

Transforming Growth Factor- β 1 Downregulates Vascular Endothelial Growth Factor-D Expression in Human Lung Fibroblasts via the Jun NH₂-Terminal Kinase Signaling Pathway

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SUPPLEMENTARY MATERIALS AND METHODS

Cell Culture

Human umbilical vein endothelial cells (HUVECs) (Lonza, Walkersville, MD) were maintained in endothelial basal medium (EBM)-2 supplemented with fetal bovine serum (FBS) and growth factors in Endothelial Growth Media (EGM)-2 SingleQuot kit (Lonza). Cells were incubated at 37°C in a humidified 5% CO₂ atmosphere and used as positive controls for VEGFR-2 and VEGFR-3 expression.

ELISA

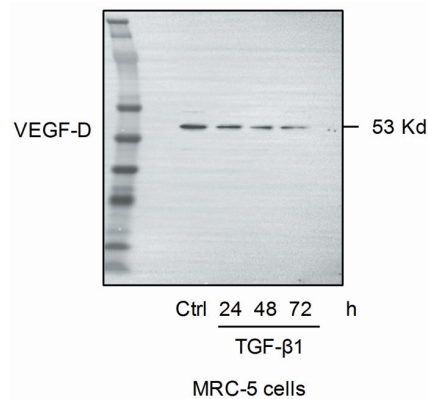
Culture supernatants from control siRNA- and VEGF-D siRNA- transfected cells were concentrated using standard acetone, trichloroacetic acid (TCA) precipitation protocols (1, 2) or Amicon Ultra centrifugal filter units (Millipore, Billerica, MA). The levels of VEGF-D in the concentrated samples were measured using a human VEGF-D DuoSet ELISA Development kit (R&D Systems) according to the manufacturer's recommendations.

Sircol Assay

Collagen concentration in culture supernatants from control siRNA- and VEGF-D siRNA- transfected MRC-5 cells was determined by Sircol assay (Bicolor, County Antrim, UK) according to the manufacturer's recommendations.

Cell Contraction Assay

Control siRNA- and VEGF-D siRNA- transfected MRC-5 cells were resuspended and mixed with collagen gel solution (Cell Biolabs, San Diego, CA) at a concentration of 2×10⁵ cells/ml. 0.5ml of the cell-collagen mixture was immediately added in 24-well tissue culture plates, incubated at 37 °C for 1 h, and then overlaid with culture media.

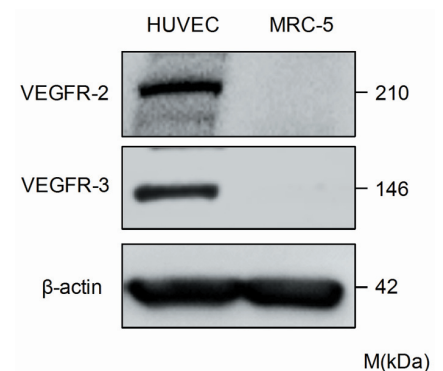


Supplementary Figure S1. Full-length unprocessed VEGF-D is the only VEGF-D isoform detected in MRC-5 cell lysates. MRC-5 fibroblasts were stimulated with TGF- β 1 (5 ng/ml) for the indicated time. Equal amounts of protein from whole cell lysates were analyzed by western blotting with antibodies against VEGF-D. Results show a single immunoreactive band of ~53Kd consistent with the full-length unprocessed VEGF-D.

The gels were subsequently released from the plates to initiate contraction. Images of gels were captured at 48h using ChemiDoc XRS+ imaging system (Bio-Rad) and analyzed using ImageJ software (NIH).

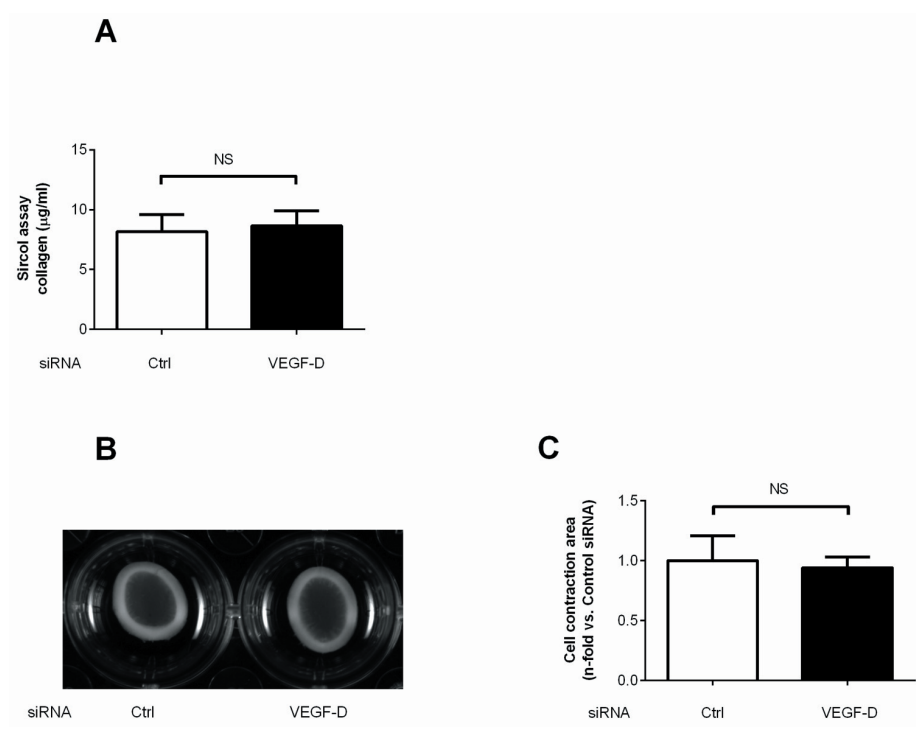
SUPPLEMENTARY REFERENCES

- Hwang BJ, Chu G. (1996) Trichloroacetic acid precipitation by ultracentrifugation to concentrate dilute protein in viscous solution. *BioTechniques* 20: 982-984.
- Fic E, Kedracka-Krok S, Jankowska U, Pirog A, Dziedzicka-Wasylewska M. (2010) Comparison of protein precipitation methods for various rat brain structures prior to proteomic analysis. *Electrophoresis* 31: 3573-3579.

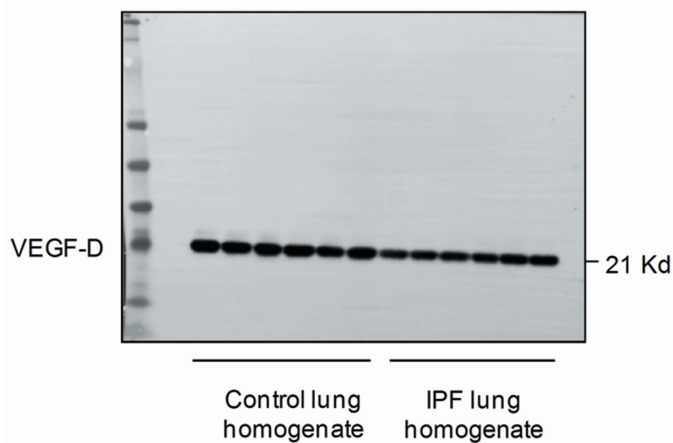


Supplementary Figure S2. MRC-5 cells do not express detectable levels VEGFR-2 and VEGFR-3. Equal amounts of protein from MRC-5 and HUVEC whole cell lysates were analyzed by western blotting with antibodies against VEGFR-2 and VEGFR-3. β -actin was used as a loading control.

TGF- β 1 DOWNREGULATES VEGF-D EXPRESSION



Supplementary Figure S3. VEGF-D knockdown in MRC-5 cells does not affect collagen synthesis and cell contractility. (A) MRC-5 cells were transfected with control siRNA or VEGF-D siRNA for 48 h. Collagen content in cell culture medium was measured using Sircol soluble collagen assay. (B) Effect of siRNA-mediated VEGF-D silencing on MRC-5 cells contractile efficiency in collagen gel. (C) Cell contraction was expressed as the fold of gel area relative to control siRNA transfection. Data represent mean \pm SEM of three independent experiments. NS, not significant, by Student *t* test.



Supplementary Figure S4. Mature VEGF-D is the only VEGF-D isoform detected in human lung homogenates. Equal amounts of protein from human lung homogenates were analyzed by western blotting with antibodies against VEGF-D (n=6 samples per group), showing a single immunoreactive band at ~21Kd, consistent with the molecular weight of the C- and N-terminally cleaved VEGF-D.