

Questionnaire

of

the microbiological procedures
of respiratory specimens
from Cystic Fibrosis patients

Please mark with a cross or use the free text field.

Address of laboratory:

Note:

The questionnaire is based on the German microbiological infectious quality standards for airway infections in CF 2006 (MiQ) and on the protocols provided by the consultant laboratories for CF microbiology (Institute of Microbiology, Medical School Hannover, Germany; Max von Pettenkofer-Institute for Hygiene and Medical Microbiology, Munich, Germany).

1. Of how many CF-patients does your laboratory regularly take care concerning microbiological testing?

Number: _____

2. Do you use special guidelines for the microbiological testing of respiratory specimens from CF-patients?

Yes No

What kind of? _____

3. What kind of materials from CF-patients do you process?

- Sputum
- Nasal swab
- Throat swab
- Bronchoalveolar lavage
- Others: _____

4. Do you homogenize the sputum before processing?

Yes No

With what kind of substance? Dithiothreitol
 NALC
 Others: _____

5. Do you streak the original material **before homogenizing and liquefaction?**

Yes No

On what kind of plate?

- Columbia blood agar
 MacConkey agar
 Chocolate agar
 Others: _____

6. Do you determine the **number of pathogens?**

Yes No

Do you make serial dilutions for the determination of pathogen number?

Yes No

How many steps of serial dilutions do you perform?

- 10^{-1}
 10^{-3}
 10^{-4}
 10^{-5}
 Others: _____

7. What are the **culture media und conditions you use to detect the following pathogens?****H. influenzae**

- Chocolate agar
 with bacitracin
 without bacitracin
 with _____
 Others:

Incubation

- under CO₂
 anaerobic

P. aeruginosa

- Tryptic Soy Agar
 Pseudomonas Isolation Agar
 MacConkey agar
 Ceftrimid agar
 Others:

B. cepacia

- BCSA
 OFPBL agar
 PC agar
 Others:

S. aureus

- Columbia blood agar
 Mannitol salt agar
 BHI-NaCl-Agar
 Baird-Parker agar
 MRSA selective agar
 S. aureus ID agar
 CHROMagar Staph aureus
 Polymyxin/Nalidixic acid agar
 Others:

S. maltophilia

- MacConkey agar
 VIA agar
 Meropenem agar
 Others:

A. xylosoxidans

- MacConkey agar
 Others:

Fungi

- Sabouraud agar
 Kimmig agar
 Others:

8. How long do you incubate the different culture media and at which temperatures?

Pathogens	Time				Incubation			
	24h	48h	5d	Other	30°C	32±1°C	36±1°C	Other temp.
H. influenzae	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>		<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	
P. aeruginosa	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>		<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	
B. cepacia	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>		<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	
S. aureus	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>		<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	
S. maltophilia	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>		<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	
A. xylosoxidans	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>		<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	
Fungi	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>		<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	

9. Do you perform a special test for cultivation of anaerobic bacteria?

Yes No

What kind of agar do you use for anaerobic cultures?

- Anaerobian blood agar
 Kanamycin Vancomycin blood agar
 Phenylethyl Alcohol agar
 Others: _____

10. How is the differentiation of pathogens performed in your laboratory?

System	for differentiation of
<input type="radio"/> API® systems	
<input type="radio"/> API Staph	
<input type="radio"/> API 20 Strep	
<input type="radio"/> RapiD 20 E	
<input type="radio"/> API 20 E	
<input type="radio"/> API 20 NE	
<input type="radio"/> VITEK® 1	
<input type="radio"/> VITEK® 2	
<input type="radio"/> BD Phoenix™	
<input type="radio"/> MicroScan®	
<input type="radio"/> MALDI-TOF	
<input type="radio"/> Others:	

11. Do you identify Gram-negative nonfermenting bacteria other than P. aeruginosa and/or B. cepacia?

- Yes No

What kind of system do you use therefore?

- API 20 NE
 VITEK® 1
 VITEK® 2
 BD Phoenix™
 MicroScan®
 MALDI-TOF
 16S Sequencing
 Others: _____

12. How is the susceptibility testing done?

- VITEK® 1
 VITEK® 2
 BD Phoenix™
 MicroScan®
 Others: _____
- Quantitative determination of the MIC via
 Agar dilution
 Microdilution
 Etest®
 Others: _____
- Qualitative susceptibility testing via
 Agar diffusion
 Others: _____

13. Do you perform antibiotic combination tests?

- Yes No

With what kind of system?

- Merlin
 Etest®
 In house
 Others: _____

14. Do you routinely investigate the following pathogens?

	Yes	No	On request
Mycobacteria	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Viruses	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>

15. Do you perform confirmatory tests for special pathogens?

Yes No

What kind of confirmatory test do you perform for which pathogen?

	PCR	16S Sequencing	Probe	Others:
MRSA	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	
B. cepacia	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	
Atypical Mycobacteria	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	
Others:	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	

16. Do you send unusual pathogens to reference laboratories for further differentiation and typing?

Yes No

What kind of pathogens?

- MRSA
 B. cepacia
 Atypical Mycobacteria
 Uncommon yeasts
 Uncommon fungi
 Others: _____

17. Do you inform the CF center to initiate special hygiene measures if B. cepacia was isolated?

Yes No

18. Do you inform the clinician if pathogens with special phenotypes were isolated?

	Yes	No
SCV	<input type="radio"/>	<input type="radio"/>
Mucoide phenotypes	<input type="radio"/>	<input type="radio"/>
Others:	<input type="radio"/>	<input type="radio"/>

19. In how many CF patients did you detect the following pathogens during the last year?

CF patients with	Number
MRSA	
B. cenocepacia	
B. multivorans	
Other Burkholderia	
Atypical Mycobacteria	
M. abscessus	
M. avium intracellulare	
M. chelonae	
Other Mycobacteria	
Pandoraea species	
Exophiala dermatitidis	
Scedosporium apiospermum	
Other uncommon fungi	

TABLE S1 Translation of the main recommendations by the MiQ 2006

Culture - Stepwise testing

Step 0 - Basis culture media

No colonization with *P. aeruginosa*/other pathogens (especially nonfermenting bacteria) → no overgrowth expected

Blood agar	E.g. Columbia blood agar, Schaedler agar with 5% sheep's blood
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Chocolate agar	Supplemented with bacitracin
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Lactose indicator agar	E.g. MacConkey agar
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CF-specific at all steps	<i>Pseudomonas</i> agar
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	<i>B. cepacia</i> selective agar
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Optional	Selective medium for <i>S. aureus</i>
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Step 1 - Extended culture media

Colonization with *P. aeruginosa*/other nonfermenting pathogens or mixed infection → probable overgrowth of slow growing pathogens or variant pathogens

Basis culture media

Selective medium for <i>S. aureus</i>	E.g. mannitol salt agar, CHROMagar Staph aureus, <i>S. aureus</i> ID agar
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Optional

Meropenem agar, quantitative culture, when indicated special anaerobic culture for *H. influenza*, fungi medium

Step 2 - Extended culture media

Colonization/infection with common pathogens for > 1 year with repeated antibiotic therapies → probable selection of resistant variants

Basis culture media

Meropenem agar

Quantitative culture

Optional

Special anaerobic culture for *H. influenza*, fungi medium

Culture media for the detection of common CF pathogens (medium of choice listed first)

S. aureus

- Columbia blood agar
- *S. aureus* ID agar
- Mannitol salt agar
- CHROMagar Staph aureus
- BHI-NaCl-Agar

-
- Polymyxin/Nalidixic acid agar
 - Baird-Parker agar
 - MRSA selective agar
- H. influenza*
- Chocolate agar with bacitracin
 - When indicated anaerobic culture
- P.aeruginosa*
- Tryptic Soy Agar
 - MacConkey agar
 - Ceftrimid agar
 - Pseudomonas Isolation Agar
- BCC
- BCSA
 - OFPBL agar
 - PC agar

Identification of pathogens

P. aeruginosa and other nonfermenting pathogens

Identification should be done to species level using

- a commercial method preferring API 20 NE or

- a molecular method e.g. 16S rDNA sequencing, species-specific PCR or FISH

Metabolic inactive nonfermenting pathogens e.g. lung adapted variants of *P. aeruginosa*, BCC amongst others can't be identified to species level in part using commercial systems

BCC isolates must be confirmed by a molecular method, obligatory when initially isolated

Method of choice: *recA* sequencing

Alternatives: RFLP of *recA* gene, species-specific PCR

S. aureus

Doubtful results should be confirmed by *S. aureus* specific PCR

Especially in case of culture of *S. aureus* SCVs, an identification at the species level is often only possible by PCR

Isolates that can't be differentiated should be determined with molecular methods e.g. 16S rDNA sequencing

Confirmatory tests for

MRSA

- Gold standard: detection of the *mecA* gene as "in-house" or commercial PCR
- Detection of PBP 2a

BCC

- RFLP of the *recA* gene
- Species-specific PCR based on polymorphisms in the rDNA und *recA* gene
- *RecA* sequencing and phylogenetic analysis of *recA* gene sequences are recommended

Susceptibility testing

Guided by CLSI or DIN 58940

Quantitative determination of the MIC via

- Dilution methods e.g. agar or bouillon dilution
- Gradient diffusion methods e.g. Etest

Qualitative susceptibility testing via Agar diffusion

Method of choice for all nonfermenting bacteria:

- Agar diffusion or
- Dilution methods preferring microdilution

Susceptibility testing of most *S. aureus* isolates can be done with commercial systems

Automated systems are not recommended for susceptibility testing of *P. aeru-*

ginosa and other nonfermenting pathogens because they are more often defective and little validated

TABLE S2 Different BCC selective agar formulations

Agar	Formulation
<i>B. cepacia</i> selective agar	10 g of lactose and 10 g of sucrose in an enriched base of casein and yeast extract with 600,000 U of polymyxin, 10 mg of gentamicin and 2.5 mg of vancomycin per liter ^a
<i>B. cepacia</i> agar	DeCicco holding medium with 300,000 U of polymyxin and 100 mg of ticarcillin per liter ^b
cepacia medium	300,000 U of polymyxin and 0.1 of ticarcillin per liter ^c
Oxoid <i>B. cepacia</i> agar	150,000 IU of polymyxin, 5 mg of gentamicin and 100 mg of ticarcillin per liter ^d
oxidative-fermentative polymyxin B-bactriacin lactose agar	oxidation-fermentation agar supplemented with 10 g of lactose, 300,000 U of polymyxin and 200 U of bacitracin per liter ^e

^a Data from reference 29.

^b Data from reference 16.

^c Data from reference 30.

^d Data from reference 31.

^e Data from reference 32.