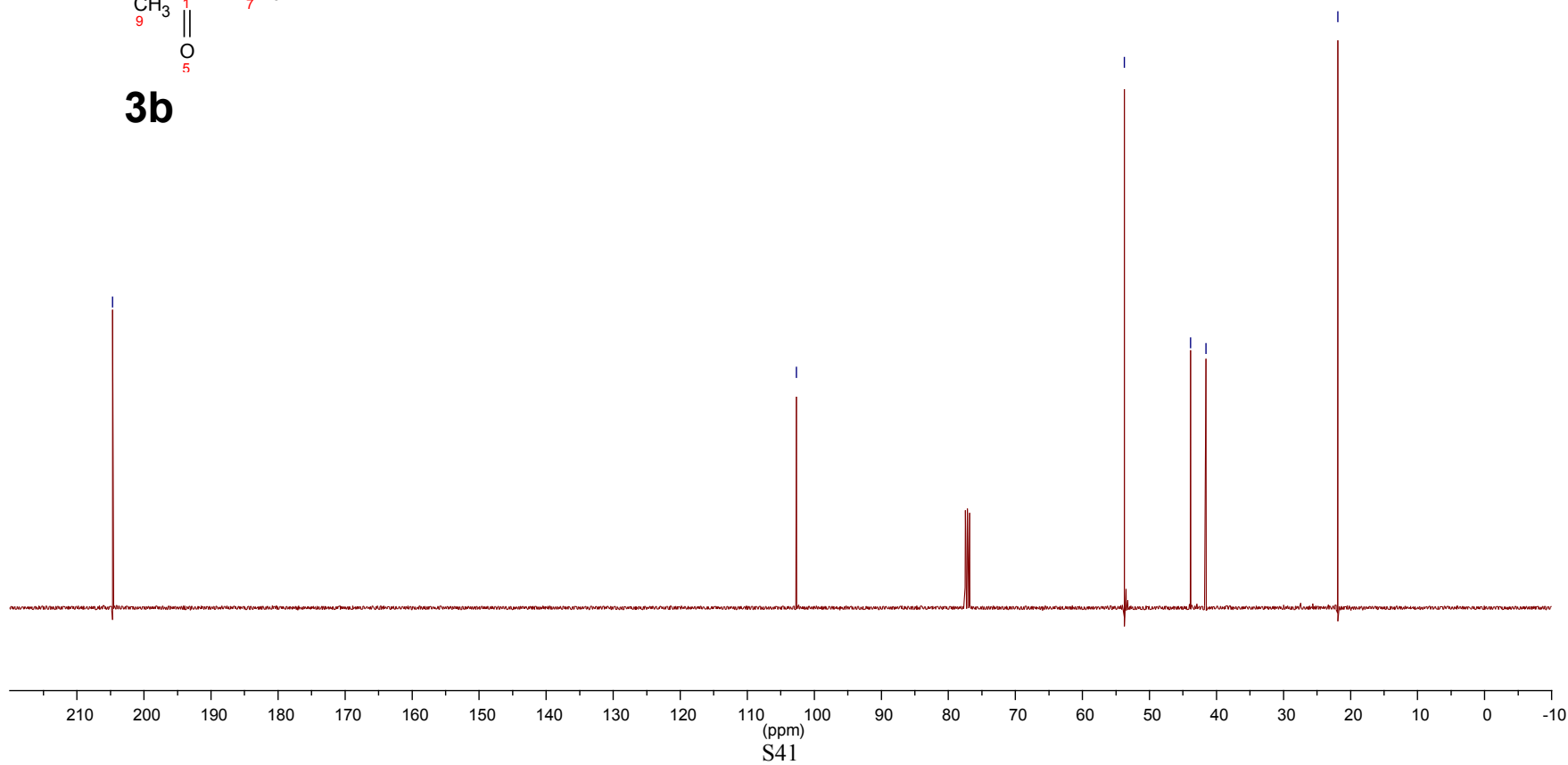
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21.902

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Number of Scans 900  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C

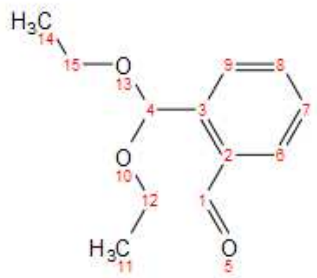


**3b**

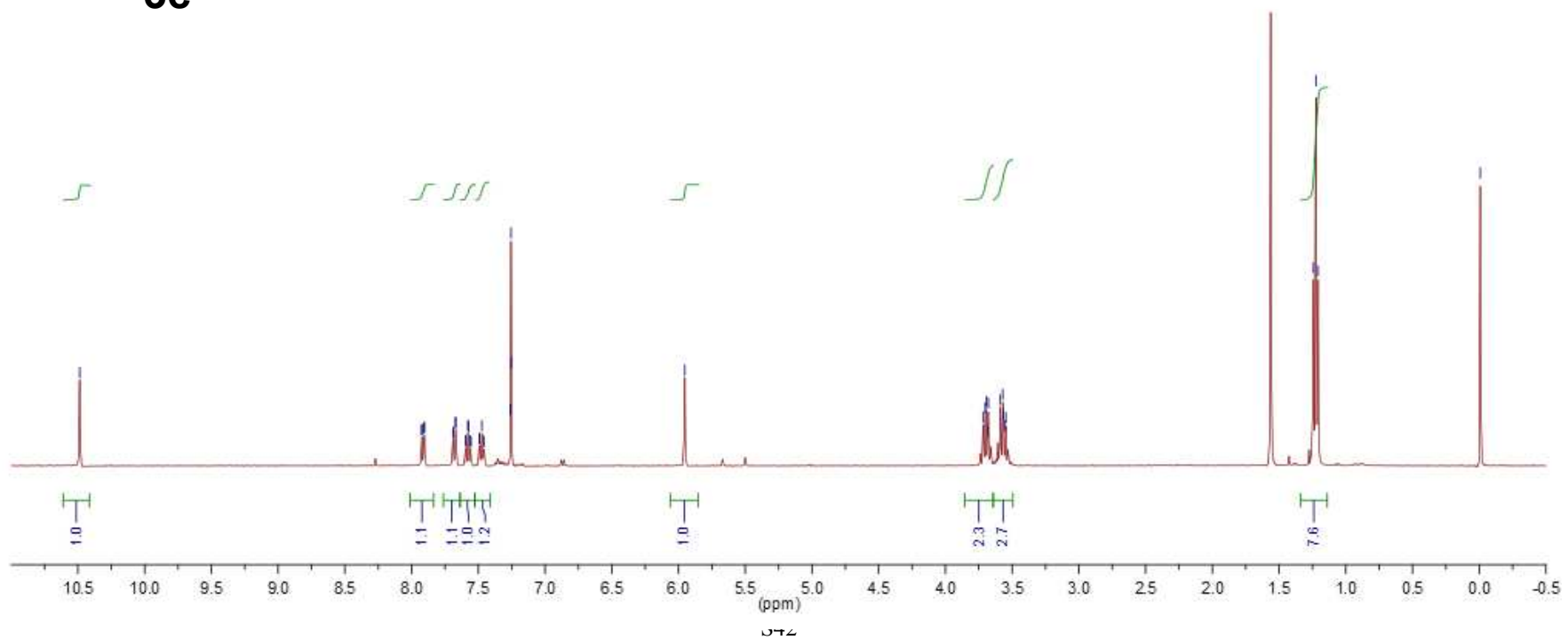




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 Spectrometer Frequency 399.78  
 Nucleus 1H



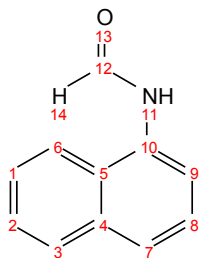
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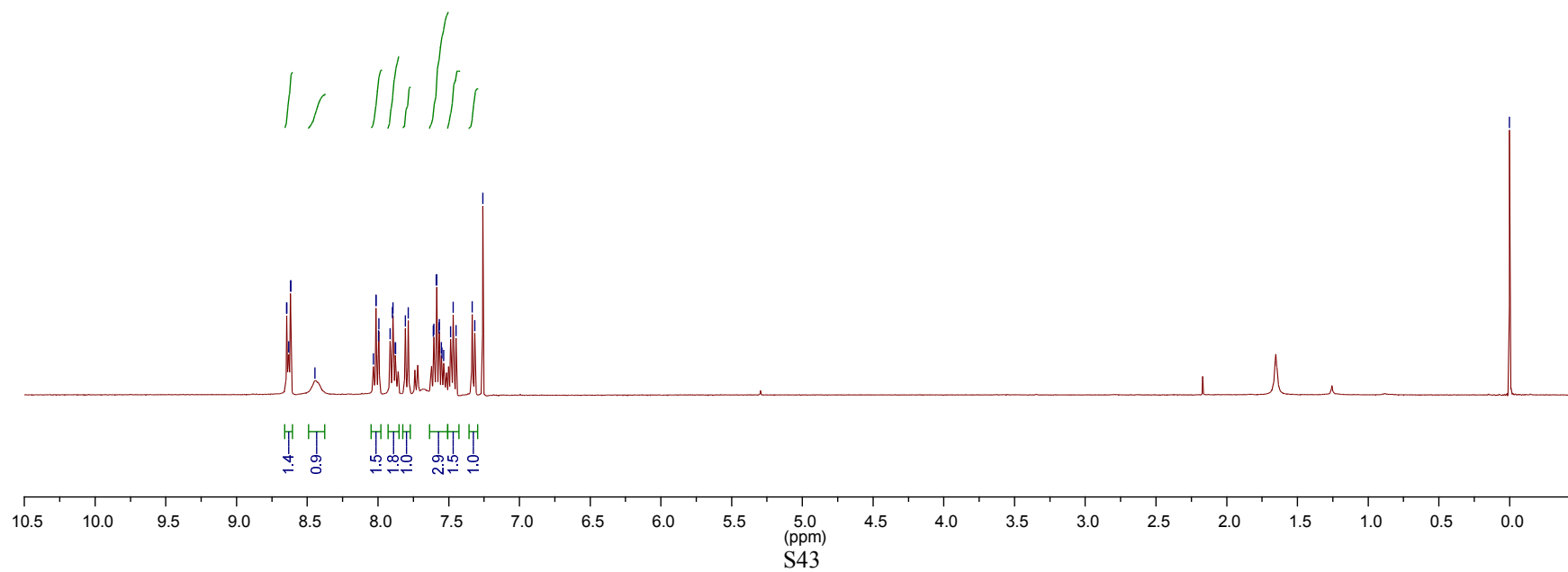
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7.315  
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Title MV-341d  
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Number of Scans 24  
Spectrometer Frequency 399.78  
Nucleus 1H



SI-7a

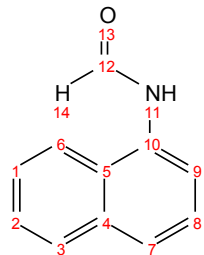


Title MV-341d  
Solvent CHLOROFORMD  
Number of Scans 12000  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C

164.101  
159.705

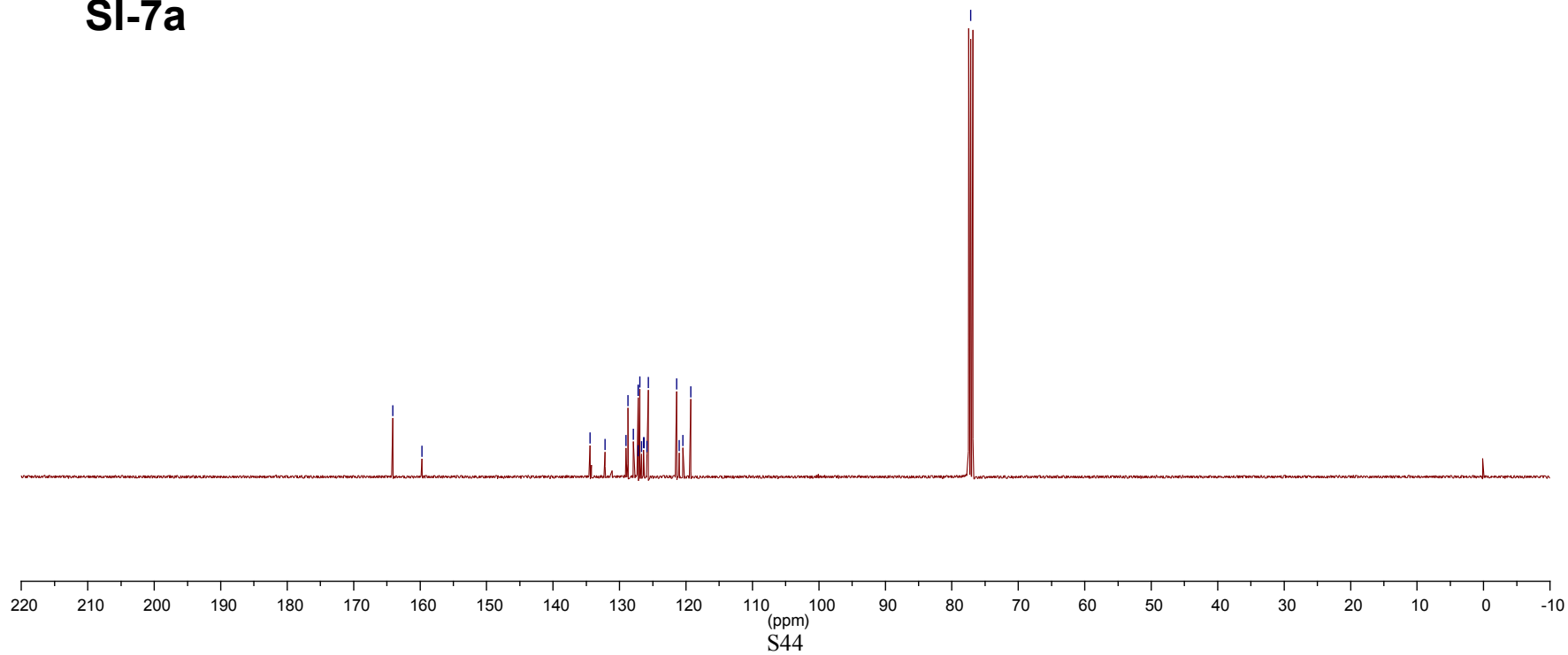
134.426  
132.183  
127.189  
125.661  
121.384  
121.030  
120.460  
119.274

77.160

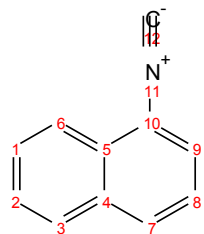


mix of geometrical isomers  
(rotamers)

**SI-7a**



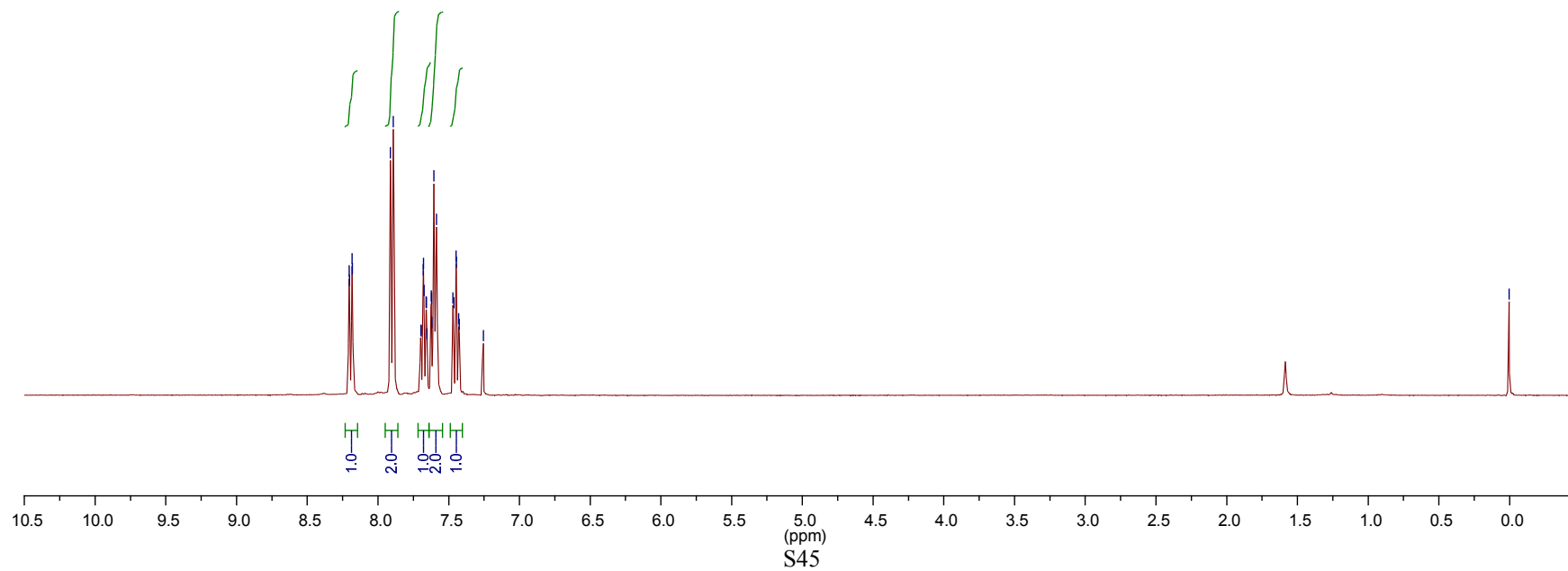
Title MV-342  
Solvent CHLOROFORMD  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



**4b**

8.204  
8.202  
8.183  
8.181  
7.893  
7.622  
7.587  
7.254

0.003

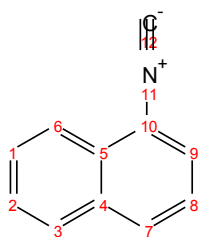


Title MV-342  
Solvent CHLOROFORMD  
Number of Scans 2000  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C

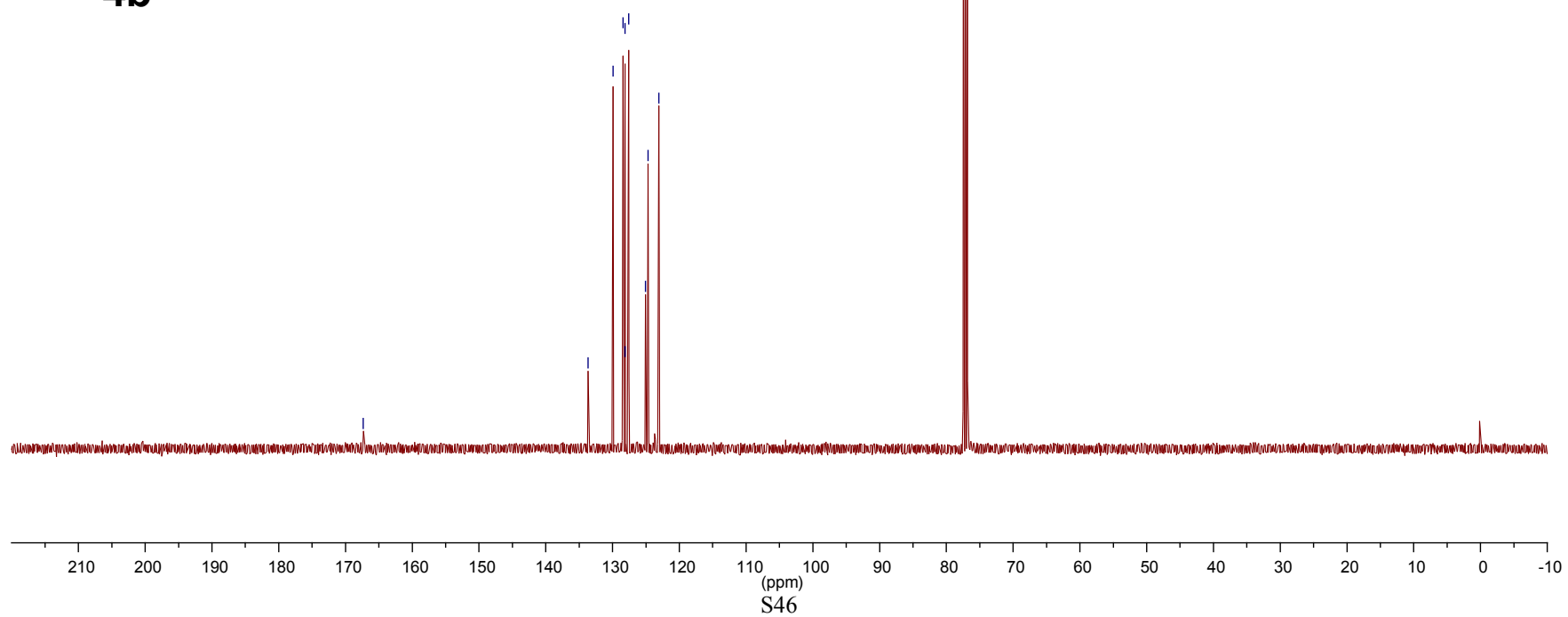
167.325

133.676  
129.918  
128.447  
128.147  
128.119  
127.557  
125.060  
124.685  
123.077

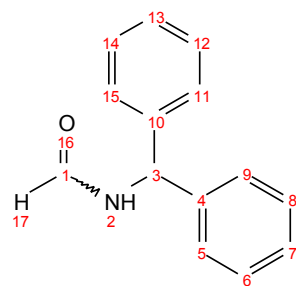
77.160



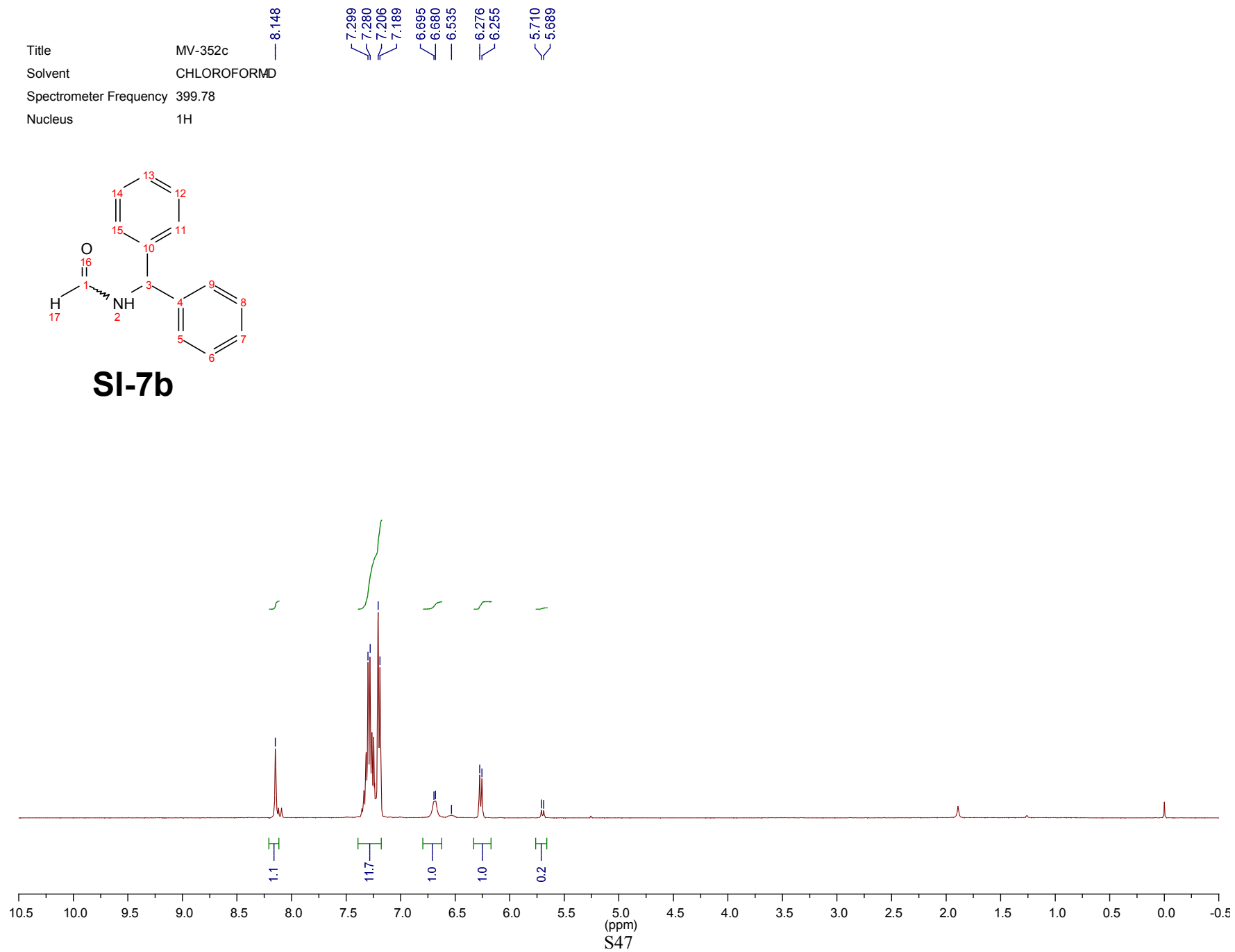
4b



Title MV-352c  
Solvent CHLOROFORMD  
Spectrometer Frequency 399.78  
Nucleus 1H

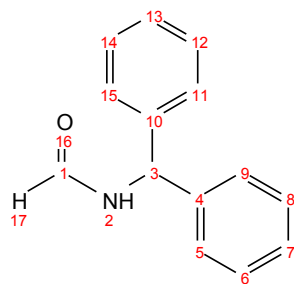


SI-7b

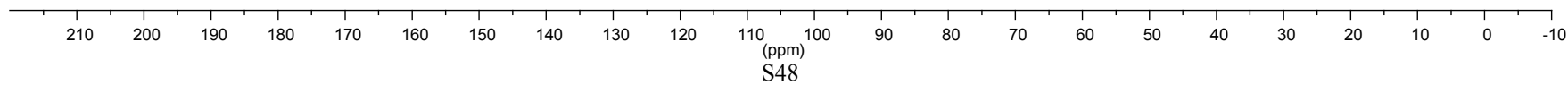
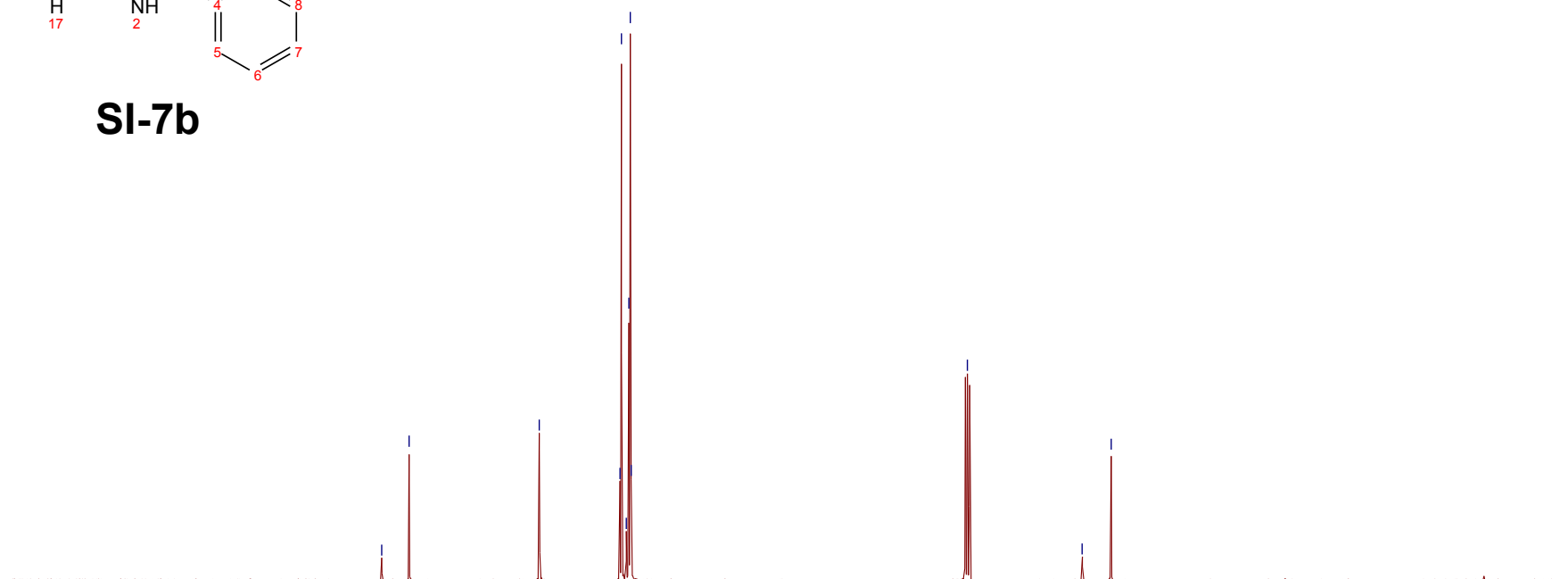


Title MV-352c  
Solvent CHLOROFORMD  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C

164.519  
160.450  
141.034  
128.992  
128.769  
128.049  
127.661  
127.464  
127.311  
77.160  
60.029  
55.714

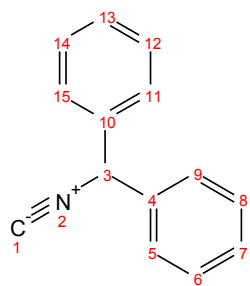


SI-7b

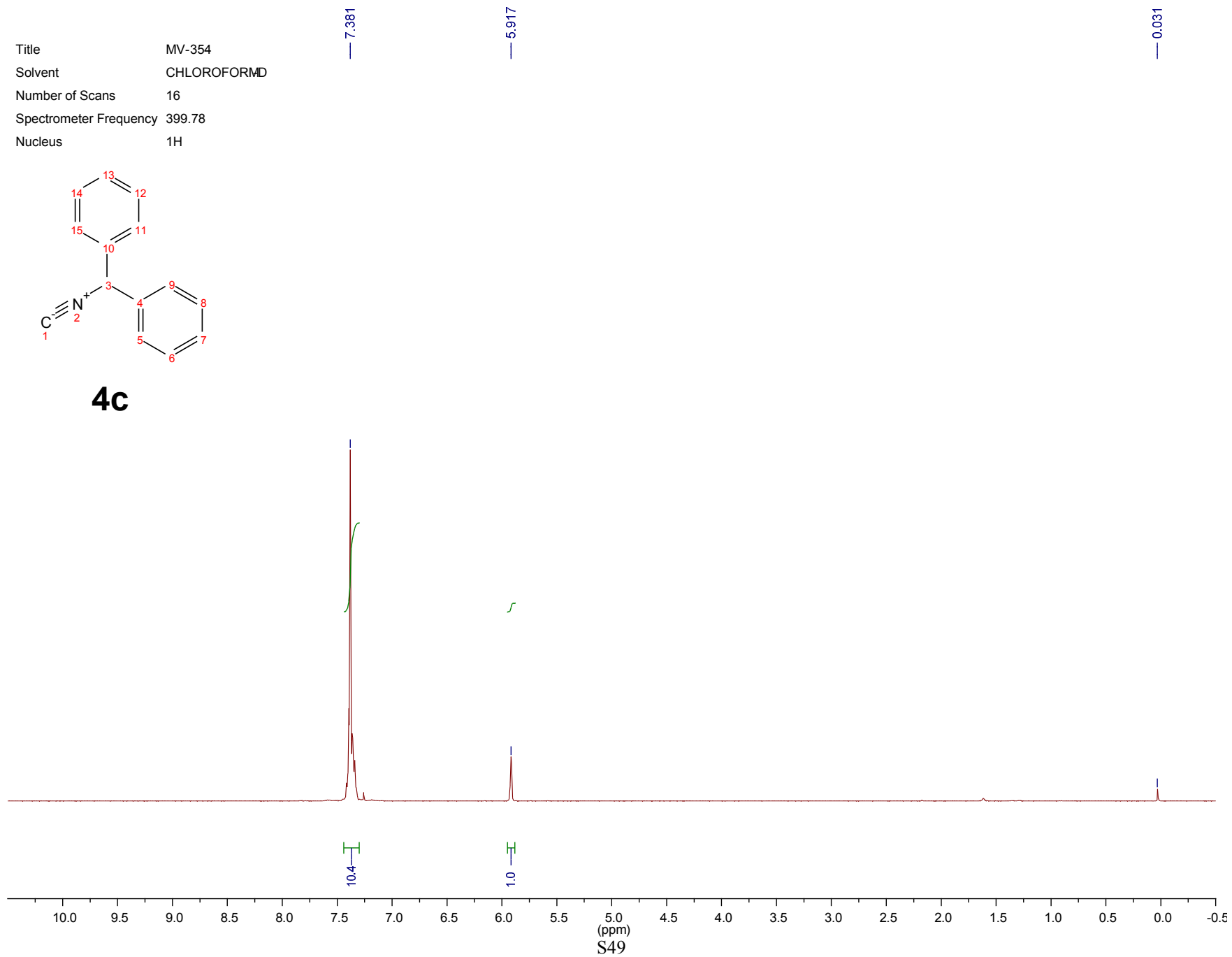




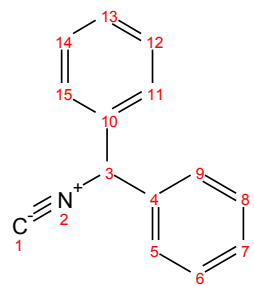
Title MV-354  
Solvent CHLOROFORMD  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



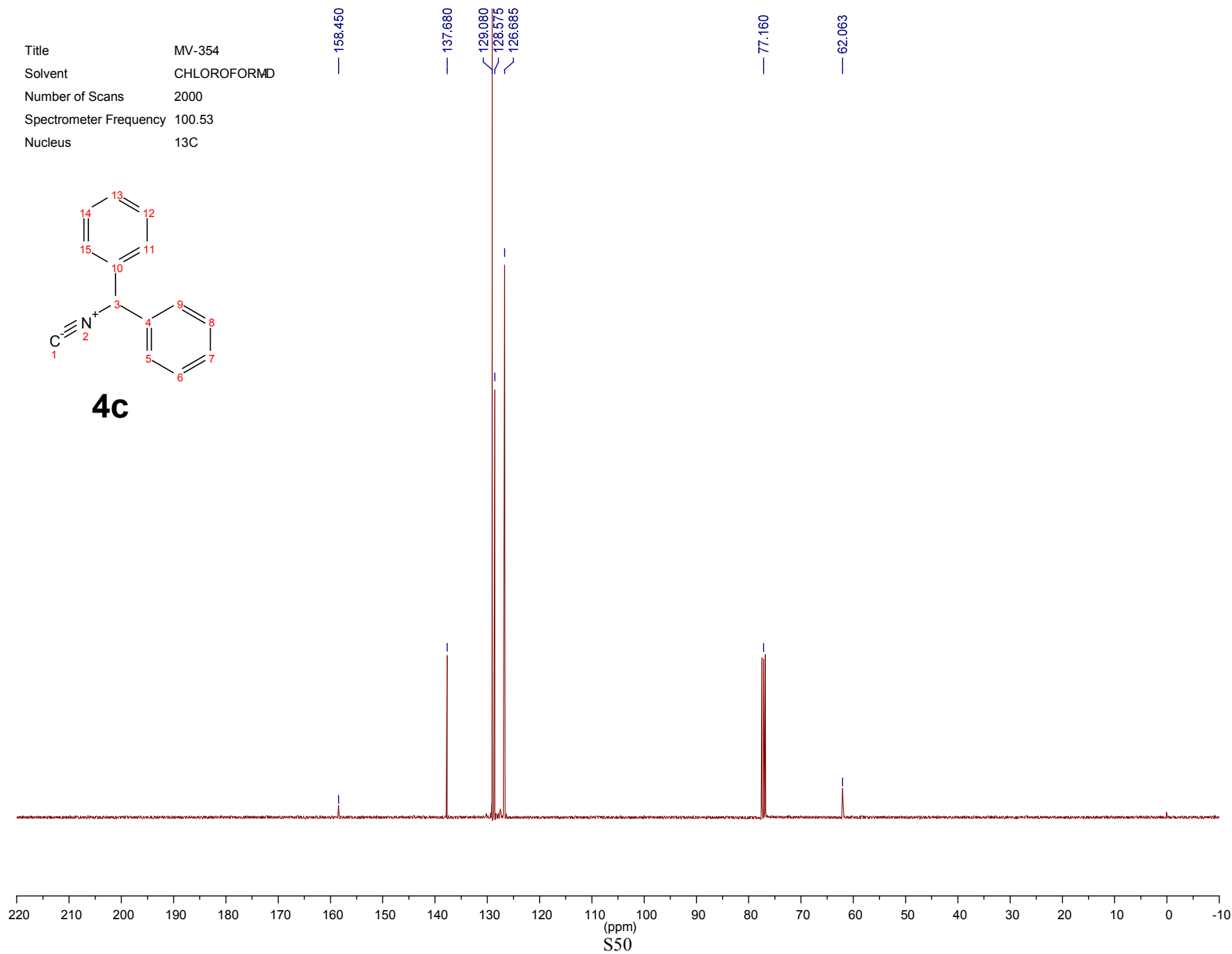
4c



Title MV-354  
Solvent CHLOROFORMD  
Number of Scans 2000  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C

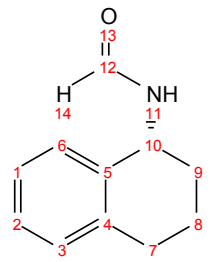


**4c**

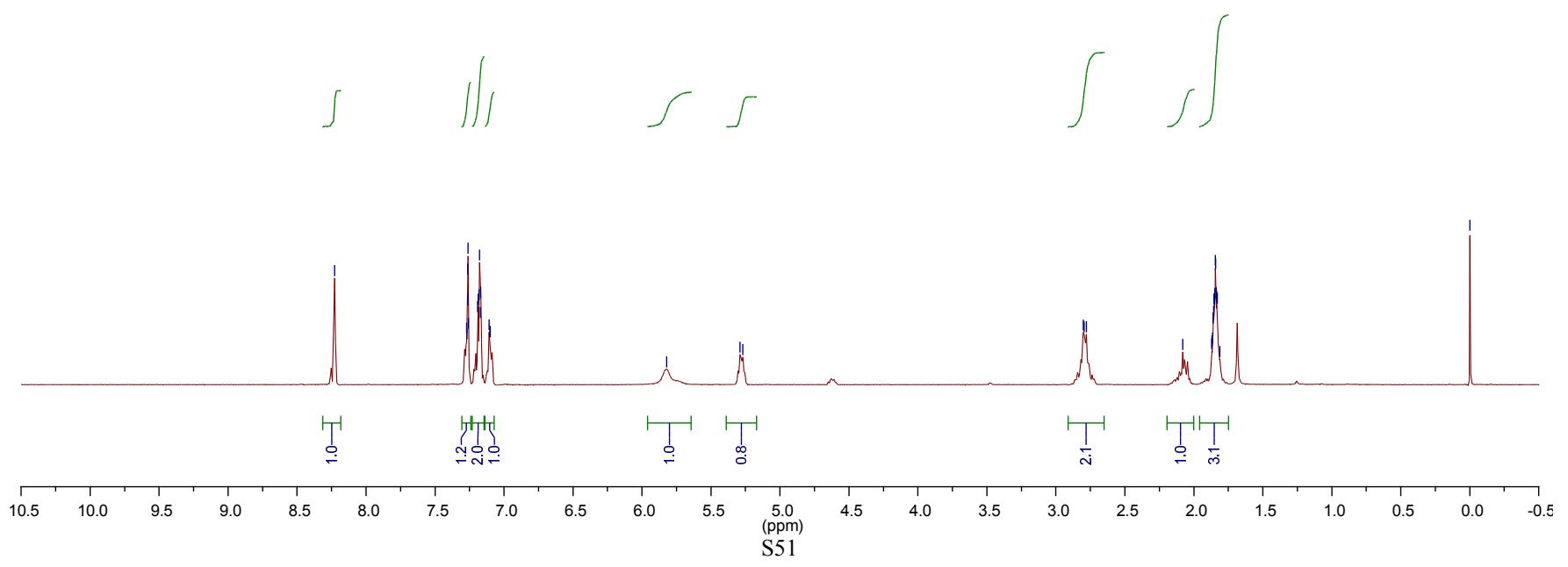


8.229  
 7.260  
 7.191  
 7.187  
 7.181  
 7.178  
 7.171  
 7.168  
 7.108  
 7.102  
 7.100  
 5.822  
 5.289  
 5.269  
 2.802  
 2.794  
 2.780  
 2.089  
 1.848  
 1.845  
 1.842  
 1.837  
 1.833  
 1.829  
 1.812  
 -0.001

Title MV-328a  
 Solvent CHLOROFORMD  
 Number of Scans 16  
 Spectrometer Frequency 399.78  
 Nucleus 1H



**SI-7c**



Title MV-328a  
Solvent CHLOROFORMD  
Number of Scans 3200  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C

160.493

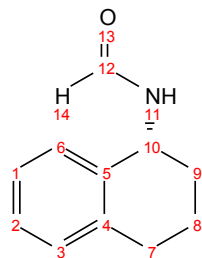
137.738  
136.127  
129.376  
128.773  
127.606  
126.471

77.160

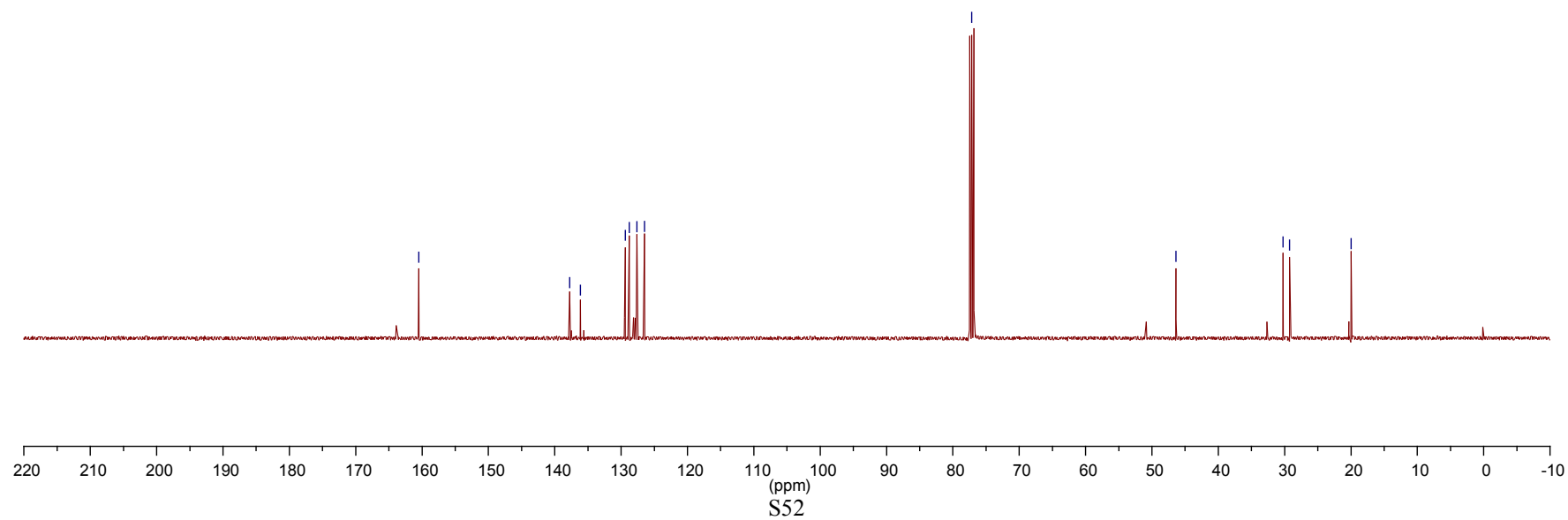
46.378

30.260  
29.263

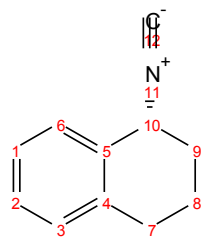
19.992



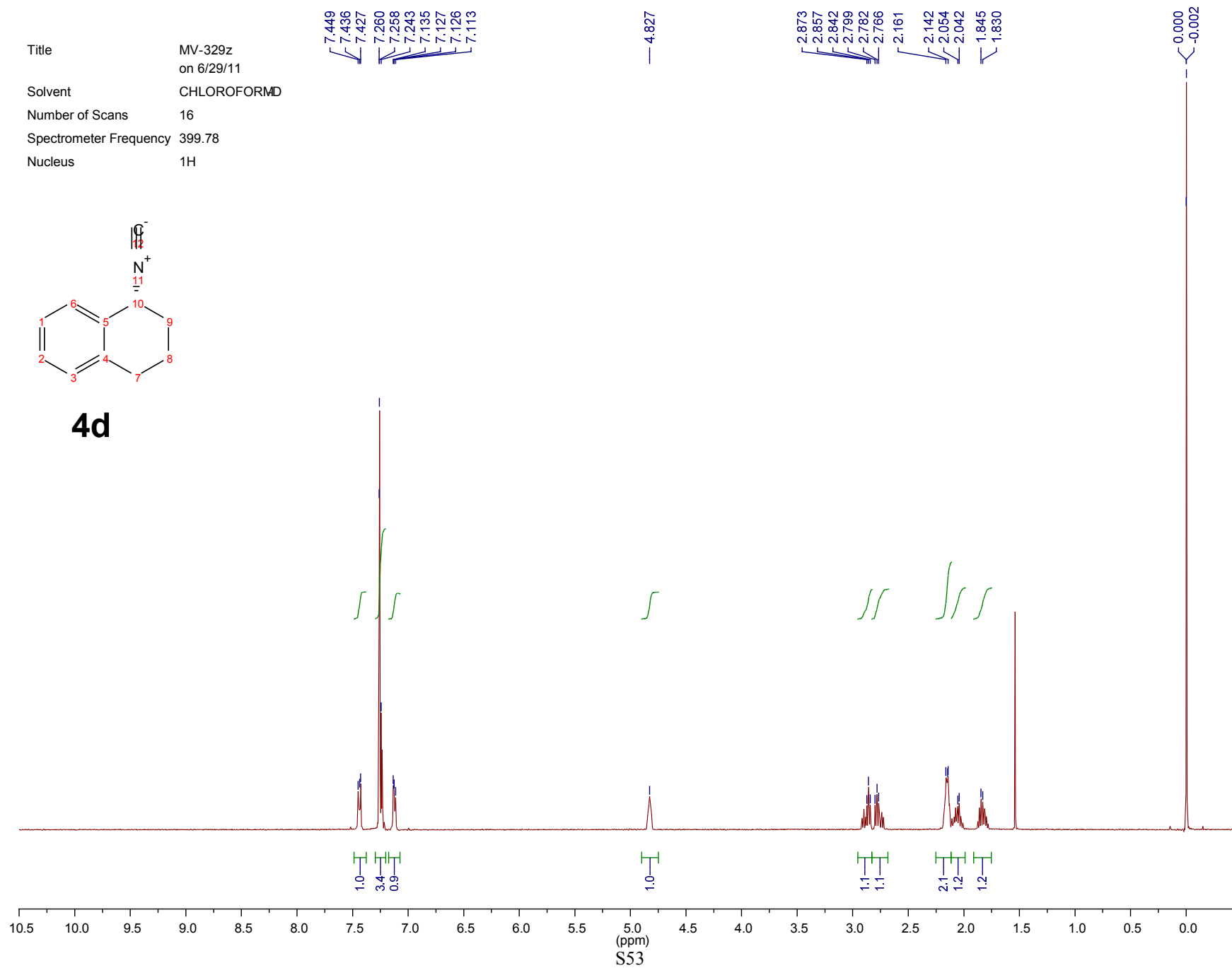
SI-7c



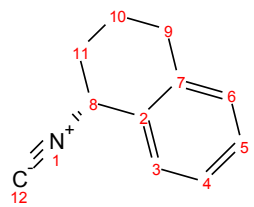
Title MV-329z  
on 6/29/11  
Solvent CHLOROFORMD  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



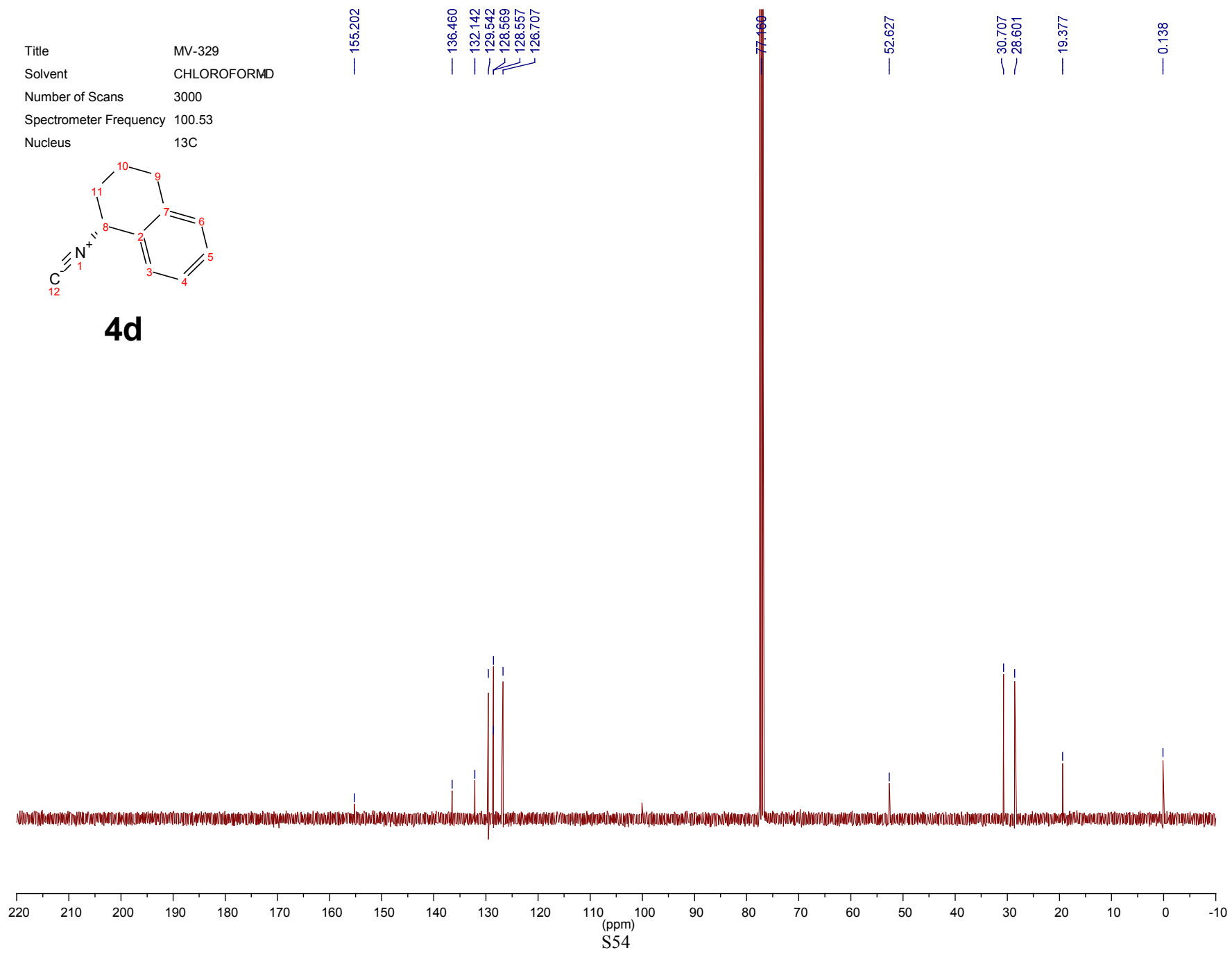
4d



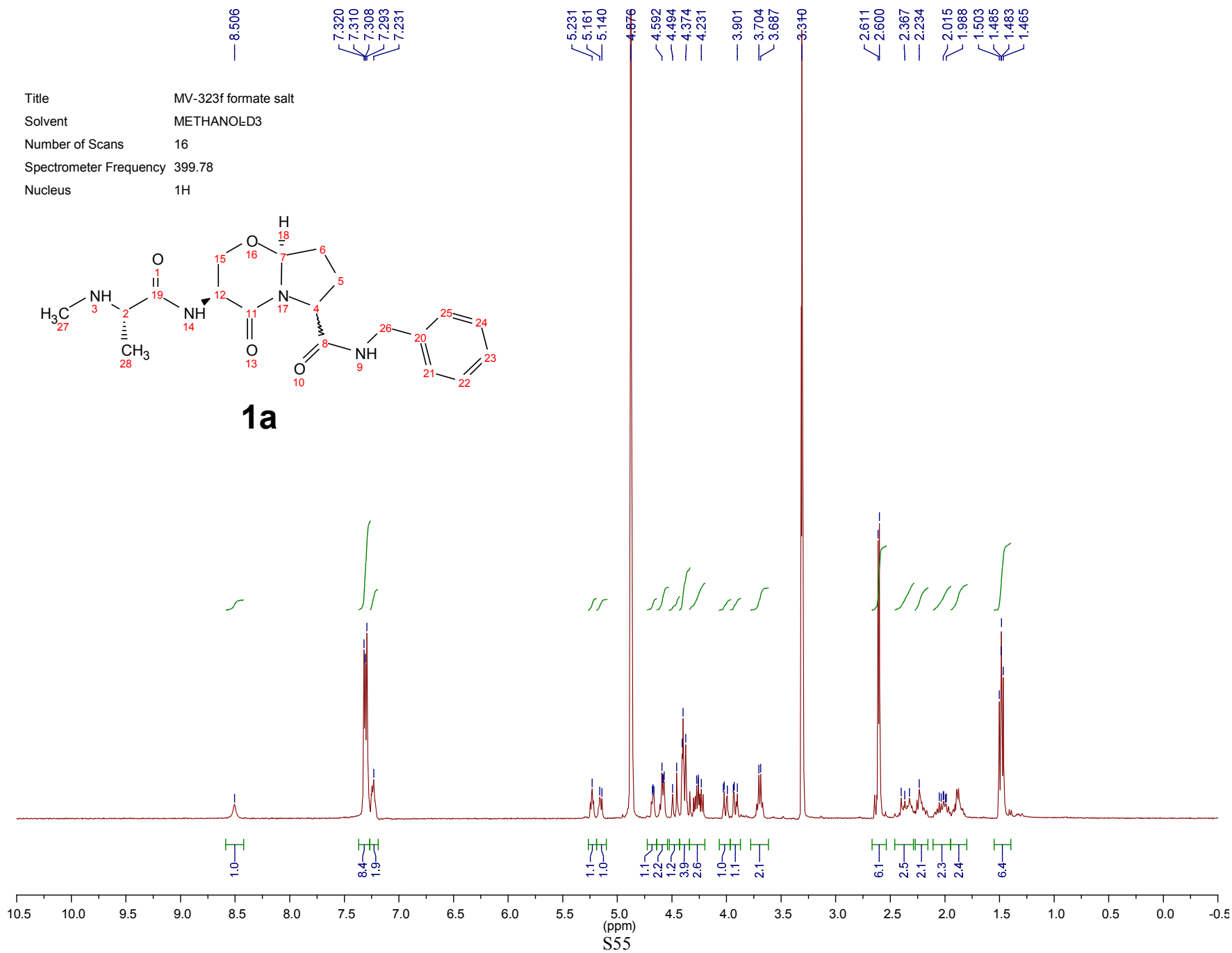
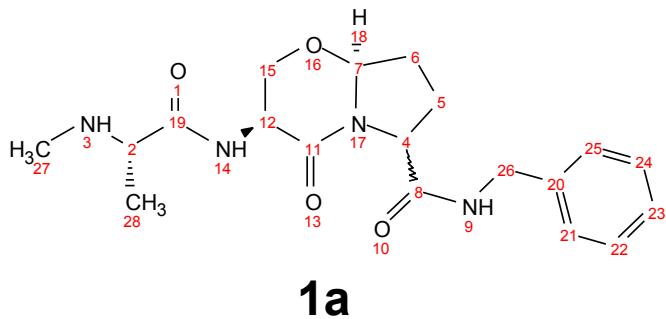
Title MV-329  
Solvent CHLOROFORMD  
Number of Scans 3000  
Spectrometer Frequency 100.53  
Nucleus  $^{13}\text{C}$



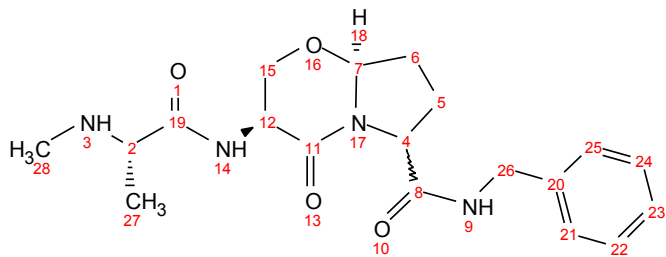
**4d**



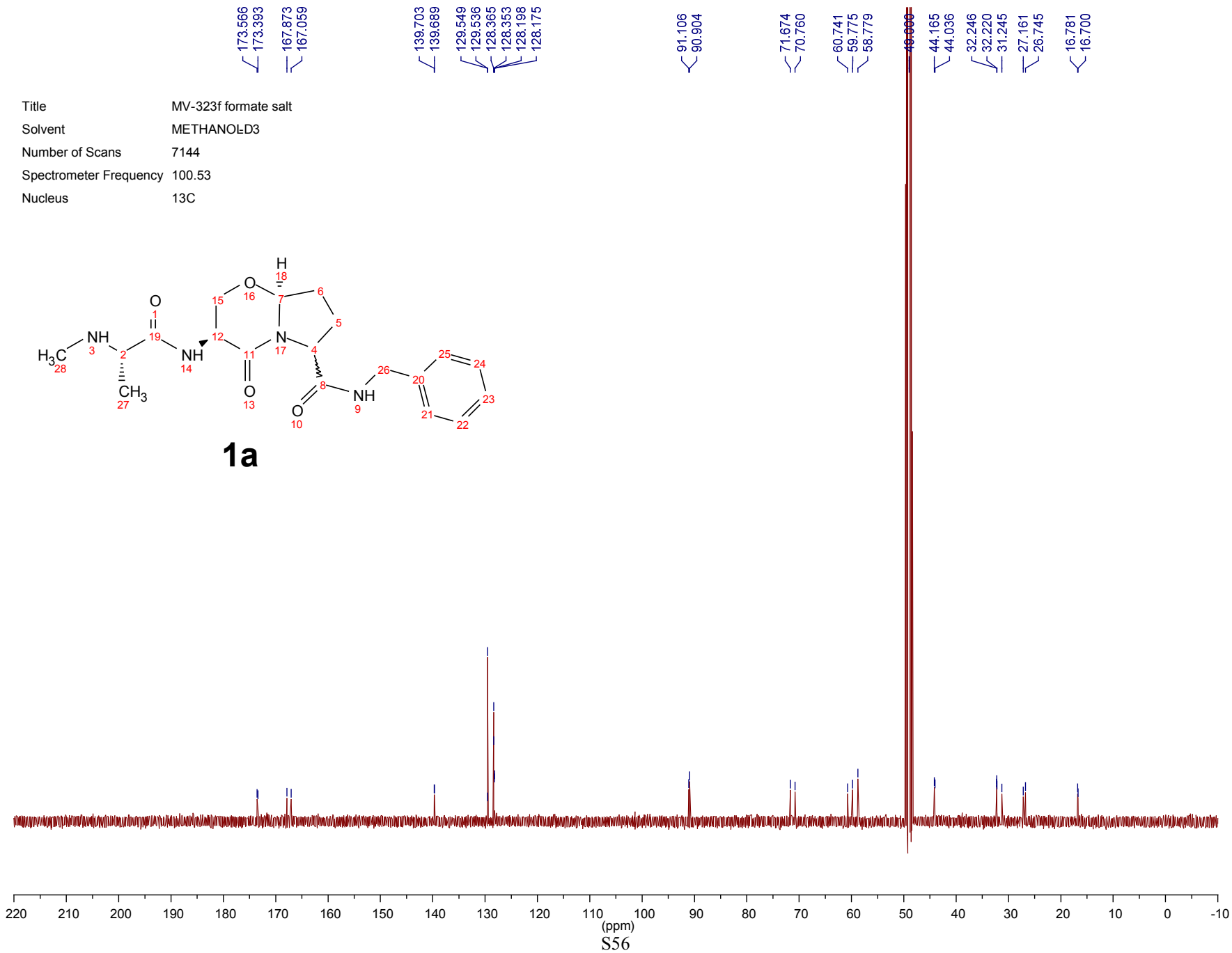
Title MV-323f formate salt  
 Solvent METHANOL-D3  
 Number of Scans 16  
 Spectrometer Frequency 399.78  
 Nucleus 1H



Title MV-323f formate salt  
 Solvent METHANOLED3  
 Number of Scans 7144  
 Spectrometer Frequency 100.53  
 Nucleus 13C

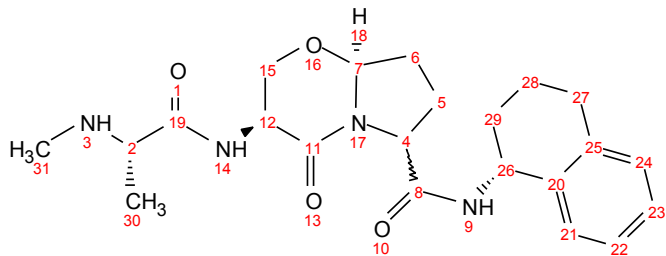


1a

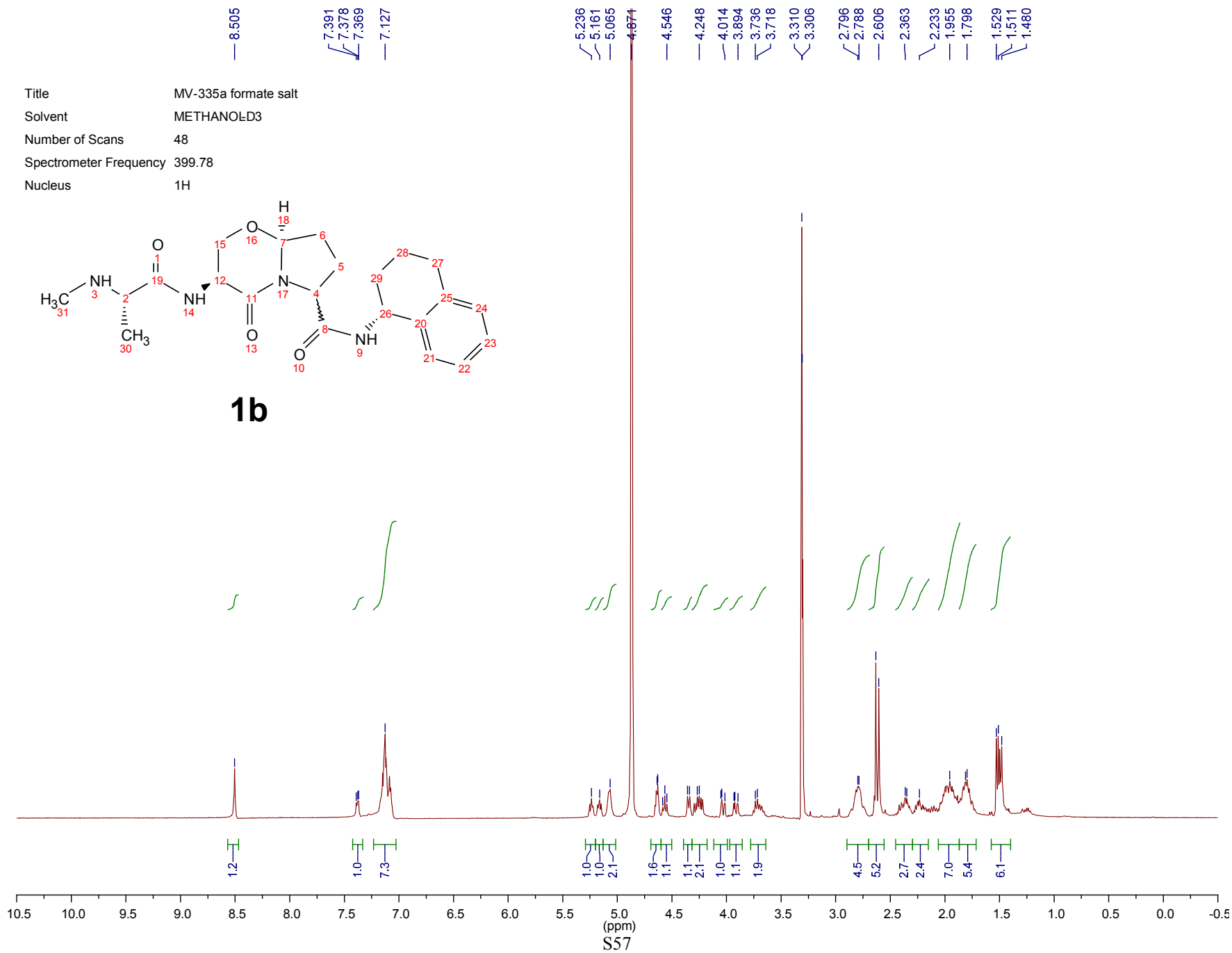




Title MV-335a formate salt  
Solvent METHANOLD3  
Number of Scans 48  
Spectrometer Frequency 399.78  
Nucleus 1H



**1b**



206.609

172.949  
172.827  
171.706  
167.736  
167.005

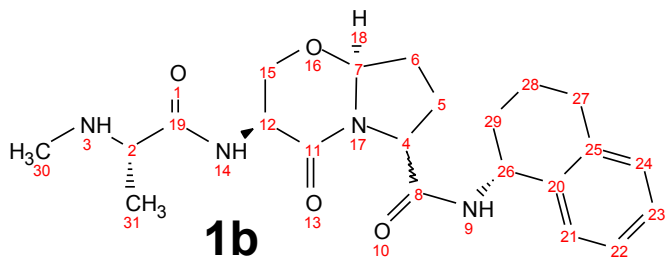
138.705  
138.471  
137.633  
137.588  
130.092  
129.893  
129.651  
129.275  
128.181  
128.112  
127.179  
127.169  
127.128

91.146  
90.963

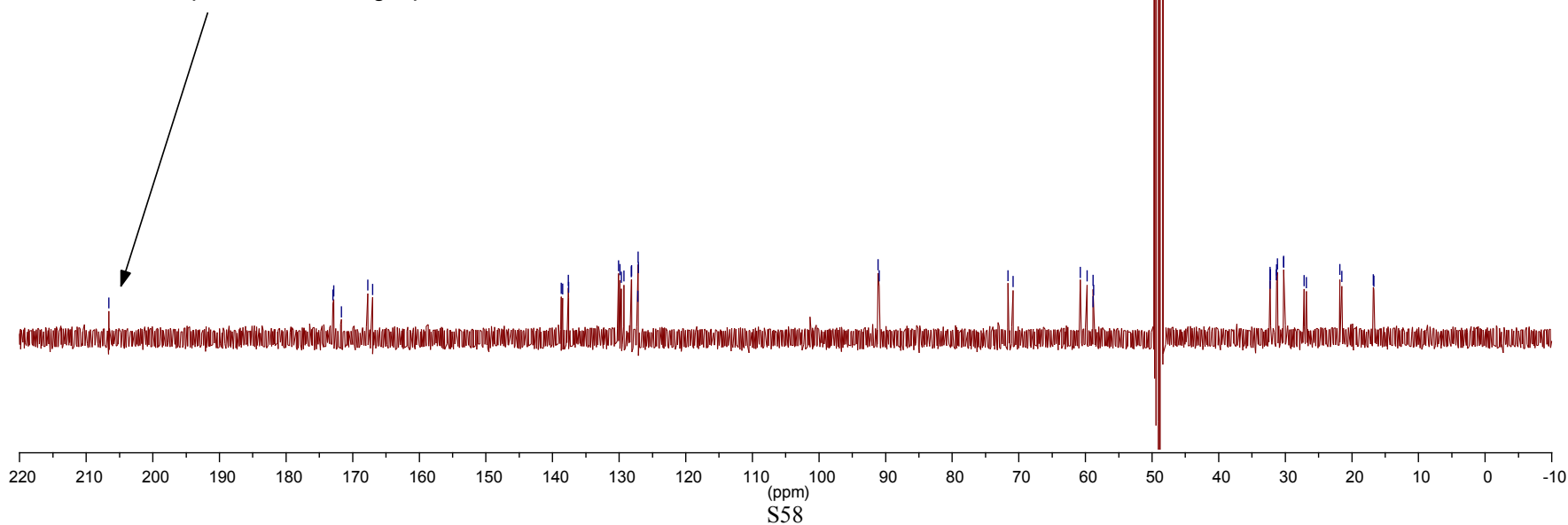
71.630  
70.873  
60.788  
59.760  
58.852  
58.841  
58.784

32.271  
31.339  
31.163  
30.231  
26.832  
21.522  
16.809  
16.723

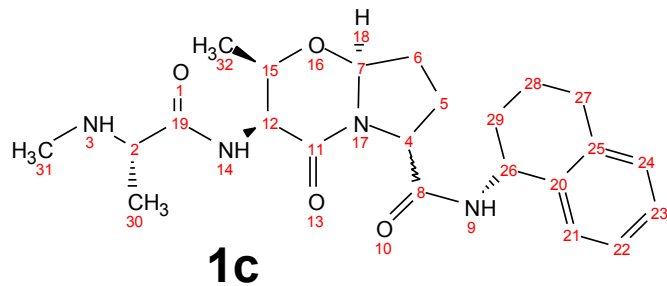
Title MV-335a formate salt  
Solvent METHANOLD3  
Number of Scans 9216  
Spectrometer Frequency 100.53  
Nucleus 13C



in equilibrium with ring- opened form?



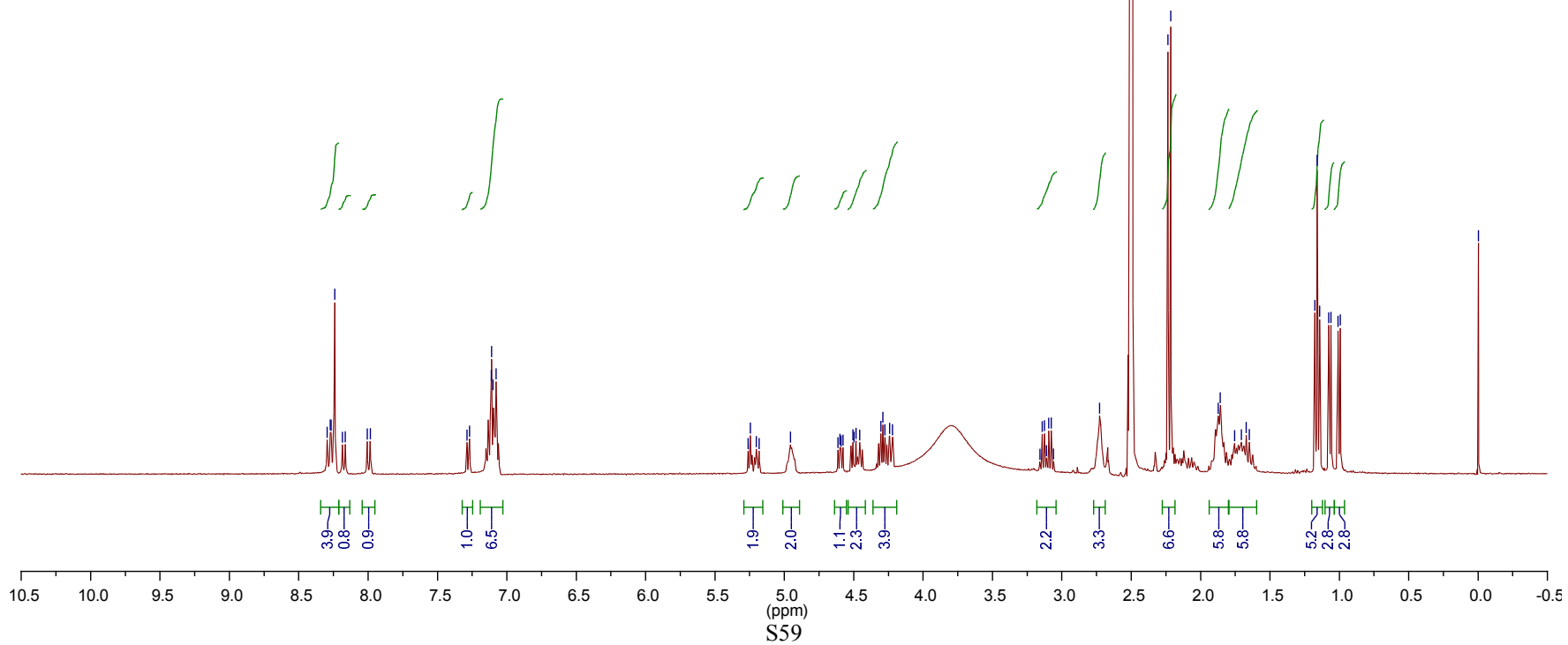
Title MV-347a formate salt  
 Solvent DMSO-D6  
 Number of Scans 24  
 Spectrometer Frequency 399.78  
 Nucleus 1H



8.296  
8.273  
8.268  
8.241  
8.186  
8.164  
8.007  
7.985  
7.287  
7.269  
7.113  
7.110  
7.100  
7.078

5.261  
5.246  
5.201  
5.182  
4.957  
4.497  
4.456  
4.304  
4.290  
4.275  
4.241  
4.220  
3.141  
3.124  
3.111  
3.094  
3.077  
3.060  
2.728  
2.500  
2.236  
2.215  
1.857  
1.706  
1.648  
1.176  
1.159  
1.142  
1.076  
1.060  
1.009  
0.994

-0.002



174.108  
169.871  
165.725  
165.238

137.596  
137.424  
137.022  
136.868  
128.705  
128.517  
128.333  
127.676  
126.673  
126.608  
125.831  
125.709

99.477

87.691  
87.573

73.374  
72.618

59.210  
58.704  
57.910

50.495  
50.258  
46.568  
46.549

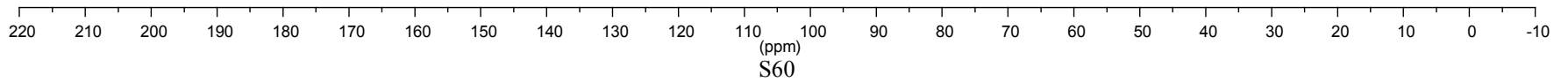
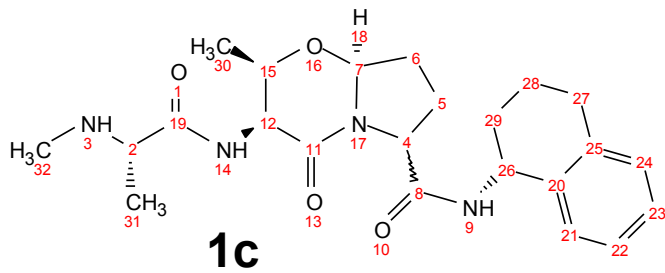
39.599

33.722

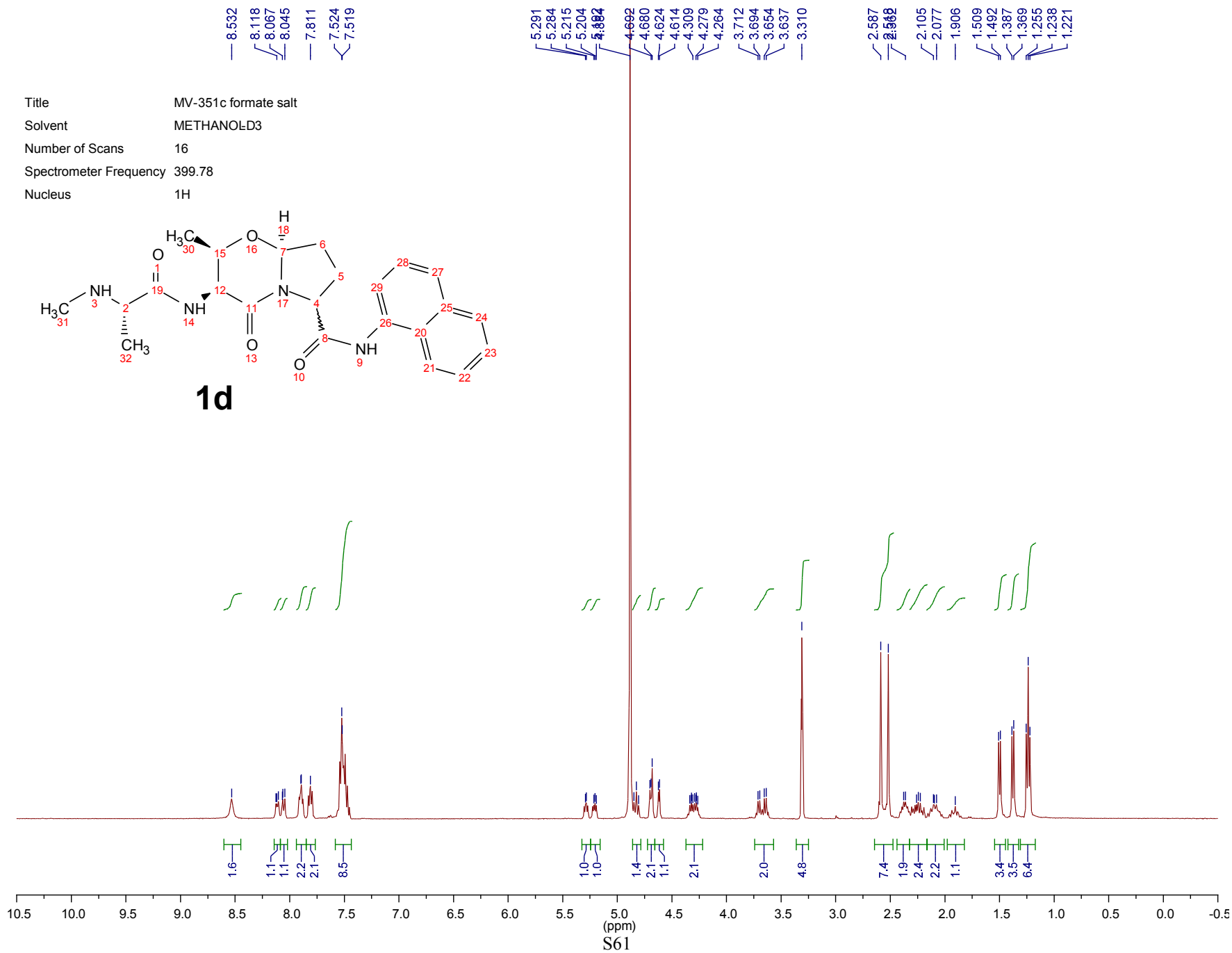
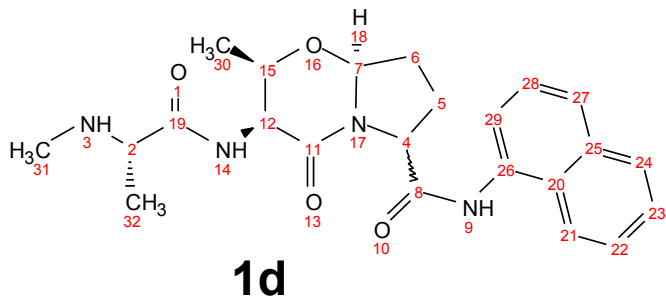
28.816  
25.669

20.459  
18.899  
18.676  
16.499

Title MV-347 formate salt  
Solvent DMSO-D6  
Number of Scans 8192  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C



Title MV-351c formate salt  
Solvent METHANOLD3  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



173.321  
173.190  
172.663  
172.352  
168.312  
167.510

135.729  
135.718  
134.023  
133.747  
130.560  
129.233  
128.147  
127.384  
127.218  
126.396  
124.252  
123.827

90.988  
90.871

76.324  
75.652

60.618  
59.541  
59.057  
58.950

52.759  
52.469

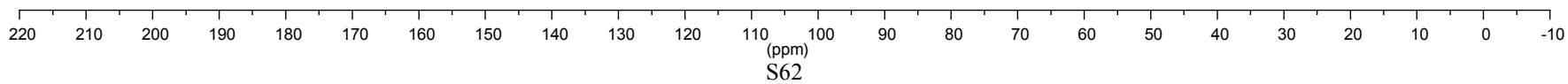
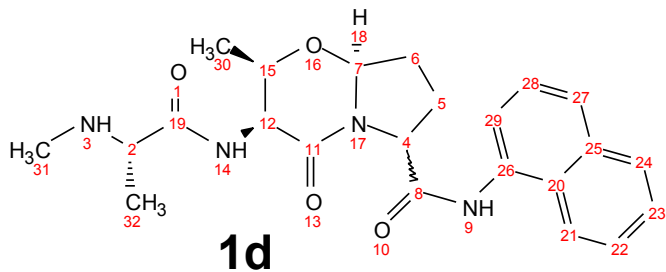
49.999

32.603  
32.432  
32.196  
31.160

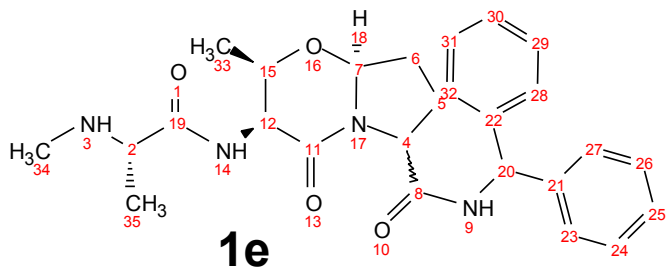
26.749  
26.157

17.311  
17.169  
16.658

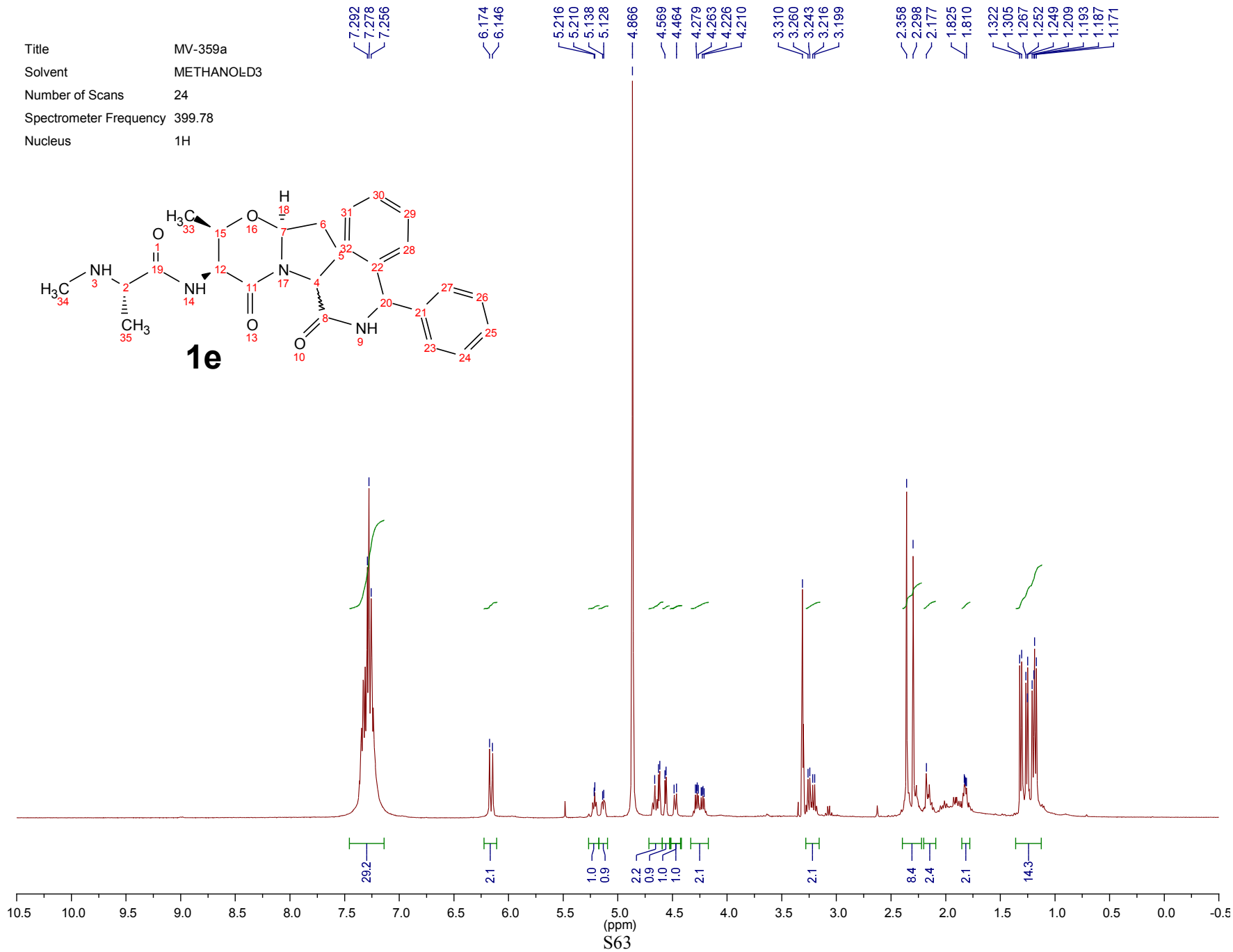
Title MV-351c formate salt  
Solvent METHANOL-D3  
Number of Scans 6180  
Spectrometer Frequency 100.53  
Nucleus 13C



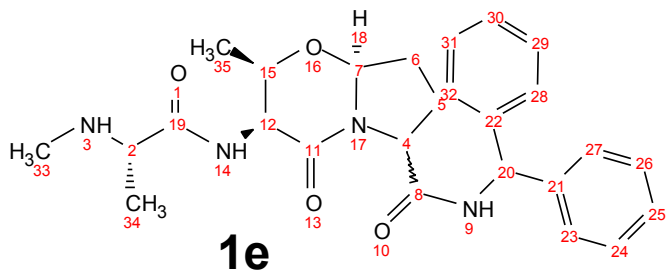
Title MV-359a  
Solvent METHANOL-D3  
Number of Scans 24  
Spectrometer Frequency 399.78  
Nucleus 1H



7.292  
7.278  
7.256  
6.174  
6.146  
5.216  
5.210  
5.138  
5.128  
4.866  
4.569  
4.464  
4.279  
4.263  
4.226  
4.210  
3.310  
3.260  
3.243  
3.216  
3.199  
2.358  
2.298  
2.177  
1.825  
1.810  
1.322  
1.305  
1.267  
1.252  
1.249  
1.209  
1.193  
1.187  
1.171



Title MV-359  
Solvent METHANOLD3  
Number of Scans 8192  
Spectrometer Frequency 100.53  
Nucleus 13C



176.759  
176.588  
172.718  
168.018  
167.551

142.970  
142.778  
142.623  
142.590  
129.486  
129.373  
128.909  
128.776  
128.735  
128.630  
128.536  
128.460  
128.311  
128.132

90.696  
90.602

76.332  
75.545

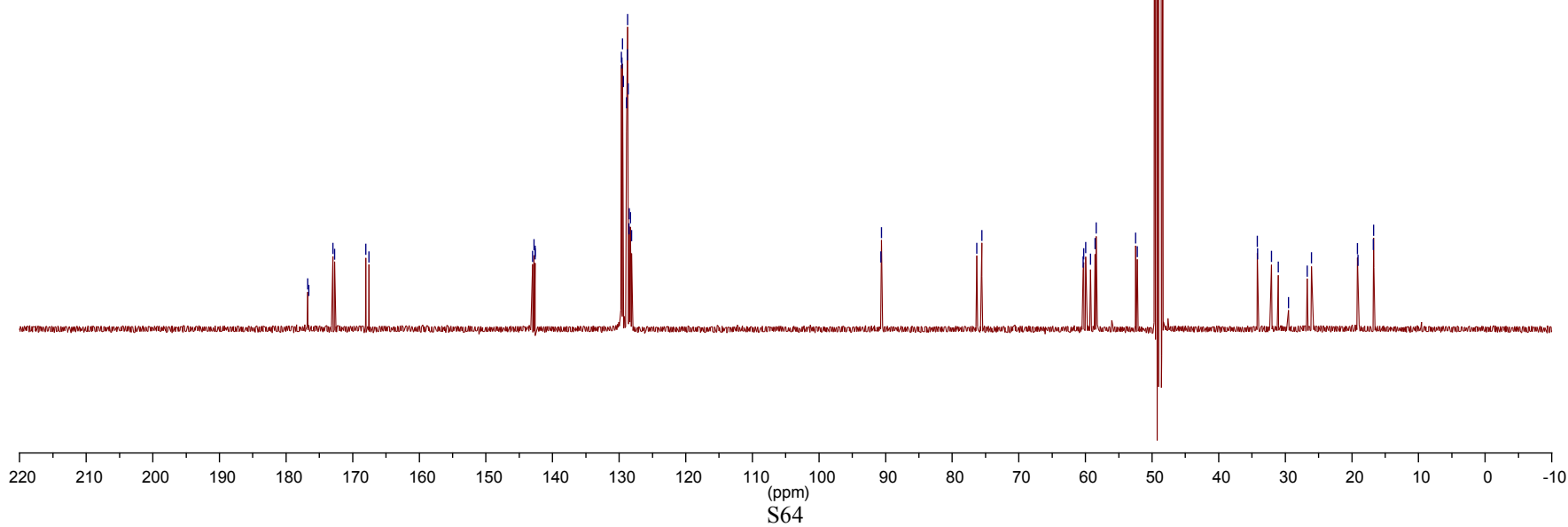
59.976  
59.224  
58.548  
58.378  
52.462  
52.195

49.609

34.224

32.063  
31.069  
29.539  
26.727  
26.073

19.163  
19.086  
16.773  
16.730





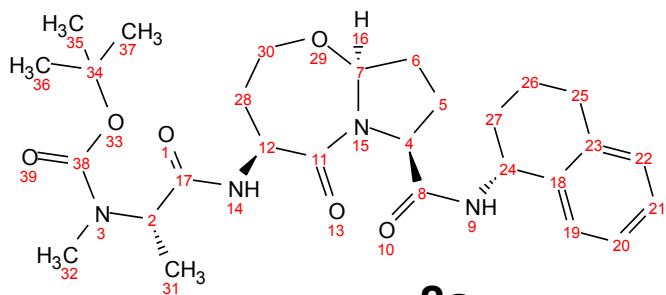
Title MV-465b low Rf  
 Solvent CHLOROFORMD  
 Number of Scans 16  
 Spectrometer Frequency 399.78  
 Nucleus 1H

7.263  
 7.260  
 7.204  
 7.154  
 7.142  
 7.121  
 7.077  
 6.848  
 6.828

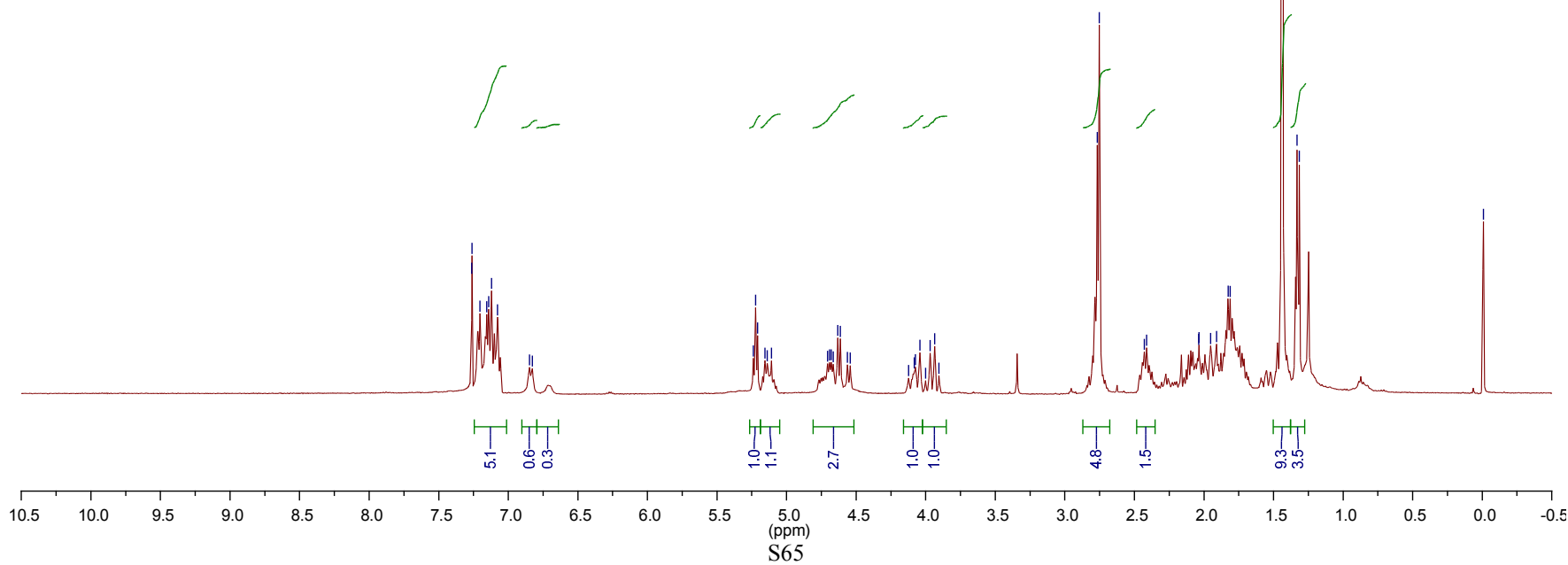
5.239  
 5.223  
 5.207  
 5.155  
 5.139  
 5.109  
 4.632  
 4.613  
 4.563  
 4.543  
 4.083  
 4.074  
 4.042  
 4.001  
 3.968  
 3.936  
 3.905

2.768  
 2.752  
 2.428  
 2.408  
 1.910  
 1.827  
 1.812  
 1.443  
 1.434  
 1.332  
 1.314

-0.008



**8a**  
 + minor isomer



171.378  
169.851  
169.757

137.559  
137.326  
136.943  
136.691  
129.285  
129.200  
128.570  
128.303  
127.362  
127.289  
126.351  
126.179

90.282  
90.027

77.158  
70.684  
70.572

61.139  
60.622

53.138  
52.608

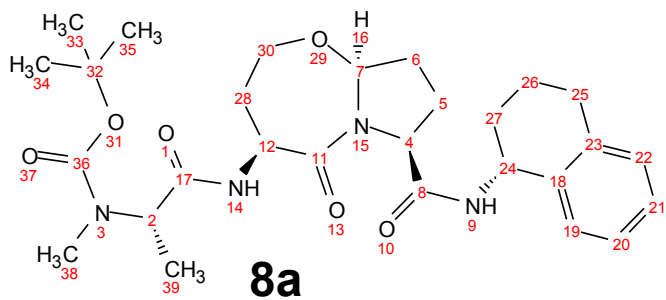
47.741  
46.516  
46.246

29.312  
29.282  
28.431  
28.402  
25.941

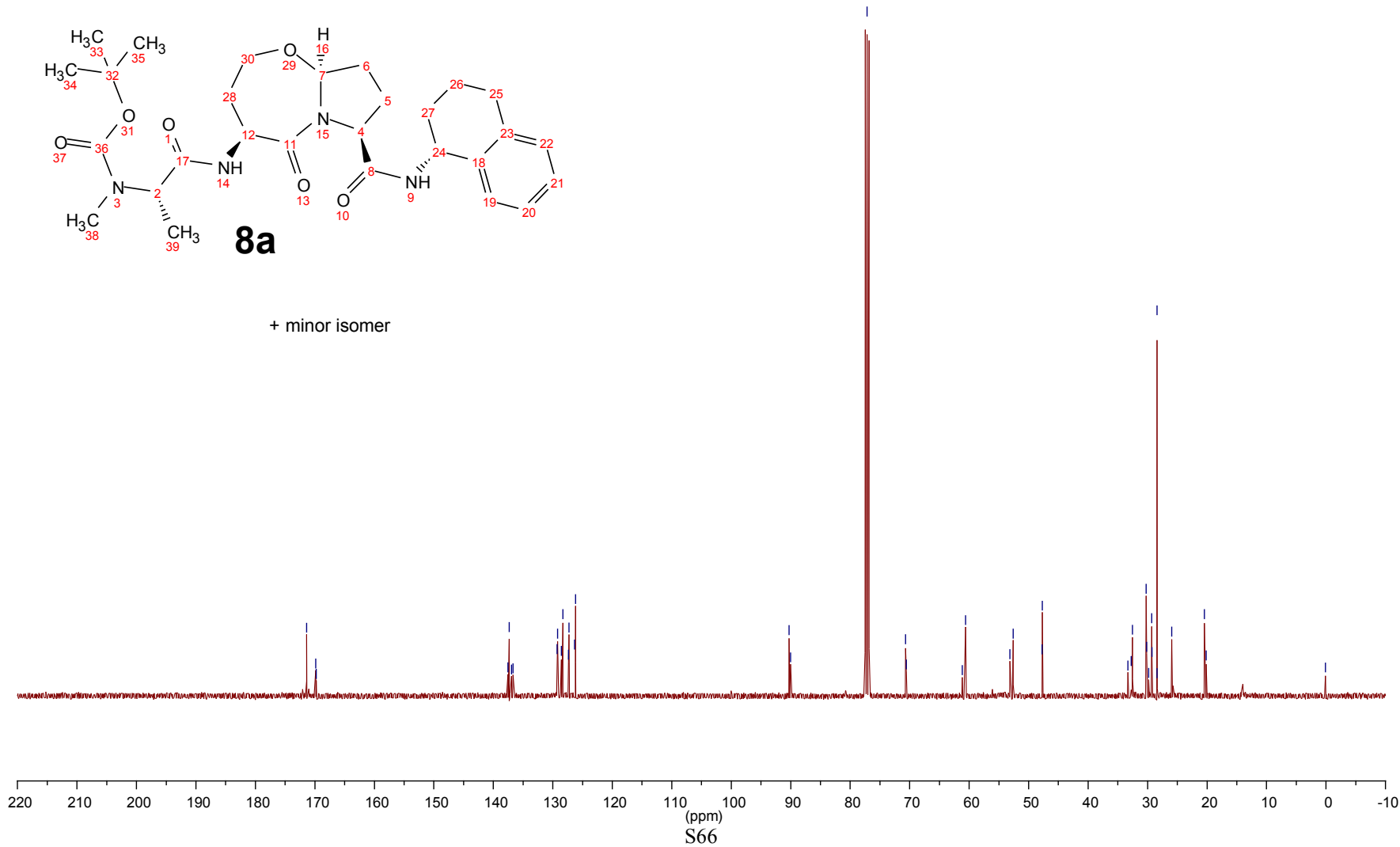
20.454  
20.140

0.113

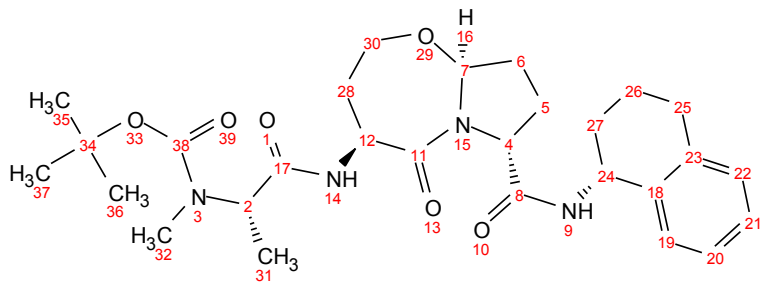
Title MV-465b low Rf  
Solvent CHLOROFORMD  
Number of Scans 6140  
Spectrometer Frequency 100.53  
Nucleus 13C



+ minor isomer



Title MV-465d\_upper\_Rf  
Solvent CHLOROFORMD  
Number of Scans 24  
Spectrometer Frequency 399.78  
Nucleus 1H



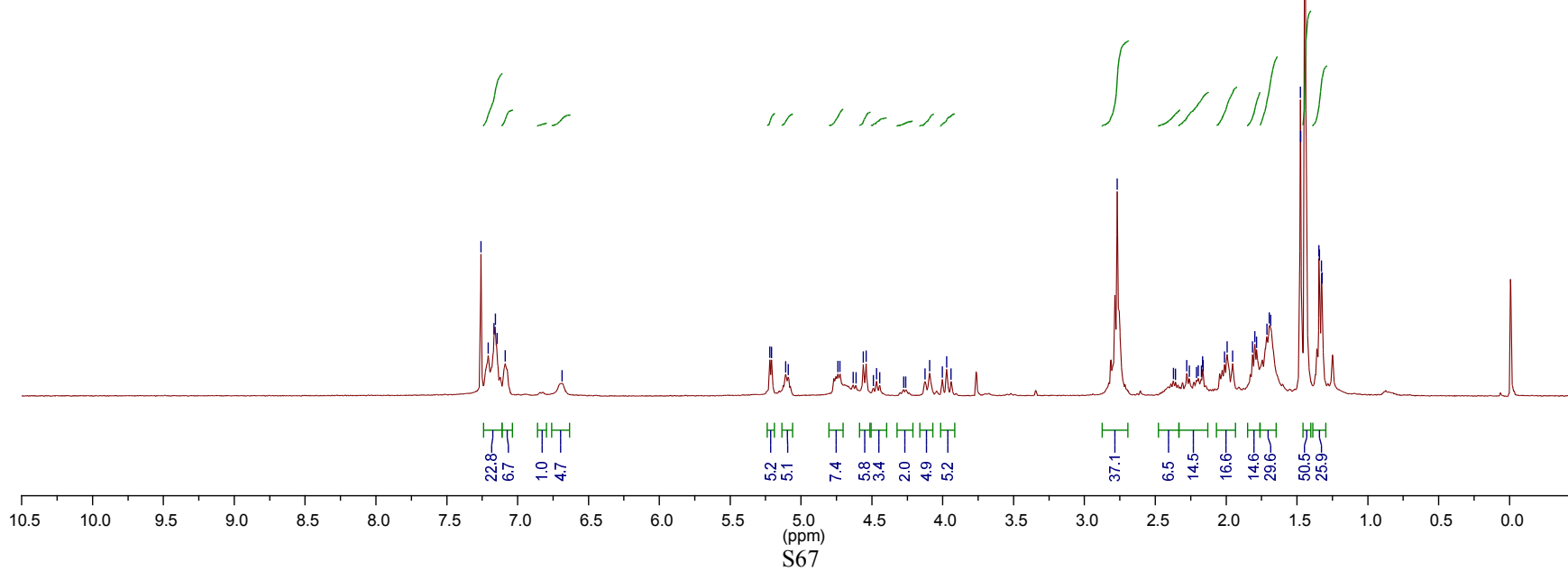
**9a**

+ minor isomer

7.260  
7.208  
7.167  
7.158  
7.145  
7.088  
6.687

5.221  
5.207  
5.108  
5.091  
4.739  
4.561  
4.490  
4.445  
4.277  
4.261  
4.125  
4.093  
4.003  
3.972  
3.941

2.769  
2.278  
2.196  
2.162  
1.953  
1.685  
1.476  
1.473  
1.445  
1.441  
1.344  
1.341  
1.327  
1.323



172.469  
172.094  
171.035  
169.757

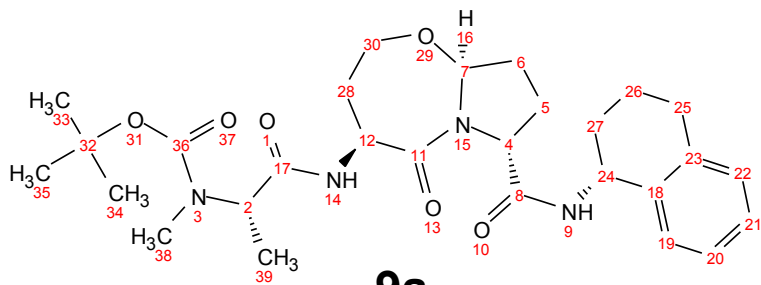
137.582  
136.706  
129.308  
128.594  
127.385  
126.373

— 90.049

— 70.597  
— 65.992  
— 61.162  
— 53.161  
— 47.761

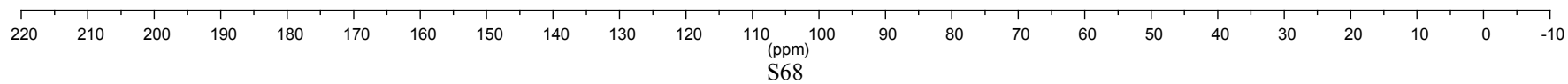
33.335  
32.703  
30.168  
29.303  
28.456  
25.751  
— 20.157  
— 14.017

Title MV-465d\_upper\_Rf  
Solvent CHLOROFORMD  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C



**9a**

+ minor isomer



(ppm)

S68

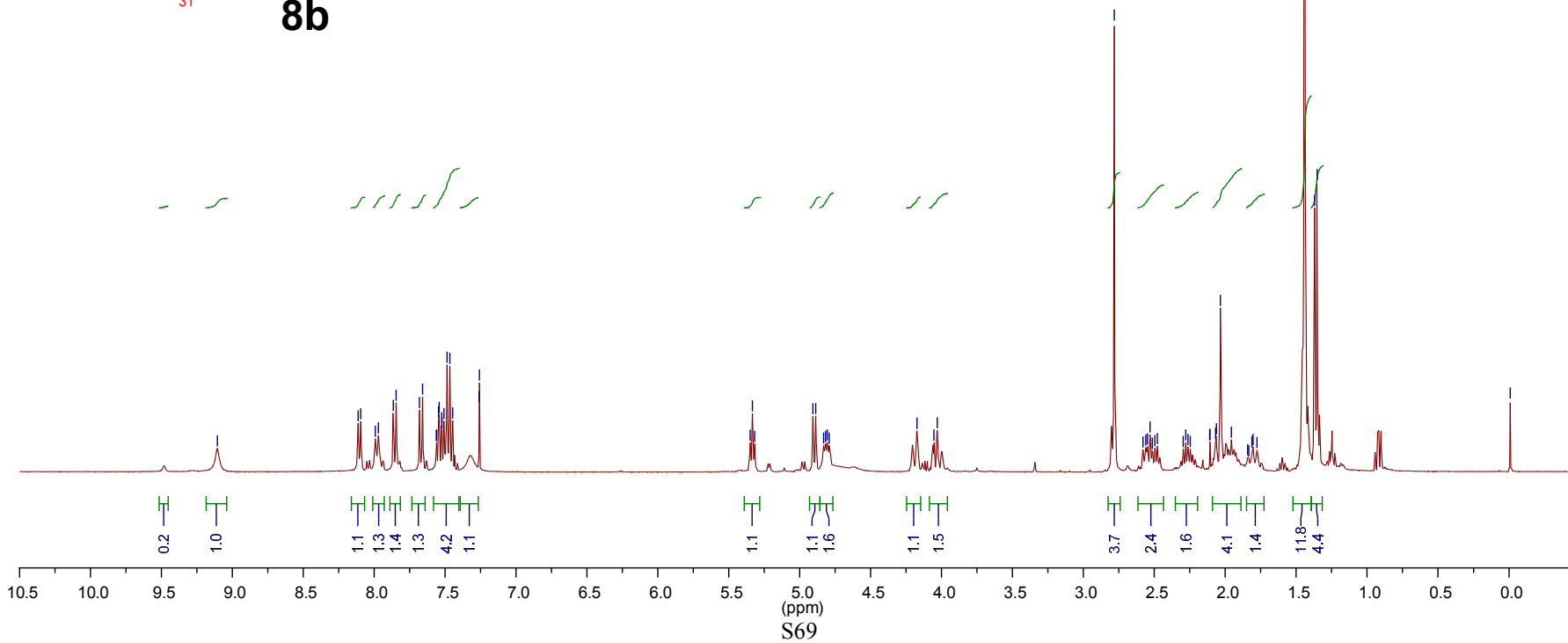
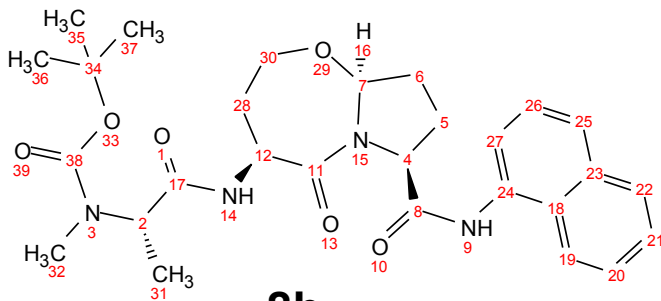
9.106  
 8.114  
 8.095  
 7.992  
 7.970  
 7.866  
 7.846  
 7.543  
 7.447  
 7.260  
 7.259

5.350  
 5.333  
 5.317  
 4.888  
 4.832  
 4.819  
 4.806  
 4.792  
 4.174  
 4.053  
 4.030

2.783  
 2.479  
 2.246  
 2.034  
 1.833  
 1.776  
 1.441  
 1.370  
 1.353

-0.008

Title MV-420a low Rf  
 Owner mitchell  
 Number of Scans 16  
 Spectrometer Frequency 399.78  
 Nucleus <sup>1</sup>H



172.051  
171.387  
171.363  
171.355  
169.062  
168.508

134.132  
132.662  
126.483  
125.475  
120.689  
119.754

90.703

77.160

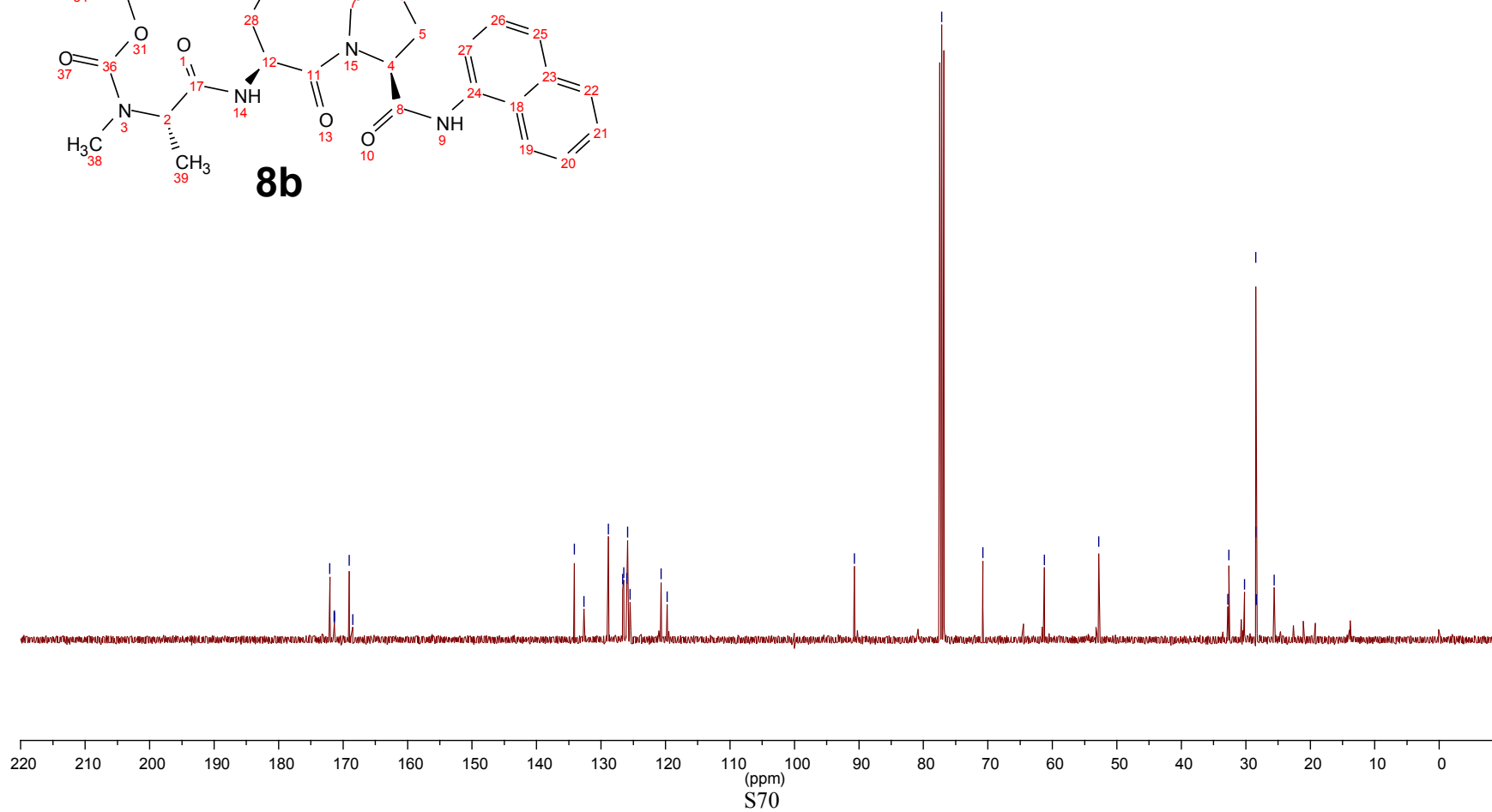
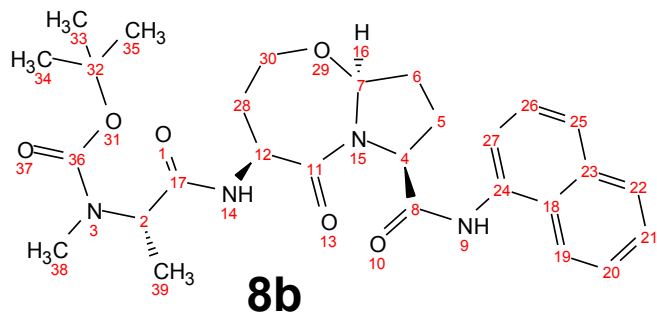
70.795

61.241

52.796

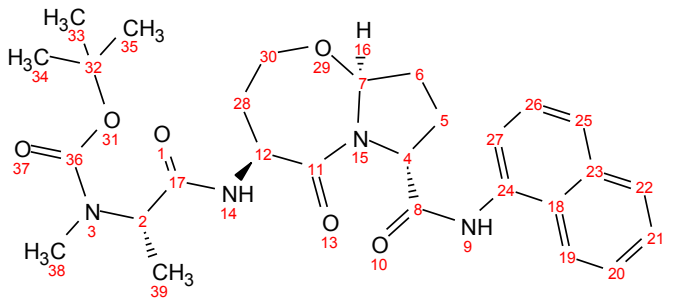
32.799  
32.617  
30.214  
28.461  
28.440  
28.419  
28.408  
25.616

Title MV-420a low Rf  
Solvent CHLOROFORMD  
Number of Scans 2048  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C

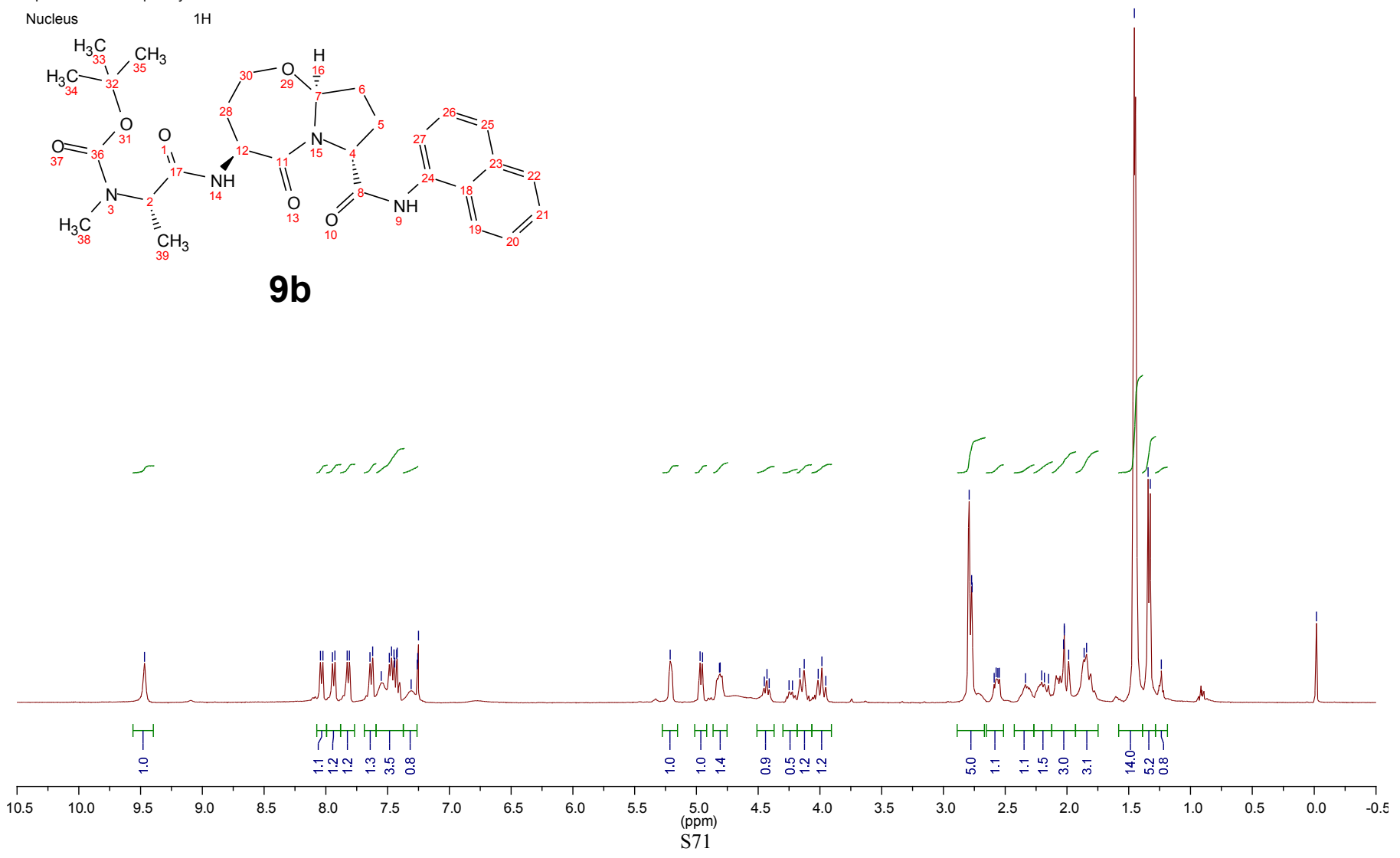


9.469  
 8.043  
 8.025  
 7.946  
 7.927  
 7.828  
 7.809  
 7.622  
 7.486  
 7.469  
 7.449  
 7.442  
 7.428  
 7.423  
 7.310  
 7.260  
 7.257  
 7.251  
 5.212  
 4.952  
 4.815  
 4.808  
 4.429  
 4.412  
 4.250  
 4.164  
 4.127  
 4.016  
 3.985  
 3.953  
 2.792  
 2.773  
 2.768  
 2.548  
 2.204  
 2.150  
 2.024  
 1.987  
 1.843  
 1.456  
 1.344  
 1.326  
 1.237  
 -0.018

Title MV-420 upper Rf  
 Solvent CHLOROFORMD  
 Number of Scans 16  
 Spectrometer Frequency 399.78  
 Nucleus 1H



9b



174.984  
173.267  
172.427  
168.507

134.126  
132.799  
126.054  
125.379  
121.007  
119.481

90.269

77.160

70.591

65.938

61.610

53.229

49.221

33.572

30.320

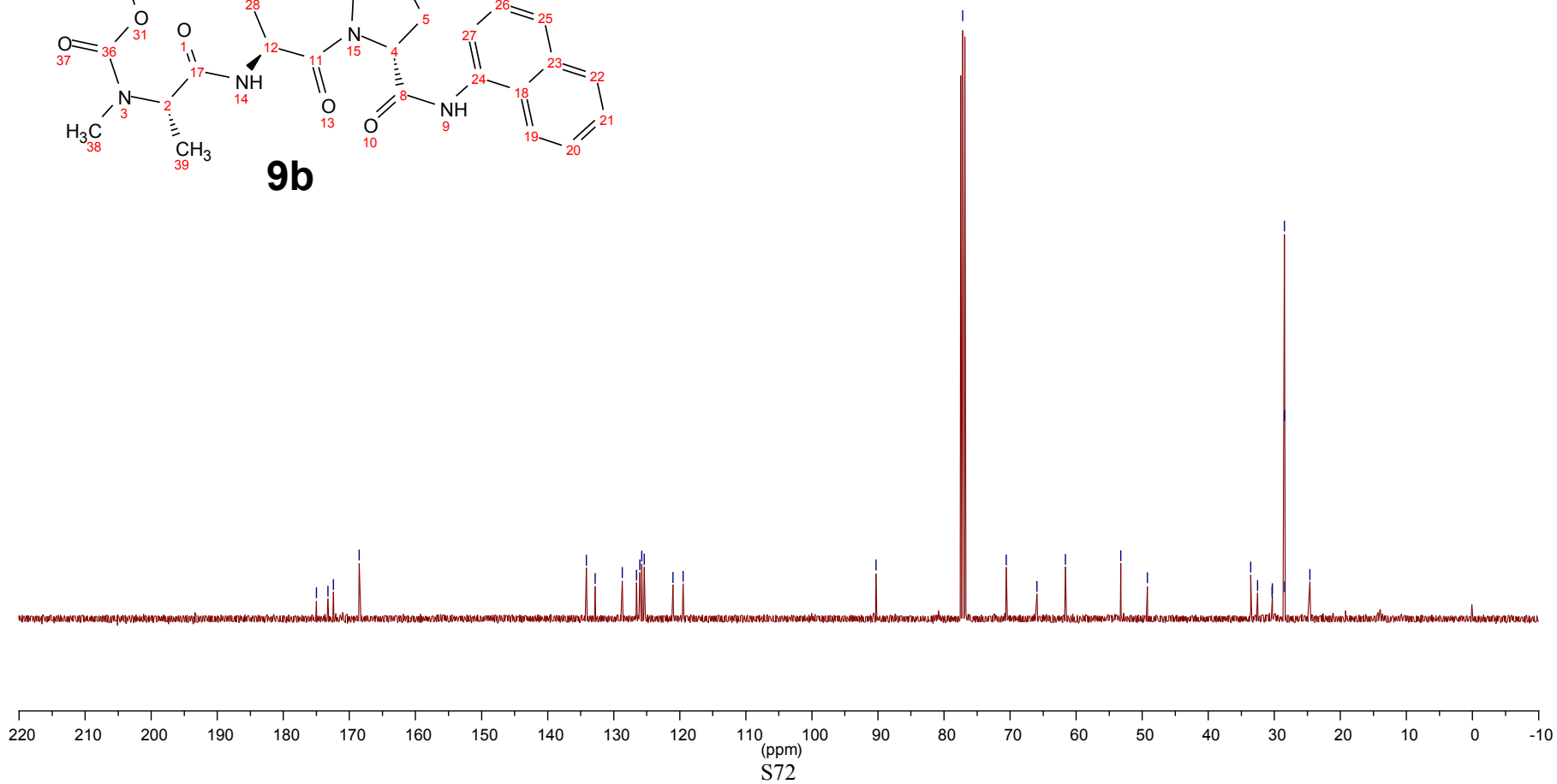
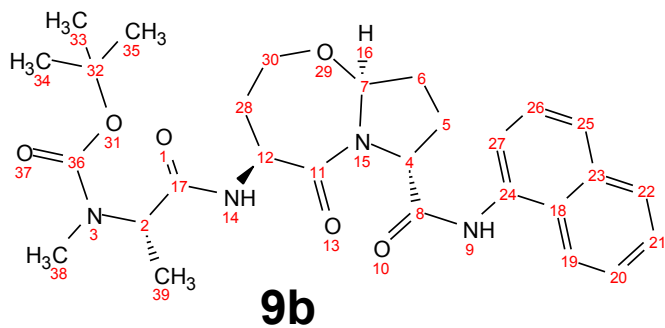
28.472

28.451

28.439

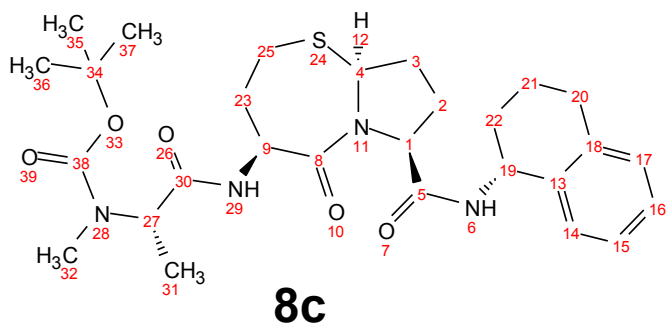
24.625

Title MV-420 upper Rf  
Solvent CHLOROFORMD  
Number of Scans 2048  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C





Title MV-466 low Rf  
Solvent CHLOROFORMD  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H

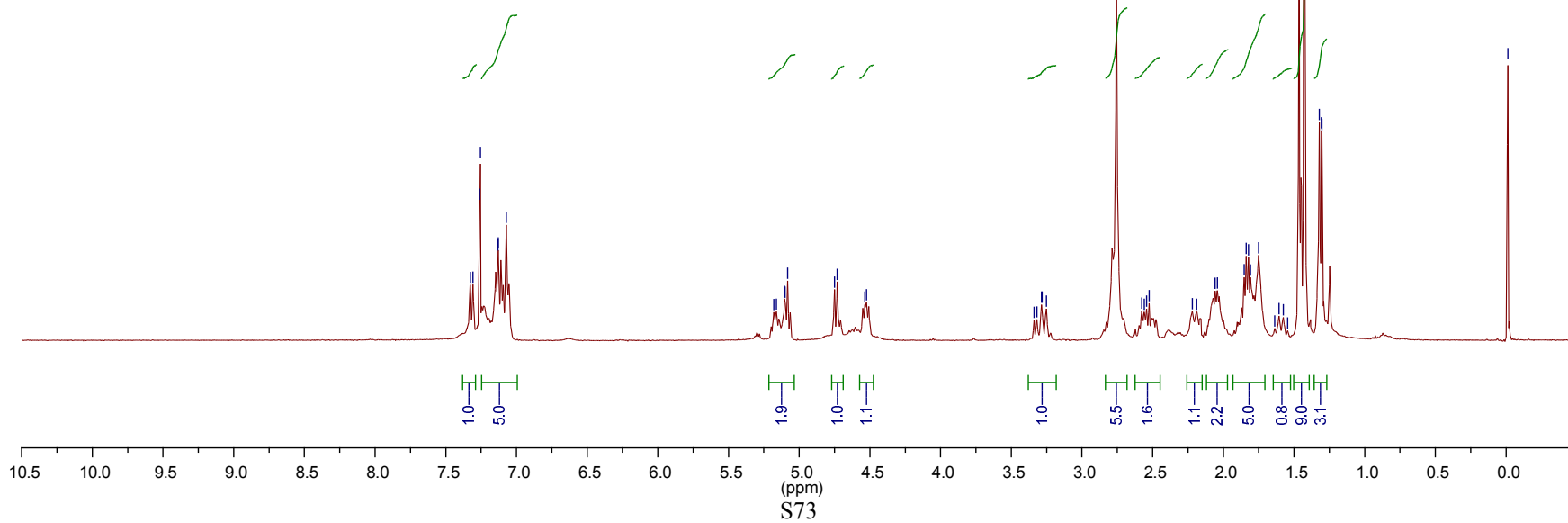


7.327  
7.308  
7.308  
7.260  
7.266  
7.131  
7.128  
7.071

5.180  
5.161  
5.105  
5.100  
5.082  
4.751  
4.731  
4.535  
4.524

3.339  
3.318  
3.287  
3.283  
3.252  
2.757  
2.754  
2.525  
2.058  
2.042  
1.751  
1.545  
1.467  
1.463  
1.430  
1.426  
1.320  
1.306  
1.302

-0.012



171.328  
169.564  
169.332

137.299  
129.172  
129.124  
128.847  
127.243  
126.057

77.466

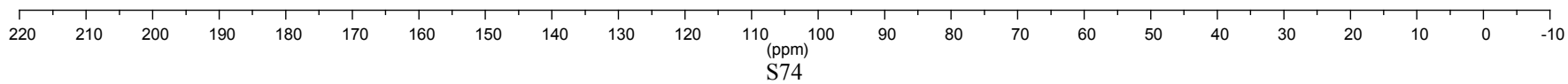
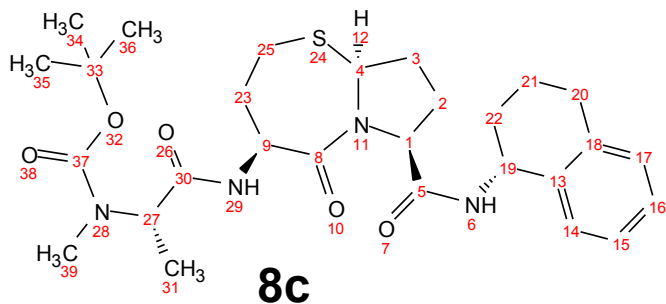
62.260  
61.754

52.827  
47.613

30.404  
30.215  
29.326  
28.445  
28.428  
26.543  
20.497

0.118

Title MV-466 low Rf  
Solvent CHLOROFORMD  
Number of Scans 1424  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C



Title MV-466d upper Rf  
 Solvent CHLOROFORMD  
 Number of Scans 16  
 Spectrometer Frequency 399.78  
 Nucleus 1H

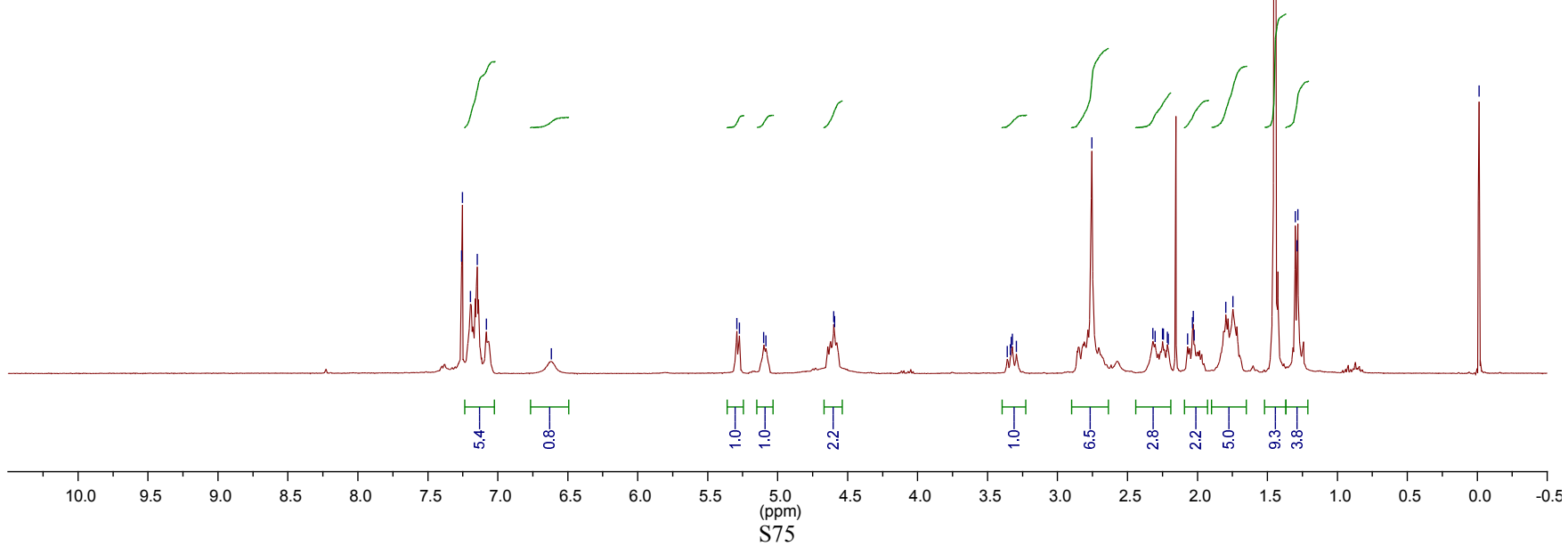
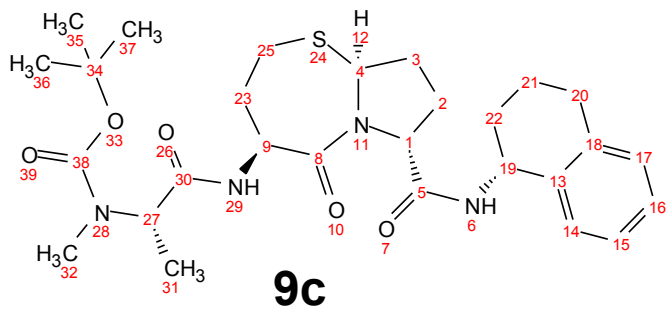
7.260  
 7.253  
 7.196  
 7.147  
 7.082  
 — 6.618

5.292  
 5.273  
 5.099  
 5.083  
 4.599  
 4.594

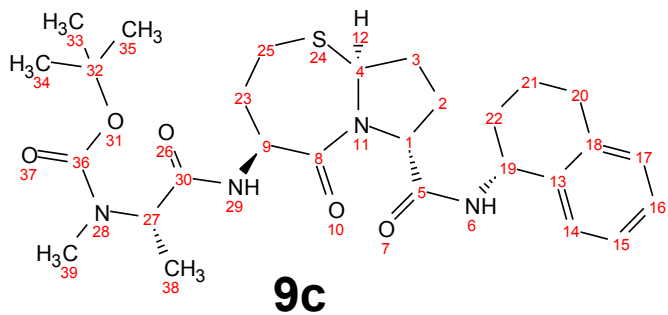
3.359  
 3.337  
 3.330  
 3.323  
 3.293

— 2.754  
 2.303  
 2.244  
 2.208  
 — 2.023  
 — 1.747  
 1.451  
 1.445  
 1.301  
 1.289  
 1.283

— -0.013



Title MV-466 upper Rf  
Solvent CHLOROFORMD  
Number of Scans 1824  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C



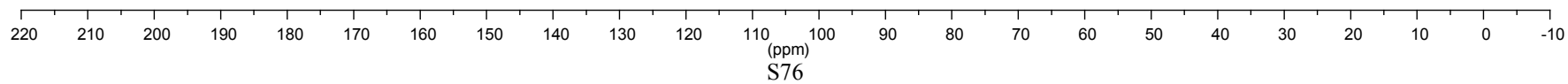
170.866  
169.631

137.582  
136.648  
129.295  
128.647  
127.373  
126.326

77.466

63.815  
61.252  
53.528  
47.743

33.662  
31.706  
30.085  
29.287  
28.486  
28.449  
20.080



S76

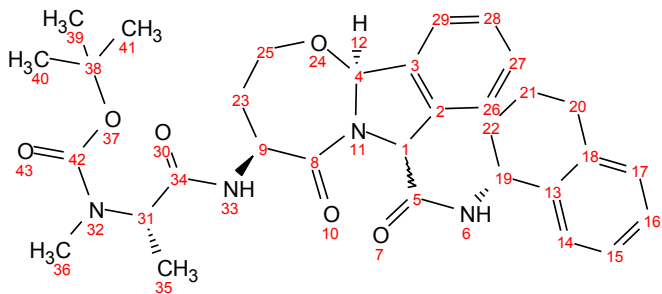
7.549  
7.478  
7.467  
7.458  
7.448  
7.438  
7.427  
7.418  
7.408  
7.403  
7.395  
7.387  
7.367  
7.293  
7.260  
7.166  
7.154  
7.144  
7.087  
7.063  
7.043  
7.041  
7.025  
7.023  
6.993  
6.976  
6.900  
6.729  
6.400  
6.367  
6.214

5.499  
5.146  
5.033  
5.019  
4.840  
4.825  
4.443  
4.417  
4.274  
4.247

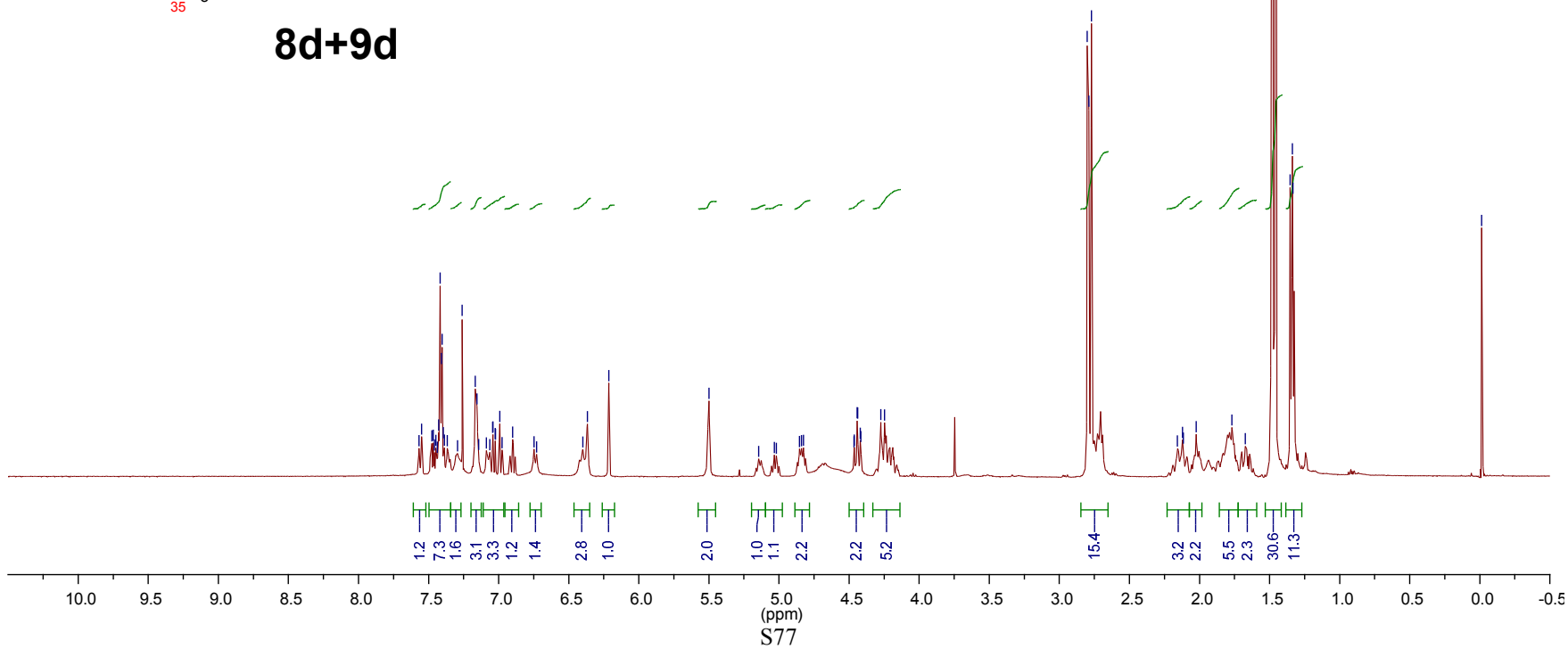
2.800  
2.789  
2.771  
2.157  
2.121  
2.116  
2.023  
1.673  
1.486  
1.459  
1.354  
1.336  
1.334

-0.014

Title MV-442a  
Solvent CHLOROFORMD  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



**8d+9d**



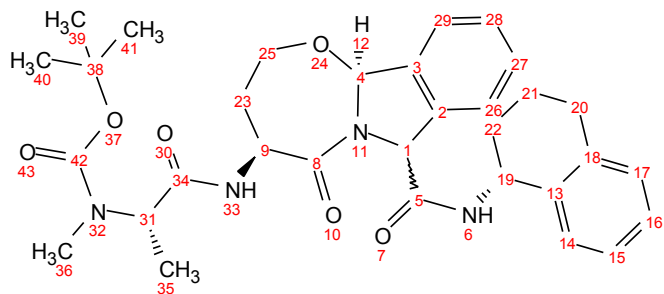
174.970  
172.420  
170.488  
168.331  
168.065

137.154  
136.506  
136.472  
135.855  
135.195  
128.987  
127.423  
127.157  
126.361  
126.183  
125.036  
125.003  
122.913

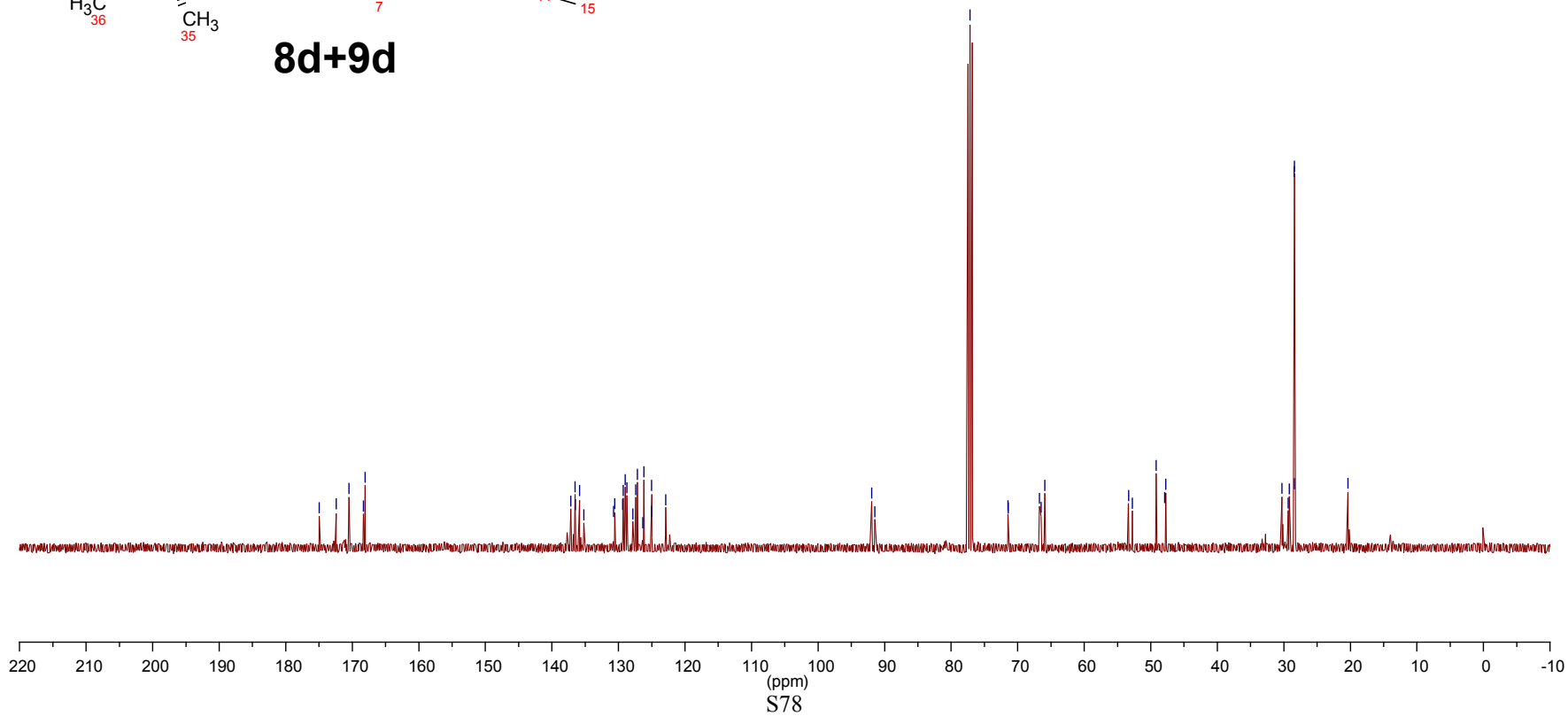
91.955  
91.473

77.160  
71.426  
66.796  
66.475  
65.925  
53.343  
52.786  
49.195  
47.934  
47.738  
30.301  
29.335  
29.173  
28.458  
28.434  
28.415  
20.383

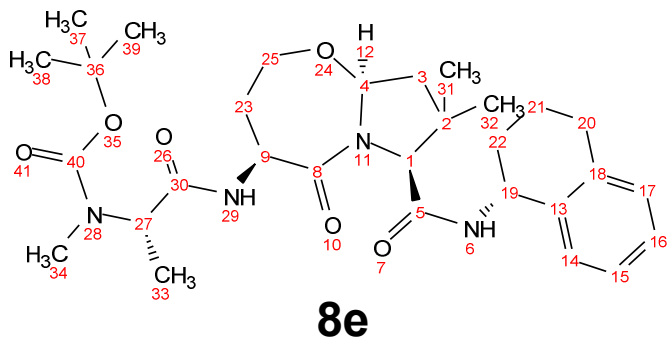
Title MV-442  
Solvent CHLOROFORMD  
Number of Scans 1024  
Spectrometer Frequency 100.53  
Nucleus 13C



8d+9d



Title MV-435 low Rf  
 Solvent CHLOROFORMD  
 Number of Scans 16  
 Spectrometer Frequency 399.78  
 Nucleus 1H



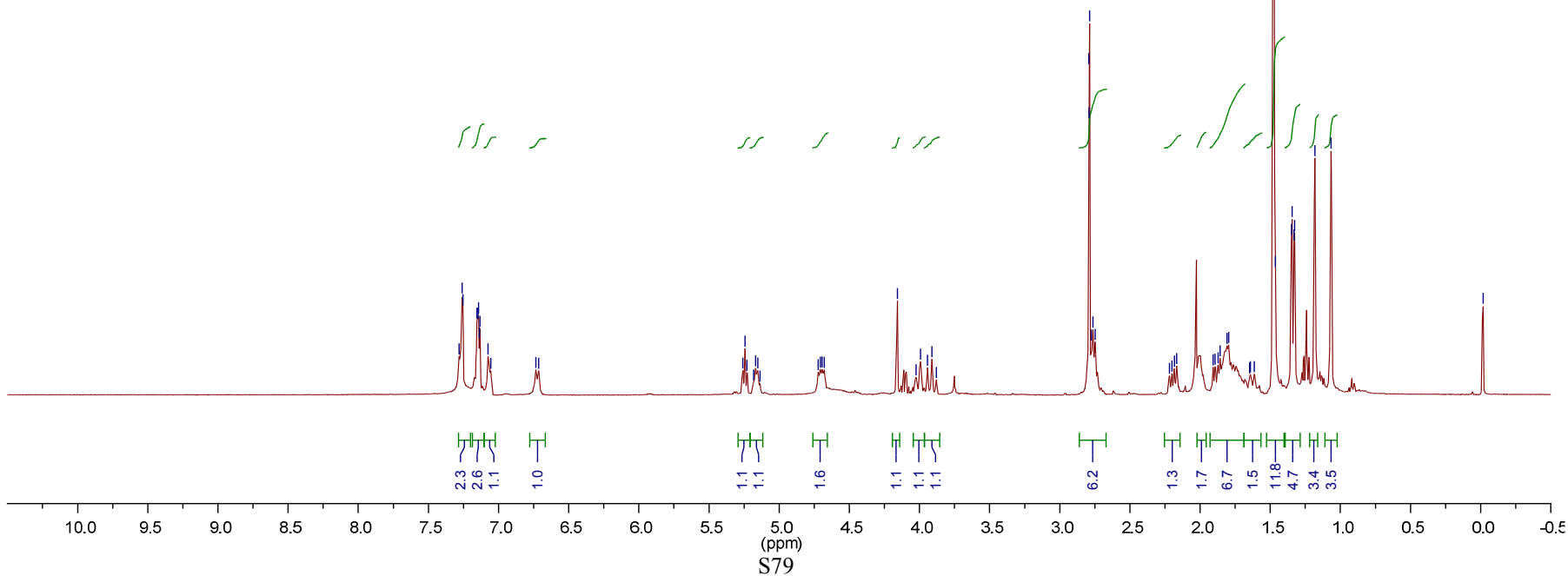
7.280  
7.260  
7.254  
7.155  
7.147  
7.142  
7.137  
7.133  
7.074  
7.056  
6.734  
6.714

5.258  
5.243  
5.229  
5.183  
5.169  
5.152  
5.137  
4.721  
4.707  
4.693  
4.679  
4.157  
4.025  
3.993  
3.942  
3.911  
3.880

2.793  
2.791  
2.788  
2.775  
2.763  
2.748  
2.748  
2.183  
2.166

1.796  
1.613  
1.348  
1.326  
1.181  
1.066

-0.017



170.720  
168.785

137.265  
136.663  
136.046  
129.226  
128.852  
127.377  
126.359

89.277  
89.248

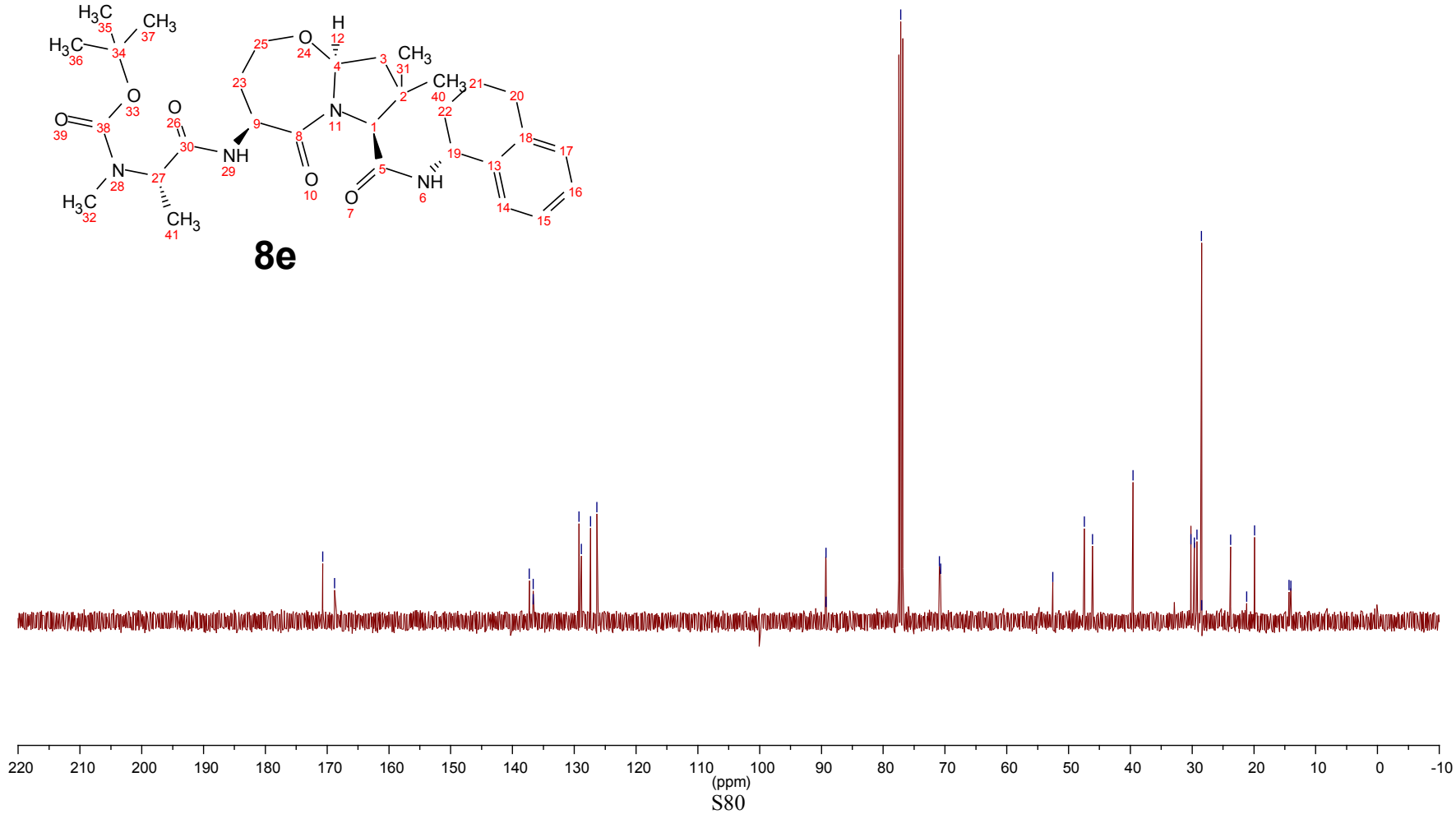
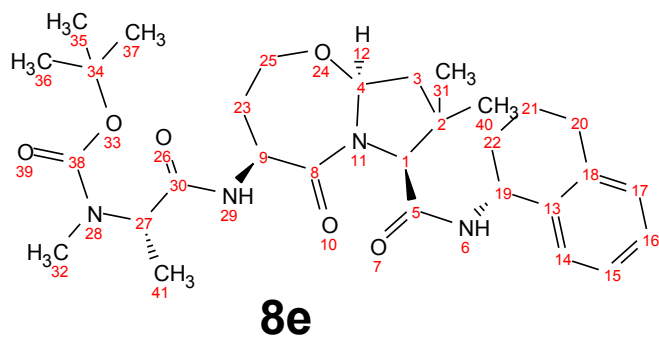
77.160  
70.899  
70.713

52.572  
47.455  
46.111

39.552  
29.234  
28.478  
28.450  
23.777

19.891  
14.317  
14.025

Title MV-435 low Rf  
Solvent CHLOROFORMD  
Number of Scans 528  
Spectrometer Frequency 100.53  
Nucleus 13C



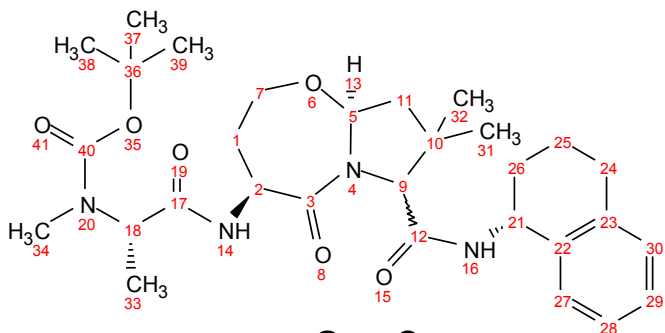


7.334  
7.288  
7.289  
7.212  
7.206  
7.179  
7.175  
7.166  
7.160  
7.157  
7.154  
7.098  
7.089  
— 6.888

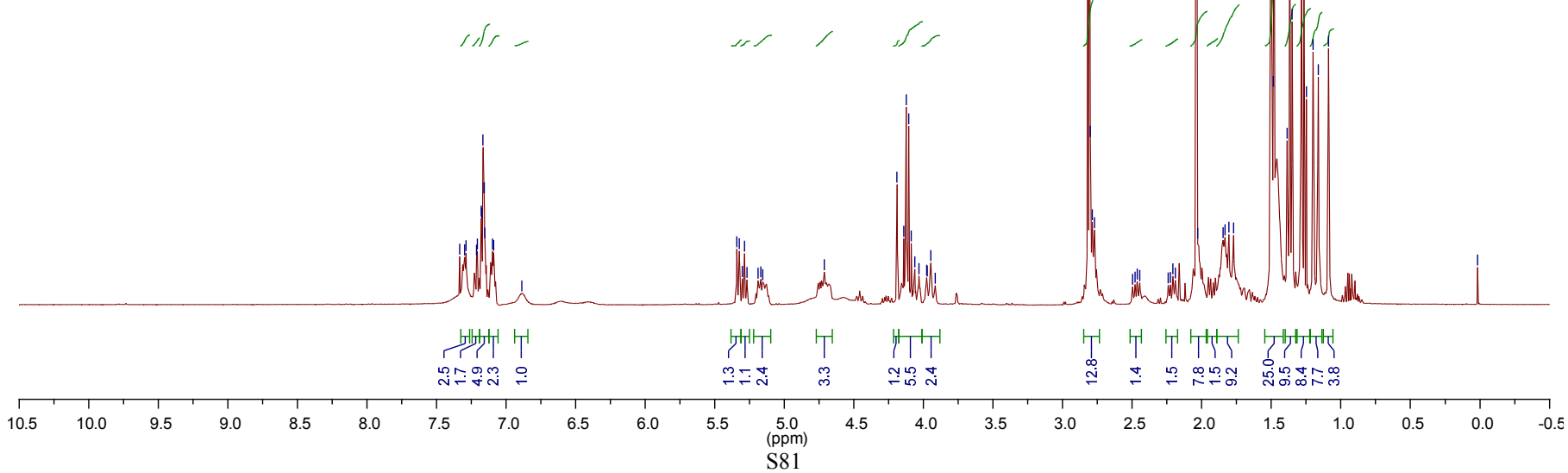
5.341  
5.324  
5.302  
5.286  
5.269  
5.190  
5.169  
5.156  
— 4.713  
4.124  
4.062  
4.032  
3.977  
3.973  
3.947  
3.915

2.819  
2.807  
2.801  
2.787  
2.779  
2.771  
2.446  
— 2.190  
— 2.029  
— 1.771  
— 1.501  
1.283  
1.265  
1.247  
1.199  
1.163  
1.088  
— 0.017

Title MV-435a mixed fractions  
Solvent CHLOROFORMD  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



8e+9e



172.279  
171.102  
170.857  
170.589  
169.407  
168.675

137.665  
137.084  
136.512  
136.224  
129.185  
129.038  
128.923  
128.720  
127.238  
127.207  
127.188  
126.184  
126.098

90.129  
89.102

77.160

70.513

65.768

60.328

53.185

52.443

47.276

45.900

39.337

39.216

33.110

28.308

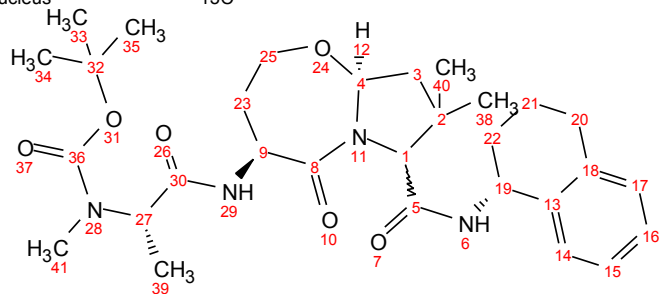
20.973

19.647

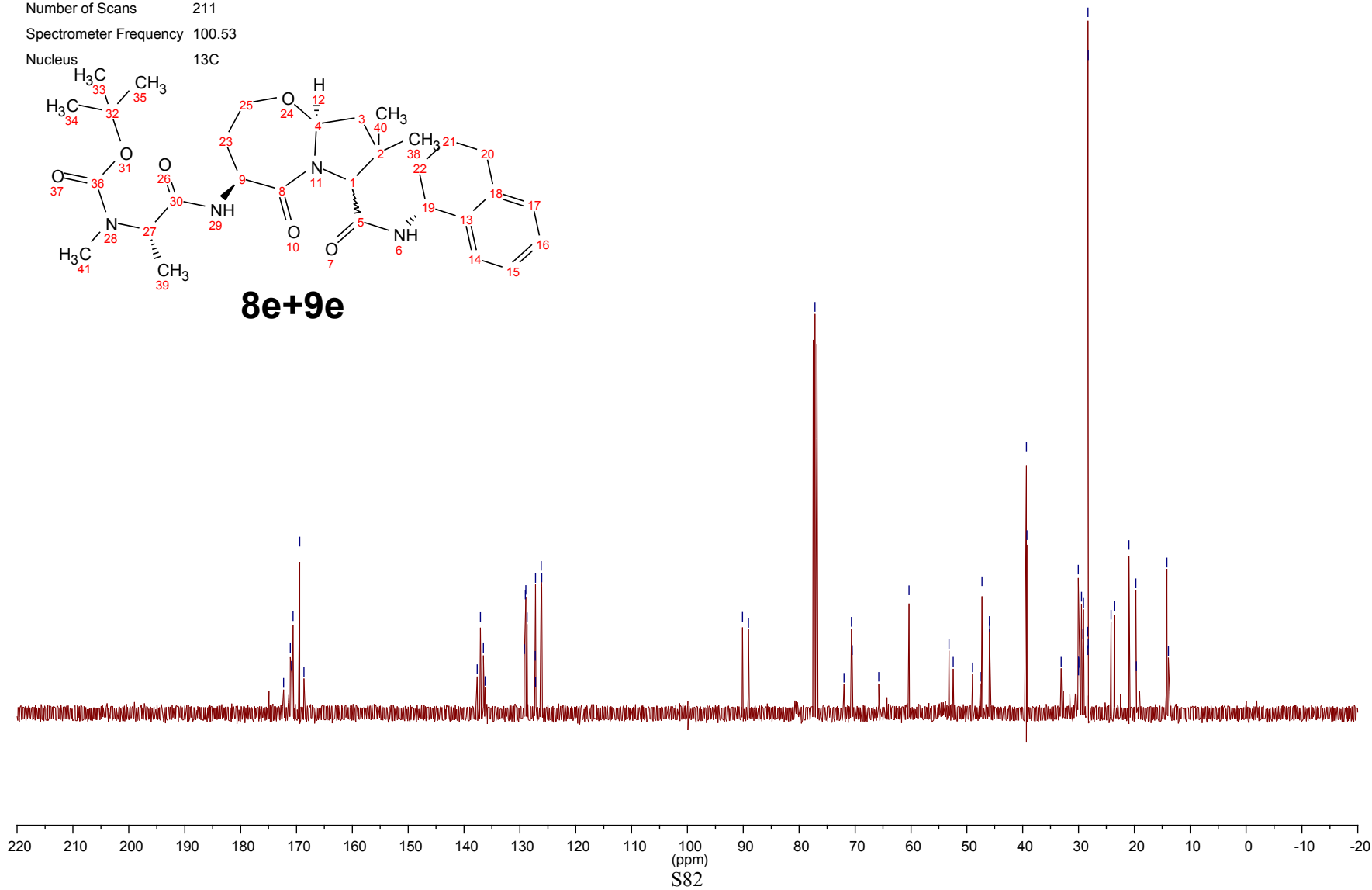
14.154

13.922

Title MV-435a mixed fractions  
Solvent CHLOROFORMD  
Number of Scans 211  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C



8e+9e



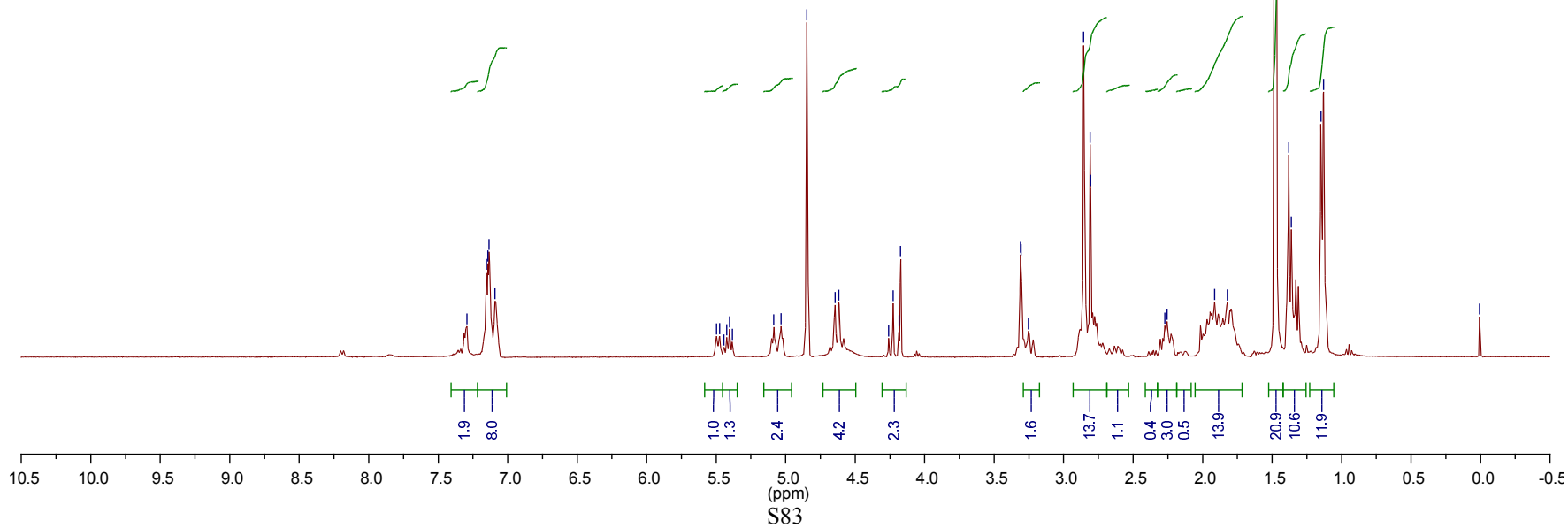
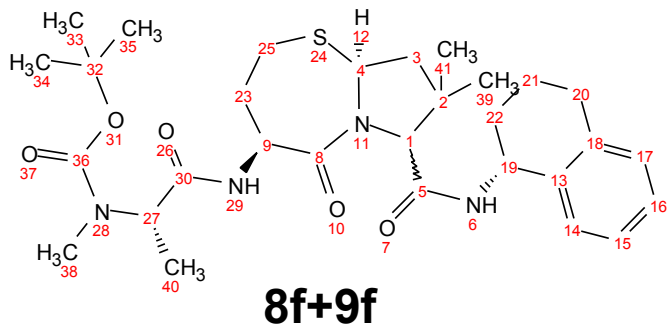
Title MV-476b mixed fractions  
Solvent METHANOL-D3  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H

7.293  
7.153  
7.144  
7.134  
7.091

5.498  
5.475  
5.444  
5.425  
5.404  
5.384  
5.085  
5.033  
4.847  
4.617  
4.258  
4.227  
4.185  
4.173

3.310  
3.306  
3.253  
2.856  
2.809  
2.806  
2.272  
2.255  
1.914  
1.822  
1.475  
1.380  
1.361  
1.149  
1.130

0.007



172.707  
171.760  
171.381

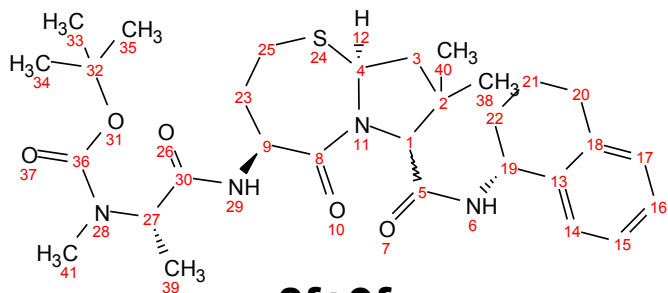
138.808  
138.513  
137.406  
137.392  
130.239  
130.114  
130.040  
130.027  
129.801  
129.776  
128.467  
128.275  
128.184  
127.156

73.309  
73.287

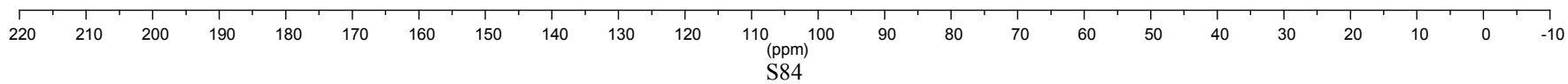
63.858  
61.949  
61.747  
54.218  
54.061

47.221  
40.943  
40.864  
40.751  
31.311  
30.220  
28.671  
25.275  
23.899  
21.310  
20.995

Title MV-476b mixed fractions  
Solvent METHANOL-D3  
Number of Scans 1200  
Spectrometer Frequency 100.53  
Nucleus 13C



8f+9f



Title MV-476a upper Rf  
 Solvent CHLOROFORMD  
 Number of Scans 16  
 Spectrometer Frequency 399.78  
 Nucleus 1H

7.314  
 7.300  
 7.254  
 7.251  
 7.173  
 7.167  
 7.163  
 7.157  
 7.146  
 7.141  
 7.092  
 7.074

6.006  
 5.984

5.345  
 5.323

5.112  
 5.099

4.513  
 4.500  
 4.490

4.056  
 4.046  
 4.039  
 4.036  
 4.023

3.314  
 3.280  
 3.248

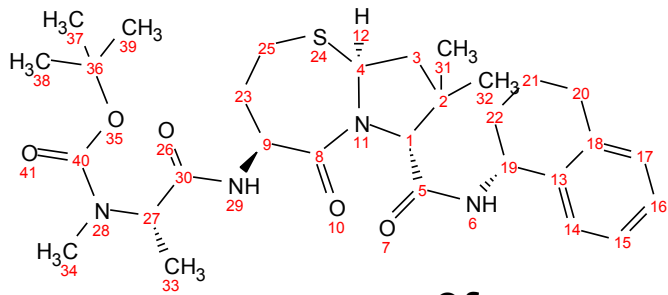
2.755

2.318  
 2.311  
 2.288

1.825  
 1.815  
 1.789  
 1.765

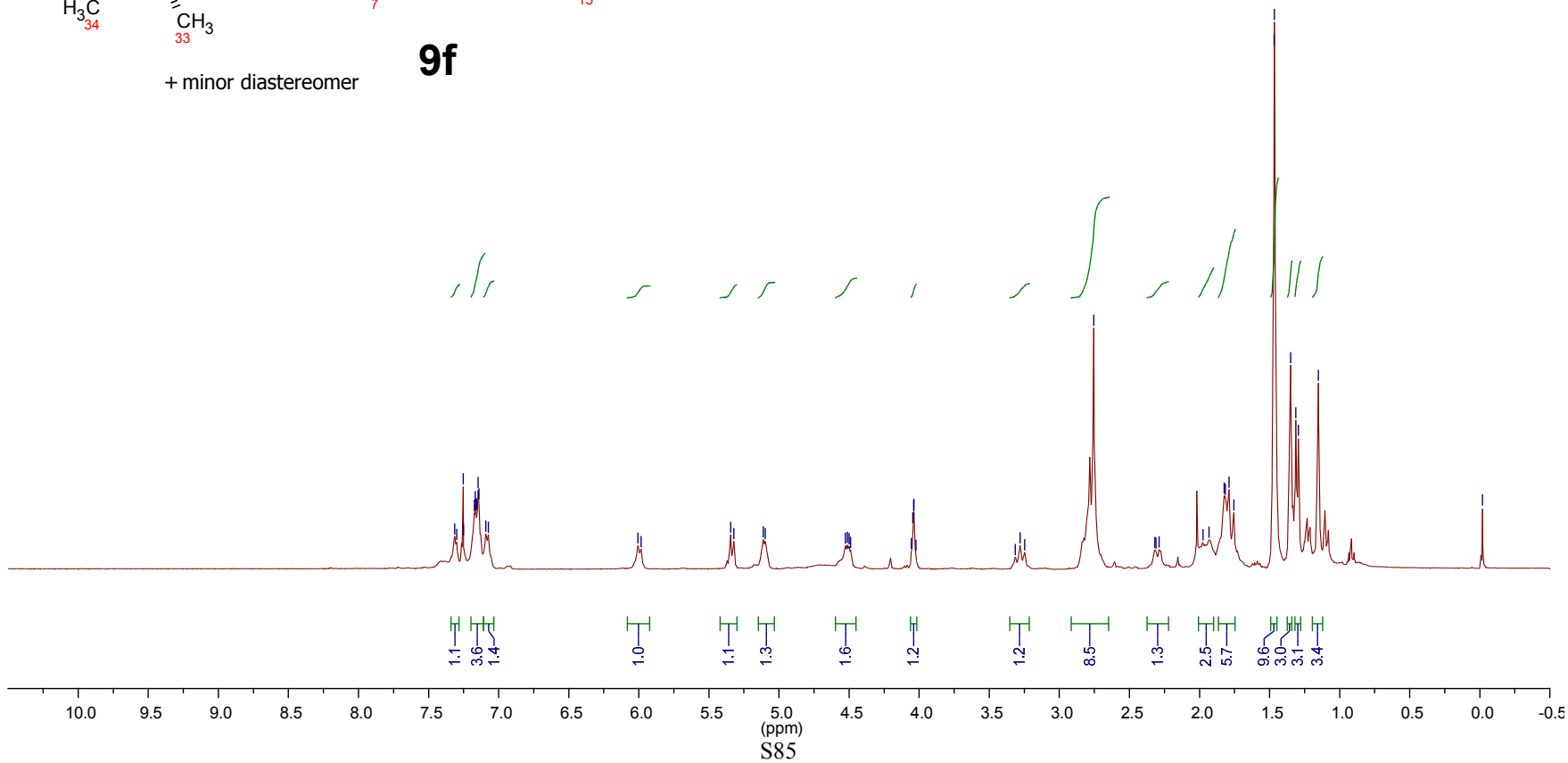
1.312  
 1.294  
 1.152

-0.018



**9f**

+ minor diastereomer



171.499  
170.644  
170.637  
169.270

137.903  
136.280  
129.406  
129.123  
129.011  
127.481  
126.296

77.160  
73.042

62.793

53.861

47.803

46.511

39.907

39.775

30.586

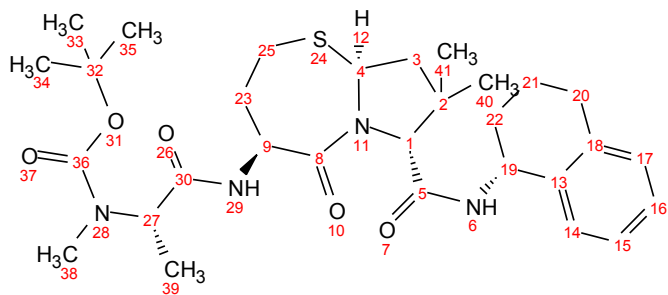
29.294

28.450

24.617

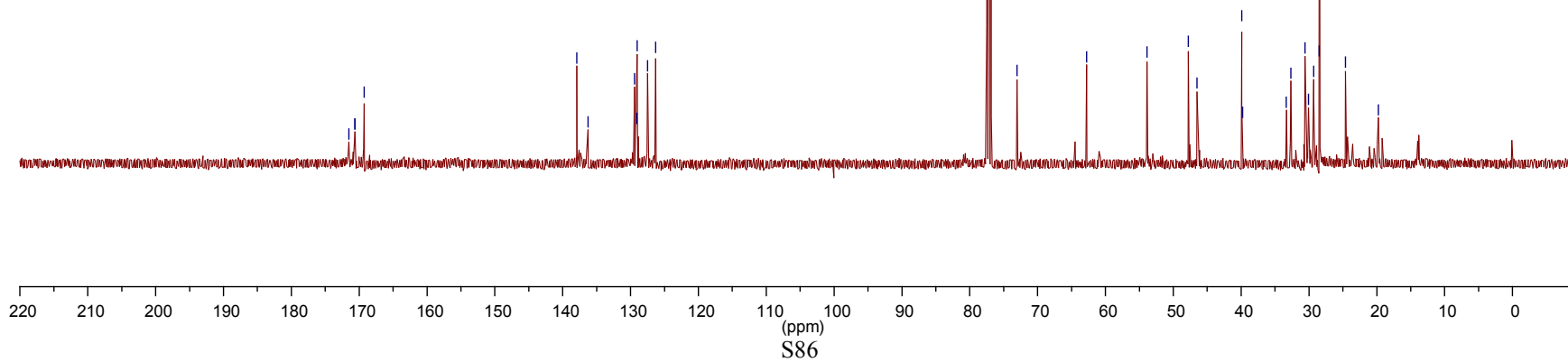
19.787

Title MV-476a upper Rf  
Solvent CHLOROFORMD  
Number of Scans 6144  
Spectrometer Frequency 100.53  
Nucleus 13C

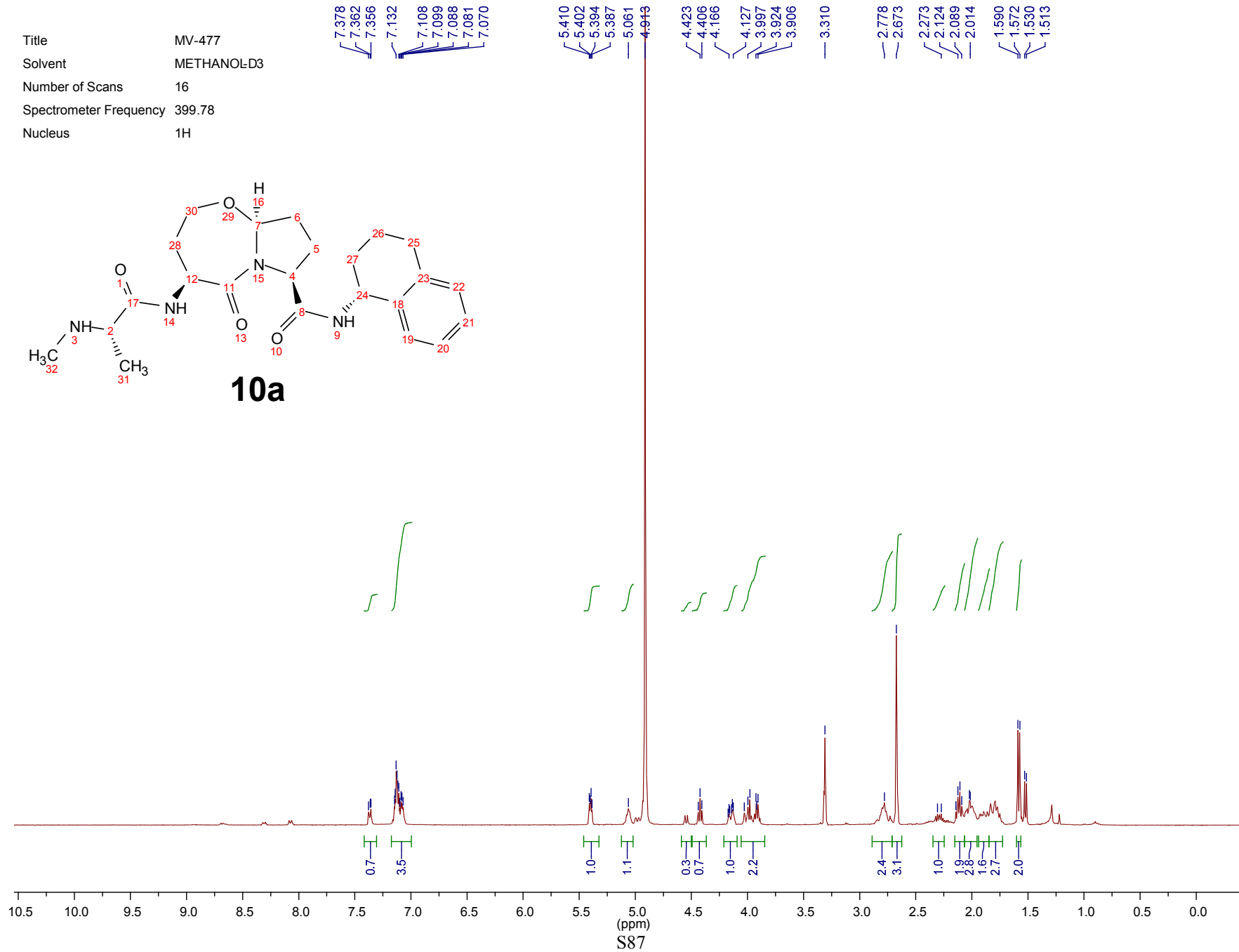
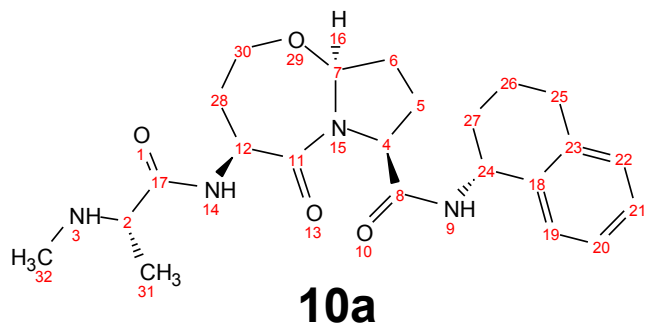


+ minor diastereomer

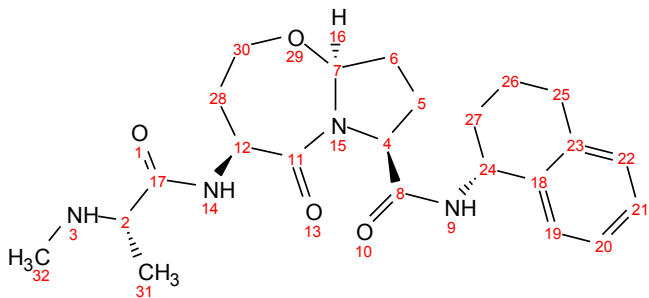
**9f**



Title MV-477  
Solvent METHANOL-D3  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



Title MV-477  
Solvent METHANOL-D3  
Number of Scans 4916  
Spectrometer Frequency 100.53  
Nucleus 13C



10a

173.379  
172.746  
172.669  
172.173  
169.648  
169.297

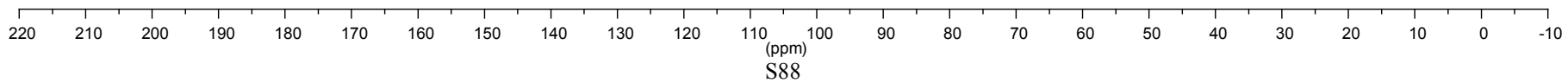
138.635  
138.492  
137.751  
137.681  
130.024  
129.992  
129.631  
129.168  
128.214  
128.067  
127.084

90.992

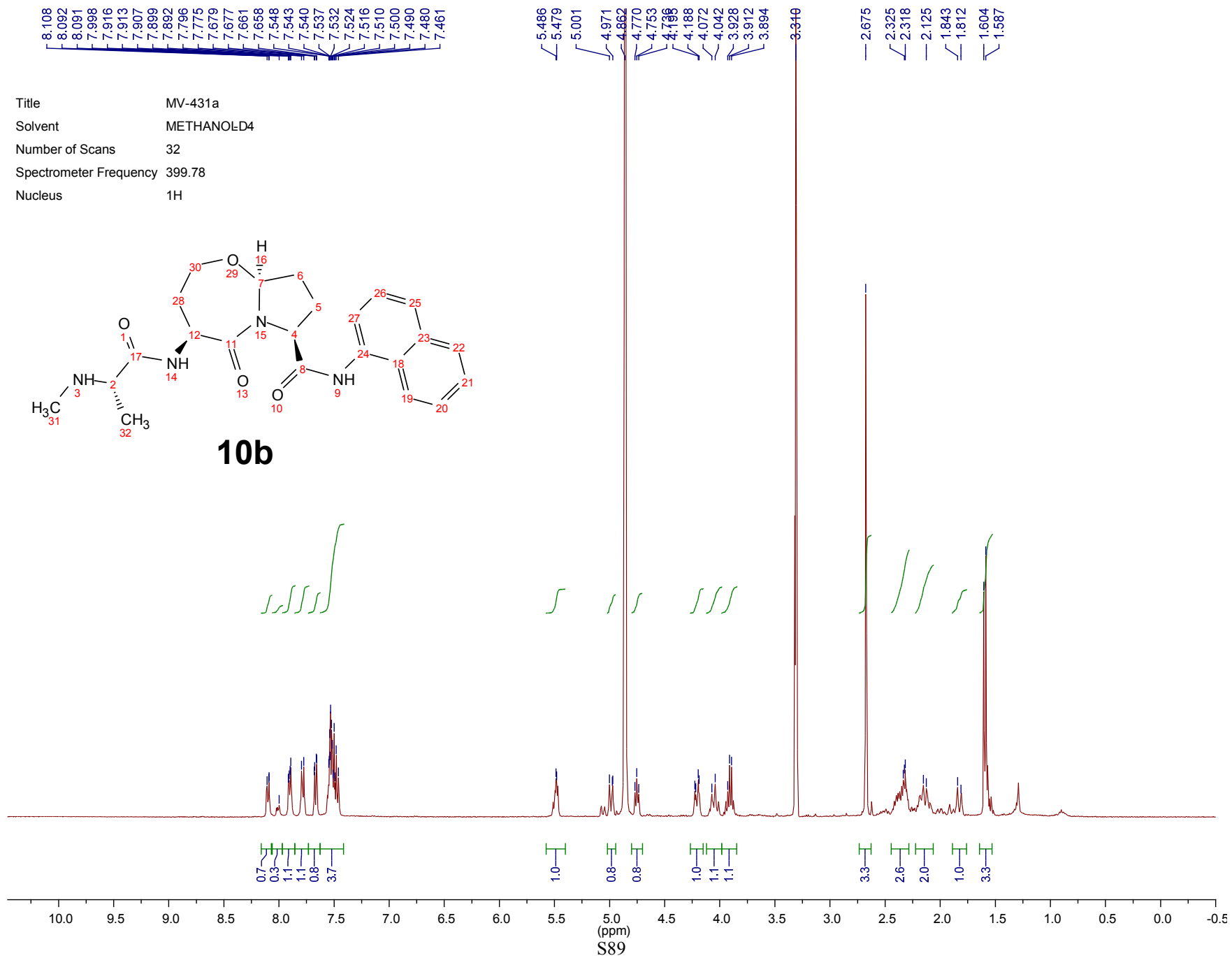
71.309  
71.192  
62.390  
62.378  
58.328  
54.225

49.000

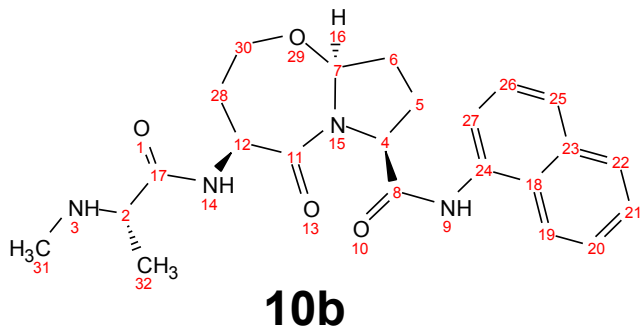
33.298  
31.755  
31.216  
30.176  
27.973  
21.684  
21.586  
16.404  
16.368







Title MV-431  
Solvent METHANOL-D4  
Number of Scans 6000  
Spectrometer Frequency 100.53  
Nucleus 13C



172.876  
172.470  
169.653

135.707  
133.956  
127.333  
126.459  
123.458

91.128

71.385

62.831

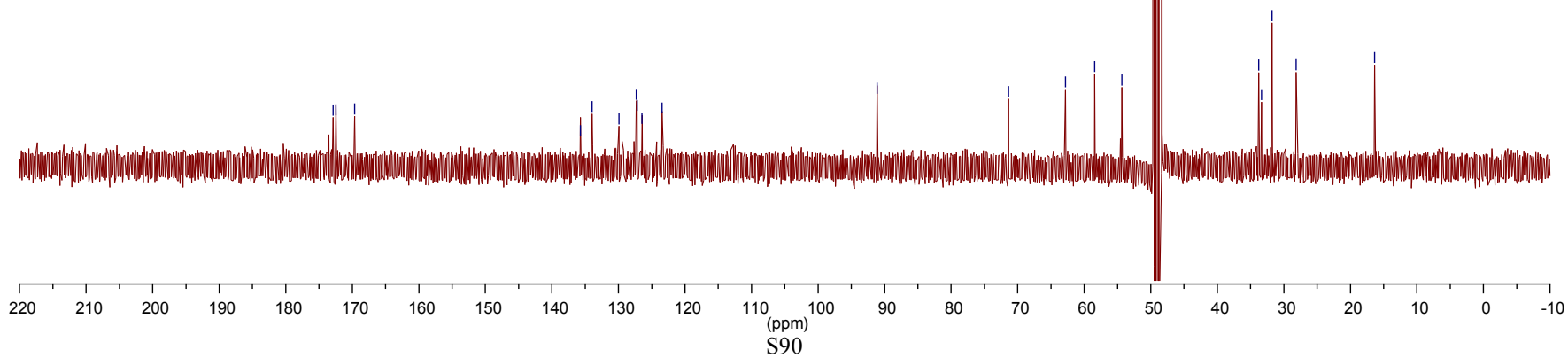
58.422

54.349

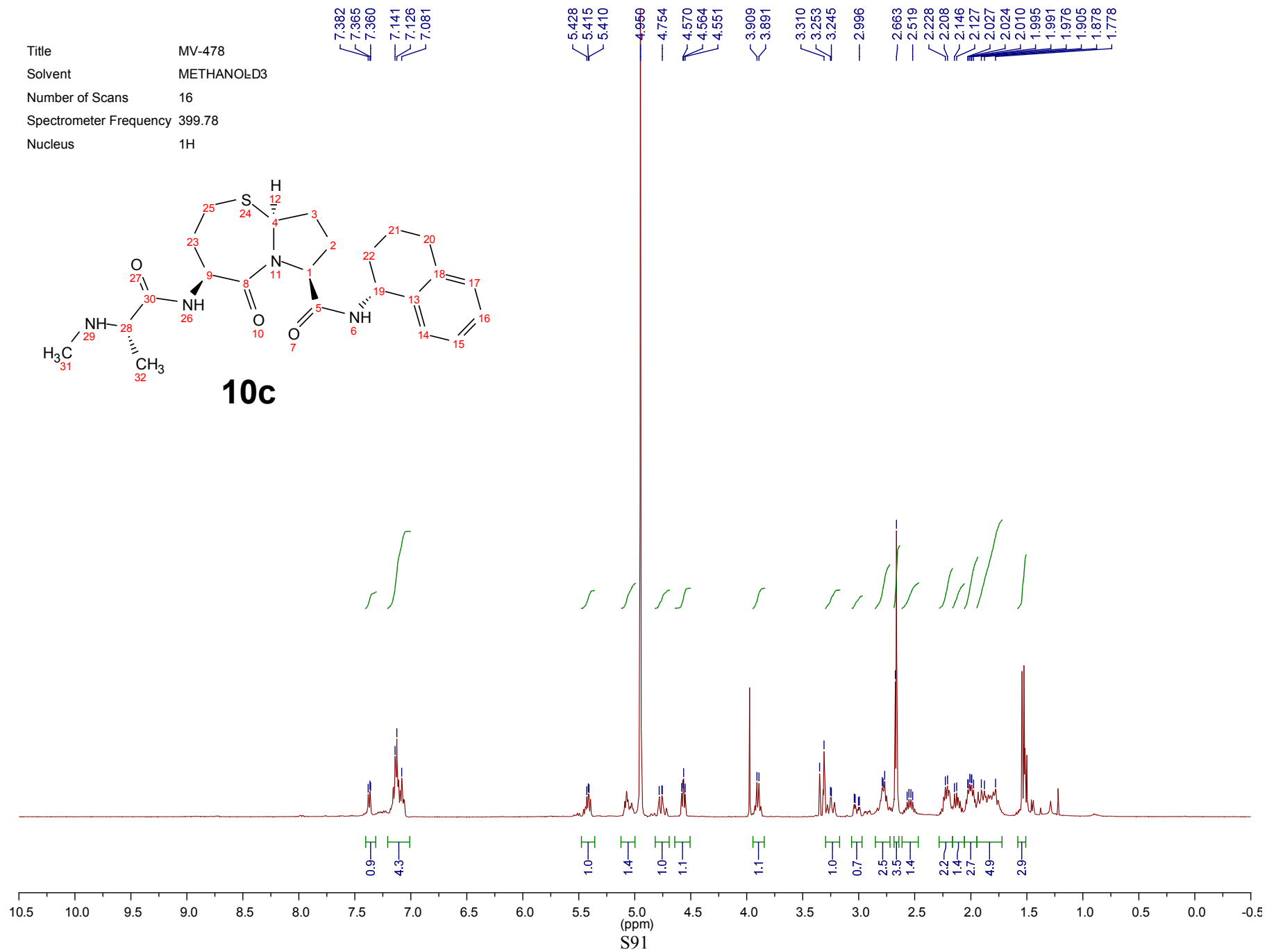
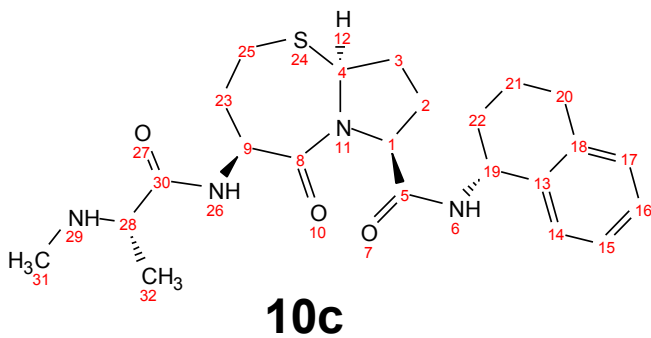
49.000

33.773  
33.355  
31.761  
28.138

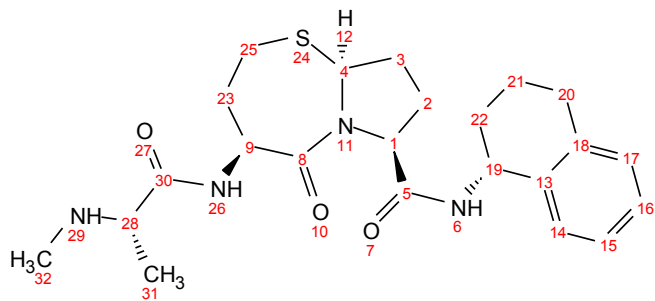
16.371



Title MV-478  
Solvent METHANOLD3  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



Title MV-478  
Solvent METHANOLD3  
Number of Scans 3600  
Spectrometer Frequency 100.53  
Nucleus 13C



10c

172.252  
171.887  
169.464

138.689  
138.451  
137.565  
137.302

130.213  
130.017  
129.856  
129.632  
128.419  
128.288  
127.208  
127.082

63.915  
63.419  
63.136

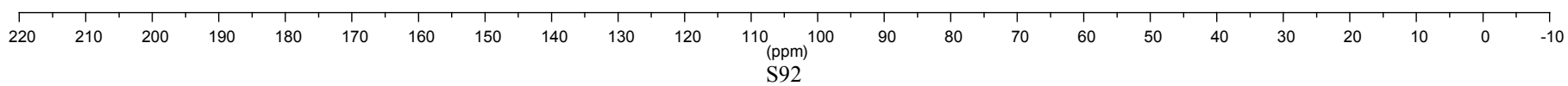
58.285  
54.094

49.000

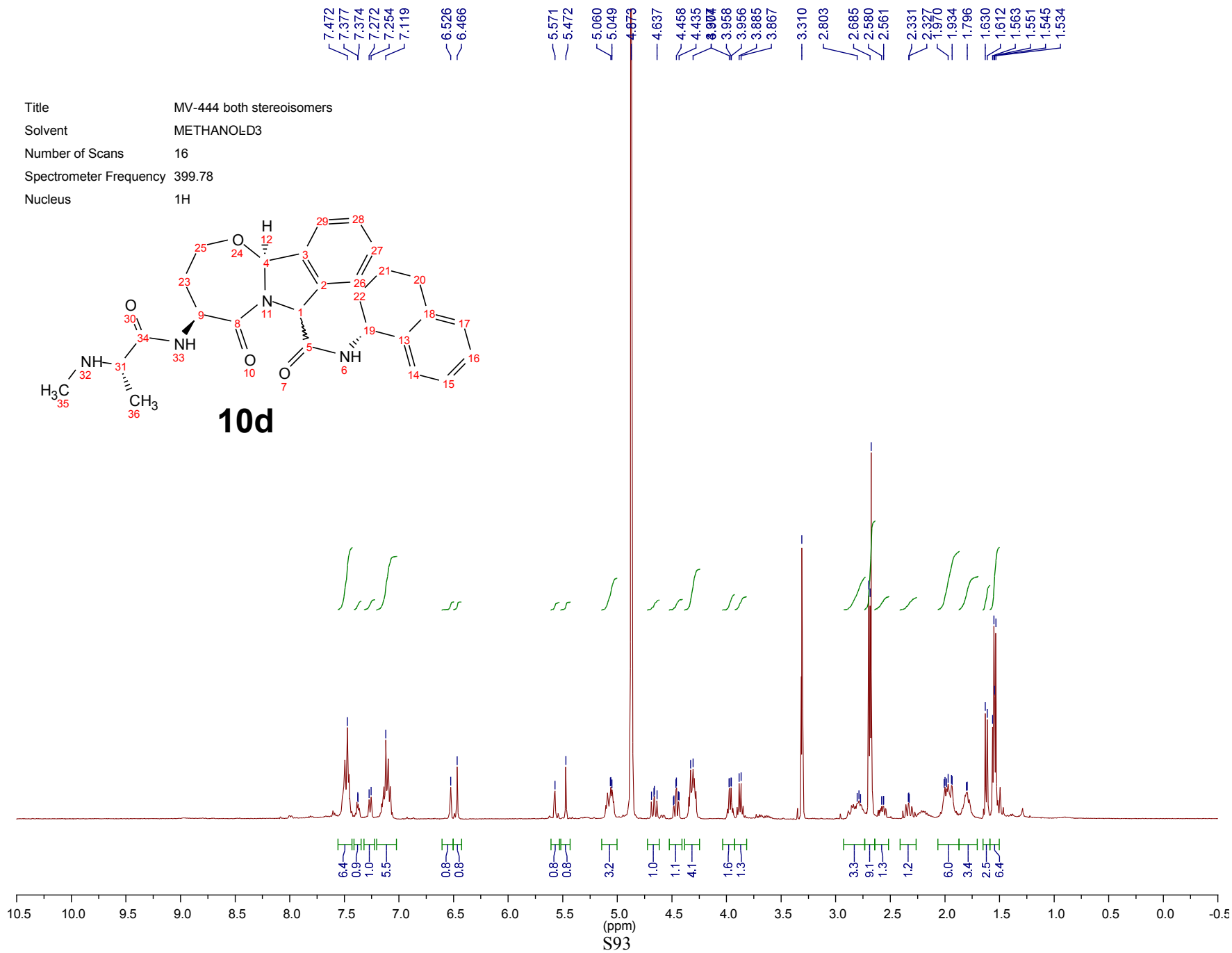
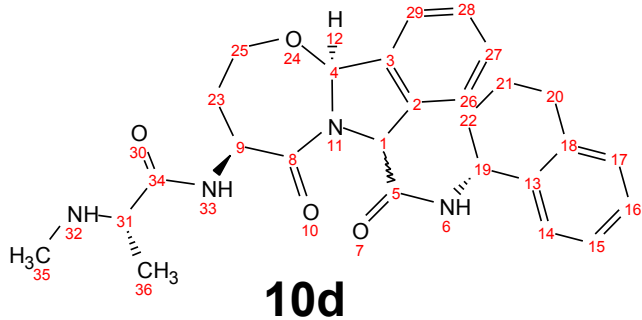
33.293  
31.753  
30.995  
30.083  
28.534

21.451  
21.112

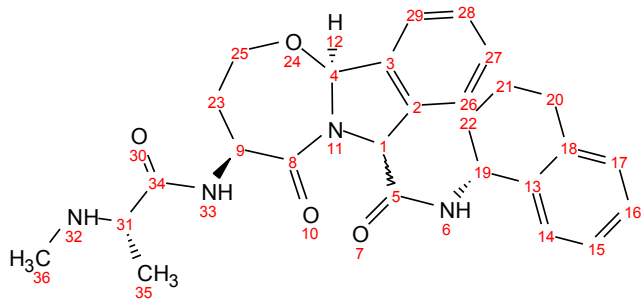
16.450  
16.278



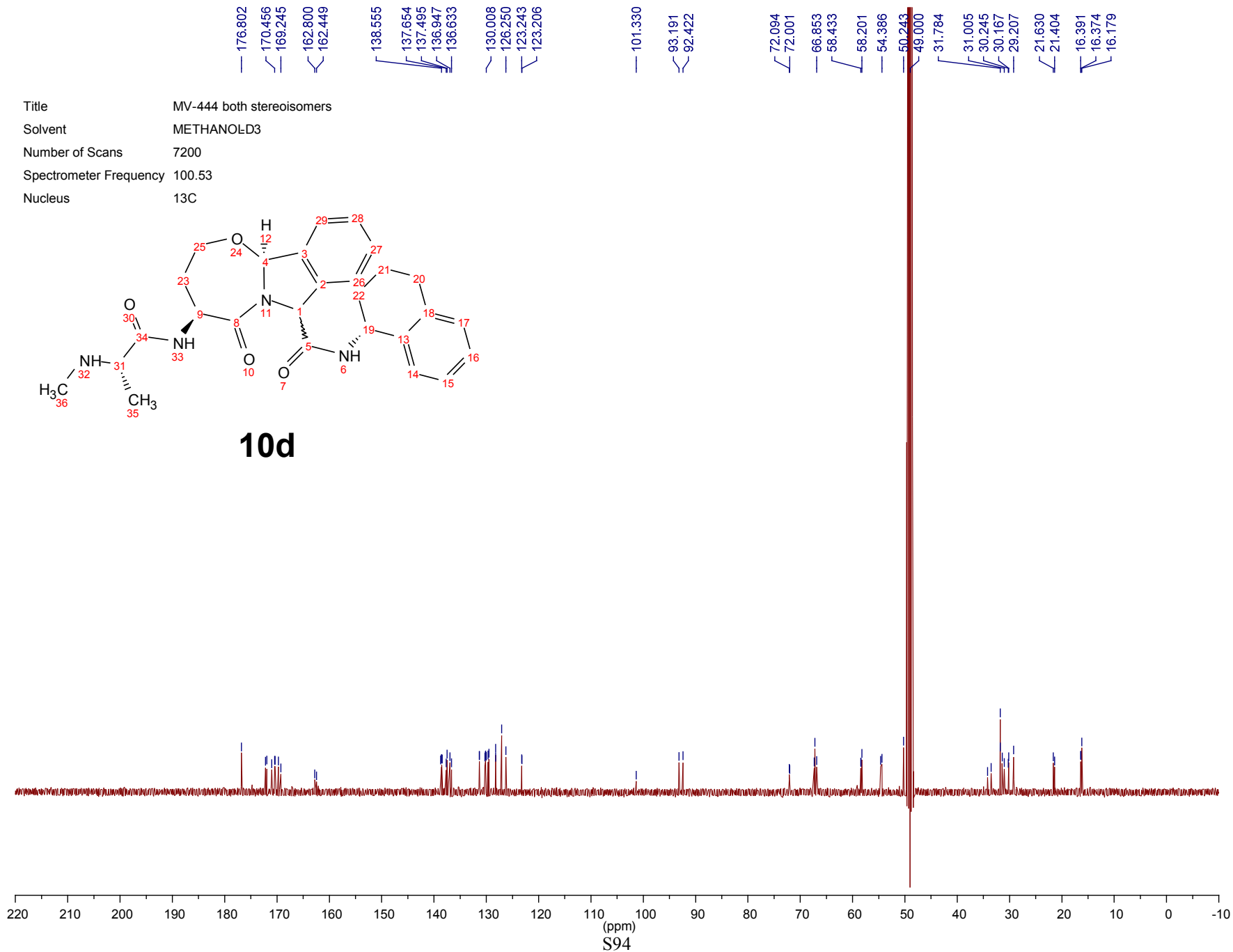
Title MV-444 both stereoisomers  
Solvent METHANOL-D3  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H

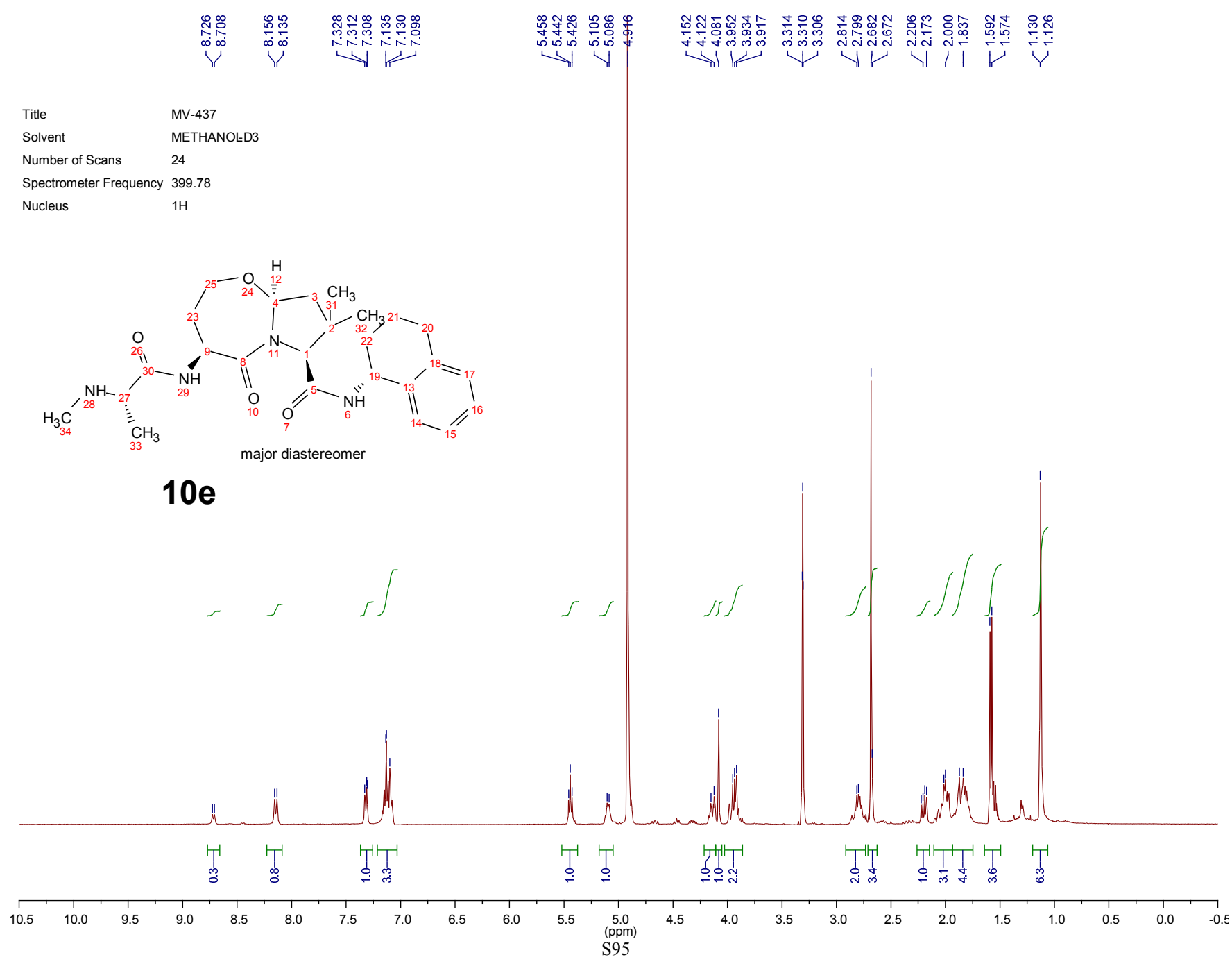


Title MV-444 both stereoisomers  
Solvent METHANOL-D3  
Number of Scans 7200  
Spectrometer Frequency 100.53  
Nucleus 13C



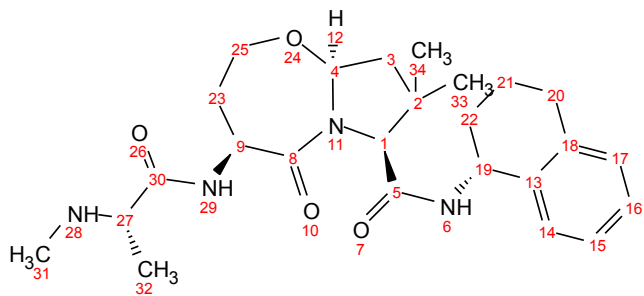
10d



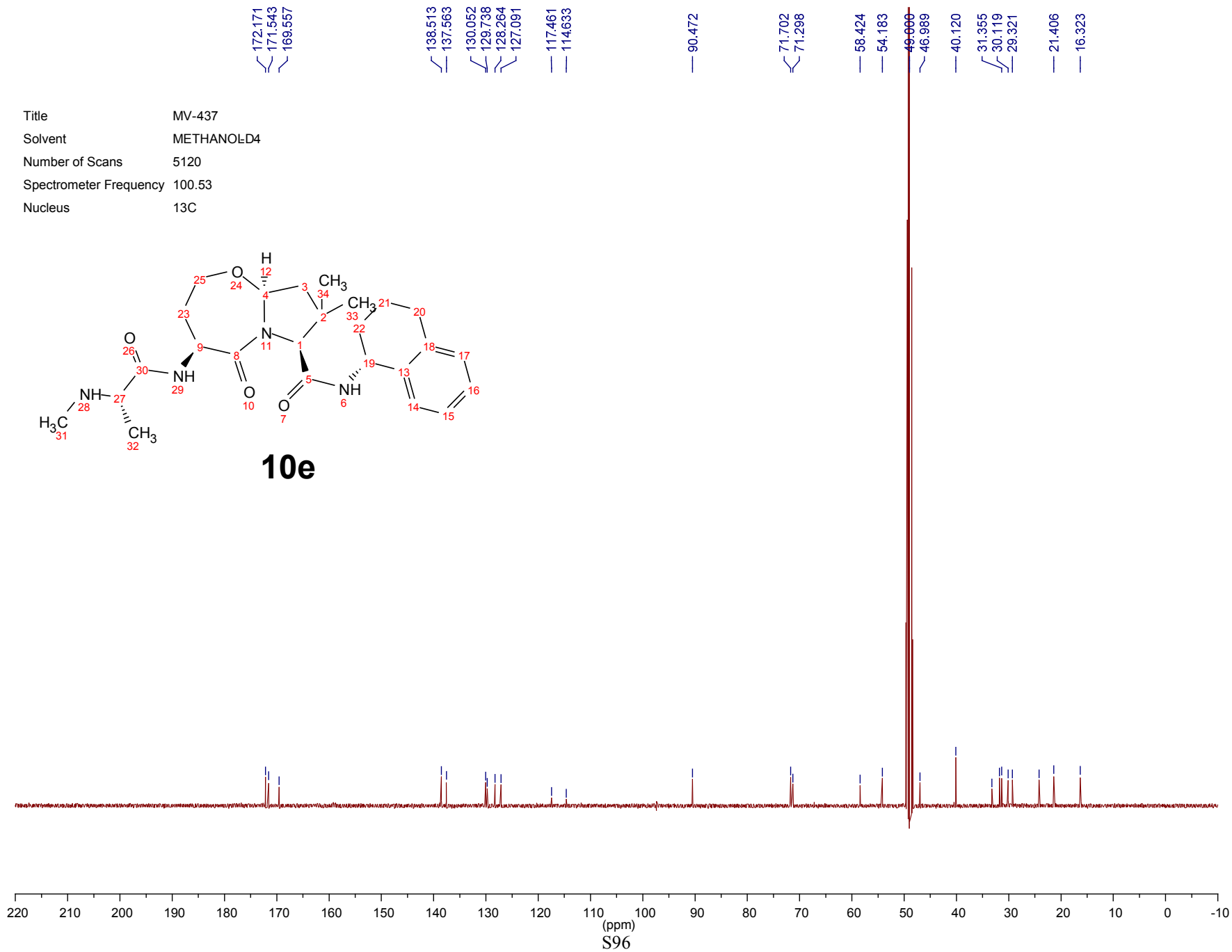


Title MV-437  
 Solvent METHANOLD3  
 Number of Scans 24  
 Spectrometer Frequency 399.78  
 Nucleus 1H

Title MV-437  
Solvent METHANOL-D4  
Number of Scans 5120  
Spectrometer Frequency 100.53  
Nucleus 13C

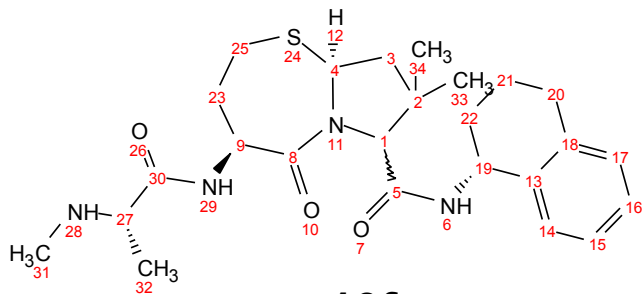


10e

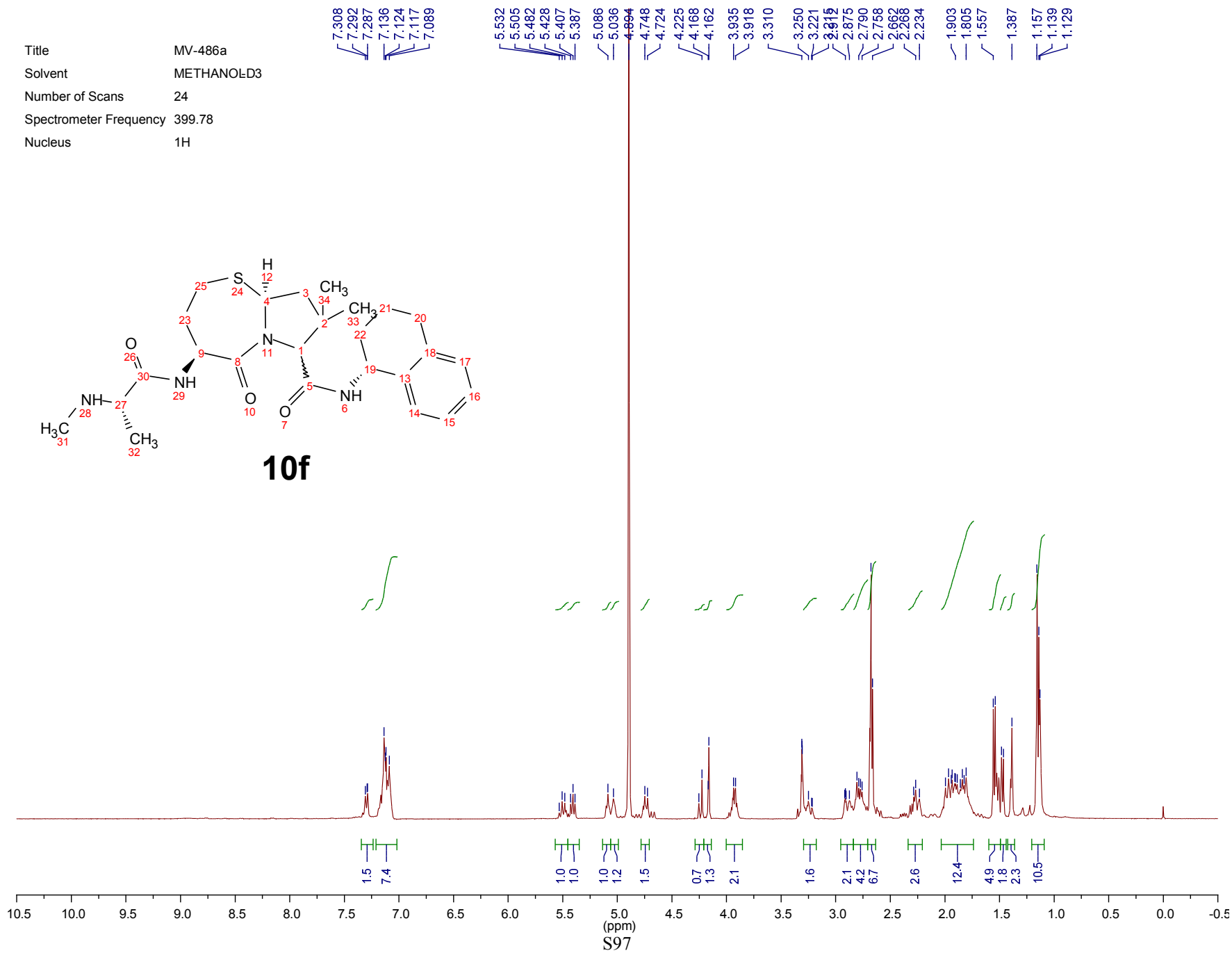




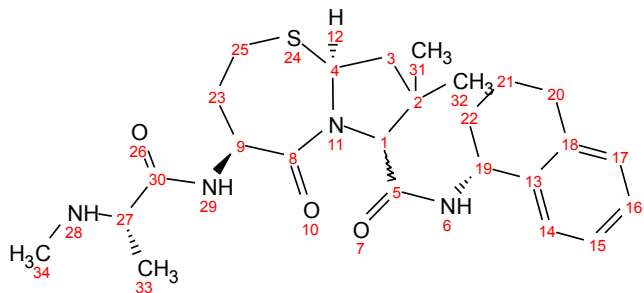
Title MV-486a  
Solvent METHANOL-D3  
Number of Scans 24  
Spectrometer Frequency 399.78  
Nucleus 1H



10f



Title MV-486a  
Solvent METHANOL-D3  
Number of Scans 7120  
Spectrometer Frequency 100.53  
Nucleus 13C



**10f**

+ small amt of +H<sub>2</sub>O adduct

172.374  
172.300  
171.841  
171.384  
169.270  
168.917

138.827  
138.533  
137.372  
137.361  
130.131  
130.074  
129.774  
128.313  
127.109  
127.007

73.353

63.781

61.844

58.329

54.439

49.666

40.871

40.742

33.627

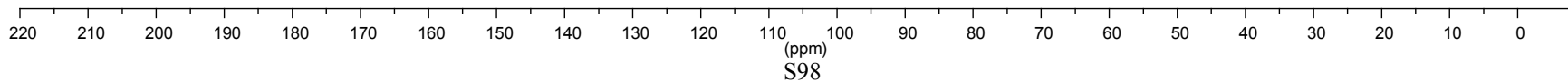
30.860

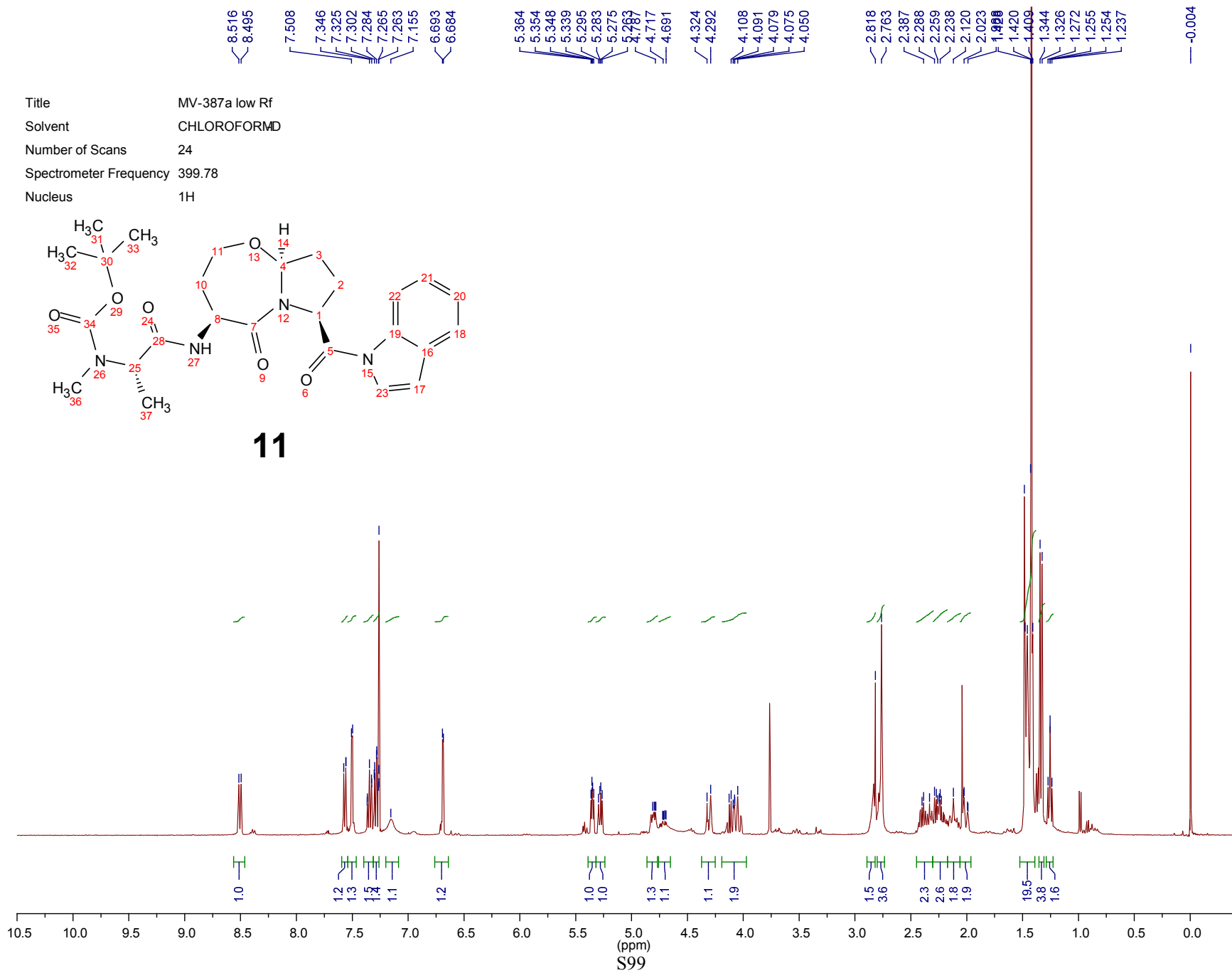
28.687

21.034

16.337

16.229





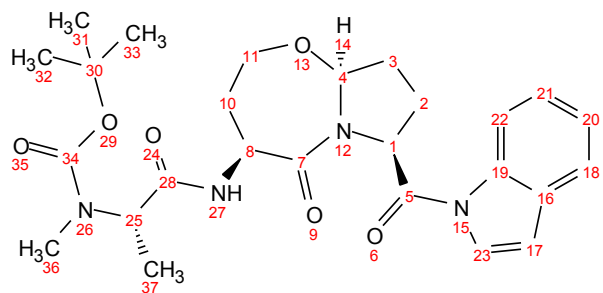
171.133  
170.817  
168.754

135.897  
130.196  
124.001  
120.837  
116.952  
109.990

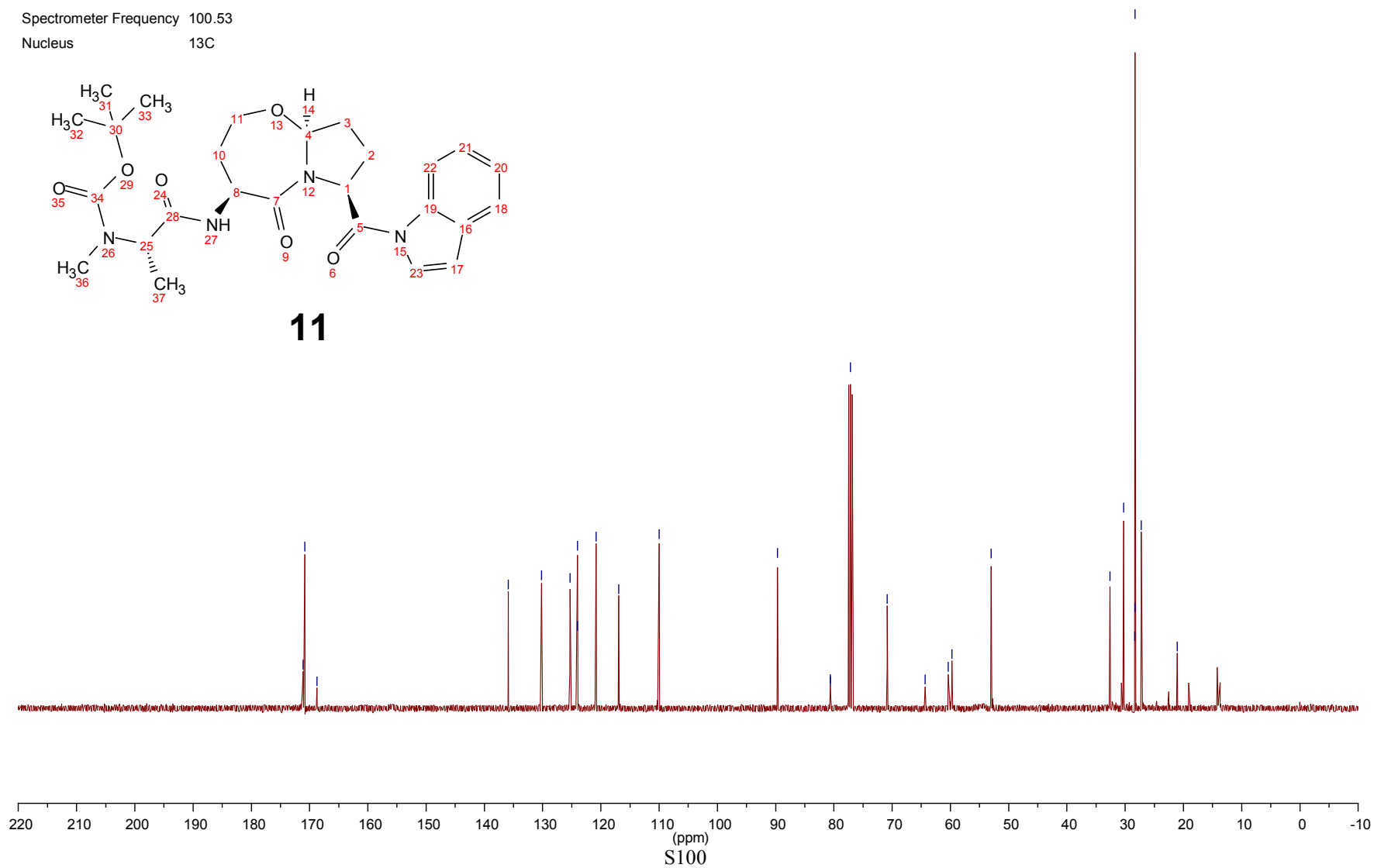
89.665  
80.600  
80.590  
77.160  
70.841  
64.338  
60.376  
59.742  
53.000

32.614  
30.284  
28.346  
28.325  
28.314  
27.230  
21.047

Title MV-419a low Rf  
Solvent CHLOROFORMD  
Number of Scans 868  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C



11



125.399  
124.111  
124.010

120.900

117.035

110.097

89.739

70.930

59.800

53.076

32.688

30.353

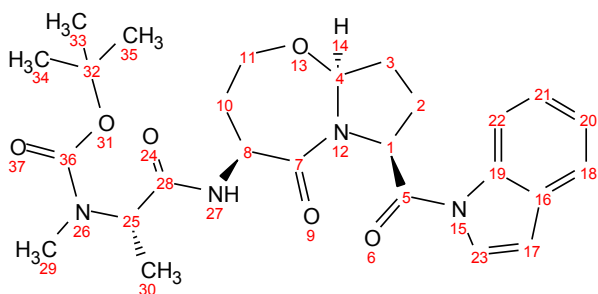
28.387

28.340

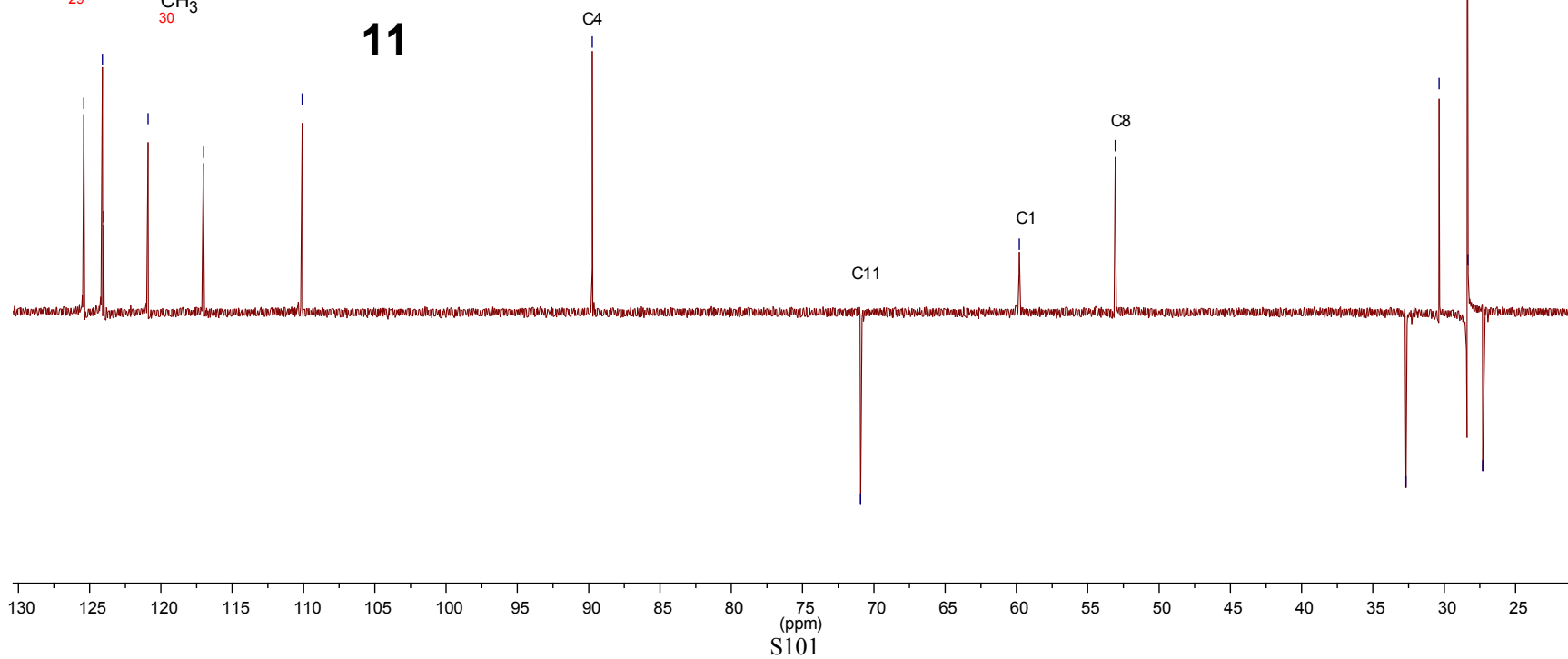
27.306

Title MV-419k low Rf  
Solvent CHLOROFORMD  
Pulse Sequence dept.ex2  
Number of Scans 453  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C

### DEPT-135



11



7.122

5.295

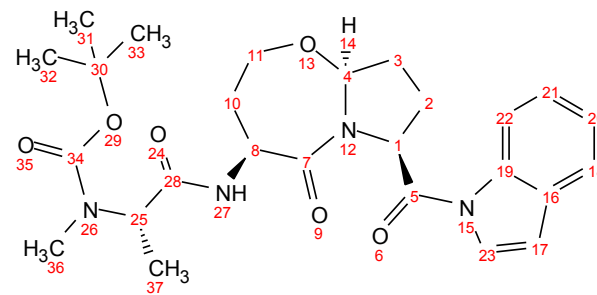
4.762

4.039  
4.008  
3.978

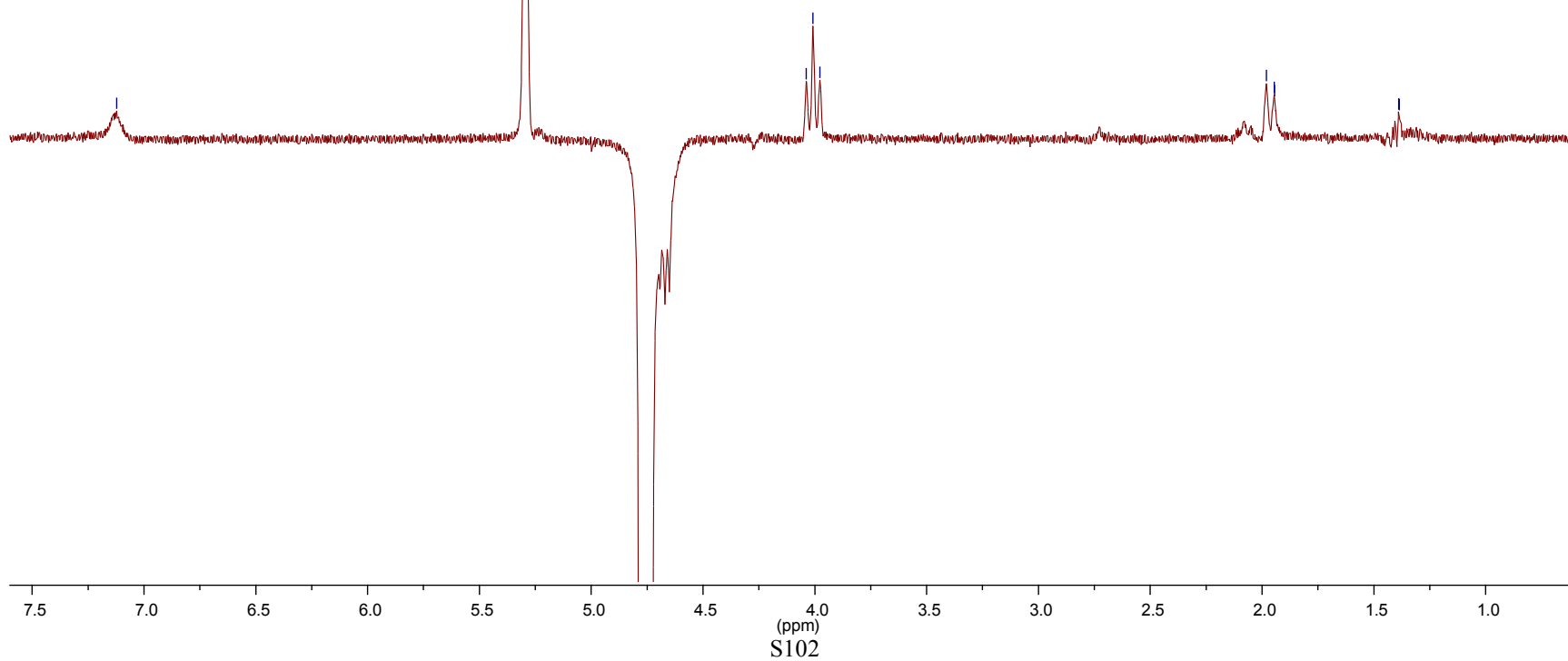
1.981  
1.946  
1.944

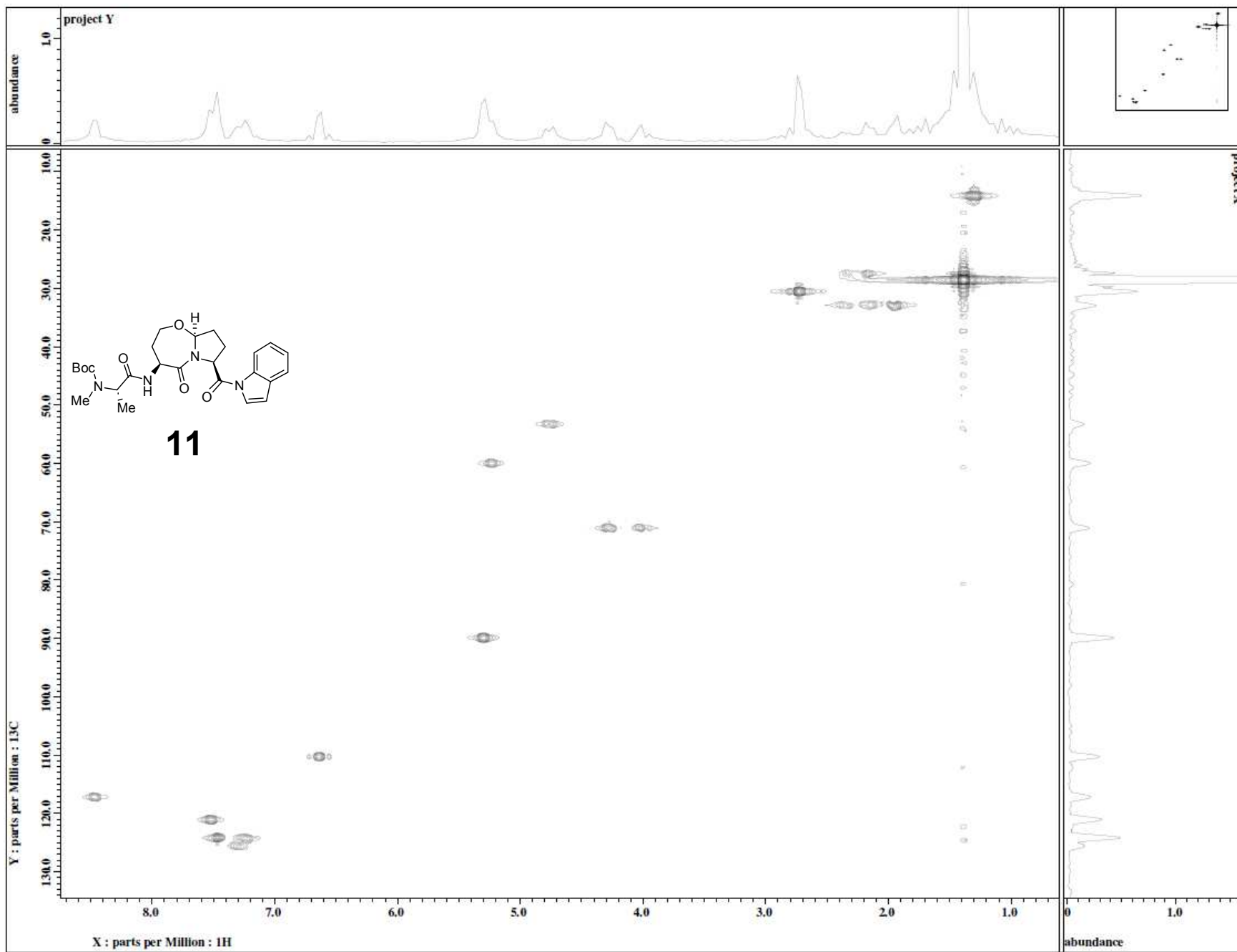
1.389  
1.386

Title MV-419q low Rf  
Solvent CHLOROFORMD  
Pulse Sequence noe\_1d\_dpfgse .ex  
Number of Scans 417  
Spectrometer Frequency 399.78  
Nucleus 1H  
x offset : 4.7 ppm



11





8.394  
8.373

7.581  
7.562  
7.495  
7.485  
7.284  
7.181

6.925  
6.711  
6.702

5.433  
5.419  
5.398

4.903  
4.879  
4.694  
4.448  
4.187  
4.075

3.688  
3.677  
3.545  
3.519

2.818  
2.787  
2.777  
2.601

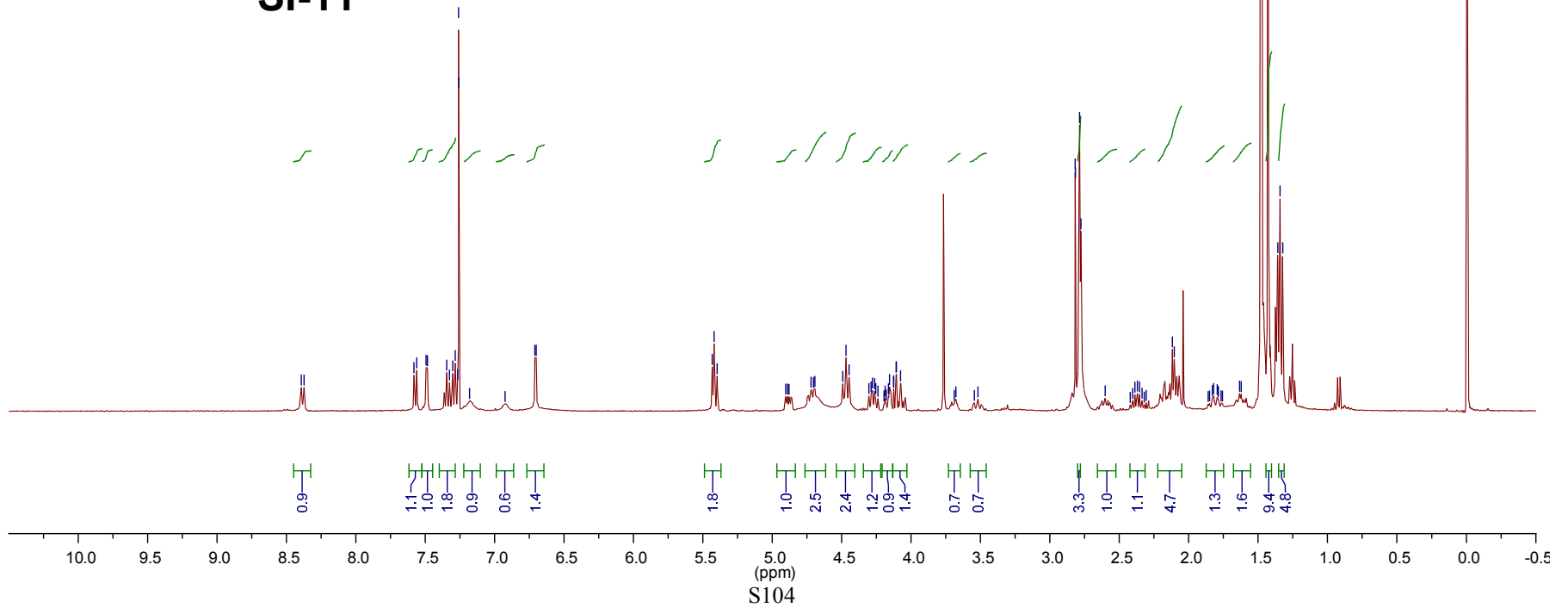
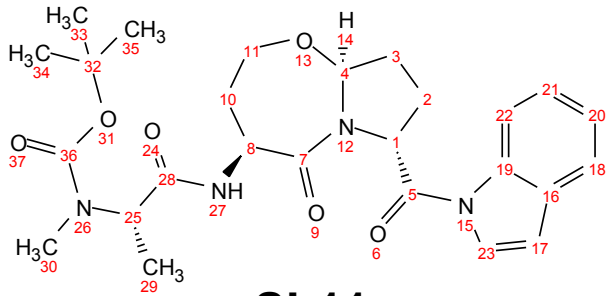
2.306  
2.104

1.795  
1.622

1.430  
1.428  
1.359  
1.341  
1.323

-0.005  
-0.007

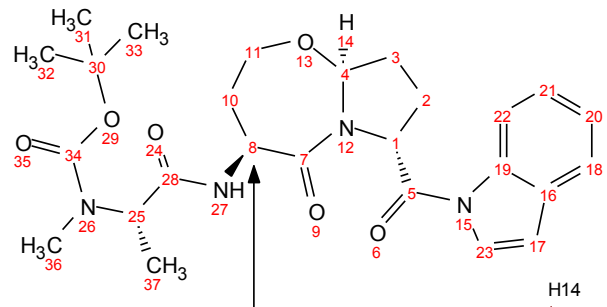
Title MV-387 upper Rf  
Solvent CHLOROFORMD  
Number of Scans 24  
Spectrometer Frequency 399.78  
Nucleus 1H



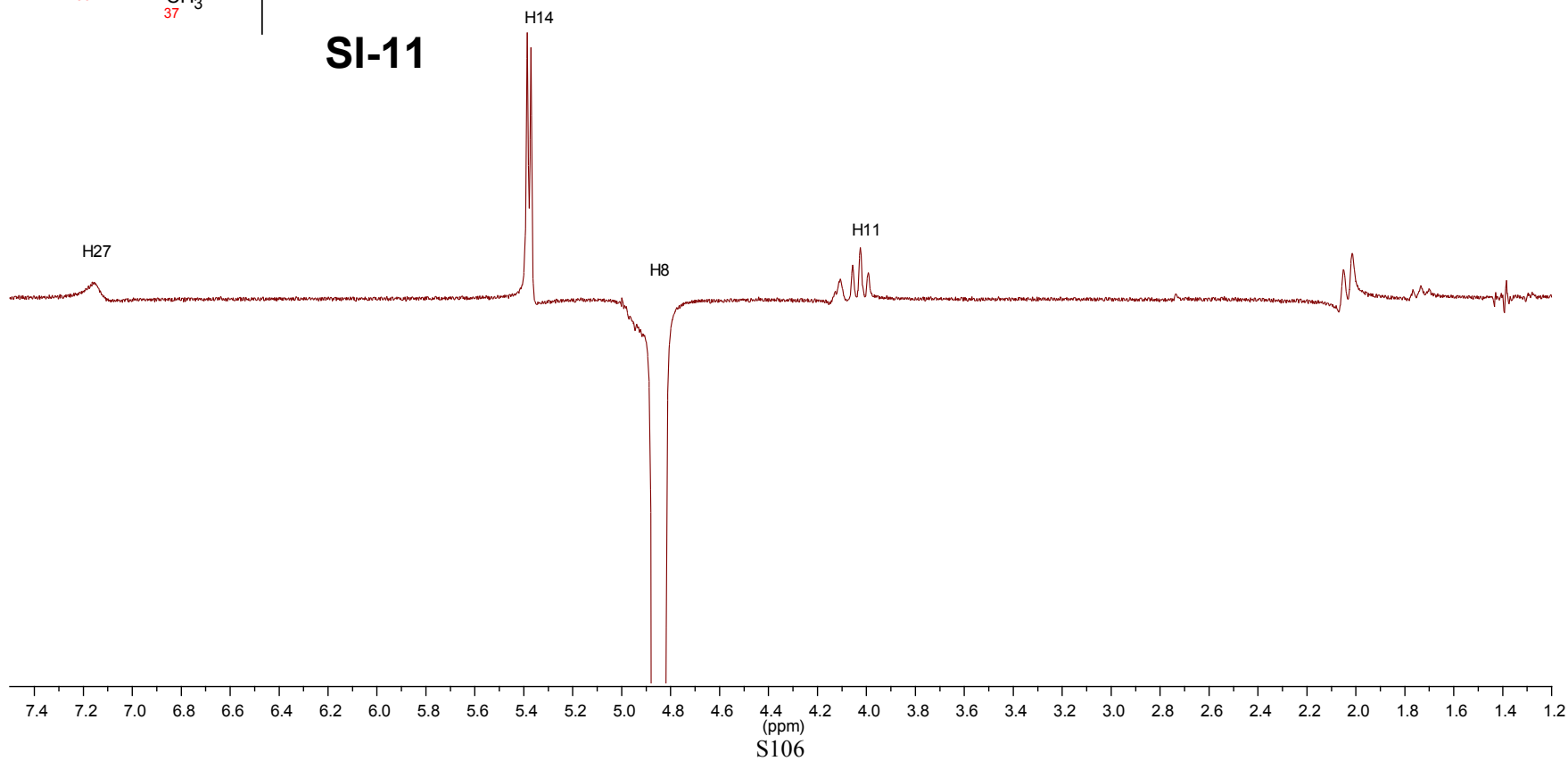




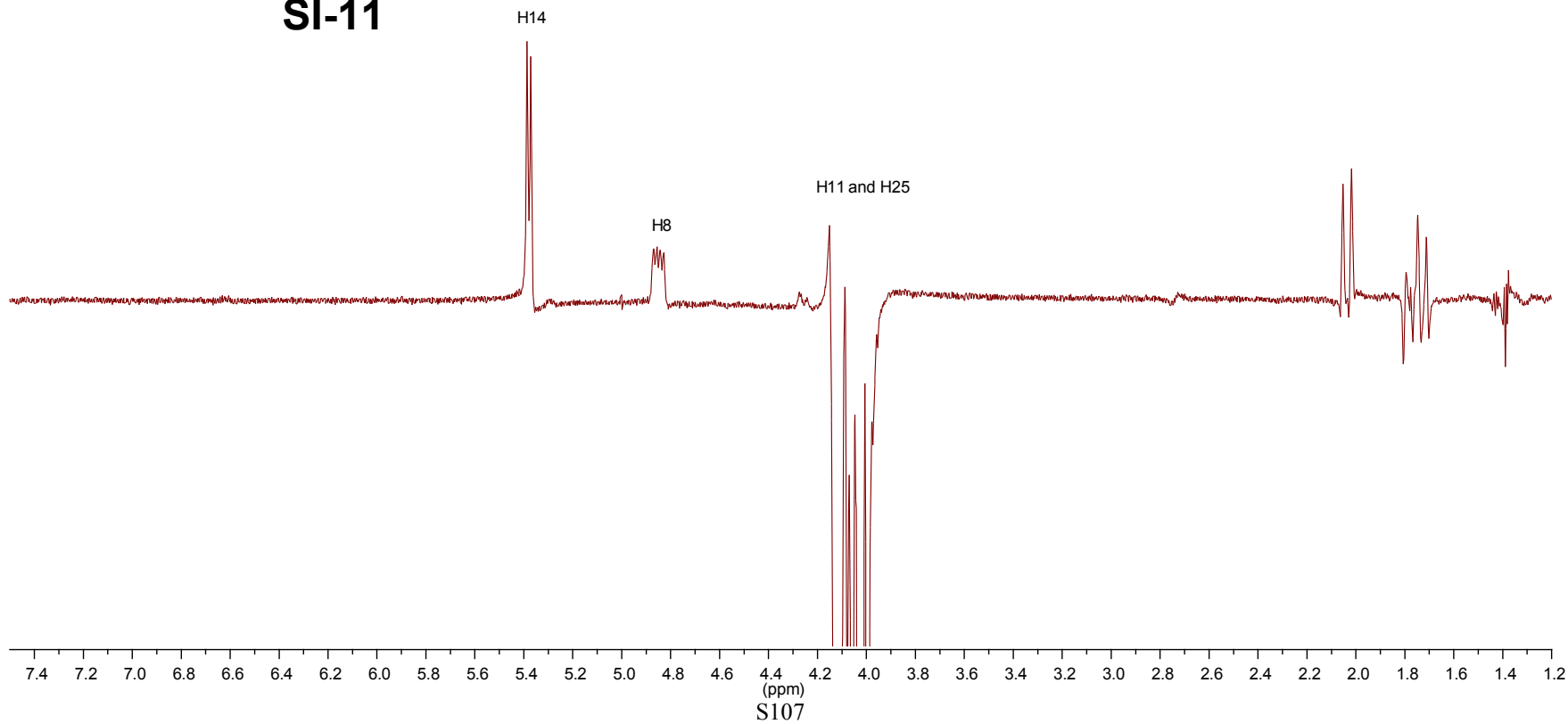
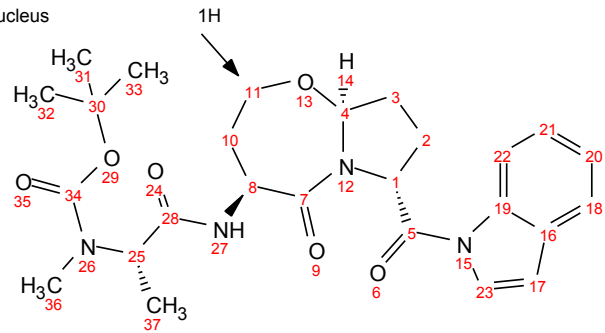
Title MV-419 upper Rf  
Solvent CHLOROFORMD  
Pulse Sequence noe\_1d\_dpfgse .ex  
Number of Scans 480  
Spectrometer Frequency 399.78  
Nucleus 1H

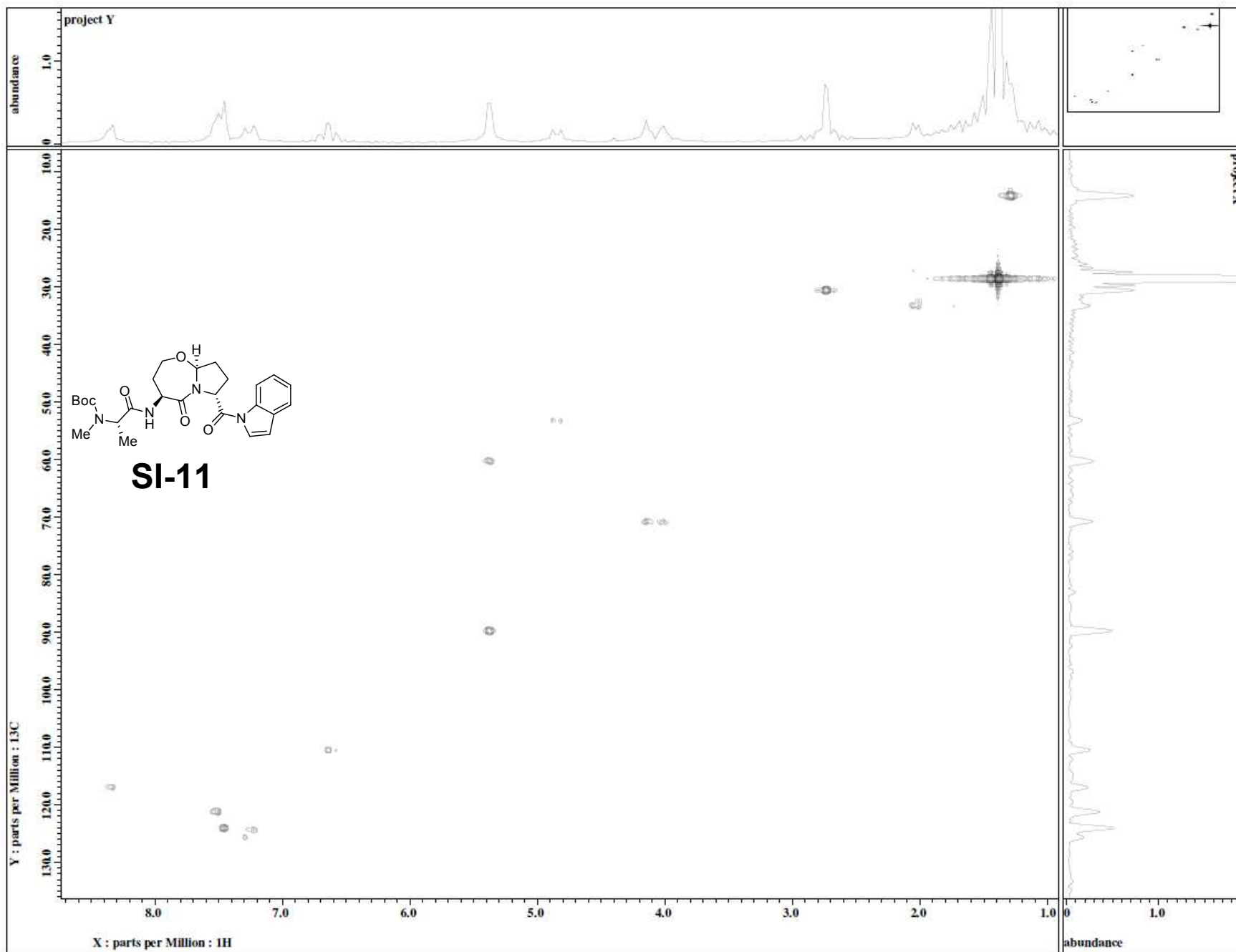


**SI-11**



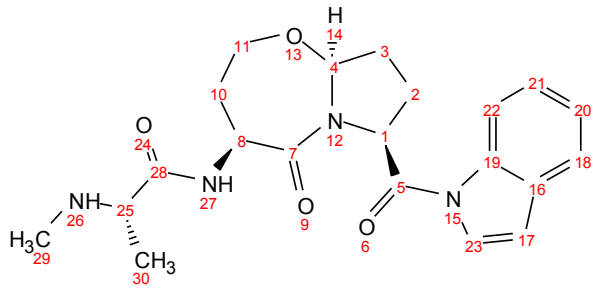
Title MV-419c upper Rf  
Solvent CHLOROFORMD  
Pulse Sequence noe\_1d\_dpfgse .ex  
Number of Scans 700  
Spectrometer Frequency 399.78  
Nucleus 1H



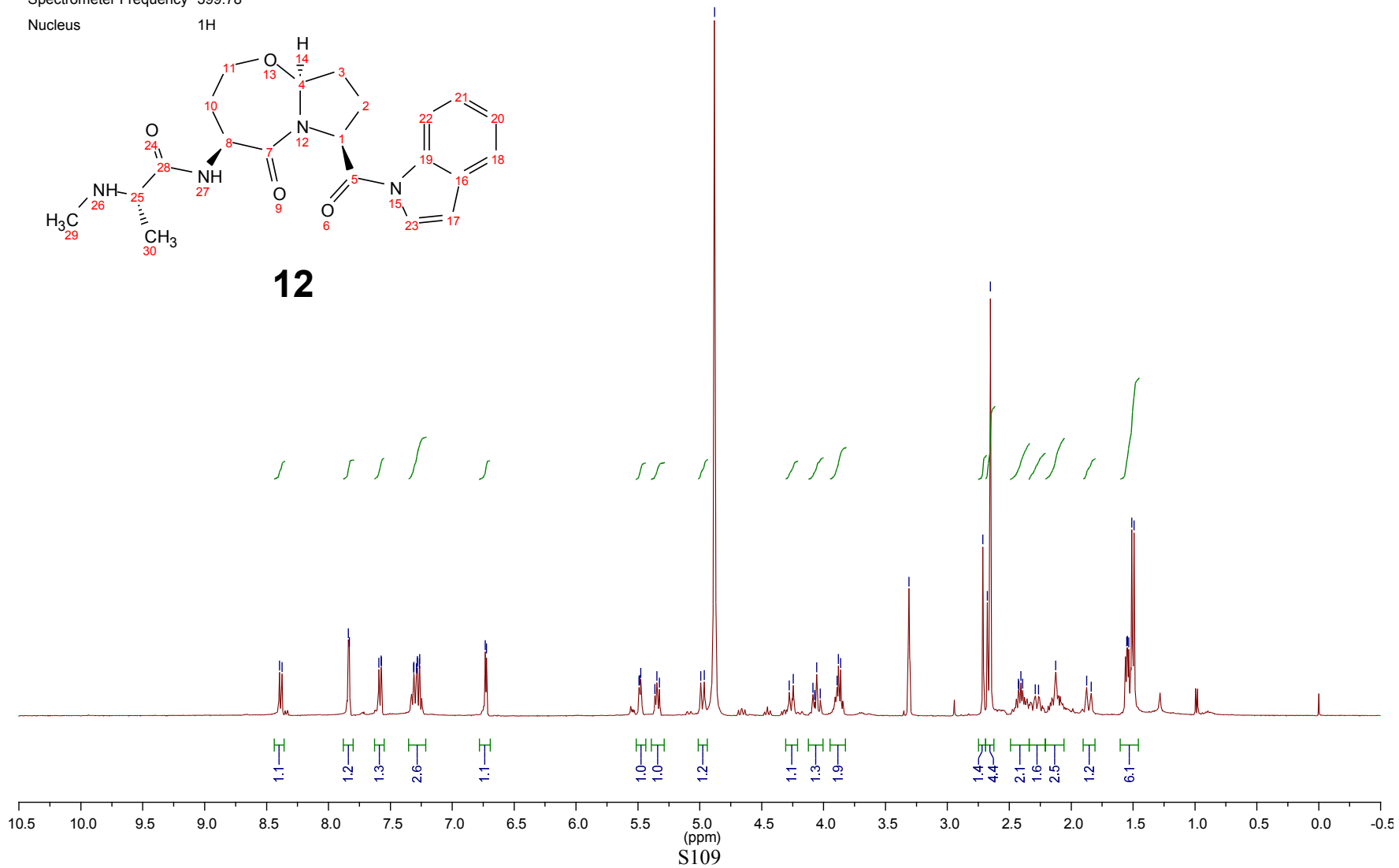


8.395  
8.375  
7.840  
7.830  
7.572  
7.312  
7.281  
7.263  
6.734  
6.724  
5.492  
5.485  
5.476  
5.362  
5.347  
5.327  
4.992  
4.963  
4.881  
4.277  
4.246  
4.055  
3.892  
3.880  
3.862  
3.310  
2.713  
2.677  
2.653  
2.406  
2.263  
2.125  
1.839  
1.551  
1.544  
1.534  
1.510  
1.492

Title MV-406  
Solvent METHANOL-D3  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



12



172.188  
170.987  
169.606

137.214  
131.929  
125.977  
125.024  
121.968  
117.418  
110.686

90.892

71.444

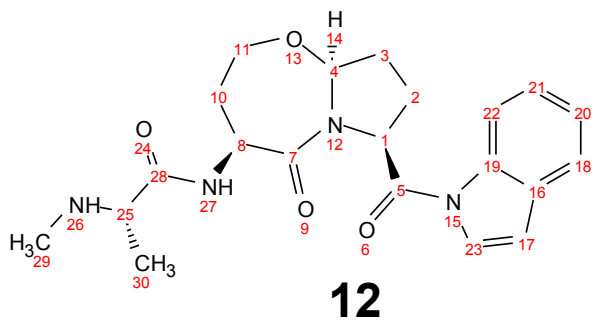
61.505  
58.338  
54.351

49.606

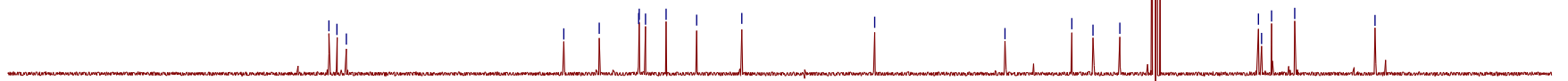
33.708  
33.239  
31.740  
28.289

16.323

Title MV-406  
Solvent METHANOL-D3  
Number of Scans 4400  
Spectrometer Frequency 100.53  
Nucleus 13C

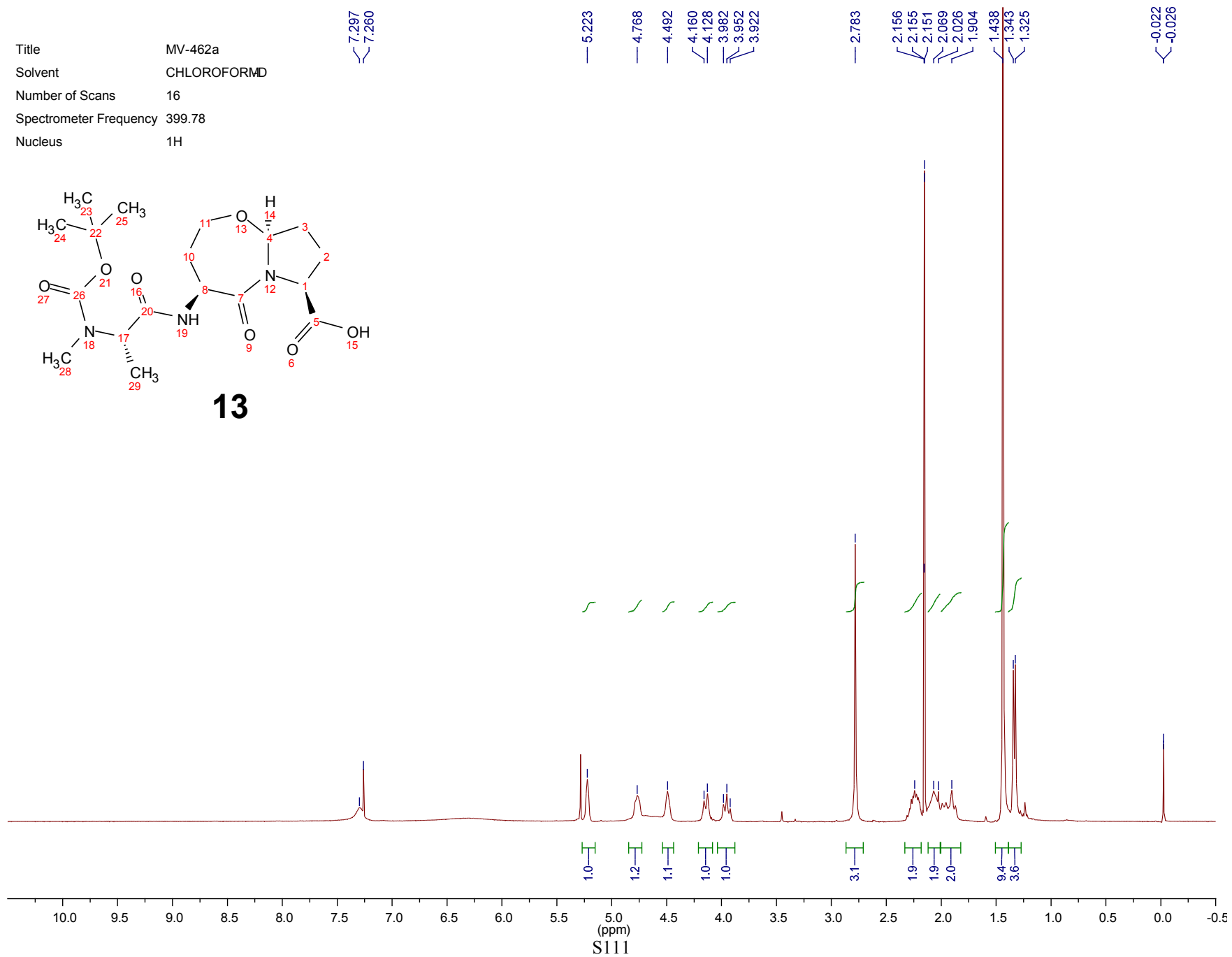
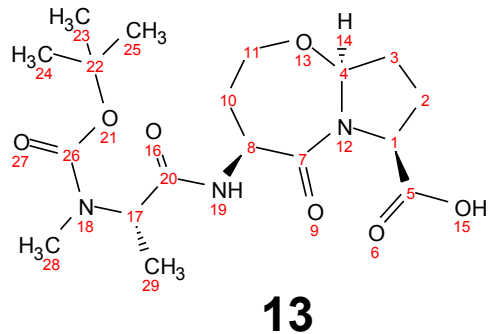


12

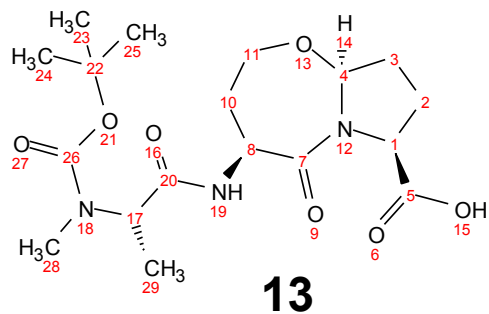


210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10  
(ppm)  
S110

Title MV-462a  
Solvent CHLOROFORMD  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



Title MV-462  
Solvent CHLOROFORMD  
Number of Scans 4096  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C



171.514  
171.486  
171.473

156.179  
156.055

89.802  
89.793

80.769  
80.739

77.466

70.746

59.683  
59.664

52.909

32.682

28.445

28.397

28.387

28.376

28.366

26.466

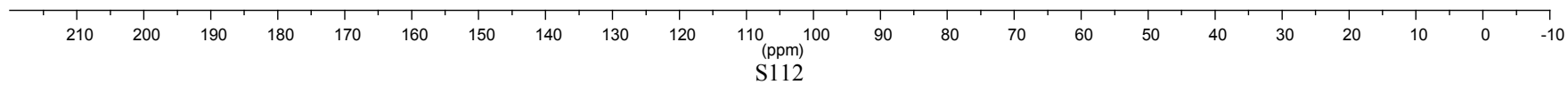
26.455

14.240

14.226

14.216

14.197





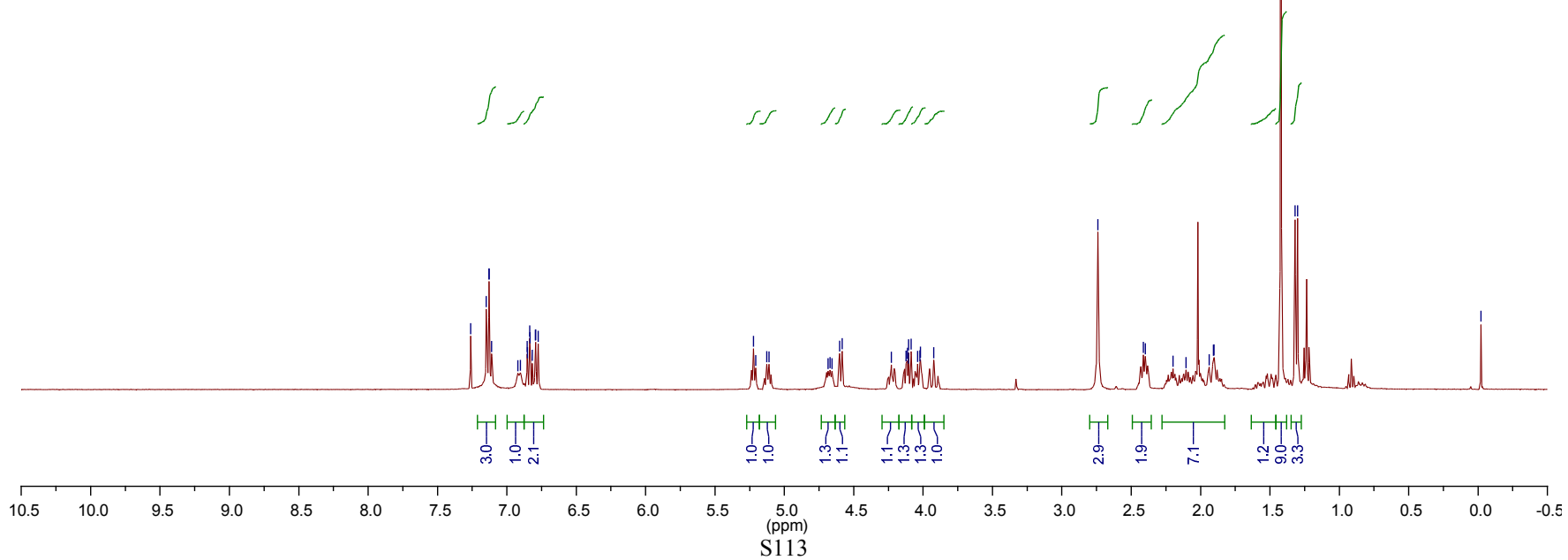
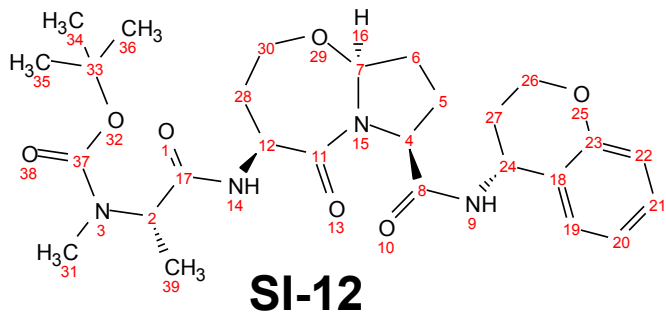
7.260  
7.148  
7.129  
7.128  
6.819  
6.835  
6.832  
6.817  
6.795  
6.793  
6.774

5.222  
5.207  
5.128  
5.111  
4.671  
4.656  
4.602  
4.584  
4.114  
4.108  
4.105  
4.087  
4.040  
4.023  
4.019  
3.924

2.740  
2.412  
2.398  
2.106  
1.908  
1.903  
1.422  
1.315  
1.301

-0.021

Title MV-470  
Solvent CHLOROFORMD  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



171.455  
170.082

154.968

129.251  
128.905

122.315  
120.724  
117.154

90.223

77.160

70.649

63.640  
60.540

52.555

43.801

32.722

32.514

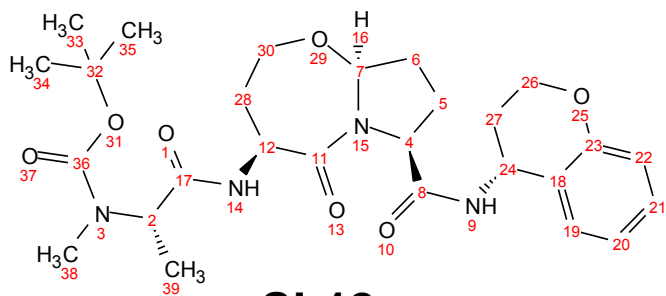
30.210

29.010

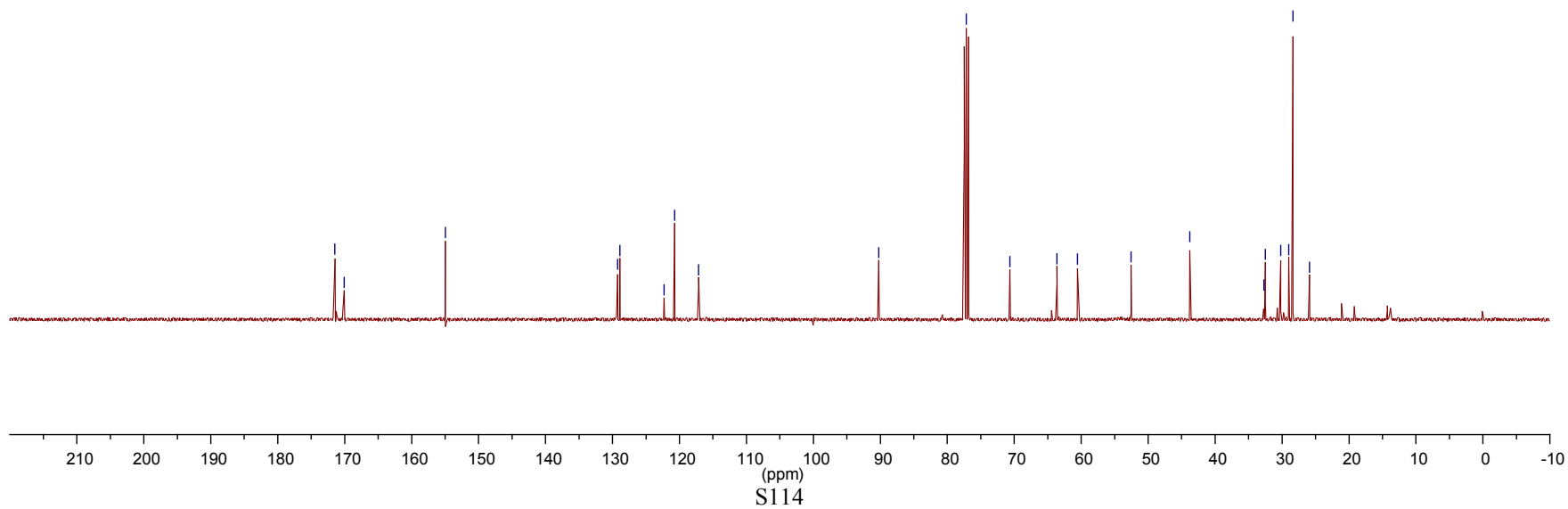
28.392

25.911

Title MV-470  
Solvent CHLOROFORMD  
Number of Scans 2800  
Spectrometer Frequency 100.53  
Nucleus 13C



SI-12

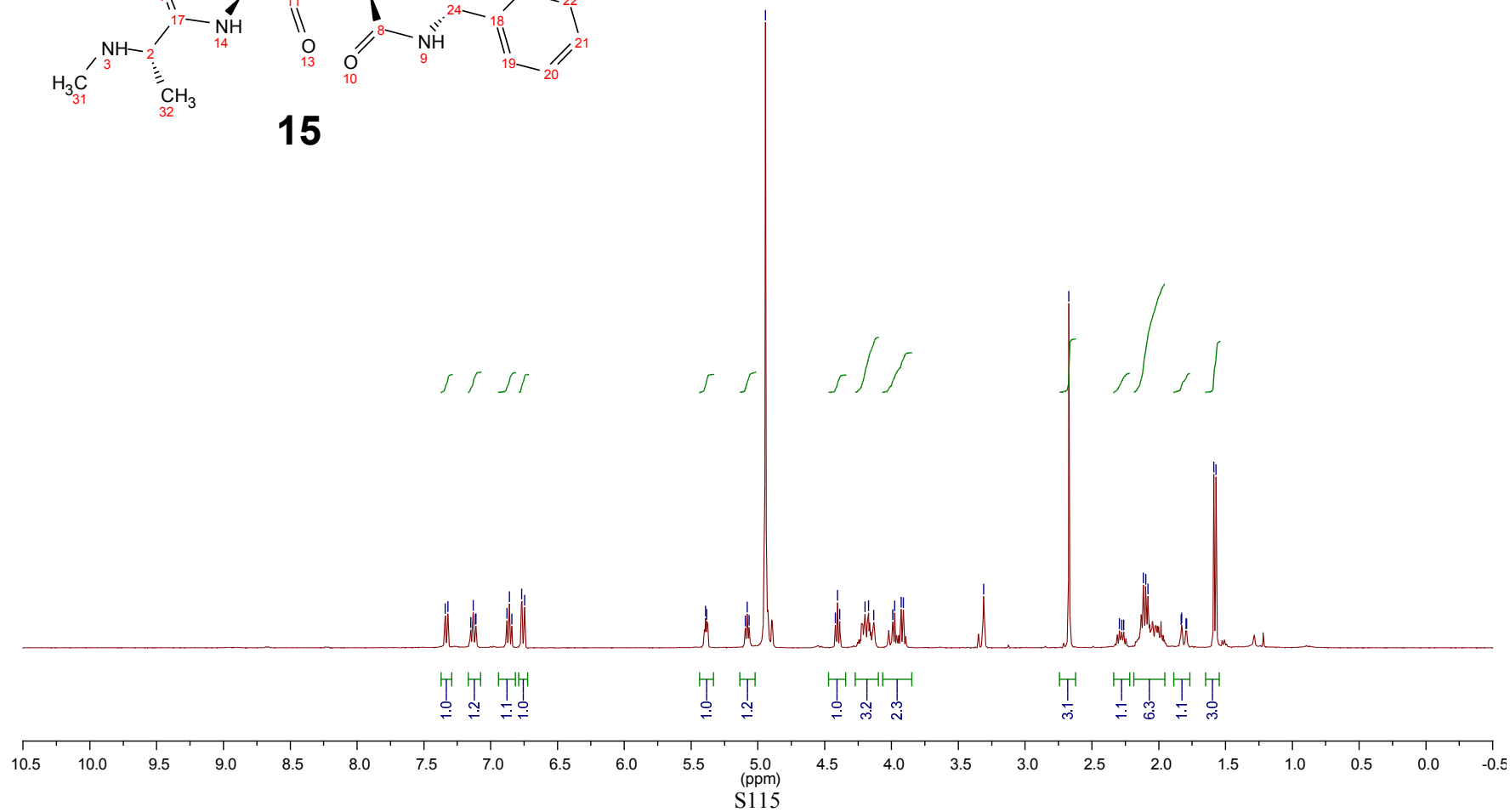
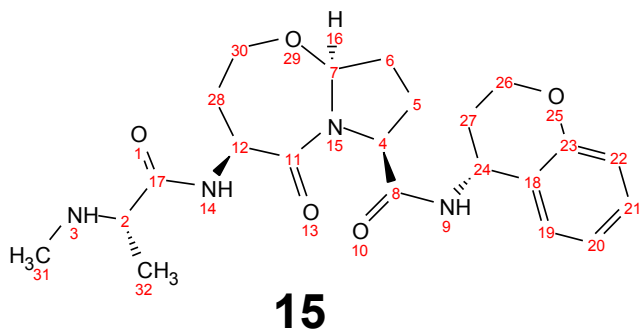


Title MV-479  
Solvent METHANOL-D3  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H

7.339  
7.320  
7.110  
6.879  
6.860  
6.842  
6.767  
6.747

5.391  
5.382  
5.095  
5.080  
5.065  
4.943  
4.420  
4.404  
4.387  
4.132  
3.976  
3.928  
3.911  
3.310

2.672  
2.295  
2.262  
2.081  
1.796  
1.588  
1.571



172.891  
172.159  
169.647

156.388

130.495  
129.986  
123.549  
121.579  
117.808

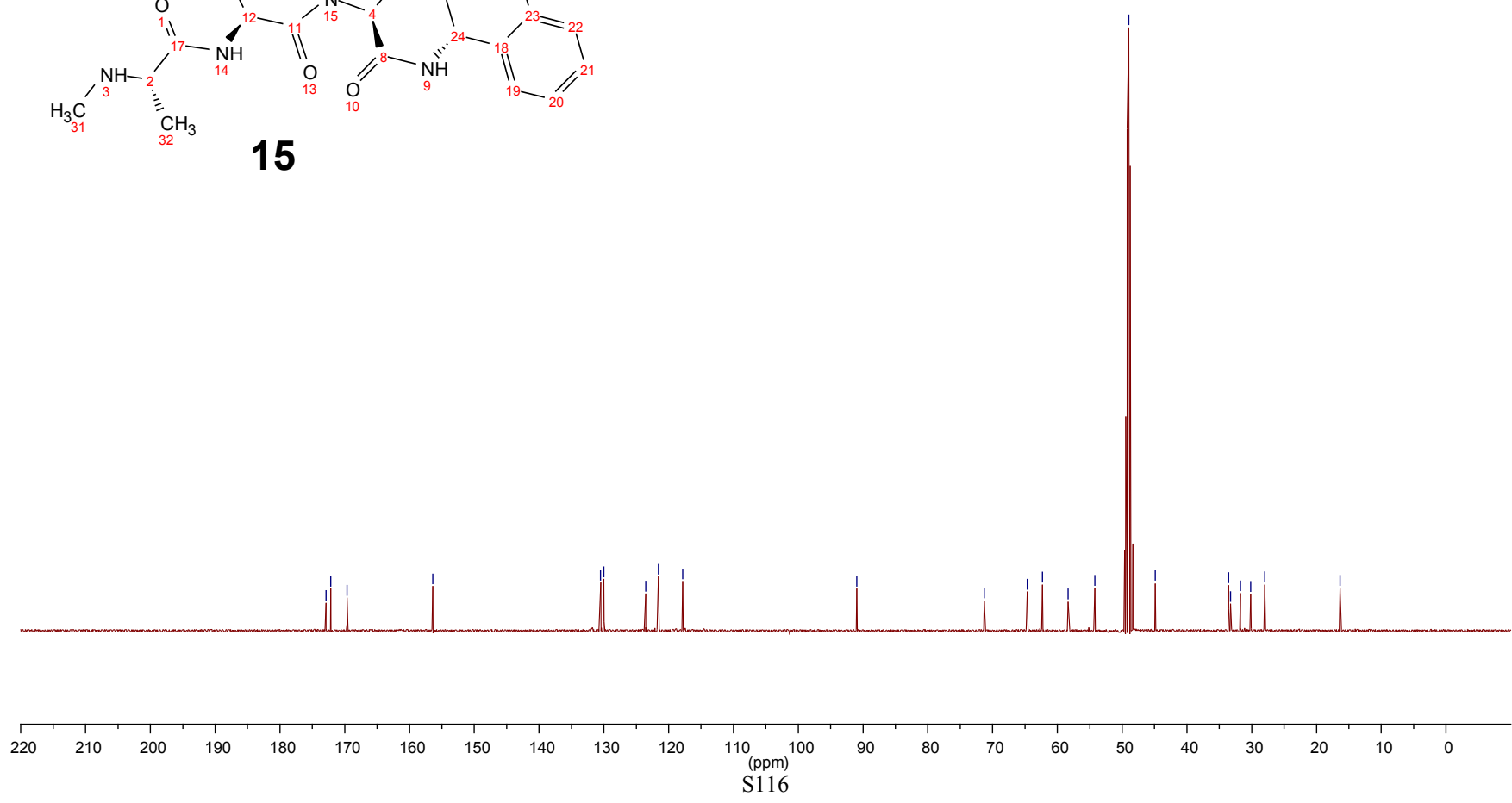
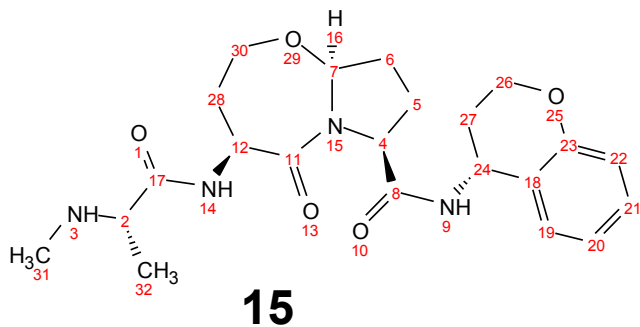
90.953

71.298  
64.632  
62.311  
58.364  
54.229  
49.000  
44.910

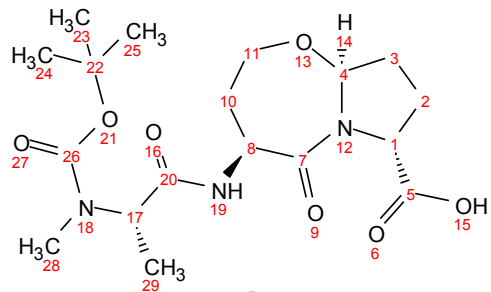
33.568  
33.263  
31.752  
30.179  
28.000

16.370

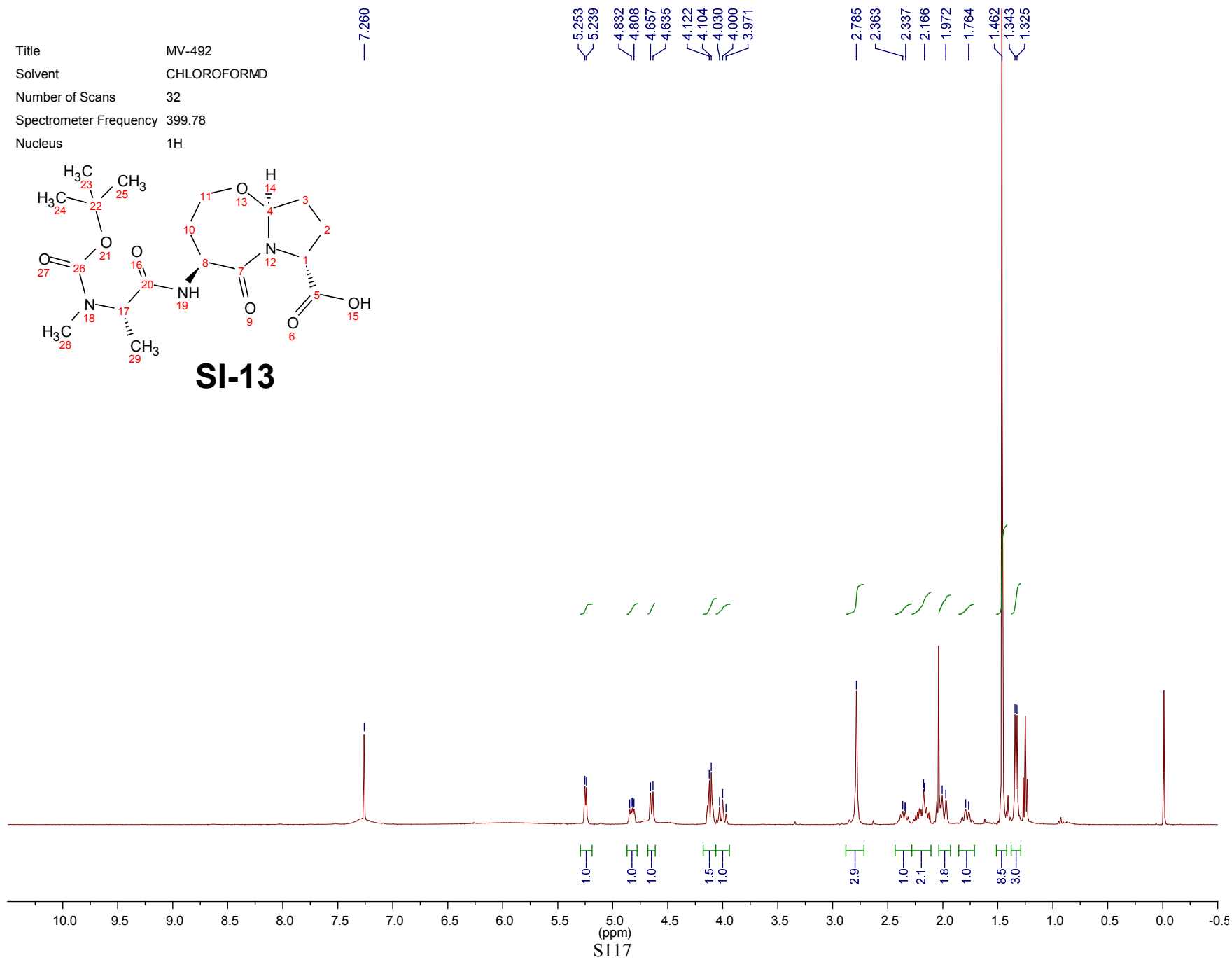
Title MV-479  
Solvent METHANOL-D3  
Number of Scans 3568  
Spectrometer Frequency 100.53  
Nucleus 13C



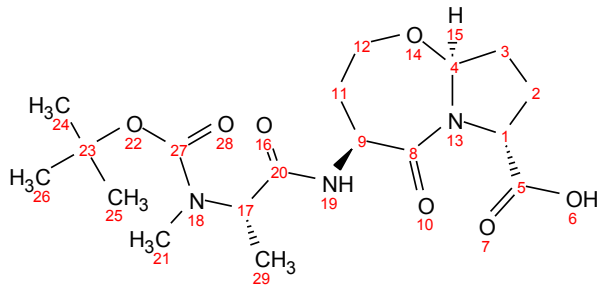
Title MV-492  
Solvent CHLOROFORMD  
Number of Scans 32  
Spectrometer Frequency 399.78  
Nucleus 1H



SI-13



Title MV-492  
Solvent CHLOROFORMD  
Spectrometer Frequency 100.53  
Nucleus 13C



SI-13

173.940  
172.002  
171.256

89.614

80.947

77.466

70.653

60.576

59.768

53.054

32.983

32.475

30.481

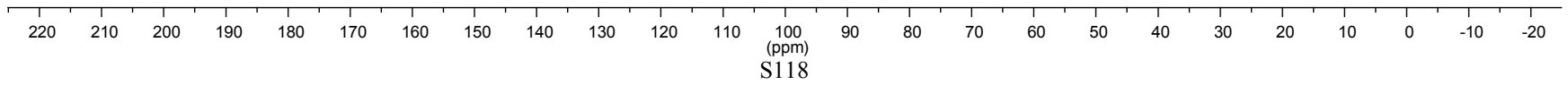
28.502

26.077

21.187

14.323

14.061

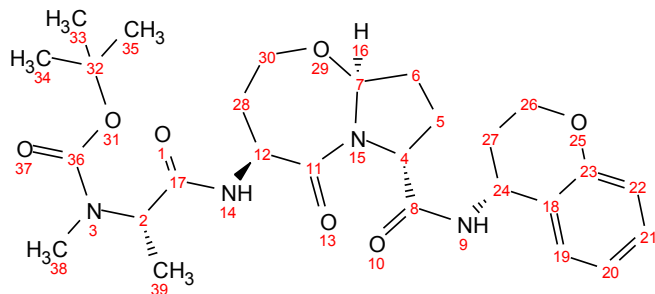


Title MV-493  
Solvent CHLOROFORMD  
Number of Scans 32  
Spectrometer Frequency 399.78  
Nucleus 1H

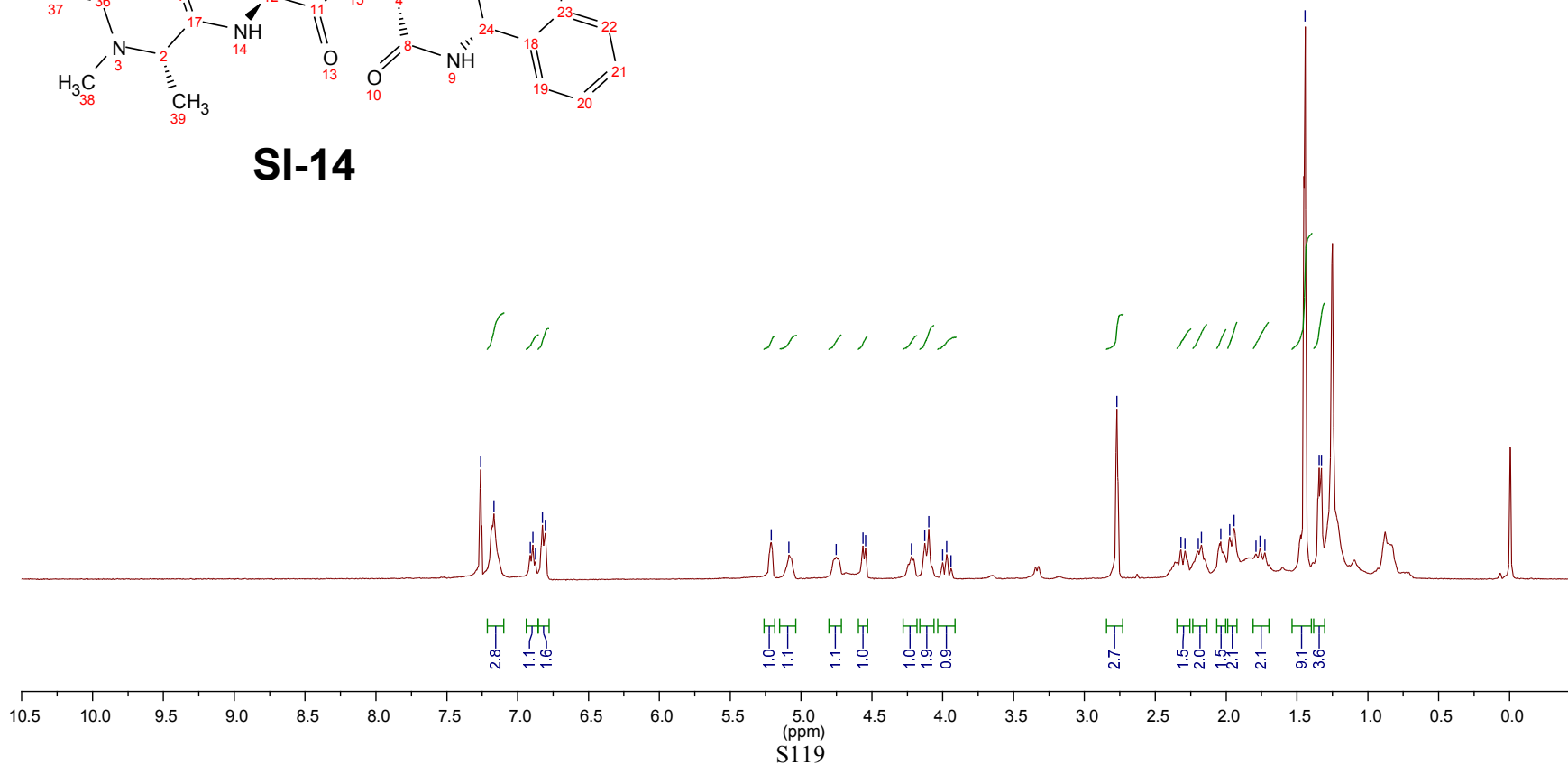
7.260  
7.168  
6.911  
6.893  
6.874  
6.826  
6.805

5.211  
5.086  
4.753  
4.543  
4.220  
4.127  
4.098  
4.001  
3.971  
3.941

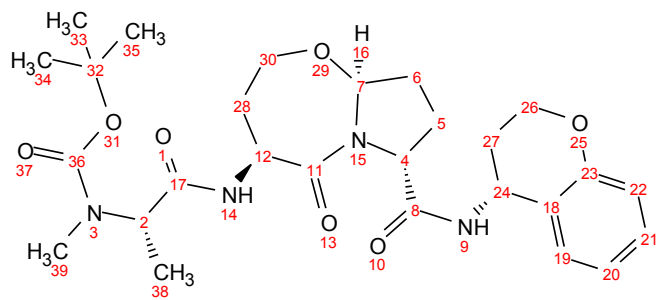
2.770  
2.319  
2.174  
2.038  
1.974  
1.945  
1.726  
1.442  
1.344  
1.326



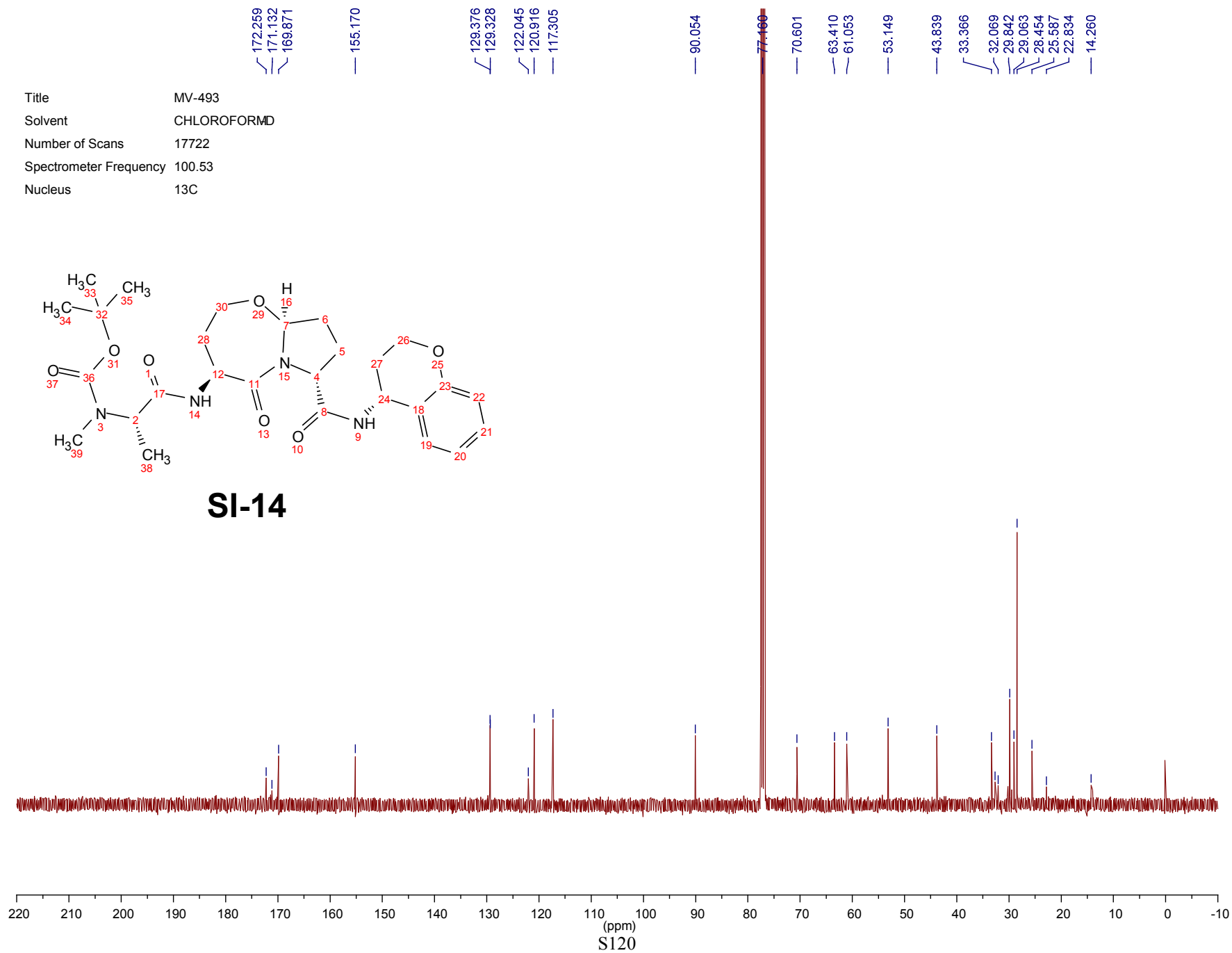
SI-14



Title MV-493  
Solvent CHLOROFORMD  
Number of Scans 17722  
Spectrometer Frequency 100.53  
Nucleus <sup>13</sup>C

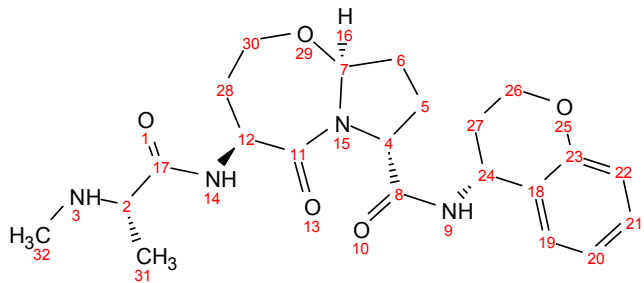


SI-14

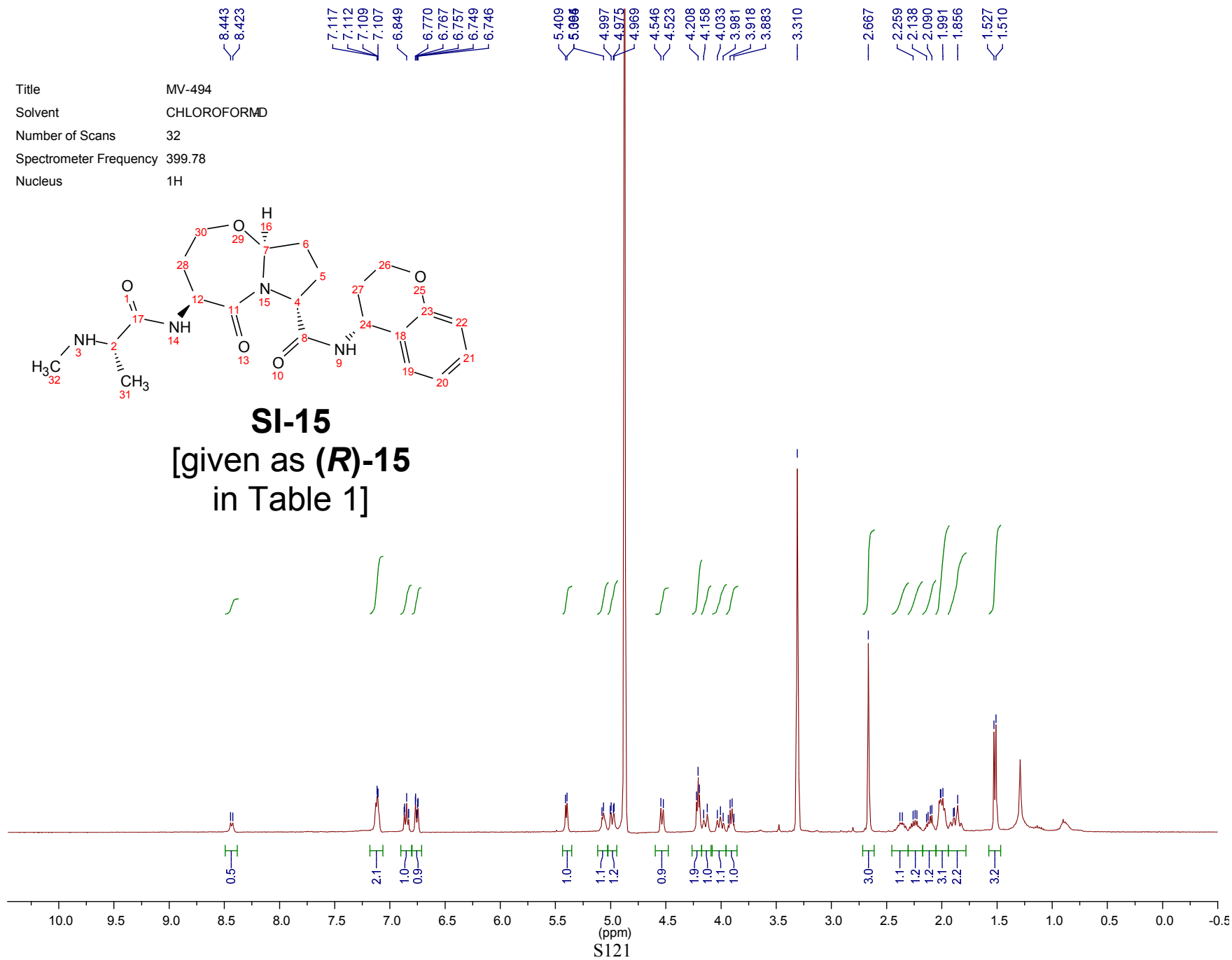




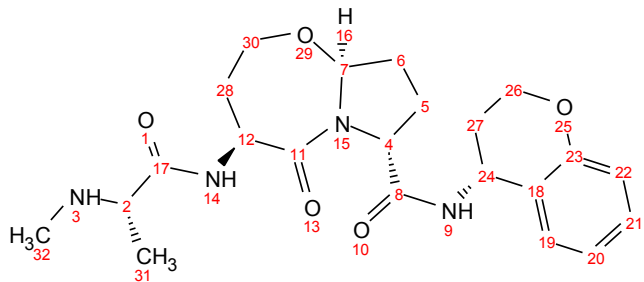
Title MV-494  
 Solvent CHLOROFORMD  
 Number of Scans 32  
 Spectrometer Frequency 399.78  
 Nucleus 1H



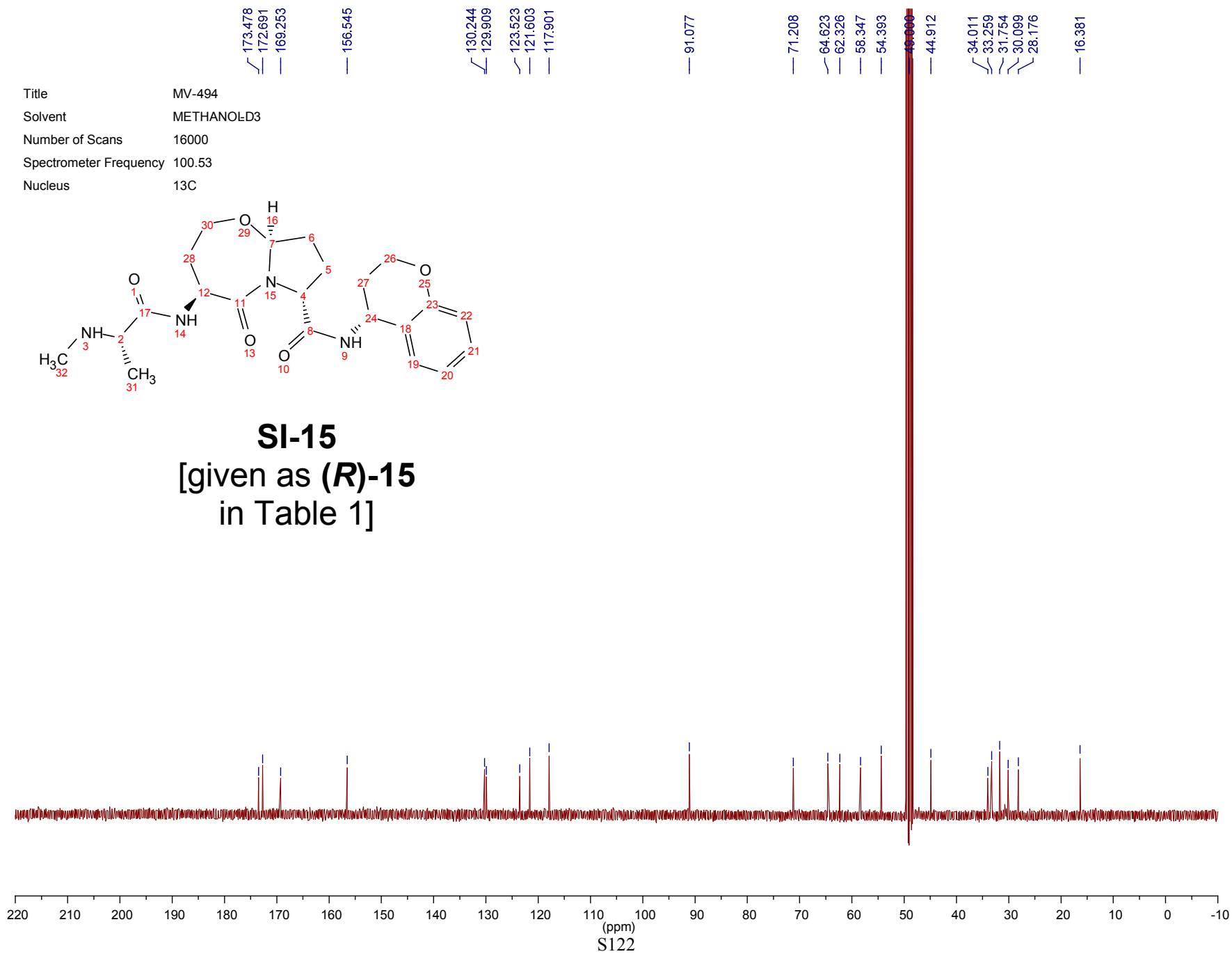
**SI-15**  
 [given as (*R*)-15  
 in Table 1]



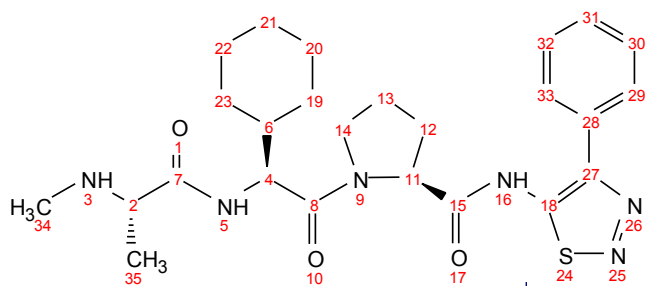
Title MV-494  
Solvent METHANOLD3  
Number of Scans 16000  
Spectrometer Frequency 100.53  
Nucleus 13C



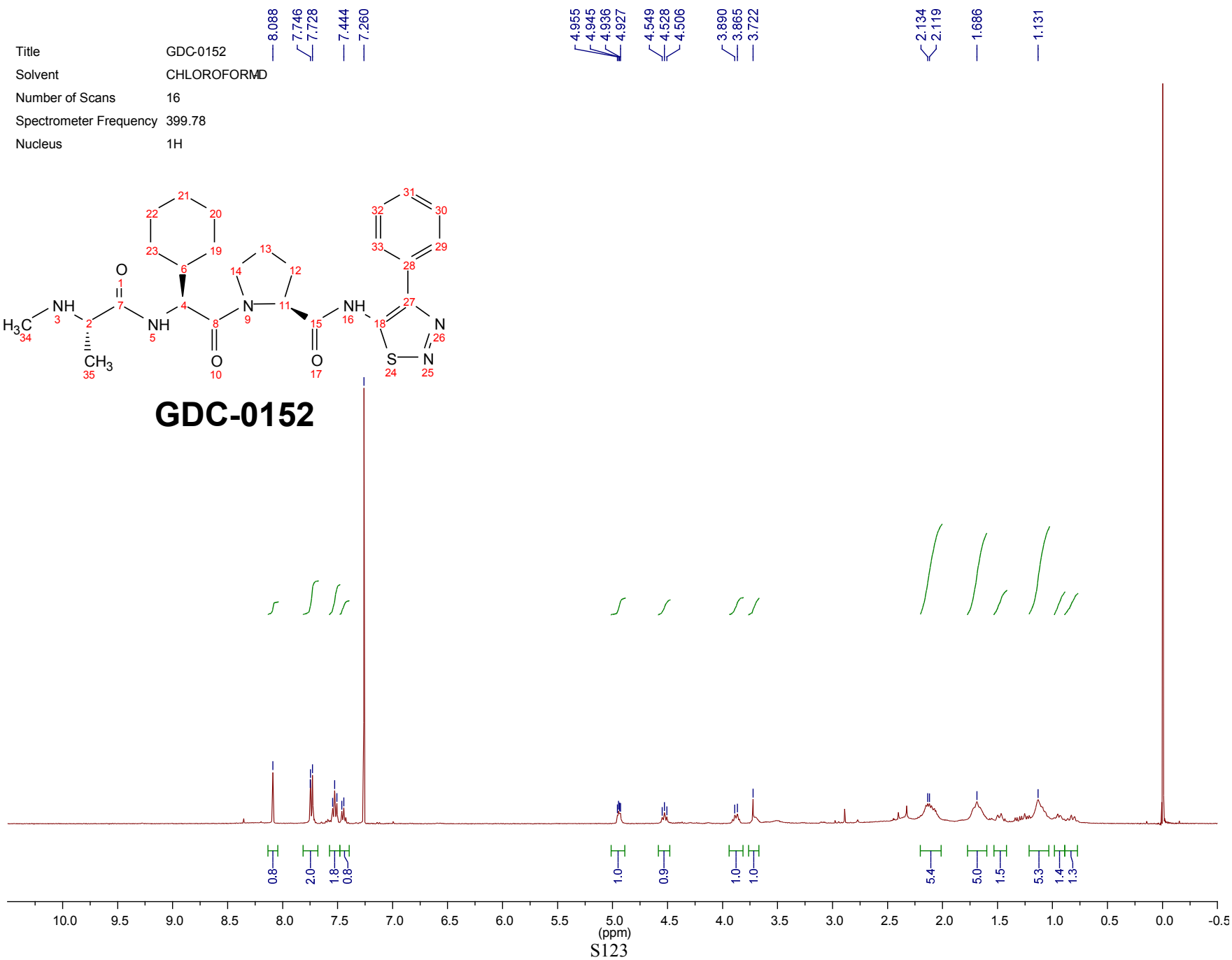
**SI-15**  
[given as (*R*)-15  
in Table 1]



Title GDC-0152  
Solvent CHLOROFORMD  
Number of Scans 16  
Spectrometer Frequency 399.78  
Nucleus 1H



**GDC-0152**



30 minutes

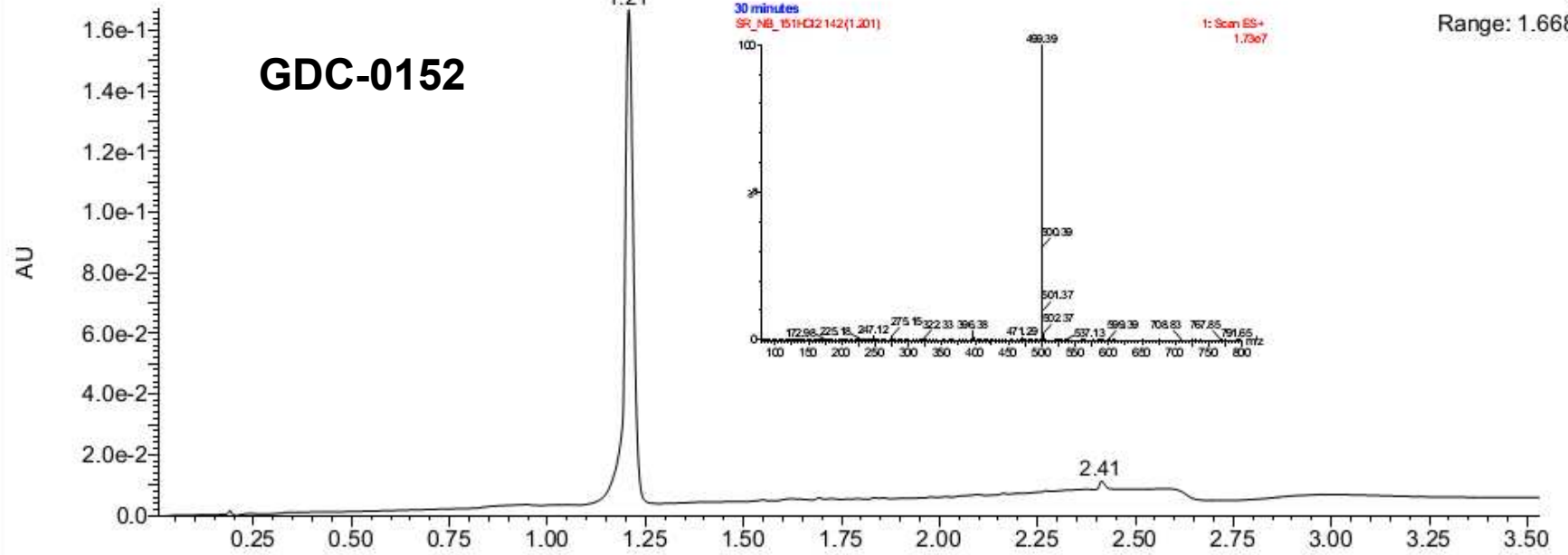
SR\_NB\_151HC12

2: Diode Array

254

Range: 1.668e-1

**GDC-0152**



SR\_NB\_151HC12

1: Scan ES+

TIC

3.07e7

