

**SUPPLEMENTAL FIGURE LEGENDS****Figure S1. IL-4 potently induces luciferase expression in THP-1 cell clones stably transfected with a MRC1 promoter-driven luciferase construct.**

A) Expression constructs with MRC1 promoter-driven luciferase activity used for generating THP-1 cell clones stably expressing these constructs. The MRC1 promoter lies within 1.0kb upstream of the MRC1 start codon. B) Luciferase activity of a THP-1 cell clone stably transfected with a 1.0kb MRC1-promoter luciferase construct. IL-4 or IL-13 induce M2-type macrophage polarization, reflected by the increased MRC1-luciferase activity (>5-fold). Increasing the dose from 20 to 40ng/ml or combining IL-4 and IL-13 does not further increase luciferase activity. C) IL-4 treatment of PMA primed THP-1 cells, induces luciferase activity, consistent with M2-type macrophage polarization (M2). IL-4-induced M2-type polarized macrophages can be repolarized to the M1-type by replacing IL-4 with IFN- $\gamma$ +LPS. After PMA treatment (100nM, for 24 hours), cells were treated with IL-4 (20ng/ml) for 2 days and changed to fresh medium for 2 days (M2); M2→M1:IL-4 treatment for 2 days and then change to IFN- $\gamma$  (20ng/ml) + LPS (10ng/ml) for another 2 days; M1: IFN- $\gamma$  + LPS for 2 days and then change to fresh medium for 2 days; “M1→M2”: IFN- $\gamma$  + LPS for 2 days and then IL-4 for 2 days. D) After priming with PMA, endogenous MRC1 proteins were further induced with IL-4 (20ng/ml) treatment, while the addition of LPS (10ng/ml) and IFN- $\gamma$  (20ng/ml) inhibited MRC1 protein levels. Densitometric band values are indicated. \*\*:  $p<0.01$ . \*\*\*:  $p<0.001$

**Figure S2. Concentrations of doxycycline that are sufficient to inhibit M2-type polarization of macrophages do not affect cell viability.**

BMDM cells isolated from 3-4 months old male C57Bl/6j mice were pretreated with Dox overnight and subsequently treated with Dox +/- IL-4 (20ng/ml) in DMEM+1% FBS+M-CSF for 3 days. Cells were stained with Live/Dead cell viability kit (Invitrogen) and assessed by a FACSCanto flow cytometer.

**SUPPLEMENTAL TABLES****Table S1. Primers used to amplify genomic DNA containing the MRC1 promoter.**

MRC1 2.5 kb (between -2383 to +116 bp), forward	ggggtaccTAGGCTACGCTGAGCCGT
MRC1 1.5 kb (between -1383 to +116 bp), forward	ggggtaccTTCCCTCCGGTCAGGATA
MRC1 1.0 kb (between -883 to +116 bp), forward	ggggtaccCCTGACTCACTGTA ACTT
MRC1, reverse	ccgctcgagGGCCCAGGGTTTATCCTT

**Table S2. Semiquantitative RT-PCR primers used.**

<b>Human</b>			
h36B4	for	GCAATGTTGCCAGTGTCTGT	
	rev	GCCTTGACCTTTTCAGCAAG	
hCD68	for	GCTACATGGCGGTGGAGTACAA	
	rev	ATGATGAGAGGCAGCAAGATGG	
hMRC1	for	TGGTTTCCATTGAAAGTGCTGC	
	rev	TTCCTGGGCTTGACTGACTGTTA	
hCXCL9	for	TTGGGCATCATCTTGCTGGTTCT	
	rev	TGGCTGACCTGTTTCTCCCACT	
hCox-2	for	CCGGGTACAATCGCACTTA	
	rev	GGCGCTCAGCCATACAG	
hCD11b	for	CAGACAGGAAGTAGCAGCTCCT	
	rev	CTGGTCATGTTGATGAAGGTGCT	
<b>Mouse</b>			
m36B4	for	TCACTGTGCCAGCTCAGAAC	
	rev	AATTTCAATGGTGCCTCTGG	
mCD115	for	ATCTGTTCCCGTCCTCACAG	
	reverse	ACTGCCATTGCTCACACATC	
mF4/80	for	GCCTATTATCTATACCCTCCAGCACATC	
	rev	TCCATCTCCCATCCTCCACATCAG	
mCD68	for	AGCTGCCTGACAAGGGACT	
	rev	AGGAGGACCAGGCCAATGAT	
mCox-2	for	TGAGTACCGCAAACGCTTCTC	
	rev	TGGACGAGGTTTTTCCACCAG	
miNOS	for	AACGGAGAACGTTGGATTTG	
	rev	CAGCACAAGGGGTTTTCTTC	
mCxcl9	for	TTTTGGGCATCATCTTCTGG	
	rev	GAGGTCTTTGAGGGATTTGTAGTGG	
mArg1	for	TGAGAGACCACGGGGACCTG	
	rev	GCACCACACTGACTCTTCCATTC	
mIL-1Ra	for	TTACAAGGACCAAATATCAAACCTAGAAG	
	rev	GGATGCCCAAGAACACACTATG	
mFizz1	for	TCCCAGTGAATACTGATGAGA	
	rev	CCACTCTGGATCTCCCAAGA	
mCcl17	for	CCCATGAAGACCTTCACCTC	
	rev	CATCCCTGGAACACTCCACT	
mCcl22	for	TGGCTCTCGTCCTTCTTGCT	
	rev	AGGCTTGCGGCAGGATT	
mCD11b	for	AAACCACAGTCCCGCAGAGA	
	rev	CGTGTTCAACAGCTGGCTTA	
mIL-1b	for	TGATGAGAHCATCCAGCTTC	
	rev	CATGAGTCACAGAGGATGGG	

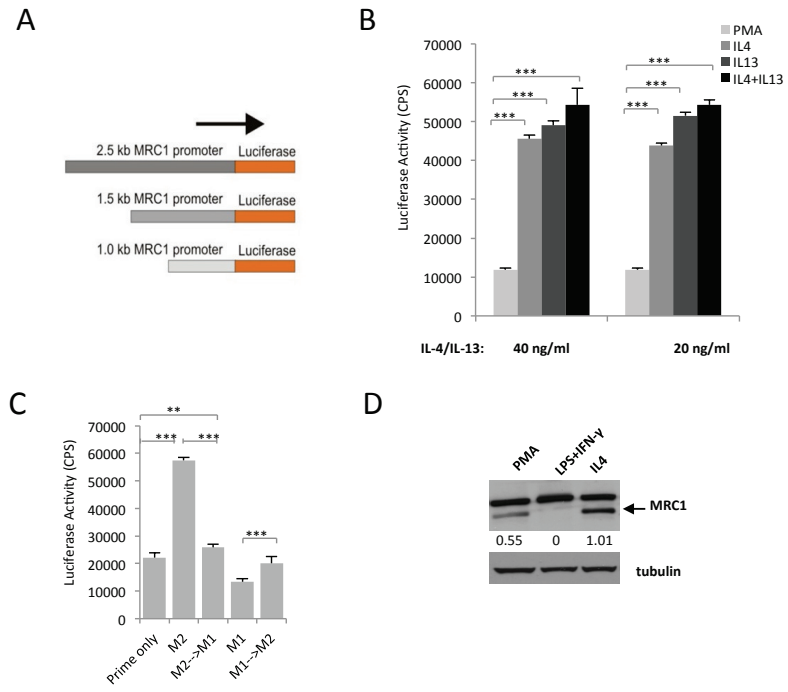


Figure S1

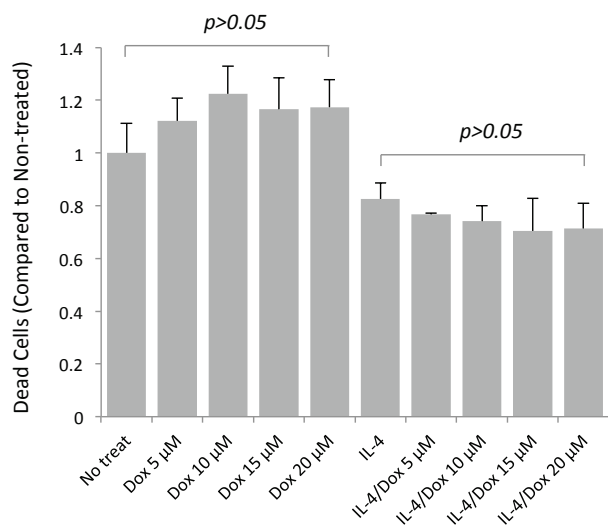


Figure S2