Supplementary Information:

Comprehensive profiling of the vaginal microbiome in HIV positive women using massive parallel semiconductor sequencing

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Supplementary Figures



Supplementary Figure S1. Overview of bioinformatics analysis strategy. HPV infections are detected through an iterative process. First, sequence reads are mapped to a combined reference consisting of the entire human genome sequence as well as references for known HPV types, resulting in a coverage profile for each of the HPVs. The reads which could not be aligned to any HPV or the human reference (unmapped reads) are processed by a *de novo* assembly, and the resulting contigs are screened for similarities to general HPV gene features. In this way we can detect novel HPV sequences. The novel HPVs are then added to the reference file after which a re-alignment is performed against the updated reference. This process continues until no more novel HPV types are detected. After the HPV analysis is complete, we search for additional non-HPV infections among the remaining unmapped reads. A *de novo* assembly is performed and the resulting contigs are used in BLAST searches against a database with all available nucleotide sequences. Reference sequences for viruses, bacteria and other infectious agents that show very high sequence similarity to the assembled contigs are extracted, and then the unmapped reads are re-aligned against those. In this way we obtain a sequence read coverage that can be used for quantification of the levels of various co-infections.



Supplementary Figure S2. Abundance of HPV types in the 20 samples. A) The gray bars show the number of infected women for each HPV type. In total 48 HPV genotypes were identified (including two novel genotypes HPV X and Y), with some being much more frequent compared to others. B) Combined coverage obtained for each HPV type in all of the women. All of the HPV infections with high coverage are present in the ROCHE Linear Assay Genotyping (marked by red asterisks) and most of them are also included in the HPVIR real-time PCR test (blue asterisks). There is a long tail of HPV types with low coverage, some of which are not detectable by the genotyping or real-time PCR.



Supplementary Figure S3. Sequence coverage for non-HPV infections. A) Coverage profile for an over 4 kb piece of the *Neisseria gonorrhoeae* plasmid in three samples (S1, S5 and S20). The plasmid is clearly present in samples S5 and S20 where the coverage reaches 112X and 281X respectively. In S1 there is not a single read mapped to the plasmid reference. SNPs in samples S5 and S20 are shown as vertical colored lines. There are clearly a number of SNPs both in S5 and S20 of which many are detected in both samples. It is thus possible to study bacterial co-infections at high resolution and even to determine genetic variation between samples. **B)** Coverage profile and individual reads for the *Bacteroides fragilis* transposon U75371.3 in samples S12, S15 and S18. The samples show high coverage across the reference except for a ~ 2kb region that appears to be deleted in all three samples.

Supplementary Tables

Sample #	Bases	>=Q20 Bases	No of reads	Mapped reads	Avg read length ^a
S1	1,123,094,246	845,921,658	9,340,619	9,293,317	120
S2	1,127,774,058	860,729,845	9,475,023	9,357,856	119
S3	1,124,994,779	866,858,532	9,355,034	9,254,266	120
S4	1,173,214,481	890,087,842	9,910,115	9,850,996	118
S 5	949,846,119	726,488,153	8,116,682	7,960,205	117
S6	1,286,638,236	901,835,493	10,370,987	10,164,937	124
S7	1,325,653,574	935,382,902	10,709,737	10,613,131	123
S8	1,228,708,905	870,224,738	9,896,537	9,789,102	124
S9	891,389,357	635,020,898	7,052,038	6,967,029	126
S10	1,359,245,507	940,175,509	10,974,157	10,833,021	123
S11	1,967,847,805	1,314,580,625	17,842,951	16,985,972	110
S12	1,862,343,610	1,276,740,852	16,246,875	15,550,508	114
S13	1,857,594,885	1,243,793,198	16,507,559	16,091,184	112
S14	1,555,333,269	1,040,171,410	13,725,450	13,482,576	113
S15	1,888,085,964	1,257,753,806	16,665,679	16,240,891	113
S16	1,180,150,019	837,426,989	9,905,939	8,715,880	119
S17	1,354,079,231	954,439,784	11,522,705	11,214,934	117
S18	1,199,675,779	853,206,452	10,142,966	9,349,135	118
S19	1,056,016,718	733,479,972	8,962,491	8,131,553	117
S20	4,400,023,141	3,094,815,089	38,899,388	38,388,505	113

Supplementary Table S1. Ion Proton run information for the 20 samples.

^a Read lengths after quality trimming

Supplementary Table S2. Sequence similarity within the L1 gene. Novel HPV types (HPV X and Y) are compared to their evolutionarily closest relatives. In the upper right corner are numbers representing the percent similarity, in the bottom right triangle are number of identical nucleotides.

	HPV 101	HPV X	HPV Y	HPV 103	HPV 108
HPV 101		78.4	65.57	67.05	67.76
HPV X	1216		65.77	66.28	67.12
HPV Y	1021	1024		67.44	66.67
HPV 103	1046	1036	1052		75.24
HPV 108	1057	1047	1040	1167	

Supplementary Table S3. Reference sequences from viral, bacterial and parasitic coinfections.

	Name	Туре	Accession	Ref length
1	Alistipes finegoldii	complete genome	CP003274.1	3.728 Mb
2	Atopobium parvulum	complete genome	CP001721.1	1.541 Mb
3	Bacteroides fragilis	transposon	U75371.3	12 kb
4	Clostridiales genomosp.	complete genome	CP001850.2	1.806 Mb
5	Clostridium tetani	complete genome	AE015927.1	2.794 Mb
6	Enterobacter agglomerans	plasmid	AF014880.1	2.492 kb
7	Fusobacterium nucleatum	complete genome	AE009951.2	2.171 Mb
8	Gardnerella vaginalis	complete genome	NC_013721.1	1.614 Mb
9	JC virus	complete genome	AF004349.1	5.112 kb
10	Lactobacillus gasseri	complete genome	CP000413.1	1.891 Mb
11	Lactobacillus johnsonii	complete genome	NC_005362.1	1.989 Mb
12	Megasphaera elsdenii	ribosomal RNA	NR_103173.1	2.907 kb
13	Neisseria gonorrhoeae	plasmid	FJ172221.1	6.053 kb
14	Neisseria meningitidis	plasmid	AF126482.1	5.589 kb
15	Porphyromonas asaccharolytica	complete genome	NC_015501.1	2.182 Mb
16	Prevotella denticola	complete genome	CP002589.1	2.932 Mb
17	Prevotella melaninogenica	complete genome	NC_014370.1	3.162 Mb
18	Roseburia hominis	ribosomal RNA	NR_076921.1	2.885 kb
19	Salmonella enterica	plasmid	NC_021157.1	4.668 kb
20	SEN virus	virus	AY183662.1	3.076 kb
21	Streptococcus agalactiae	complete genome	NC_021485.1	2.135 Mb
22	Streptococcus oligofermentans	complete genome	NC_021175.1	2.138 Mb
23	Streptococcus pneumoniae	transposon	KC488256.1	57 kb
24	Trichomonas vaginalis	hypothetical protein	XM_001319996.1	3.298 Mb
25	Torque teno virus	virus	AM712004.1	3.754 kb

Supplementary Table S4. Long contigs (at least 4kb) for which a blastn analysis gives no hit

to any known reference sequence.

Sample	Contig#	Contig	Contig	total	nr ORF	closest relative	Highest amino
		length	reads	ORFs ^a	hits		acid similarity
85	00001	12179	19262	10	2	Bacillus	54
<u>88</u>	00001	4163	461	7	3	Eubacterium	60
<u>88</u>	00002	4298	2712	10	2	Metascardovia	54
<u>S13</u>	00001	9648	14501	10	8	Streptococcus	/4
<u>S13</u> S15	00002	4902	527	4	5	Lactobacillus	95
S15 S15	00001	5556	733	9	5	Closifiaium Puminococcus	55
S15 S15	00002	4148	1145	6	2	Stroptococcus	55
<u>\$15</u>	00003	5321	496	10	<u>2</u> <u>1</u>	Lachnospiraceae	60
<u>815</u>	00005	4383	573	7	2	Clostridium	54
<u>815</u>	00005	4256	441	5	3	Rhodococcus	54
S16	00001	25555	30141	24	13	Veillonella	62
S16	00010	9868	991	14	6	Clostridium	59
S16	00011	9716	1366	17	10	Clostridium	68
S16	00020	7835	1087	13	6	Clostridium	70
S16	00021	7613	788	12	4	Clostridiales	68
S16	00022	7561	931	13	7	Clostridium	63
S16	00024	7380	1023	19	2	Lactobacillus	50
S16	00025	7269	832	9	2	Enterococcus	41
S16	00026	6961	897	9	5	Clostridium	63
S16	00028	6586	649	9	5	Clostridium	68
S16	00029	6490	998	7	3	Streptococcus	66
S16	00030	6425	31051	6	4	Prevotella	87
S16	00032	6274	577	7	6	Paenibacillus	61
S16	00033	6248	722	9	4	Clostridium	60
S16	00036	6035	667	5	4	Clostridium	59
S16	00037	5982	516	11	4	Gardnerella	69
S16	00038	5849	853	7	4	Eubacterium	67
S16	00039	5834	540	7	3	Clostridiales	48
S16	00041	5732	604	9	6	Lachnospiraceae	67
S16	00042	5717	613	7	6	Clostridium	71
<u>S16</u>	00043	5632	831	6	1	Actinomyces	44
<u>S16</u>	00045	5612	690	8	5	Oribacterium	73
S16	00052	5269	480	12	5	Lactobacillus	68
<u>S16</u>	00056	5149	463	6	5	Ruminococcus	67
S16	00057	5107	586	5	4	Streptococcus	58
<u>S16</u>	00059	4981	331	5	3	Clostrialales	/0
S10 S16	00061	4927	427	5	4	Firmicules Clostridium	50
S16	00003	4001	407	6	4	Clostridium	56
<u>\$16</u>	00067	4754	566	8	5	Clostridium	75
<u>\$16</u>	00068	4746	403	6	<u>J</u>	Eusobacterium	77
S16	00069	4738	580	8	2	Parvimonas	67
<u>\$16</u>	00072	4656	490	7	3	Lactobacillus	62
<u>\$16</u>	00075	4581	437	5	4	Clostridium	69
S16	00076	4502	394	7	5	Clostridium	58
<u>\$16</u>	00078	4419	393	7	5	Clostridium	62
S16	00079	4391	627	7	3	Clostridium	65
S16	00080	4387	413	6	4	Clostridiales	61
S16	00085	4283	438	7	5	Fusobacterium	65
S16	00087	4219	459	6	5	Oribacterium	73
S16	00088	4210	387	7	3	Clostridium	60
S16	00089	4208	285	7	3	Clostridium	58
S16	00091	4153	3549	5	2	Parabacteroides	49
S16	00093	4134	356	4	4	Clostridium	64
S16	00094	4105	502	8	4	Lachnospiraceae	54
S16	00096	4027	350	8	2	Clostridium	61
S16	00097	4030	415	6	3	Clostridium	66

S16	00098	4010	370	5	4	Clostridium	70
S18	00001	5009	10848	4	3	Streptococcus	75
S19	00001	4832	1060	8	4	Prevotella	84
S19	00004	4151	1045	5	2	Streptococcus	64

^a Only ORFs encoding proteins of at least 100 aa are considered ^b Number of translated ORFs with protein similarity to the organism in the closest relative column, as predicted by blasts